# Bulletin of the Museum of Comparative Zoölogy AT HARVARD COLLEGE. Vol. LXXIII

## CLASSIFICATION OF INSECTS

A Key to the Known Families of Insects and Other Terrestrial Arthropods

BY

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			17	AB	LE	i Oi	F CC	MII	PW I	0					
															PAGE
Preface .								•	٠.						5
Introduction															8
						PA	RT	I							
						In	SECT	A							
Conspectus of the	Hio	her	·G	r011	ns	of T	nsect	:9							17
Classes of Arthrop			. С.	iou	.ps	01 1	115000		•	•			•		26
Orders of Insects		•		•		•	•	•	•	•			•		28
Protura .	•	•		•		•	•	•	•						40
Thysanura .	•	٠		•		•	•	•	•						41
Entotrophi .	•	•		•		•	•	•	•	·					43
Collembola .	•	•		•		•	•	•	•						44
Grylloblattodea	•	•		•			•	•	•						47
Orthoptera .	•	•		•		•	•	•	•		· ·				48
Phasmatodea	•	•		•		•	•	•	•						61
Dermaptera .	•	•		•		•	•	•	·	Ī					65
Diploglossata	•	•		•		•	·	·	-				_		71
Thysanoptera	•	•		•			•	·	•						72
Blattariæ .	•	•		•		•	•								77
Mantodea .	•	•		·		•	•	•	•	•					83
Embiodea .	•	•		•		·	•			-					91
Isoptera .	•	•		•		•	•								93
Corrodentia .	•	•		Ī			•						-		96
Zoraptera .	•	•		•		•	•	•	·						100
Mallophaga .	•	٠		•		•	•	•	-						101
Anoplura .	•	•		•		•	•	•		•				4.	104
Homoptera .	•	•		•		•	•	•	•	•					106
Hemiptera .	•	·		•		•	•	•	•						140
Odonata .	•	•		•		•	·								159
Plectoptera .	•	•		•		•	•	•	·			_			169
Plecoptera .	•	•		•		•	·	•							176
Megaloptera .		•		•		•	•	•	•	•					179
Raphidiodea	•	•		•		•	•	•	•	•			•		180
Neuroptera .	•	•		•		•	•	•	•	•				•	181
Mecoptera .	•	•		•		•	•	•	•	•					189
Trichonters	•	•		•		•	•	•							191

					,	J.W.I. 11		VE Z		GI.		
Lepidoptera .				•								198
Diptera .					•		•					264
Siphonaptera							•		•		•	404
Coleoptera .												408
Strepsiptera .							•			•	•	467
Hymenoptera	•	•		•		•	•				•	471
				PA	RT :	II						
		Отн	ER T	ERRES	STRIAI	L Arı	HROE	ODA				
Conspectus of the	ne H	igher	Grou	$\mathbf{ps}$ .					•			529
Onychophora			•							•	•	532
Crustacea, Isop	oda				•							534
Arachnida .												535
Microthelyphon	ida	٠.			•		. •	•	•			539
Pedipalpida .	٠. •							•		•		540
Ricinulei .	٠.			٠.					. •		•	542
Scorpionida .				•	. •			٠.		•		543
Solpugida .	•				•	•	•					546
Chelonethida	•	•	•			•		•	•	•		548
Phalangida .	•	•	•		•	•	•	•	•		•	551
Araneida .	•	•	•	•	•	•	•	•			• -	<b>55</b> 4
Acarina .		•	٠.	•	٠.	*. •	•	•		•	•	565
Pentastomida	٠.	•	•		. •			•		•		579
Tardigrada .	•	•		•		•.		, ·		•		581
Pauropoda .	• •		•	•		. •	. •			•		583
Diplopoda .	•	•	•		•	•		•	. •	. •	• ,	585
Chilopoda .	•	• •	•	•	•		. •	•	•	. •	•	599
Symphyla .	•	•	•	•	•	•	. •	•	•	•	•	603
												ela t L
				PA	RT I	II						
Glossary .	•	•	•	•	•	•	•		٠.	•	•	605
Index	•	•		•	•	•	•			5	•	617

## PREFACE

Due to the great abundance and diversity of insects their taxonomy presents many difficulties and complexities. Classification is consequently a discouraging aspect of entomology to the young student who must attempt in a short space of time to gain a sufficient knowledge of relationships that may enable him to recognize the more common and important types of insect life.

Long teaching experience has served clearly to demonstrate to the authors that the fundamental principles of classification cannot be fully appreciated through the study of descriptive text-books, nor can any comprehension of the infinite variety of nature be acquired except by close observational contact with the things themselves. Moreover, the study of a few selected types by the laboratory method fails to give more than a very superficial view of the organic world, and as biological knowledge advances, tends to emphasize the similarities between animals and to minimize the differences that exist between them. It is, therefore, highly desirable that the principles of taxonomy be presented to the student in a practical way to demonstrate particularly certain of its more fundamental and important phases.

The present volume is the outcome of a series of steps, developed by the authors during the course of many years to train students in the practical taxonomy of insects and to provide a manual for the identification of the larger groups of insects; one which would prove useful and reliable in the hands of professional entomologists and other persons, especially biologists working in other fields, for the actual identification of specimens of insects.

The precursor of the present manual was published by the authors in 1915 and printed privately as a "Key to the Families of North American Insects." It has proved to be very useful and workable in the teaching laboratory both by ourselves and others and has, we believe, proved to be a valuable aid to entomologists and other interested persons who have occasion to identify specimens of insects. We have regretted the geographical limitations of the original book which have made it less useful to workers in other parts of the world, and also the almost entire omission of aids for the identification of the immature stages. The lack of lists of literature was also recognized as a fault as well as the omission of keys which would serve to identify specimens of the various other terrestrial arthropods which are frequently collected by those interested in insects.

The new volume has, therefore, been entirely rewritten to include the families of the entire world, and in many cases subfamilies of the larger or more important groups. A selected list of literature on the several groups has been added, and so far as possible, keys for the identification of the immature stages. We have also prepared an additional part dealing with the numerous other groups of terrestrial Arthropoda.

The keys aim to reproduce as accurately as possible the most generally accepted system of classification of each group. They are necessarily to some extent heterogeneous as they represent the accumulated opinions and revisions of hundreds of workers during the course of many decades. Taxonomy is the oldest branch of biological science and as it has in the past had many more devotees than the recently developed branches, it has a much more complex background. The present treatment can in no sense be considered as original, nor does it follow in detail any previous comprehensive system. There are numerous changes from the arrangement in our previous book, although this has served in a general way, more closely in some groups than others, as a model for the present one. In a few groups where recent extensive revisions have been published, these have been quite generally followed. In others, where there is considerable disagreement among authorities, we have attempted so far as possible to present what seems to be the most generally acceptable classification or the one which appears to be most rational. The arrangement of the Hymenoptera and Diptera represents mainly our own viewpoint, and this is to some extent true of the Coleoptera. The systems followed in certain other groups are indicated from time to time in the text. Where recent monographs of certain groups have appeared, the classification there proposed has been accepted with few changes, although we have by no means made it a rule to regard the newest arrangement as the most satisfactory.

The illustrations have been derived from many sources and are in great part redrawn from published figures, although it must be stated that many have been simplified, differently lettered or otherwise modified to adapt them to the purpose of the present manual. In each case, where not original, the source of each is indicated by the name of the author in parentheses.

Most of the figures from our previous book are reproduced here. They were drawn mainly by Beirne Barrett Brues, the wife of one of the authors. To these have been added an extensive series prepared by Anna Scholl O'Connor, a considerable number by Selina Tetzlaff

Johnson, and a few by others, including the writers. The figures have been selected principally to represent the general appearance of the species of the various groups and the special structures used in classification.

The marking of accents and the indications for the proper pronunciation of the names of genera, families and higher groups has been done in great part by Alice M. Brues, daughter of one of the authors.

The manuscript was transferred into typewritten form chiefly by Mrs. O'Connor who has also aided greatly in the reading of the proof

and preparation of the index.

For advice and criticism on numerous occasions we are greatly indebted to a number of friends and co-workers, as well as to several students, especially Mr. R. P. Dow, who was of great assistance in the preparation of the section on Hymenoptera.

Professor W. M. Wheeler has examined the section on ants, but far more, has offered encouragement and criticism during the tedious

process of preparation.

Finally, we are deeply grateful to Dr. Thomas Barbour for issuing the volume as a contribution from the Museum of Comparative Zoölogy.

No one is more keenly aware than the writers of the great chance that errors of statement or omission may creep into a work of this kind. We have diligently guarded against the occurrence of such errors during the course of preparation, but for any that may remain we trust that we may be forgiven.

## INTRODUCTION

Approximately half a million species of insects have so far been described and named, and their number is being gradually increased from year to year. So far as those competent to judge are able to estimate, it seems probable that this number represents perhaps one-tenth or one-twentieth of those which actually exist upon our planet at the present time. Their descriptions fill libraries and their final identification requires the knowledge of specialists. Obviously no single volume can provide means for their complete determination, and attempts to deal with selected series of abundant species from particular regions are always disappointing and untrustworthy. It is possible to deal quite fully with the families into which the insects are divided within a reasonable space and it is with their recognition that the present work is concerned.

Identification of the families has been effected by means of analytical keys, which have been arranged as dichotomies. There are also provided similar keys, first for the determination of the classes of Arthropoda, and later others for the determination of the orders of each class. The families are then tabulated under each order. Unless the class or order is known, the student should, therefore, first consult the "Key to Classes of Arthropoda" on page 26. If it be known that the specimen is an insect or an arachnid, or that it surely belongs to a particular class or order, the appropriate key (as indicated in the "Table of Contents") will form the starting point for the work of identification.

Each key contains a number of couplets, or pairs of contrasting statements which indicate clearly two characters or groups of characters, one of which will agree with the specimen in hand. In the first couplet, for example, two contrasting descriptions are given, one of which should agree with the insect to be determined. The number at the end of the line following this description indicates the couplet which should next be studied, and so on until the final name is reached. All the keys have been arranged in this way, as the writers experience in the classroom shows that specimens can be most easily, rapidly and accurately classified with a key of this type where the contrasting descriptions appear together on the page and may very easily be compared.

Another type of identification key has the pairs of contrasting characters distinguished from other pairs by differences of indention on the page. Keys thus arranged can be constructed to follow natural relationships more closely, but they require much more space for printing

and in long keys are exceedingly difficult to follow. We have, therefore, not adopted such an arrangement.

While the dichotomies frequently represent natural relationships or lines of phyletic development, we have not attempted to indicate natural divisions wherever the convenience and practical arrangement of the keys would have been sacrificed. It is at best not possible to express the natural relationships of animals or plants by any linear arrangement. It may be said, however, that a carefully arranged dichotomy, in most cases, may be made to conform quite closely to such linear arrangement as may seem most appropriate.

The keys present, therefore, to some extent an artificial, as opposed to a natural sequence, although wherever possible a natural order has been retained. The conspectus of families, on the other hand, aims to present a natural classification as nearly as this can be expressed by a linear sequence. As a linear arrangement is the only one which can be used in a list of this sort, it must be understood that it cannot actually indicate the relationship of the several units by their proximity in the list. We must regard the present fauna as representing a horizontal section of the "phyletic tree" which we see only as a multitudinous series of sections through its twigs and branchlets. The linear arrangement of such a complex cannot even suggest the equally irregular branches to which these twigs were attached in the past. Many of the latter represent extinct groups, and these are not considered in the present book, although the researches of palæontologists have served greatly to clarify our views on the relationships of modern insects.

There is a very pronounced tendency continually to restrict the extent of families and consequently increase their number. This is greatly deplored by many zoölogists and regarded as inevitable, if not highly undesirable by others. The reasons for such a change are manifold. The continual discovery of new forms rapidly increases the number of known species and this in itself merely on the basis of numbers makes it easier to deal with classification if we have a greater number of units of convenient size. The more careful study of anatomical structure frequently leads to the discovery that certain groups are polyphyletic, that is to say, are not of common origin, but represent a convergence or parallelism in the possession of certain closely similar characteristics. If we are to develop a natural classification, such groups must, of course, be divided into a number of smaller families, each easily distinguishable from the others, in fact, more easily characterized than the original large family. Such changes are reasonable and will without doubt be acceptable in the future to all who view them without prejudice. Many such changes represent merely the elevating of groups from subfamily to family rank, and they have been made much more extensively by workers in some groups than in others. For this reason, if for no other the value of family rank varies in the several groups. There is another reason for such differences which relates to the age of the several groups. Those of greater evolutionary age usually present more constant, clear-cut characters and can be grouped into smaller numbers of families while those now in an active state of evolution or differentiation show so many permutations and intergrades that the value of the family has been cheapened and numerous very closely related series have been accorded family rank. As already indicated we have endeavored to follow the general custom of workers in each group in regard to the number of families that should be recognized at the present time.

The characters useful for the separation of family and other major groups are frequently of very minor nature, due to the fact that characteristics of apparently trivial importance commonly persist over long periods of time, presumably because they are not acted upon by natural selection or other evolutionary factors. The value of such characters is everywhere evident, but in the keys many correlated characters of more noticeable and conspicuous kinds have been added, even though, as indicated, they are not invariably present, and cannot be relied on implicitly. When in the keys, such correlation characters are encountered, there is need for judgment in deciding which way to proceed.

As families and other major groups, as well as genera, are concepts and not percepts they are constantly at the mercy of a changing viewpoint and the characters used to define their limits are valid only as accepted by present students. It is hence impossible to standardize the family concept.

Even the selection of family names is not done in a uniform way by all entomologists. This is very unfortunate as it leads to the use in some cases of several names for the same group. Thus the family name may be formed from the oldest contained genus, or it may represent the oldest usage, or it may be still another name long in use, but originally formed by neither of the first two methods. We have not been entirely consistent in the selection of these family designations, but have tried to follow the most general present usage in each group. Synonyms that are or have been in general use are given in italic capital letters placed in parentheses just preceding the accepted family names in the keys. If uniformity is ever secured in the use of family names,

there will be a number of changes in the ones here used, and the citation of synonyms here must consequently be understood to indicate that there is or has recently been a division in usage. Wherever a family name has been suppressed and the group which it designates has been merged with another, the word "Including" has been added in the parenthesis together with the name thus suppressed.

A short, selected list of genera is given for each family. These represent large or important genera, those characteristic of particular regions or those containing common or important species. In most cases the geographical range is given in greatly abbreviated form. The following abbreviations have been used, all of which should be easily understood without explanation: cosmop., cosmopolitan; tropicopol., tropicopolitan; holarc., holarctic (northern hemisphere); palæarc., palæarctic (Europe and northern Asia); nearc., nearctic (America, north of Mexico); neotrop., neotropical (America, south of the United States); Am., new world; ethiop., ethiopian (Africa, south of the Sahara); ind., India; indomal., indomalayan (India and east Indian Islands); austr., Australian. A few variants of these, as indoaustr. and malay., need no explanation. Where genera occur in most of the regions named or in several widely separated ones, their range is indicated as widespr., widespread.

A number of vernacular names for common or important species or groups are given from place to place, enclosed in parentheses. These are mainly ones that are actually in use among persons not acquainted with entomology, although certain others that have been adopted by the American Association of Economic Entomologists as designations for specific insects are listed also. However, no attempt has been made to include a great many that are newly coined, cumbersome, or otherwise of such origin or formation that they may never be expected to come into general popular use as vernacular names.

In addition to the keys to families we have added a considerable number of keys which serve to distinguish the subfamilies of many of the more important or extensive families. These are printed in more condensed form than the family keys and the couplets are numbered a, b, c, etc. to distinguish them from the main keys in which they are inserted next to the family to which they pertain. Thus, although inserted directly into the family keys they remain entirely independent so far as couplet numbers are concerned.

In all cases we have endeavored to cite the family names in exact form. Family names are by rule formed by adding the suffix "idæ" to the Latin or Latinized stem of the name of the type genus. The com-

pound when thus formed should properly include all of the stem. Unless attention is given to the declension of such words according to Latin rules, errors may occur and unfortunately a few incorrectly formed family names have been used from time to time in publications. sometimes over long periods. We have attempted to correct all such errors of orthography, and trust that we have not overlooked any others. Thus family names based on generic names ending in -cera become -ceratida: in -cerus become -cerida: in -ceros become -cerotida, etc. The names of superfamilies have been consistently treated in a similar way, by adding -oidea to the Latin stem of the name of the type genus of the typical family name. No fixed rule is generally accepted for the formation of group names and we have not always corrected the spelling of such names. For example, Nematocera is in general use to replace Nemocera as a suborder of Diptera. Properly this should be Nematocerata and the well known designation of Rhopalocera for the butterflies should be Rhopalocerata. Whether such well established names should be emended is certainly open to question on the basis of long usage.

The pronunciation of all family, group and generic names is indicated in the text, following a very simple and quite satisfactory method which was apparently first used by Asa Gray in 1848 in the first edition of his "Text Book of Botany." Later revisions of Grav's manual have continued the same method but for some reason it has not been very generally adopted by others. An accent mark is placed over the yowel in the accented syllable, thus indicating whether the accent should be placed on the penultimate or antepenultimate syllable. A long vowel is indicated by a grave accent (e.g. è. È) and a short vowel by an acute accent (e.g. \(\ell.\) E). Thus with the length of the important vowel and the accent indicated, the word may be pronounced with small chance of error. Usage is not consistent in the pronunciation of the character "oi" which occurs in the names of superfamilies, etc. We are advocating the sound given to the diphthong in the word "coin," rather than the pronunciation of the two vowels separately. The difficulty of indicating this by means of accented type has led us to mark the diphthong oi with a grave accent over the "o," thus: oi, OI.

As the names are regarded as Latin, the rules for the pronunciation of Latin words are followed, although most generic and family names are actually of Greek derivation. Many new Latin words or those of non-classical origin have to be treated by analogy, for example, those based on the names of entomologists (Winthemia, after Winthem, Meigenia, after Meigen). In some cases, where names do not readily

lend themselves to Latin pronunciation, none is indicated. One of the authors has already dealt at some length with the rules governing the pronunciation of insect names.'

A selected list of literature on the classification of insects and the other arthropods dealt with in the present volume is included in the text. This is intended to introduce the student to the taxonomic literature as completely as is possible in the space which could be devoted to that purpose and since the amount of published material is enormous, only those papers which may be most generally useful have been cited. During the past ten years, approximately 30,000 separate publications on entomology have been issued, by far the greater part of these dealing with taxonomy. Many other earlier monographs and synopses have not vet been supplanted, either wholly or in part by more recent studies. It has, therefore, not been an easy task to determine exactly what ought to be included in such a list. In general we have listed monographs, revisions or synopses of families and larger groups, relating to the fauna of the whole world, or to extensive regions. Often such treatments relating to quite restricted areas are cited where the circumstances seem to warrant their inclusion. Similar publications relating to subfamilies or smaller groups in some cases appear in the lists also where the abundance or importance of the groups concerned render them of special interest. Large works relating to the faunas of extensive regions are included and to some extent the parts of such works are listed separately under the several groups for more ready reference.

Catalogues and bibliographies are cited rather extensively as they serve to introduce the student to the smaller, highly specialized papers which he will find it necessary to consult for the serious study of any particular group. The literature of some of the larger orders like the Coleoptera and Lepidoptera has been more frequently listed than that of the other groups and here we have generally restricted the citations to the most useful works, catalogues, bibliographies and those containing keys for the identification of genera and species. Almost no references to papers containing only scattered descriptions or very incomplete synopses are included since these will be found by an examination of the catalogues and specialized bibliographies.

In many instances long titles have been abbreviated or paraphrased to save space and to render the scanning of the lists easier, but we have tried in all cases to do this in such a way as not to interfere with

<sup>&</sup>lt;sup>1</sup> A. L. Melander. The Pronunciation of Insect Names. Bull. Brooklyn Entom. Soc., **11**, pp. 93-101 (1916).

the usefulness of the lists, for the present purpose. The references are given with sufficient completeness to insure the ready location of each paper.

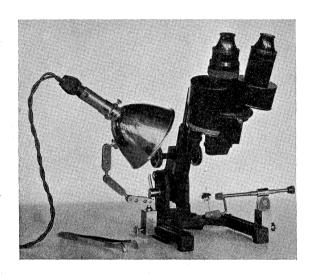
Several languages are necessarily represented, although nearly all of the papers cited are in one of the widely used languages, *i.e.* English, German, French, Italian, Latin, Spanish or Portuguese. The language of the individual papers may be ascertained from the titles, which although abbreviated are given in the language used by the author.

The literature relating to each order is inserted at the end of the keys of that order. In the case of the larger orders, general references are placed together, followed by other lists relating to superfamilies or other groups as indicated in each case, and every individual list is arranged in alphabetical order by authors. The more general works relating to more than a single order are placed at the end of the key to the Orders of Insects on page 38. The literature relating to the Arachnida and other groups of terrestrial Arthropods is arranged in a similar way, in parts following the several keys.

It is impossible to avoid the use of highly technical terms, but we have reduced their number as greatly as clarity permits. A special glossary will be found on page 605, containing definitions of such special terms as are not defined in the text or by reference to figures, or of those that cannot be readily understood by the use of a good dictionary. The nomenclature applied to the body parts, wings, veins, etc., is that used generally in the literature, and represents the current usage in each group. Until quite recently the workers in nearly every group of insects made use of special terminologies for the wing-veins but within the last few years a more or less uniform system of nomenclature is coming into use. This change has been adopted in the present book as representing the most rational method. The student will, however, encounter some difficulties in consulting the older, and even some of the more recent, treatises and papers. Unfortunately this change is one that cannot be avoided.

The equipment necessary for the identification of specimens of insects is quite simple and inexpensive, at least that required for the study of the larger species. Extremely small forms present greater difficulties on account of the very minute size of the structures which must be examined.

For species of moderate or large size, all that is necessary to observe most structural details is a good hand lens. This should be preferably one having a rating of 12x or 14x magnification. Such lenses are made of several types. The best, known as an achromatic triplet, which gives a clear, sharp image, is made of three lenses cemented together into a single piece. Such a lens may be purchased for about six or



AN ENTOMOLOGICAL PRISM BINOCULAR MICROSCOPE Built by the Spencer Lens Company from specifications of A. L. Melander

An inclination joint permits tilting the prism body forward for restful observation and allows reversal of the usual position of the stand, thus affording a free working space unobstructed by the upright support. At the right is an insect holder adjustable on a universal joint, with cork-tipped elbow for holding the insect pin so that the specimen can be rotated constantly in focus. At the left on a gooseneck support is the spotlight reflector, operating on low voltage.

seven dollars; or a pair, giving 8x and 12x magnification respectively, mounted in a single holder for about ten dollars. Cheaper types, known as Coddingtons or doublets may be purchased at considerably lower prices, but their optical properties are comparatively poor and they are very unsatisfactory, except for examining large or very flat

structures. For small insects and for others as well, a binocular microscope is the most satisfactory. Such an instrument, provided with two or three paired objectives and two pairs of eyepieces made by any of several first class manufacturers may be purchased for \$150 or slightly more. With an automobile spotlight, lighted by a storage battery or by a small transformer such as is used for the running of electrical toys, set above the table so as to throw a spot of light on the object, the most minute details of structure may be observed with surprising clearness. Such an apparatus is so much better than any other that its use is strongly to be advised.

As a rule no special method of preparation is necessary since most of the characters made use of in the keys are readily observed on dried pinned specimens. However, in a few cases some previous treatment is necessary. The wings of Lepidoptera must usually be freed from their scaly covering before the venation can be made out. For this, they may be soaked in eau de Javelle to loosen the scales which may then be removed by a camel's hair brush, after which they may be dehydrated, cleared and mounted in balsam or varnish, on slides. Certain small insects such as Thysanoptera may also be mounted very satisfactorily on slides in balsam or spar varnish, preferably the latter as it does not clear transparent structures so completely. Scale insects should be boiled in a solution of caustic potash, washed, dehydrated, cleared and mounted in the same way to show the minute structures upon which these insects are classified.

For the methods of collecting, preparing and preserving insects the reader is referred to a very comprehensive account by Professor Nathan Banks, entitled "Directions for Collecting and Preserving Insects," published as Bulletin No. 67 by the United States National Museum in Washington in 1909.

Directions for the preparation of chitinous structures for study are contained in Lee's "Microtomist's Vade-Mecum," published by P. Blakiston's Sons & Co., Philadelphia (9th edit., 1928), as well as in other less inclusive and pretentious works.

## PART I

## INSECTA

## CONSPECTUS OF THE HIGHER GROUPS OF INSECTA

## Subclass APTERYGOTA

## Order PROTURA

(Eosentomidæ, Acerentomidæ)

#### Order THYSANURA

(Machilidæ, Lepismatidæ)

#### Order ENTOTROPHI

Campodeoidea (Campodeidæ)

Japygoide (Japygide, Projapygide)

#### Order COLLEMBOLA

Suborder **Arthropleona** (Entomobryidæ, Onychiuridæ, Poduridæ) Suborder **Symphypleona** (Sminthuridæ, Neelidæ)

#### Subclass PTERYGOTA

## Order GRYLLOBLATTODEA

(Grylloblattidæ)

#### Order ORTHOPTERA

## Suborder **Tettigoniodea**

Tettigonioidea (Tettigoniidæ, Gryllacridæ, Gryllidæ, Stenopelmatidæ, Phasmodidæ)

Gryllotalpidæ, Tridactylidæ, Cylindrachetidæ)

## Suborder Acridodea

ACRIDOIDEA (Acrididæ, Tetrigidæ, Proscopiidæ)

PNEUMOROIDEA (Pneumoridæ)

## Order PHASMATODEA

Phasmatoidea (Bacillidæ, Phyllidæ, Phasmatidæ)

Bacteriodea (Bacunculidæ, Bacteriidæ)

#### Order DERMAPTERA

#### Suborder Forficulina

#### PROTODERMAPTERA

Pygidicranoidea (Echinosomatidæ, Pyragridæ, Pygidicranidæ, Karschiellidæ, Anateliidæ, Diplatyidæ)

#### PARADERMAPTERA

LABIDUROIDEA (Platylabiidæ, Allostethidæ, Esphalmenidæ, Psalididæ, Labiduridæ, Parisolabidæ, Brachylabidæ)

#### EUDERMAPTERA

Labioidea (Pericomidæ, Nesogastridæ, Vandicidæ, Strongylopsalididæ, Sparattidæ, Spongiphoridæ, Labiidæ)

Forficuloidea (Chelisodochidæ, Cheliduridæ, Anechuridæ, Forficulidæ, Eudohrniidæ, Neolobophoridæ, Aneistrogastridæ, Opisthocosmiidæ, Diaperasticidæ)

Suborder **Arixenina** (Arixeniidæ)

## Order DIPLOGLOSSATA

(Hemimeridæ)

#### Order Thysanoptera

#### Suborder Terebrantia

Жолотня родова (Æolothripidæ, Orothripidæ, Melanothripidæ, Franklinothripidæ)

Thripoidea (Heterothripidæ, Thripidæ, Panchætothripidæ, Ceratothripidæ, Merothripidæ)

## Suborder Tubulifera

Phlceothripidæ, Chlceothripidæ, Ecacanthothripidæ, Eupatithripidæ, Pygothripidæ, Hystrichothripidæ, Megathripidæ, Idolothripidæ, Chirothripoididæ)

UROTHRIPOIDEA (Urothripidæ)

## Order BLATTARIÆ

Phyllodromioidea (Blaberidæ, Oxyhaloidæ, Panchloridæ, Epilampridæ, Nyctiboridæ, Perisphæriidæ, Diplopteridæ, Panesthesiidæ, Cryptocercidæ, Phyllodromiidæ, Areolariidæ, Chorisoneuridæ, Ectobiidæ, Nothoblattidæ, Attaphilidæ)

Blattoidea (Blattidæ, Archiblattidæ, Nocticolidæ)

CORYDIOIDEA (Latindiidæ, Homœogamiidæ, Polyphagidæ, Corydiidæ, Atticolidæ, Euthyrrhaphidæ)

## Order MANTODEA

(Mantidæ)

#### Order EMBIODEA

(Embiidæ, Oligotomidæ)

#### Order ISOPTERA

(Mastotermitidæ, Calotermitidæ, Hodotermitidæ, Rhinotermitidæ, Termitidæ)

#### Order CORRODENTIA

Suborder **Parapsocida** (Phyllipsocidæ, Perientomidæ, Lepidopsocidæ, Psoquillidæ, Atropidæ, Troctidæ, Archipsocidæ)

Suborder **Eupsocida** (Mesopsocidæ, Myopsocidæ, Cæciliidæ, Psocidæ, Amphientomidæ, Thyrsophoridæ)

# Order ZORAPTERA

(Zorotvpidæ)

#### Order MALLOPHAGA

Suborder **Amblycera** (Gyropidæ, Boopiidæ, Trimenoponidæ, Menoponidæ, Læmobothridæ, Ricinidæ)

Suborder **Ischnocera** (Trichodectidæ, Trichophilopteridæ, Nesiotinidæ, Philopteridæ)

Order **ANOPLURA** (Hæmatomyzidæ, Echinophthiriidæ, Hæmatopinidæ, Hæmatopinoididæ, Pediculidæ, Phthiriidæ)

#### Order HOMOPTERA

## Suborder Auchenorrhyncha

CICADOIDEA (Cicadidæ)

Membracoidea (Membracidæ)

Cercopoidea (Machærotidæ, Tomaspididæ, Clastopteridæ, Cercopidæ)

Jassoidea (Cicadellidæ, Gyponidæ, Penthimiidæ, Thaumastoscopidæ, Ledridæ, Bythoscopidæ, Jassidæ, Paropiidæ, Stenocotidæ, Kæbeliidæ, Ulopidæ, Nirvanidæ, Signoretiidæ, Euacanthidæ, Pythamidæ, Æthialoniidæ)

Fulgoroidea (Tettigometridæ, Cixiidæ, Delphacidæ, Derbidæ, Achilixiidæ, Meenoplidæ, Kinnaridæ, Dictyopharidæ, Fulgoridæ, Achilidæ, Tropiduchidæ, Nogodinidæ, Flatidæ, Acanaloniidæ, Issidæ, Ricaniidæ, Lophopidæ, Eurybrachidæ)

## Suborder Sternorrhyncha

Chermoidea (Chermidæ)

ALEYRODOIDEA (Aleyrodidæ)

APHIDOIDEA (Aphididæ, Phylloxeridæ)

Coccoidea (Ortheziidæ, Monophlebidæ, Diaspididæ, Conchaspididæ, Lecaniidæ, Lacciferidæ, Apiomorphidæ, Asterolecaniidæ, Phenacoleachidæ, Coccidæ, Cylindrococcidæ, Eriococcidæ, Kermesidæ)

Suborder Coleorrhyncha (Peloridiidæ, Pleidæ)

#### Order HEMIPTERA

#### Suborder Gymnocerata

Scutelleroidea (Corimelænidæ, Plataspididæ, Cydnidæ, Scutelleridæ, Pentatomidæ, Podopidæ)

COREOIDEA (Corizidæ, Coreidæ, Alydidæ)

GERROIDEA (Gerridæ, Veliidæ)

Aradoidea (Isodermidæ, Aradidæ, Termatophylidæ, Dysodiidæ, Saldidæ, Thaumastotheriidæ, Mesoveliidæ, Hebridæ, Hydrometridæ)

Lygæoidea (Lygæidæ, Hyocephalidæ, Neididæ, Colobothristidæ, Pyrrhocoridæ)

TINGIDOIDEA (Piesmidæ, Tingididæ)

Reduvioidea (Henicocephalidæ, Phymatidæ, Reduviidæ, Ploiariidæ, Nabidæ, Joppeicidæ)

POLYCTENOIDEA (Polyctenidæ)

Cimicoidea (Aepophilidæ, Cimicidæ, Velocipedidæ, Anthocoridæ, Miridæ, Microphysidæ, Isometopidæ)

DIPSOCOROIDEA (Dipsocoridæ, Schizopteridæ)

HELOTREPHOIDEA (Helotrephidæ)

Suborder **Cryptocerata** (Nerthridæ, Ochteridæ, Naucoridæ, Nepidæ, Belostomatidæ, Notonectidæ, Corixidæ)

## Order ODONATA

## Suborder Zygoptera

Cœnagriouea (Hemiphlebiidæ, Protoneuridæ, Cœnagriidæ, Synlestidæ, Megapodagriidæ, Lestidæ, Pseudostigmatidæ)

AGRIOIDEA (Amphipterygidæ, Libellaginidæ, Epallagidæ, Polythoridæ, Agriidæ)

## Suborder Anisozygoptera

Ергорныевгогова (Epiophlebiidæ)

#### Suborder Anisoptera

ÆSCHNOIDEA (Petaluridæ, Gomphidæ, Cordulegastridæ, Æschnidæ)

LIBELLULOIDEA (Cordulidæ, Libellulidæ)

## Order PLECTOPTERA

EPHEMEROIDEA (Palingeniidæ, Polymitarcidæ, Ephemeridæ, Potamanthidæ, Prosopistomatidæ)

Baetoidea (Baetidæ, Caenidæ, Leptophlebiidæ, Ephemerellidæ, Oligoneuridæ)

SIPHLUROIDEA (Baetiscidæ, Siphluridæ, Ametropodidæ, Heptageniidæ)

## Order PLECOPTERA

(Perlidæ, Pteronarcidæ, Eustheniidæ, Austroperlidæ, Leptoperlidæ, Capniidæ, Nemouridæ)

## Order MEGALOPTERA

CORYDALOIDEA (Corydalidæ)

Sialoidea (Sialidæ)

# Order RAPHIDIODEA (Raphidiidæ)

## Order NEUROPTERA

ITHONOIDEA (Ithonidæ)

Немековіопеа (Hemerobiidæ, Sympherobiidæ, Dilaridæ, Psychopsidæ, Osmylidæ, Polystæchotidæ, Sisyridæ, Chrysopidæ, Apochrysidæ, Berothidæ, Trichomatidæ, Mantispidæ)

NEMOPTEROIDEA (Nemopteridæ)

MYRMELEONTOIDEA (Myiodactylidæ, Nymphidæ, Myrmeleontidæ, Stilbopterygidæ, Ascalaphidæ)

CONIOPTERYGOIDEA (Coniopterygidæ)

#### Order MECOPTERA

Suborder Protomecoptera (Notiothaumidæ, Meropidæ)

Suborder **Eumecoptera** (Panorpidæ, Boreidæ, Bittacidæ)

## ${\bf Order} \ \ {\bf TRICHOPTERA}$

# Suborder Æquipalpia

RHYACOPHILOIDEA (Rhyacophilidæ, Philopotamidæ, Polycen-

tropodidæ, Hydropsychidæ, Psychomyiidæ, Calamoceratidæ, Odontoceridæ, Leptoceridæ, Molannidæ)

HYDROPTILOIDEA (Hydroptilidæ)

#### Suborder Inæquipalpia

Phryganoidea (Phryganeidæ, Limnephilidæ)

Sericostomatoidea (Sericostomatidæ)

#### Order LEPIDOPTERA

## Suborder Jugatæ

MICROPTERYGOIDEA (Micropterygidæ, Eriocraniidæ, Mnesarchæidæ)

Hepialoidea (Hepialidæ, Prototheoridæ, Anomosetidæ, Palæosetidæ)

#### Suborder Frenatæ

Cossoidea (Zeuzeridæ, Cossidæ, Argyrotypidæ, Metarbelidæ, Hypoptidæ, Ratardidæ)

Castnioidea (Castniidæ, Tascinidæ)

Psycholdea (Psychidæ, Talæporiidæ, Heterogynidæ, Zygænidæ, Charidæidæ, Himantopteridæ, Pyromorphidæ, Eucleidæ, Megalopygidæ, Dalceridæ, Chrysopolomidæ, Mimallonidæ)

TORTRICOIDEA (Olethreutidæ, Tortricidæ, Phaloniidæ, Carposinidæ)

Pterophoroidea (Pterophoridea, Orneodidea, Agdistidea, Oxychirotidea)

Pyralidoidea (Pyralididæ, Tineodidæ, Hyblæidæ)

Uranioldea (Epiplemidæ, Uraniidæ, Lonomiidæ, Epicopeiidæ) Bombycoidea (Lasiocampidæ, Bombycidæ, Lemoniidæ, Citheroniidæ, Saturniidæ, Eupterotidæ, Brahmæidæ, Oxytenidæ, Cercophanidæ)

Geometriolea (Œnochromatidæ, Boarmidæ, Geometridæ, Sterrhidæ, Larentiidæ, Hemitheidæ, Acidaliidæ, Brephidæ)

Sphingoidea (Sphingidæ)

Tineoidea (Adelidæ, Incurvariidæ, Nepticulidæ, Ashinagidæ, Amphitheridæ, Agonoxenidæ, Acrolophidæ, Setomorphidæ, Tineidæ, Amydriidæ, Tischeriidæ, Lyonetiidæ, Opostegidæ, Œnophilidæ, Prodoxidæ, Epipyropidæ, Coleophoridæ, Momphidæ, Elachistidæ, Phyllocnistidæ, Heliozelidæ, Douglasiidæ, Œcophoridæ, Ethmiidæ, Xyloryctidæ, Stenomidæ, Gelechiidæ, Metachandidæ, Epimarptidæ, Blastobasidæ, Hyposmocomidæ, Cosmopterygidæ, Scythrididæ, Epermeniidæ, Argyresthesiidæ, Chlidanotidæ, Yponomeutidæ, Cecidosidæ, Ridiaschinidæ, Plutellidæ, Cyclotornidæ, Anomologidæ, Glyphipterygidæ, Heliodinidæ, Copromorphidæ, Ægeriidæ)

Noctuoidea (Endromidide, Anthelide, Drepanide, Thyatiride, Hypside, Lymantriide, Noctuide, Lithosiide, Plusiide, Nolide, Hylophilide, Cocytiide, Arctiide, Euchromiide,

Agaristidæ, Pericopidæ, Apoprogenidæ, Sematuridæ, Cymatophoridæ, Callidulidæ, Pterothysanidæ)

## Suborder Rhopalocera

HESPERIOIDEA (Euschemonidæ, Megathymidæ, Hesperiidæ)

Papilionoidea (Asciidæ, Papilionidæ, Parnassiidæ, Danaidæ, Eueididæ, Ithomiidæ, Satyridæ, Brassolidæ, Morphoidæ, Nymphalidæ, Libytheidæ, Riodinidæ, Lycænidæ)

#### Order DIPTERA

#### Suborder Nematocera

TIPULOIDEA (Trichoceratidæ, Tipulidæ, Limoniidæ)

PSYCHODOIDEA (Psychodidæ, Tanyderidæ, Ptychopteridæ)

BLEPHAROCERATOIDEA (Blepharoceratidæ, Deuterophlebiidæ)

Chironomoidea (Chironomidea, Ceratopogonidea, Thaumaleidea, Simuliidea)

Culicoidea (Culicidæ, Dixidæ)

Anisopodidea (Anisopodidea, Mycetobiidea, Pachyneuridea)

BIBIONOIDEA (Hesperinidæ, Bibionidæ, Scatopsidæ)

Мусеторні (Mycetophilidæ, Sciophilidæ, Bolitophilidæ, Ditomyiidæ, Ceroplatidæ, Macroceratidæ, Diadocidiidæ, Sciaridæ, Cecidomyiidæ)

## Suborder Brachycera

## Division ORTHORRHAPHA

Stratiomyioidea (Stratiomyiidee, Pantophthalmidee, Chiromyzidee, Xylomyiidee)

Tabanoidea (Rhagionidæ, Xylophagidæ, Cœnomyiidæ, Tabanidæ)

Therevoidea (Therevidæ, Scenopinidæ, Nemestrinidæ, Acroceratidæ)

ASILOIDEA (Bombyliidæ, Asilidæ, Apioceratidæ, Mydaidæ)

Емриопрем (Empididæ, Dolichopodidæ)

#### Division CYCLORRHAPHA

#### Series Aschiza

LONCHOPTEROIDEA (Lonchopteridæ)

Рнової DEA (Phoridæ, Termitoxeniidæ, Thaumatoxenidæ, Braulidæ)

PLATYPEZOIDEA (Platypezidæ, Sciadoceratidæ)

Syrphoidea (Pipunculidæ, Syrphidæ)

#### Series Schizophora

Section Myodaria

Muscoidea Thecostomata (Tachinidæ, Dexiidæ, Phasiidæ, Rhinophoridæ, Sarcophagidæ, Calliphoridæ; Muscidæ, Glossinidæ, Anthomyiidæ, Cordyluridæ, Æstridæ)

Muscoidea Haplostomata (Gastrophilide, Ctenostylide, Conopide, Pyrgotide, Phytalmiide, Platystomatide, Pterocallide, Ortalide, Ulidiide, Richardiide, Rhopalomeride, Rhinotoridæ, Trypetidæ, Tachiniscidæ, Lonchæidæ, Pallopteridæ, Tanypezidæ, Nothybidæ, Micropezidæ, Neriidæ, Sepsidæ, Piophilidæ, Thyreophoridæ, Megamerinidæ, Diopsidæ, Clusiidæ, Tetanoceratidæ, Dryomyzidæ, Neottiophilidæ, Lauxaniidæ, Celyphidæ, Periscelidæ, Cælopidæ, Helomyzidæ, Trichoscelidæ, Chyromyiidæ, Psilidæ, Anthomyzidæ, Opomyzidæ, Ochthiphilidæ, Tethinidæ, Agromyzidæ, Odiniidæ, Carnidæ, Milichiidæ, Cryptochætidæ, Drosophilidæ, Diastatidæ, Astiidæ, Aulacogastridæ, Canaceidæ, Ephydridæ, Oscinidæ, Borboridæ, Leptoceratidæ)

Section Pupipara

HIPPOBOSCOIDEA (Hippoboscidæ, Streblidæ, Nycteribiidæ)

#### Order SIPHONAPTERA

Suborder **Fractocipita** (Ischnopsyllidæ, Hystrichopsyllidæ, Macropsyllidæ)

Suborder **Integricipita** (Pulicidæ, Ctenophthalmidæ, Dolichopsyllidæ, Uropsyllidæ, Hectopsyllidæ, Tungidæ)

## Order COLEOPTERA

## Suborder Adephaga

Caraboidea (Cicindelidæ, Carabidæ, Pseudomorphidæ, Omophronidæ, Amphizoidæ, Hygrobiidæ, Haliplidæ, Dytiscidæ)

GYRINOIDEA (Gyrinidæ)

Paussoidea (Paussidæ)

CUPOIDEA (Cupidæ)

Rhysodoide, Jacobsoniide)

## Suborder Polyphaga

HYDROPHILOIDEA (Hydrophilidæ, Hydroscaphidæ)

STAPHYLINOIDEA (Silphidæ, Thorictidæ, Sphæritidæ, Leptinidæ, Clambidæ, Scaphidiidæ, Scydmænidæ, Brathinidæ, Staphylinidæ, Platypsyllidæ, Pselaphidæ, Clavigeridæ)

CUCUJOIDEA (Cucujidæ, Parandridæ, Helotidæ, Passandridæ, Silvanidæ)

Cantharoidea (Lampyridæ, Cantharidæ, Lycidæ, Drilidæ, Lymexylidæ, Micromalthidæ, Atractoceridæ, Telegeusidæ, Dasytidæ, Malachiidæ, Cleridæ, Corynetidæ)

MORDELLOIDEA (Cephaloidæ, Œdemeridæ, Mordellidæ, Rhipiphoridæ, Meloidæ, Eurystethidæ, Pythidæ, Pyrochroidæ, Pedilidæ, Anthicidæ, Euglenidæ)

ELATEROIDEA (Cerophytidæ, Cebrionidæ, Plastoceridæ, Rhipiceratidæ, Elateridæ, Melasidæ, Throscidæ, Buprestidæ)

Dryopoidea (Psephenidea, Dryopidea, Helmidea, Heteroceridea, Georyssidea)

DASCYLLOIDEA (Dascyllidæ, Helodidæ, Chelonariidæ, Dermestidæ, Byturidæ, Byrrhidæ, Nosodendridæ, Ptinidæ, Ectrephidæ, Gnostidæ, Eucinetidæ)

HISTEROIDEA (Histeridæ, Niponiidæ, Synteliidæ)

Colydioidea (Colydiidea, Murmidiidea, Ostomatidea, Monotomidæ, Cryptophagidæ, Nitidulidæ, Brachypteridæ, Rhizophagidæ, Ciidæ, Mycetophagidæ, Erotylidæ, Languriidæ, Catopochrotidæ, Phalacridæ, Lvctidæ, Anobiidæ, Bostrychidæ, Lathridiidæ, Monotomidæ, Derodontidæ, Endomychidæ, Mycetæidæ, Coccinellidæ, Orthoperidæ, Phænocephalidæ, Sphæriidæ, Ptiliidæ, Hydroscaphidæ, Discolomidæ, Cyathoceridæ)

Tenebrionoidea (Tenebrionidea, Cossyphodidea, Cistelidea, Petriidæ, Lagriidæ, Elacatidæ, Nilionidæ, Sphindidæ, Melandryidæ, Scraptiidæ, Monommatidæ, Trictenotomidæ)

CERAMBYCOIDEA (Prionidæ, Lamiidæ, Cerambycidæ, Donaciidæ, Megascelidæ, Sagridæ, Crioceridæ, Cryptocephalidæ, Chlamydidæ, Megalopodidæ, Clytridæ, Chrysomelidæ, Lamprosomatidæ, Eumolpidæ, Galerucidæ, Halticidæ, Hispidæ, Cassididæ, Mylabridæ)

CURCULIONOIDEA (Brentidæ, Cyladidæ, Rhinomaceridæ, Curculionidæ, Platypodidæ, Chapuisiidæ, Scolytidæ, Scolytoplatypodidæ, Ipidæ, Anthribidæ, Aglycideridæ, Proterhinidæ)

Scarabæoidea (Passalidæ, Lucanidæ, Sinodendridæ, Trogidæ, Scarabæidæ, Geotrupidæ, Pleocomidæ, Glaphyridæ, Orphnidæ, Ochodæidæ, Hybosoridæ, Aphodiidæ, Ægialiidæ, Melolonthidæ, Euchiridæ, Rutelidæ, Pachypodidæ, Dynastidæ, Phænomeridæ, Cetoniidæ, Trichiidæ)

#### Order STREPSIPTERA

Mengeoidea (Mengeidæ, Mengenillidæ)

STICHOTREMATOIDEA (Stichotrematidæ)

Xenoidea (Callipharixenidæ, Myrmecolacidæ, Stylopidæ, Hylecthridæ. Xenidæ)

Halictophagoidea (Diozoceratidæ, Halictophagidæ)

Elenchoidea (Elenchidæ)

## Order HYMENOPTERA

# Suborder Chalastogastra

Pamphilioidea (Pamphiliidea Xvelidea)

Tenthredinoidea (Megalodontidæ, Cimbicidæ, Pergidæ, Argidæ, Blasticotomidæ, Tenthredinidæ, Diprionidæ)

Siricoidea (Xiphydriidæ, Cephidæ, Siricidæ)

Oryssoidea (Oryssidæ)

# Suborder Clistogastra

## Division TEREBRANTIA

ICHNEUMONOIDEA (Megalyridæ, Stephanidæ, Aulacidæ, Gasteruptionidæ, Evaniidæ, Ichneumonidæ, Braconidæ, Agriotypidæ, Alysiidæ, Trigonalidæ)

Chalcidoidea (Callimomidæ, Agaontidæ, Leucospididæ, Or-

myridæ, Eurytomidæ, Perilampidæ, Eucharididæ, Cleonymidæ, Encyrtidæ, Pteromalidæ, Miscogastridæ, Elasmidæ, Lepto'cenidæ, Eulophidæ, Trichogrammatidæ, Mymaridæ)

Serphoidea (Pelecinidæ, Monomachidæ, Heloridæ, Roproniidæ, Vanhorniidæ, Serphidæ, Belytidæ, Diapriidæ, Scelionidæ, Calliceratidæ, Platygastridæ)

Cynipoidea (Ibaliide, Cynipide, Figitide)

Division ACULEATA

Bethyloidea (Rhopalosomatidæ, Bethylidæ, Dryinidæ, Embolemidæ, Sclerogibbidæ)

Chrysidide, Chrysidide, Alienide)

Scoliodea (Sapygidæ, Plumariidæ, Scoliidæ, Tiphiidæ, Anthoboscidæ, Sierolomorphidæ, Thynnidæ, Methocidæ, Myrmosidæ, Mutillidæ, Apterogynidæ)

FORMICOIDEA (Formicidæ)

Vespoidea (Vespidæ)

PSAMMOCHAROIDEA (Psammocharidæ)

Sphecoidea (Ampulicidæ, Sphecidæ, Mellinidæ, Gorytidæ, Nyssonidæ, Trypoxylidæ, Stizidæ, Dimorphidæ, Bembicidæ, Larridæ, Miscophidæ, Philanthidæ, Cerceridæ, Alysonidæ, Pemphredonidæ, Crabronidæ, Oxybelidæ)

Apoidea (Colletidæ, Prosopididæ, Andrenidæ, Megachilidæ, Panurgidæ, Podaliriidæ, Bombidæ, Euglossidæ, Xylocopidæ, Ceratinidæ, Stelididæ, Chrysanthedidæ, Nomadidæ, Melectidæ, Apidæ)

# KEY TO THE CLASSES OF ARTHROPODA

1.	Antennæ absent; often four or five pairs of legs, sometimes more, very rarely less
	One or two pairs of antennæ present; legs variable, often three or
	many pairs9
2.	Marine animals, occasionally living near the tidal zone3
	Terrestrial, rarely aquatic species living in fresh water; never
	living in the ocean below the tidal zone4
3.	Respiratory organs well developed, consisting of blood gills; four
	anterior pairs of legs chelate at tip; large animals, the body
	covered by a convex carapace. King crabs, Horseshoe crabs.
	XIPHOSŪRA
	Respiratory organs absent or vestigial; none of the legs chelate at
	tip; body small, with extremely long legs PYCNOGÓNIDA
4.	Four pairs of legs in the adult, sometimes with another more
	anterior pair of appendages (pedipalpi) resembling walking
	legs
_	Three pairs of legs or less6
5.	With a well developed respiratory system composed of book lungs
	or of tubular tracheæ opening by spiracles, or of both in the
	same animal; reproductive organs opening near the base of the
	abdomen below; integument usually heavily chitinized at least
	on part of the body. Spiders, Scorpions, Mites, Ticks, etc.
	ARÂCHNIDA (Page 535) Without any special respiratory organs, reproductive organs open-
	ing into the alimentary canal; very small or microscopic animals
	with weakly chitinized integument, living in damp places or
	aquatic. Water bears, Bear animalcules.
	TARDÌGRADA (Page 581)
6	With three pairs of legs, or with only two pairs, in the latter case
•	the body is vermiform and shows many minute transverse
	wrinkles or annulations; small or minute animals7
	Without legs in the adult which is vermiform and shows many
	minute transverse wrinkles or annulations, or with two pairs
	in the embryo or young larva which is short and not thus an-
	nulate; internal parasites of vertebrates (see couplet 5). (Order
	Pentastomida)
7.	Three pairs of legs8
	Two pairs of legs; body long, tapering, the integument with

	minute transverse rings or wrinkles; living in plant or animal
	tissues. Some Acarina (see couplet 5).
	ARÁCHNIDA, part. (Page 535)
8.	Body elongate; abdomen consisting of eleven segments, its basal
	three segments each bearing a pair of vestigial legs. (Order
	Protura. (If internal parasites, cf. Strepsiptera, p. 467).
	insécta, part. (Page 28)
	Body short, rounded or oval; abdomen not segmented and without
	appendages below. The young of some Acarina.
	ARÁCHNIDA, part. (Page 535)
9.	Only one pair of antennæ10
	Two pairs of antennæ; five or more pairs of legs; aquatic (very
	rarely terrestrial) animals living in the sea or in fresh water,
	provided with true gills, except in some parasitic forms.
	CRUSTÂCEA (Page 534)
10.	With three pairs of legs in the adult, and usually with wings; legs
	reduced in size and structure or often completely absent in the
	larvaINSÉCTA (Page 28)
	More than three pairs of legs; no wings; body usually very elon-
	gate
11.	0 / 1
	of distinct joints, but of a different type from the more posterior
	pairs which are not clearly jointed. Larvæ of some insects.
	INSÉCTA, part. (Page 28) Legs essentially similar, at most only the first pair strikingly
	different from the more posterior ones
19	Legs composed of distinct, heavily chitinized segments which are
14.	articulated by sharply differentiated joints; integument usu-
	ally heavily chitinized
	Legs fleshy with numerous annulations, but without any clearly
	articulated joints; long cylindrical, soft-bodied animals. Tropi-
	calONYCHÓPHORA (Page 532)
13.	
10.	Two pairs of legs on some, usually on most of the segments of the
	body (as indicated by the tergites); body usually more or less
	cylindricalDIPLÓPODA (Page 585)
14.	Antennæ simple, not branched
	Antennæ branched apically, terminating in three multiarticulate
	setiform appendages; nine pairs of legs; minute animals without
	respiratory organsPAURÓPODA (Page 583)
15.	Legs terminating in a single tarsal claw; first pair of legs modified

to form jaw-like poison fangs; at least nineteen body segments and fifteen or more pairs of legs in the adult. Centipedes.

CHILÓPODA (Page 599)

Legs each bearing two tarsal claws; first pair of legs more or less reduced in size, not jaw-like and without poison gland; 15 to 22 body segments and twelve pairs of legs.

SÝMPHYLA (Page 603)

## CLASS INSÉCTA

# $(HEX \acute{A} PODA)$

Small or moderate-sized, frequently very small or minute, never very large. Arthropoda. Body of adult and sometimes also of the immature stages more or less clearly divided into three groups of segments, head, thorax and abdomen: those forming the head fused into a single piece. Three thoracic segments, each bearing a pair of legs; the first segment often much more freely articulated than the others; second and third each usually bearing a pair of wings, sometimes absent on the third or entirely wanting. Abdomen composed of eleven segments or less, frequently six to eight, often terminated by a pair of cerci. One pair of antennæ almost always present; three pairs of mouthparts; a pair of mandibles fitted for chewing or piercing, and two pairs of maxillæ, the latter usually bearing a jointed palpus and the first pair usually biramose. Compound eyes and three simple ocelli usually present. Legs almost always terminating in claws, usually nine- or eight-jointed, occasionally less. Wings usually supplied with a series of branched veins or chitinous thickenings, fore pair often thicker than the hind ones. Respiration by branched, tubular tracheæ opening by segmentally arranged pairs of spiracles. Development direct in the primitive forms or with a metamorphosis often involving great changes in form and habits. Younger stages without functional wings; in the forms undergoing metamorphosis with the legs and antennæ much reduced and the body grub-like, caterpillar-like, or vermiform. Insects.

# KEY TO THE ORDERS OF INSECTA

- 2. The wings of the mesothorax (the fore wings) horny, leathery or parchment-like, at least at the base; differing materially in texture from the membranous hind wings which exceptionally

	may be absent. Prothorax large and not fused with the meso-
	thorax (except in the rare Strepsiptera, which have minute
	fore wings)
	The mesothoracic wings membranous
3.	Mesothoracic wings (called tegmina or hemelytra) containing
	veins, or at least the metathoracic wings not folded crossways
	when hidden under the upper wings4
	Mesothoracic wings (called elytra) veinless, of uniform, horny
	consistency, the metathoracic wings, when present, folded cross-
	ways as well as lengthwise when at rest and hidden beneath the
	elytra; mouth mandibulate11
4.	
-	ing
	Mouthparts with mandibles fitted for chewing and moving later-
	ally
5.	Head usually horizontal and with the beak arising from the under
•	part so as to project downwards, the gula well developed; meso-
	thoracic wings usually leathery at the base and abruptly mem-
	branous on the apical portion, the membranous parts usually
	overlapping one another and lying flat over the abdomen when
	at rest. True bugs
	Head usually vertical and with the beak arising from the back
	part so as to project backward between the front legs; gula
	absent, or represented by a small membrane.
	HOMÓPTERA (Page 106)
6.	Hind wings not folded, similar to the fore wings; social species,
	living in colonies. Termites ISÓPTERA (Page 93)
	Hind wings folding, fan-like, broader than the fore wings7
7.	Usually rather large or moderately large species; antennæ usually
	lengthened and thread-like; prothorax large and free from the
	mesothorax; cerci present; fore wings rarely minute, usually
	long8
	Very small active species; antennæ short, with few joints, at least
	one joint bearing a long lateral process; no cerci; fore wings
	minute; prothorax small. Rare, short-lived insects, parasites
	of other insects, usually wasps and bees.
	Males of STREPSIPTERA (Page 467)
8.	Hind femora not larger than the fore femora; mute species; body
	more or less flattened with the wings superposed when at rest;
	tergites and sternites subequal9
	Hind femora almost always much larger than the fore femora,

jumping species, if not (Gryllotalpidæ) the front legs broadened

	for burrowing; species usually capable of chirping or making a
	creaking noise; body more or less cylindrical, the wings held
	sloping against the sides of the body when at rest; tergites usu-
	ally larger than the sternites. Grasshoppers, Katydids, Crickets.
	ORTHÓPTERA $(Page 48)$
9.	Body elongate; head free, not concealed from above by the pro-
	thorax; deliberate movers10
	Body oval, much flattened; head nearly concealed beneath the
	oval pronotum; legs similar and fitted for rapid running, the
	coxæ large. RoachesBLATTÀRIÆ (Page 77)
10.	Prothorax much longer than the mesothorax; front legs almost
	always heavily spined, formed for seizing prey; cerci usually
	with several joints. Mantes, Leaf insects.
	MANTÒDEA (Page 83)
	Prothorax short; legs similar, formed for walking; cerci one-
	jointed. Stick insects, Walking sticks.
	PHASMATÒDEA (Page 61)
11.	Abdomen terminated by movable, almost always heavily chiti-
	nized forceps; antennæ long and slender; fore wings short,
	leaving most of the abdomen uncovered, hind wings nearly
	circular, delicate, radially folded from near the center; elongate
	insects. Earwigs DERMÁPTERA (Page 65)
	Abdomen not terminated by forceps; antennæ of various forms
	but usually eleven-jointed; fore wings usually completely
	sheathing the abdomen; generally hard-bodied species. Beetles,
	Weevils
12.	With four wings
	With but two wings (the mesothoracic) usually outspread when
	at rest
13.	Wings long, very narrow, the margins fringed with long hairs, al-
	most veinless; tarsi one- or two-jointed, with swollen tip
	mouthparts asymmetrical, without biting mandibles, fitted
	for lacerating and sucking plant tissues; no cerci; minute spe-
	cies. Thrips
	Wings broader and most often supplied with veins, if rarely some-
	what linear the tarsi have more than two joints and the last
	tarsal joint is not swollen
14	Hind wings with the anal area folded in plaits, fan-like, in repose
	I very rarely (one African genus) with apical forceps-like appendages, the wings are not a ermaptera.

	larger than the fore wings; antennæ prominent; veins usually
	numerous; larvæ aquatic
	Hind wings not folded, not larger than the fore wings, the anal
	area small and not separated17
5.	Tarsi five-jointed; cerci not pronounced
	Tarsi three-jointed; body rather flattened, with jointed cerci;
	wings at rest overlapping the abdomen, prothorax large, free;
	species of moderate to large size. Stone-flies.
	PLECÓPTERA (Page 176)
16.	Costal area with few crossveins; wing with the surface hairy; pro-
	thorax small; species of small to moderate size. Caddice flies.
	TRICHÓPTERA (Page 191)
	Costal area with many crossveins; prothorax rather large; species
	of moderate to large size MEGALÓPTERA (Page 179)
17.	Antennæ short and inconspicuous; wings netveined with numerous
	crossveins; nymphs aquatic. (SUBULICÓRNIA)18
	Antennæ larger, distinct, if rarely small the crossveins are few;
	larvæ almost always terrestrial19
18.	Hind wings much smaller than the fore wings; abdomen ending in
	long, thread-like processes; tarsi normally four- or five-jointed;
	sluggish fliers. May flies PLECTÓPTERA (Page 169)
	Hind wings nearly like the fore wings; no caudal setæ; tarsi three-
	jointed; vigorous, active fliers, often of large size. Dragon flies,
	Damsel flies
19.	Head produced into a mandibulate beak (Fig. 354), hind wings
	not folded; wings usually with color pattern, the crossveins
	numerous; male genitalia usually greatly swollen, forming a
	reflexed bulb. Scorpion flies MECOPTERA (Page 189)
	Head not drawn out as a mandibulate beak; male abdomen not
	forcipate
20.	Mouth mandibulate
	Mouth haustellate, the mandibles not formed for chewing; no
	cerci; crossveins few
21.	Tarsi five-jointed, if rarely three- or four-jointed, the hind wings
	are smaller than the front ones and the wings lie flat over the
	body; no cerci22
	Tarsi two-, three- or four-jointed; veins and crossveins not numer-
	ous
<b>2</b> 2.	Prothorax small or only moderately long. (In Mantispidæ the
	prothorax is very long, but the front legs are strongly rap-
	torial)

	Prothorax very long and cylindrical, much longer than the head;
	front legs normal; antennæ with more than eleven joints; cross-
	veins numerous
23.	Wings similar, with many veins and crossveins; prothorax more or less free. If the neuration is very rarely reduced (Coniopteryx) the wings are powdered
	Wings with relatively few angular cells, the costal cell without
	crossveins; hind wings smaller than the fore pair; prothorax
	fused with the mesothorax; abdomen frequently constricted
	at the base and ending in a sting or specialized ovipositor.
	Ants, Wasps, Bees, etc
24.	
	crossveins
	Costal cell without crossveins MECÓPTERA (Page 189)
25.	Wings equal in size, or rarely the hind wings larger, held super-
	posed on the top of the abdomen when at rest; media fused
	with the radial sector for a short distance near the middle of the
	wing; tarsi three-, four-, or five-jointed
	Hind wings smaller than the fore wings; wings held at rest folded
	back against the abdomen; radius and media not fusing; tarsi two- or three-jointed
26	Tarsi apparently four-jointed; cerci usually minute; social species,
	living in colonies ISÓPTERA (Page 93)
	Tarsi three-jointed, the front metatarsi swollen; cerci conspicu-
	ous; usually solitary species
<b>27</b> .	Cerci absent; tarsi two- or three-jointed; wings remaining attached
	throughout life; radial sector and media branched, except
	when fore wings are much thickened.
	CORRODÉNTIA (Page 96)
	Cerci present; tarsi two-jointed; wings deciduous at maturity,
	the neuration greatly reduced; radial sector and media simple,
	unbranchedZORÁPTERA (Page 100)
28.	Wings not covered with scales, not outspread when at rest; pro-
	thorax large; antennæ with few joints; mouthparts forming a
	jointed piercing beak
	Wings and body covered with colored scales which form a definite
	pattern on the wings; antennæ many-jointed; mouthparts when
	present forming a coiled tongue. Moths and Butterflies.
	LEPIDÓPTERA (Page 198)
20	Real arising from the heal of the head

HOMÓPTERA (Page 106)

	Beak arising from the front part of the head. <b>HEMÍPTERA</b> (Page 140)
30.	Mouth not functional; abdomen furnished with a pair of caudal filaments
	Mouthparts forming a proboscis, only exceptionally vestigial; abdomen without caudal filaments; hind wings replaced by
	knobbed halteres. Flies, Mosquitoes, Midges. <b>DÍPTERA</b> (Page 264)
31.	No halteres; antennæ inconspicuous; crossveins abundant. A few
ы.	rare May flies
	Hind wings represented by minute hook-like halteres; antennæ
	evident; neuration reduced to a forked vein; crossveins lacking;
	minute delicate insects. Males of Scale insects.
	HOMÓPTERA (Page 106)
32.	Body more or less insect-like, i.e. with more or less distinct head,
	thorax and abdomen, and jointed legs, and capable of locomo-
	tion
	Without distinct body parts, or without jointed legs, or incapable
	of locomotion
33.	Terrestrial, breathing through spiracles; rarely without special
	respiratory organs
	Parasites on warm-blooded animals
34.	Mouthparts vestigial, retracted in the head and scarcely or not
JT.	at all visible; underside of the abdomen with styles or other ap-
	pendages; very delicate small or minute insects. (APTERY-
	GOTA)
	Mouthparts mandibulate, formed for chewing (if body is covered
	with scales, see Thysanura)
35.	· · · · · · · · · · · · · · · · · · ·
JJ.	at its base, no terminal springing apparatus
	Abdomen consisting of six segments or less, with a forked
	sucker at base below and usually with a springing apparatus
	(furcula) near the tip beneath; cerci absent.
	COLLÉMBOLA (Page 44)
36.	Basal three segments of the abdomen with ventral styles; antennæ
	absent; no cerci, but a short anal tube present; head pear-shaped;
	prothorax short; abdomen with twelve segments; minute,
	cylindrical species. (Class MIRIENTÓMATA).
	PROTURA (Page 40)

	Ventral styles occurring to the seventh segment; antennæ thread
	like; cerci present; prothorax not short
37.	Body never scaly; mouthparts concealed except for the palpi
	apex of the abdomen without a median process.
	ENTÓTROPHI (Page 43)
	Body usually covered with minute scales; tips of the mouthparts
	visible; abdomen with a median cerciform appendage.
	THYSANÙRA $(Page 41)$
38.	Underside of abdomen entirely without legs39
	Abdomen bearing false legs beneath which differ from those of
	the thorax; body caterpillar-like, cylindrical, the thorax and
	abdomen not distinctly separated; larval forms55
39.	Antennæ long and distinct40
	Antennæ short, not pronounced; larval forms
40.	Abdomen terminated by strong movable forceps; prothorax free.
	Earwigs DERMÁPTERA (Page 65)
	Abdomen not ending in forceps41
41.	Abdomen not strongly constricted at the base, broadly joined to
	the thorax42
	Abdomen strongly constricted at the base; prothorax fused with
	the mesothorax. Ants, etc HYMEN OPTERA (Page 471)
42.	Head not prolonged into a beak43
	Head produced into a mandibulate beak.
	MECÓPTERA (Page 189)
43.	Very small (three millimeters), louse-like jumping species; pro-
	thorax inconspicuous. Book lice. CORRODENTIA (Page 96)
	Larger, or at least not louse-like species; prothorax large
44.	
	larvæ when present in inverse position, the metathoracic over-
	lapping the mesothoracicORTHÓPTERA (Page 48)
	Hind legs not enlarged for jumping; wing-pads, if present, in
٠.	normal position
45.	Prothorax much longer than the mesothorax; front legs fitted for
	grasping prey MANTÒDEA (Page 83)
	Prothorax not greatly lengthened46
46.	Cerci present; antennæ usually with more than fifteen joints, often
	many-jointed
	No cerci; body often hard-shelled; antennæ usually with eleven
	joints
47.	Cerci with more than three joints
	Cerci short, with one to three joints

48.	Body flattened and oval; head inflexed; prothorax oval. Roaches.
	BLATTÀRIÆ (Page 77)
	Body elongate; head nearly horizontal49
49.	Cerci long; ovipositor chitinized, exserted; tarsi five-jointed.
	GRYLLOBLATTÒDEA (Page 47)
	Cerci short; no ovipositor; tarsi four-jointed.
	ISÓPTERA (Page 93)
50	Tarsi five-jointed; body very slender and long. Walking-sticks.
00.	PHASMATÒDEA (Page 61)
	Tarsi two- to four-jointed; body not linear
51.	Front tarsi not enlarged
	Front tarsi with the first joint swollen EMBIODEA (Page 91)
52.	Tarsi apparently four-jointed; cerci with several joints; antennæ
	with nine to thirty joints ISÓPTERA (Page 93)
	Tarsi two-jointed; cerci one-jointed; antennæ nine-jointed; minute
	species ZORÁPTERA (Page 100)
53.	Body cylindrical, caterpillar-like MECÓPTERA (Page 189)
	Body more or less depressed, not caterpillar-like54
54.	Mandibles united with the corresponding maxillæ to form sucking
	jaws (Fig. 975)Larvæ of NEURÓPTERA
	Mandibles almost always separate from the maxillæ (Figs. 972,
	973, 976, 977, 979, 980, 983, 984, 986, 992).
	Larvæ of COLEÓPTERA, RAPHIDIÒDEA, STREPSÍPTERA
55.	
	nal segments; but not on the second; the false legs (prolegs)
	tipped with many minute hooks. (Figs. 978, 982).
	Larvæ of LEPIDÓPTERA (Page 240)
	False legs numbering from six to ten pairs, one pair of which
	occurs on the second abdominal segment; the prolegs not tipped
	with minute hooks
56.	Head with a single ocellus on each side. (Figs. 966–971).
	Larvæ of some <b>HYMENÓPTERA</b> (Page 408)
	Head with several ocelli on each side. Larvæ of MECÓPTERA
57.	Body bare or with few scattered hairs, or with waxy coating58
	Body densely clothed with hairs or scales; proboscis if present
	coiled under the head. Moths. <b>LEPIDÓPTERA</b> (Page 198)
58.	Last tarsal joint swollen; mouth consisting of a triangular un-
,	jointed beak; minute species. Thrips.
	THYSANÓPTERA (Page 72)
	Tarsi not bladder-like at the tip, and with distinct claws59
59.	Prothorax distinct

	Prothorax small, hidden when viewed from above.
<b></b>	DÍPTERA (Page 264)
60.	
	HEMÍPTERA (Page 140)
	Beak arising from the back part of the head.
01	HOMÓPTERA (Page 106) Mouth mandibulate
61.	Mouth haustellate, forming a strong pointed inflexed beak.
	Nymphs of <b>HEMÍPTERA</b>
62.	
04.	
	responding maxillæ to form piercing jaws.  Larvæ of some <b>NEURÓPTERA</b>
an.	Mandibles normal, moving laterally to function as biting jaws63
63.	Body not encased in a shell made of sand, pebbles, leaves, etc 64
	Case-bearing forms; tracheal gills usually present. Periwinkles,
	Caddis-worms. (Fig. 974)Larvæ of TRICHÓPTERA
64.	Abdomen furnished with external lateral gills or respiratory pro-
	cesses (a few Coleoptera and Trichoptera here also)
	Abdomen without external gills
65.	Abdomen terminated by two or three long caudal filaments.
	Larvæ of PLECTÓPTERA
	Abdomen with short end processes. Larvæ of MEGALÓPTERA
66.	Lower lip strong, extensile, and furnished with a pair of opposable
	hooksLarvæ of ODONATA
	Lower lip not capable of being thrust forward and not
<u>-</u>	hooked
67.	Abdomen without false legs
	Abdomen bearing paired false legs on several segments.
00	A few larvæ of LEPIDÓPTERA
68.	The three divisions of the thorax loosely united; antennæ and caudal filaments long and slenderLarvæ of <b>PLECÓPTERA</b>
	Thoracic divisions not constricted; antennæ and caudal filaments
	short (also a few Trichoptera here). (See couplet 54).  Larvæ of COLEÓPTERA
3U .	
39.	Body flattened
	jumping species. FleasSIPHONÁPTERA (Page 404)
70	Mandibulate mouthparts formed for chewing71
70.	Haustellate mouthparts formed for piercing and sucking73
71.	Mouth inferior; cerci long
11.	Mouth interior; cerci long
	Mouth afterior; no cerci; generally elongate-oval insects with

	somewhat triangular head; parasites of birds or mammals. Biting lice
72	Cerci straight, eyes absent; antennæ short; external parasites of
	rodentsDIPLOGLOSSATA (Page 71)
	Cerci strongly bent or angulate at the middle; eyes present; an-
	tennæ nearly as long as the body; external parasites of bats.
	Some <b>DERMÁPTERA</b> (Page 65)
73.	Antennæ exserted, visible, though rather short74
	Antennæ inserted in pits, not visible from above.
	Pupiparous <b>DÍPTERA</b> (Page 264)
74.	Beak unjointed; tarsi formed as a hook for grasping the hairs of
	the host; permanent parasites. Lice. ANOPLURA (Page 104)
	Beak jointed; tarsi not hooked; temporary parasites.
75	HEMÍPTERA (Page 140) Legless grubs, maggots or borers; locomotion effected by a squirm-
10.	ing motion. Larvæ of Strepsiptera, and of some Coleoptera
	(see couplet 54), Diptera, Lepidoptera and Hymenoptera.
	(If living in the body of wasps or bees, with the flattened head
	exposed, compare the females of Strepsiptera).
	Sedentary forms, incapable of locomotion
<b>7</b> 6.	Small degraded forms bearing little superficial resemblance to
	insects, with a long slender beak, and usually covered with a
	waxy scale or powder or cottony tufts; living on various plants.
	Scale insects
	Body quiescent, but able to bend from side to side; not capable
	of feeding, enclosed in a skin which is tightly drawn over all the members, or which leaves the limbs free but folded against the
	body; sometimes free; sometimes enclosed in a cocoon or in a
	shell formed from the dried larval skin
77.	The skin encasing the legs, wings, etc., holding the members
	tightly against the body; prothorax small; a proboscis show-
	ing
	Legs, wings, etc., more or less free from the body; biting mouth-
	parts showing79
<i>7</i> 8.	Proboscis long; four wing cases; sometimes in a cocoon. (Figs.
	990, 996)
	Proboscis short; two wing cases. (Figs. 991, 997).
70	Pupæ of <b>DÍPTERA</b> (Page 368)
19.	Prothorax small, fused into one piece with the mesothorax; sometimes enclosed in a loose cocoon. Pupe of HYMENÓPTERA
	Prothorax larger and not closely fused with the mesothorax80
	and and another than the medical and the medic

80. Wing cases with few or no veins. (Fig. 994).

Pupæ of **COLEÓPTERA** 

Wing cases with a number of veins. (Fig. 993).

Pupæ of NEUROPTEROID ORDERS

## LITERATURE ON INSECTS

General Works on Insects and References Dealing with at least Several Orders Together

- Balfour-Browne, F. Keys to the Orders of Insects. Cambridge Univ. Press, 58 pp. (1920).
- Banks, N. Synopsis, Catalogue and Bibliography of Neuropteroid Insects of North America. Trans. American Entom. Soc., 19, pp. 328–373 (1892).
   (Includes Neuroptera, Odonata, Plectoptera, Plecoptera, Trichoptera, Mecoptera, Isoptera, Embiodea, Megaloptera).

A List of Works on North American Entomology. Bull. Bur. Entom., U. S. Dept. Agric., No. 81, 120 pp. (1910). (A most valuable list of selected works, classified by groups, including also the Arachnida).

Berlese, A. Gli Insetti. 2 vols., 2018 pp., Milan (1909–25).

- BIOLOGICAL ABSTRACTS. Published by the Union of American Biological Societies, University of Pennsylvania, Philadelphia, 1–4 (1926–30). An index to biological literature beginning with 1926, with an abstract of those papers to which reference is made.
- Britton, W. E. Check List of the Insects of Connecticut. Bull. State Geol. Nat. Hist. Surv., Connecticut, No. 31, 397 pp. (1920).
- Brohmer, P., Ehrmann, P. and Ulmer, G., Editors. Die Tierwelt Mitteleuropas, 1928; vol. 2 contains the lower Arthropoda; vol. 3, Arachnida; vol. 4, Insects. (The parts are by numerous contributors, and so far as published are listed under the separate groups).
- Brues, C. T. and Melander, A. L. Key to the Families of North American Insects. 140 pp., Boston, Mass. and Pullman, Wash. (1915).
- Carus, J. V. Bibliographia Zoologica. Vols. 1–24 issued with the Zoologischer Anzeiger, Leipzig (1900–13); vols. 25–40 issued separately by the Concilium Bibliographicum, Zurich (1913–30).
- Cockerell, T. D. A. Zoölogy of Colorado. 262 pp., Boulder, Colo., (1927). Concilium Bibliographicum. Zurich, Switzerland. Index cards to zoölogical literature, 1896—. (Deals with the whole animal kingdom; the cards issued, one card for each title, usually with brief indication of contents.)
- Comstock, J. H. The Wings of Insects. 423 pp., Comstock Pub. Co., Ithaca, N. Y. (1918).

An Introduction to Entomology. Comstock Pub. Co., Ithaca, N. Y., 1044 pp. (1924).

- Essig, E. O. Insects of Western North America. 1035 pp. New York, Mac-Millan (1926).
- Ewing, H. E. Manual of External Parasites. 225 pp. Springfield, Ill., C. C. Thomas (1929).

- Felt, E. P. Key to American Insect Galls. Bull. New York State Mus., No. 200, 310 pp. (1917). (Contains an extensive bibliography.)
- FERRIS, G. F. The Principles of Systematic Entomology. Stanford Univ. Publ. Biol. Sci., 5, pp. 103-269 (1928).
- FLETCHER, T. B. South Indian Insects. 587 pp. Madras (1914).

Tentative Keys to the Orders and Families of Indian Insects. Bull. Agric. Res. Inst. Pusa, No. 162, 101 pp. (1926).

Froggatt, W. W. Australian Insects. 463 pp. Sydney (1908).

- Godman, F. D. and Salvin, O., Editors. Biologia Centrali-Americana, Contributions to a Knowledge of the Fauna and Flora of Mexico and Central America. (Contains Arachnida, Diplopoda, Chilopoda and Insects in many volumes, some of which we have listed separately). London (1896–1915).
- Hagen, H. A. Bibliotheca Entomologica. 2 vols. Leipzig (1862–63). (Literature up to the year 1863).
- HANDLIRSCH, A. Die fossilen Insekten und die Phylogenie der rezenten Formen. Leipzig (1906–08).

Geschichte, Litteratur, Technik, Paläontologie, Phylogenie und Systematik der Insekten. Schröder's Handbuch der Entomologie, **3**, 1201 pp. Jena (1925).

Insecta. Kükenthal, Handbuch der Zoologie, 4, Berlin (1929).

- Horn, W. and Schenkling, S. Index Litteraturæ Entomologicæ. (Before 1863). Berlin-Dahlem, 4 vols. (1928–29).
- HOULEBERT, C. Bibliothèque de Zoologie. In Encyclopédie Scientifique, Paris, Doin. (Insects form part 5 and numerous volumes.)
- IMMS, A. D. A General Textbook of Entomology. E. P. Dutton, New York and Methuen and Co., London (1926).
- Leonard, M. D. List of the Insects and Spiders of New York State. Mem. Cornell Agric. Expt. Sta., No. 101, 1121 pp. (1926).
- LUTZ, F. E. Field Book of Insects. (Northeastern North America.) 509 pp. New York (1918).
- MAXWELL-LEFROY, H. and HOWLETT, F. M. Indian Insect Life. 798 pp. Calcutta (1909).
- RILEY, C. V. Enumeration of the Published Synopses, Catalogues and Lists of North American Insects (1888).
- ROUSSEAU, E. Les larves et nymphes aquatiques des Insectes d'Europe. 1, 967 pp. (1921).
- Schulze, F. E., Editor. Das Tierreich. Berlin (1897- ). (The parts are listed separately.)
- SHARP, D. Insects. Cambridge Natural History, vols. 5 and 6, MacMillan & Co., New York and London (1895–99).
- Sharp, D., Editor. Fauna Hawaiiensis. 3 vols. Honolulu (1899–1913).
- SMITH, J. B. Catalogue of the Insects of New Jersey. Ann. Rept. New Jersey State Mus., 1909, 888 pp. (1910).
- Taschenberg, O. Bibliotheca Zoologica. Vol. 1 (1863); vols. 2 and 3 (1889–90). (Literature of Entomology to the year 1880).

- Thompson, M. T. Illustrated Catalogue of American Insect Galls. 116 pp. Nassau, N. Y. (1916).
- TILLYARD, R. J. The Insects of Australia and New Zealand. 560 pp. Angus and Robertson, Sydney (1926).
- Tullgren, A. and Wahlgren, E. Svenska Insekter. Stockholm, 812 pp. (1920–1922).
- Weiss, H. B. Some Old Classifications of Insects. Canadian Entom., 47, pp. 369–376 (1915).
- WYTSMAN, P., Editor. Genera Insectorum. Brussels (1902—). (The many fascicles of this work are listed separately under the families and subfamilies; each contains a synoptic key to genera and a catalogue of species of the family or subfamily treated.)
- Zoölogical Record. Zoölogical Society of London (1865—). (Complete review of all literature in Zoölogy, year by year. The most complete and useful bibliographic index extant.)

# ORDER PROTÙRA

# (MIRIENTÓMATA)

Minute, slender, delicate, wingless, terrestrial, colorless or yellowish, blind species. Body bare; head pear-shaped, eyes and ocelli both absent; antennæ absent or reduced to a tubercle; abdomen comprising eleven segments, the last three very short, the basal three segments furnished with styles, no cerci; mouthparts formed for sucking, but retracted within the head, the mandibles long and styliform; legs short, tarsi one-jointed, with a single claw. Younger stages with only nine abdominal segments.

Spiracles absent; no tracheal system; second and third pairs of abdominal appendages one-jointed. (Aceréntomon (Fig. 4), Aceréntulus (Fig. 12), widespr.)......ACERENTÓMIDÆ

### LITERATURE ON PROTURA

Berlese, A. Monografia dei Myrientomata. Redia, 6 (1909).

EWING, H. E. New Genera and Species of Protura. Proc. Biol. Soc. Washington, 23, pp. 193–202 (1921).

Silvestri, F. Descrizione di un nuovo genere di insetti apterigoti rappresentanti di un nuovo ordine. Boll. Lab. Zool. Gen. Agrar. Portici., 1, (1907).

STACH, J. Eosentomon armatum. (With list of known Protura.) Spraw. Kom. fizyogr., Krakow., **61**, pp. 205–215 (1927).

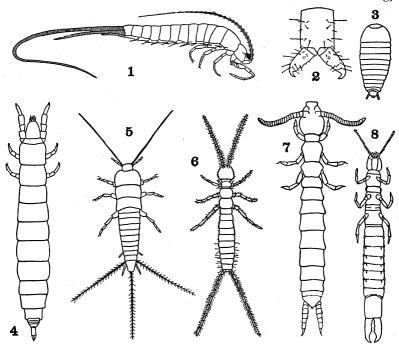
Womersley, H. Notes on the British Species of Protura. Entom. Monthly Mag., 63, pp. 140-148 (1927).

Further Notes on the British Species of Protura. Entom. Monthly Mag., **64**, pp. 113–115 (1928).

# ORDER THYSANÙRA

(ECTÓTROPHI, ECTÓGNATHA)

Moderate-sized or small, wingless, terrestrial insects of active habits. Body tapering behind, generally clothed with scales. Antennæ long,



Figs. 1-8. Protura, Thysanura, Entotrophi

- 1. Nesomachilis (Tillyard) Machilidæ.
- 2. Parajapyx, apex of abdomen (Silvestri) Japygidæ.
- 3. Gastrotheus (Silvestri) Lepismatidæ.
- 4. Acerentomon (Silvestri) Acerentomidæ.
- 5. Lepisma (Butler) Lepismatidæ.
- 6. Campodea (Maxwell-Lefroy) Campodeidæ.
- 7. Anajapyx (Silvestri) Projapygidæ.
- 8. Japyx, from below (Berlese) Japygidæ.

slender, tapering, many-jointed. Head closely articulated with the thorax; compound eyes present, large, reduced, or absent. Mouth-parts external, their tips projecting from a mouth cavity; mandibles large, toothed; maxillæ long. Thorax composed of three similar, freely articulated segments. Legs similar, the coxæ large; tarsi two- or three-jointed, with two or three claws. Abdomen composed of eleven segments, bearing at tip a pair of long, filiform, many-jointed cerci and a similar long median cerciform appendage; some ventral segments, usually the second to seventh, each with a pair of styles and generally also with a median sac; eighth and ninth ventral segments of female with paired, sometimes jointed appendages. No metamorphosis. Bristle tails. (Figs. 1, 3, 5, 9 and 10).

Eyes large, composed of very many minute facets, extending over the front and usually touching above on the vertex; maxillary palpi very long, seven-jointed; eleventh tergite not covered by tenth; four posterior coxæ with a style or hooked process. Body strongly convex above. A cosmopolitan family more abundant in the tropics. (Máchilis, Petròbius, Machilòides, Machilìnus; Dílta). (Superfamily MACHILÒIDEA).

## MACHÍLIDÆ

Eyes small or wanting, when present composed of a few large facets, set on the sides of the head and not extending over the front; maxillary palpi five- or six-jointed; eleventh tergite partly covered by the tenth; coxæ without style or process. Body more or less flattened. A widespread group. (Lepísma, Thermòbia, Maindrònia, Atelùra, Nicolètia). (Superfamily LEPISMATÒIDEA) . . . . . . . . . . . LEPISMÁTIDÆ

### LITERATURE ON THYSANURA AND ENTOTROPHI

(These are combined since many citations refer to both groups)

- Bär, H. Beitrag zur Kenntnis der Thysanuren. Jenaische Zeitschr., 48, pp. 1–92 (1912).
- Bartholin, T. Catalogue of Danish Apterygota (Danish). Nath. Medd., 67, pp. 155-209 (1916).
- Dalla-Torre, K. W. Die Gattungen und Arten der Apterygogenea. Innsbruck, 23 pp. (1895).
- ESCHERICH, K. Das System der Lepismatiden. Zoologica, Heft 43 (1905). HANDSCHINN, E. Urinsekten oder Apterygota. In Tierwelt Deutschlands, Lief. 16 (1929).
- JACKSON, C. F. Key to the families and genera of the Thysanura. Ohio Naturalist, 6, pp. 545–549 (1906).

- Lubbock, J. Monograph of the Collembola and Thysanura. Roy. Soc. London (1873).
- MacGillivray, A. D. Catalogue of North American Thysanura. Canadian Entom., 23, pp. 267–276 (1891).
- Schött, H. North American Apterygogenea. Proc. California Acad. Sci., 6, pp. 169–196 (1896).
- SILVESTRI, F. Nuova contribuzione alla consocenza dell' Anajapyx. Ann. Lab. Zool. Gen. Agrar., Portici, 6, 15 pp. (1905).

Contribuzione alla conoscenza dei Campodeidæ. Boll. Lab. Zool. Gen. Agrar., Portici, **6**, pp. 110–147 (1912).

Nuovi generi e specie di Campodeidæ. Boll. Lab. Zool. Gen. Agrar., Portici. 6. pp. 5-25 (1912).

Thysanura. Fauna süd-west Australiens, 2, pt. 4, pp. 47–68 (1908).

SWENK, M. H. A Synopsis of the North American Species of Japyx. Journ. New York Entom. Soc., 11, pp. 129–132 (1903).

Verhoeff, K. W. Zur vergleichende Morphologie und Systematik der Japygiden. Arch. f. Naturg., Jahrg. 71, Heft 1, pp. 63-114 (1904).

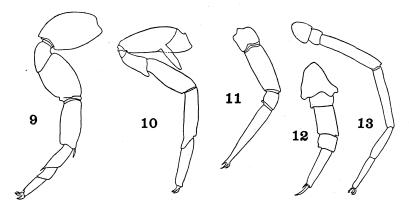
WAHLGREN, E. Svenska Insektfauna, I. Apterygota. Entom. Tidskr., 27, pp. 233–270 (1906).

## ORDER ENTÓTROPHI

# (ENTÓGNATHA, CAMPODEÒIDEA, DIPLÙRA)

Small, slender, wingless insects, incapable of springing and of sluggish habits. Body almost always without scaly covering. Head large, freely articulated with the thorax; mouthparts concealed in a cavity within the head. Antennæ long, many-jointed. Eyes absent. Mandibles toothed; maxillæ and labium reduced, their palpi vestigial. Segments of thorax similar, freely articulated. Legs similar; coxæ small; tarsi one-jointed, with two claws. Abdomen consisting of eleven segments, the last segment either more or less covered by the preceding or fused with it; no ventral tube, but some of the ventral segments, usually the second to seventh, provided with ventral styles; apex of abdomen without median process; cerci long and many-jointed, or short and indistinctly jointed; appendages of eighth and ninth segments greatly reduced. No metamorphosis. (Figs. 2, 6, 7, 8 and 13).

2. Cerci long, slender, many-jointed, of approximately the same



Figs. 9-13. Protura, Thysanura, Entotrophi

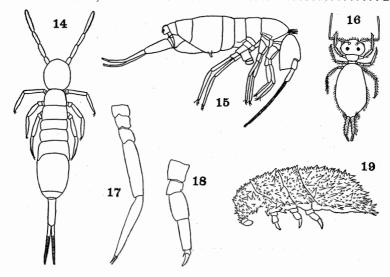
- 9. Thermobia, middle leg. Lepismatidæ.
- 10. Machilis, hind leg. Machilidæ.
- 11. Hanseniella, front leg. Scutigerellidæ.
- 12. Acerentulus, middle leg. Acerentomidæ.
- 13. Campodea, middle leg. Campodeidæ.

# ORDER COLLÉMBOLA

Small or minute, soft-bodied, wingless, springing insects, the body sometimes clothed with scales or hairs. Head freely movable. Antennæ four- to six-jointed. Eyes of a degenerate compound type, composed of a few separated facets; sometimes entirely absent. Mouthparts retracted within the head; mandibles well developed; palpi vestigial. Thorax with the segments fused into a single mass. Legs similar, slender; tarsi one-jointed, with a single claw and a smaller claw-like empodium. Abdomen consisting of six segments; without cerci, and never terminated by caudal filaments or pincers-like appendages; usually with a leaping appendage which is often forked at apex and

arises from the fourth or fifth segment. Ventral tube always present as a simple or bifid process developed under the base of the abdomen. No metamorphosis. Spring tails. (Figs. 14–19).

 Abdomen consisting of six evident segments or at least with only two of these indistinctly separated; body lengthened, subcylindrical, frequently clothed with scales. (Suborder ARTHRO-PLÈONA)



Figs. 14-19. Collembola

- 14. Isotoma (Imms) Entomobryidæ.
- 15. Tomocerus (Folsom) Entomobryidæ.
- 16. Sminthurus (Sharp) Sminthuridæ.
- 17. Tomocerus, hind leg. (Ewing) Entomobryidæ.
- 18. Onychiura, hind leg. (Ewing) Onychiuridæ.
- 19. Anoura (Tillyard) Poduridæ.

- - Pronotum concealed from above; body often with scales; furca when present usually arising from the fifth segment; antennæ with four to six joints. (Entomòbrya, Orchesélla, Sinélla, Tomócerus, Isótoma, Sìra)......ENTOMOBRŸIDÆ
- 4. Front with false ocelli present; eyes absent; furcula usually absent; sluggish species with tuberculate integument. (Onychiùra (= Aphorùra) holarc.; Tetrodontóphora, palæarc.). (APHO-RÙRIDÆ)......ONYCHIÙRIDÆ
- 5. Coxe long; last joint of antennæ short, not divided into false ringjoints; antennæ not longer than the head, inserted at the middle of the head or below; thorax longer than the abdomen. (Neèlus, Megalothòrax, holarc.). (MEGALOTHORÁCIDÆ).

### **NEÉLIDÆ**

Coxæ of the usual length; last joint of antennæ long, usually divided into false ring-joints; antennæ much longer than the head; inserted above the middle of the head. (Sminthurus, Sminthurinus, Dicýrtoma (= Papírius), Corynephòria, Bourletiélla). (Including CORYNEPHORIDÆ = PAPI-RÌIDÆ).....SMINTHÜRIDÆ

### LITERATURE ON COLLEMBOLA

BÖRNER, C. Neelidæ. Gen. Insectorum, fasc. 45, 5 pp. (1906).

Das System der Collembolen. Mitt. naturh. Mus. Hamburg, 23, pp. 147–188 (1906).

Die Familien der Collembolen. Zool. Anz., 16, pp. 315–322 (1913).

CARPENTER, G. H. and Evans, W. Collembola and Thysanura of Scotland. Proc. Roy. Phys. Soc. Edinburgh, 14, pp. 221–266 (1899).

Folsom, J. W. Tomocerinæ of North America. Proc. U. S. Nat. Mus., **46**, pp. 451–472 (1914).

North American Collembolous Insects of the Subfamilies Achoreutinæ, Neanurinæ and Podurinæ. Proc. U. S. Nat. Mus., **50**, pp. 477–525 (1916). Subfamily Onychiurinæ. Proc. U. S. Nat. Mus., **53**, pp. 637–659 (1917).

GUTHRIE, J. E. The Collembola of Minnesota. Geol. Nat. Hist. Surv. Minnesota, ser. 4, 110 pp. (1903).

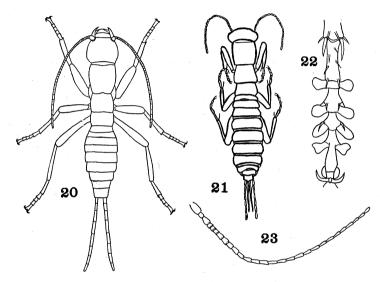
LINNANIEMI, W. M. Die Apterygoten-fauna Finlands. Acta Sci. Faun. Flor. Fennica, **40**, pp. 1–361 (1907–12).

Lubbock, J. Monograph of the Collembola and Thysanura. Roy. Soc. London (1873).

# ORDER GRYLLOBLATTÒDEA

# (GRYLLOBLATTÒIDEA, NOTÓPTERA)

Elongate, more or less depressed, wingless insects, measuring about an inch in length. Head rather large, free, horizontal or slightly inclined; eyes small or absent, no ocelli; antennæ long, filiform, with about 30 to 40 joints, inserted at the sides of the front of the head near



Figs. 20-23. Grylloblattodea

- 20. Ishiana (Silvestri) Grylloblattidæ.
- 21. Grylloblatta (Walker) Grylloblattidæ.
- 22. Grylloblatta, tarsus of male (Walker) Grylloblattidæ.
- 23. Galloisiana, antenna (Crampton) Grylloblattidæ.

the mandibles; mandibles large and strong. Prothorax quadrate or somewhat longer than wide, not expanded laterally; free and larger than the meso- or metathorax. Legs similar, formed for running, the coxæ close together; tarsi five-jointed, with two claws, those of the adult male with a pair of membranous lobes beneath each joint. Abdomen elongated, tergites more or less equal, transverse, extending down at the sides of the abdomen; cerci long, eight- or nine-jointed; ovipositor exserted, sword-shaped, composed of six paired pieces.

Metamorphosis very slight, the nymphs very much like the adults and apparently of slow growth; terrestrial in all stages. (Figs. 20–23).

### LITTERATURE ON GRYLLOBLATTODEA

CAUDEL, A. N. Grylloblatta in California. Canadian Entom., 53, pp. 148–150 (1923).

Notes on Grylloblatta. Journ. Washington Acad. Sci., 14, pp. 369–371 (1924).

CAUDELL, A. N. and King, J. L. A New Genus of Grylloblattidæ from Japan. Proc. Entom. Soc. Washington, **26**, pp. 53–60 (1924).

Silvestri, F. Grylloblattidæ. Boll. Zool. Gen. Agrar. Portici, **20**, pp. 107–121 (1928).

WALKER, E. M. A New Species of Orthoptera Forming a New Genus and Family. Canadian Entom., 46, pp. 93–97 (1914).

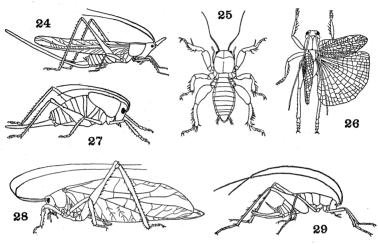
The Male and Immature Stages of Grylloblatta. Canadian Entom., **51**, pp. 131–139 (1919).

## ORDER ORTHÓPTERA

# (ULONÀTA part; SALTATÒRIA)

Small to large, nearly always jumping species, often possessing a device on the wings for making a creaking or chirping sound; hind femora almost always very much stouter basally, or longer, or both, than the middle femora; wings of adults reposing over the abdomen, the fore wings toughened, narrower and thicker than the membranous, plaited hind pair; sometimes the wings vestigial or completely absent; head usually vertical, sometimes conically produced forward; ovipositor almost always free, often long, sword- or needle-shaped; mouthparts conspicuous, mandibulate; antennæ long and many-jointed or short with few joints; tarsi usually four- or three-jointed; prothorax large, free, often much enlarged; cerci short. Metamorphosis gradual, the young resembling the adults, but with the small wings in a reversed position in the last two nymphal stages, the hind wings then overlapping the fore wings.

Antennæ shorter, with less than 30 joints, filiform, rarely clubbed or serrate, but not delicately tapering; auditory organ if present, near the base of the abdomen; ovipositor of female never elongated; tarsi three-jointed, rarely with the front and middle ones two-jointed. (Suborder ACRIDODEA)............9



Figs. 24-29. Orthoptera

- 24. Conocephalus (Blatchley) Tettigoniidæ.
- 25. Stenopelmatus (Saussure) Stenopelmatidæ.
- 26. **Gryllacris** (Karny) Gryllacridæ.
- 27. Anabrus (Caudell) Tettigoniidæ.
- 28. Stilpnochlora (Saussure) Tettigoniidæ.
- 29. Ceuthophilus (Blatchley) Stenopelmatidæ.

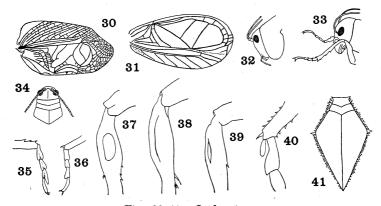
This family comprises five subfamilies, separable as follows:

a. Front tibiæ with an auditory organ....b

Front tibiæ without an auditory organ....c

rated; wings large, with a chirping organ. (Prophala (= Tárraga), ind.)	parated; nacrida, ATINÆ nabove; philus, us, pal- LINÆ). ORINÆ nabelow;d (Mim- &MINÆ s, Am.). ATINÆ
Cerci long; first and second tarsal joints distinctly se usually wingless. (Anostóstoma, ethiop., Austr.; Dem Austr.; Magréttia, ethiop., As.) ANOSTOSTOM.  c. Hind femora at base more sharply produced below that legs rather long and slender. Cave crickets. (Ceuthó Am. (Fig. 29); Dolichópoda, palæarc.; Troglóphil æarc.; Rhaphidóphora, indoaustr.). (CEUTHOPHIL RHAPHIDOPHO Hind femora at base more sharply produced above that legs stouter	parated; nácrida, ATÌNÆ n above; philus, us, pal- LÎNÆ). ORÌNÆ n below;d (Mim- &MÌNÆ s, Am.).
usually wingless. (Anostóstoma, ethiop., Austr.; Dem Austr.; Magréttia, ethiop., As.) ANOSTOSTOM.  c. Hind femora at base more sharply produced below that legs rather long and slender. Cave crickets. (Ceuthó Am. (Fig. 29); Dolichópoda, palæarc.; Troglóphil æarc.; Rhaphidóphora, indoaustr.). (CEUTHOPHIL RHAPHIDOPHO Hind femora at base more sharply produced above that legs stouter	nácrida, ATÌNÆ n above; philus, us, pal- LÌNÆ). ORÌNÆ n below;d (Mim- &MÌNÆ s, Am.).
Austr.; Magréttia, ethiop., As.) ANOSTOSTOM.  c. Hind femora at base more sharply produced below that legs rather long and slender. Cave crickets. (Ceuthó Am. (Fig. 29); Dolichópoda, palæarc.; Troglóphil æarc.; Rhaphidóphora, indoaustr.). (CEUTHOPHIL RHAPHIDOPHO Hind femora at base more sharply produced above that legs stouter	ATÎNÆ a above; philus, us, pal- LÎNÆ). ORÎNÆ a below;d (Mim- &MÎNÆ s, Am.). ATÎNÆ
c. Hind femora at base more sharply produced below that legs rather long and slender. Cave crickets. (Ceuthó Am. (Fig. 29); Dolichópoda, palæarc.; Troglóphil æarc.; Rhaphidóphora, indoaustr.). (CEUTHOPHI RHAPHIDOPHO Hind femora at base more sharply produced above that legs stouter	a above; philus, us, pal- LINÆ). ORINÆ a below;d (Mim- EMINÆ s, Am.).
legs rather long and slender. Cave crickets. (Ceutho Am. (Fig. 29); Dolichópoda, palæarc.; Troglóphil æarc.; Rhaphidóphora, indoaustr.). (CEUTHOPHI RHAPHIDOPHO Hind femora at base more sharply produced above that legs stouter	philus, us, pal- LINÆ). ORÌNÆ a below; d (Mim- EMÌNÆ s, Am.).
Am. (Fig. 29); Dolichópoda, palæarc.; Troglóphil æarc.; Rhaphidóphora, indoaustr.). (CEUTHOPHI.  RHAPHIDOPHO  Hind femora at base more sharply produced above that legs stouter	us, pal- LINÆ). ORÌNÆ n below; d (Mim- AMÌNÆ s, Am.).
earc.; Rhaphidóphora, indoaustr.). (CEUTHOPHIL RHAPHIDOPHO Hind femora at base more sharply produced above that legs stouter	LÎNÆ). ORÎNÆ n below; d (Mim- RMÎNÆ s, Am.). ATÎNÆ
Hind femora at base more sharply produced above that legs stouter	ORÎNÆ n below;d (Mim- &MÎNÆ s, Am.). ATÎNÆ
Hind femora at base more sharply produced above that legs stouter	n below; d ( <b>Mim-</b> <b>&amp;MÌNÆ</b> s, Am.). <b>ATÌNÆ</b>
legs stouter	d (Mim- RMÌNÆ s, Am.). ATÌNÆ
<ul> <li>d. Front coxe armed in front with a tooth-like projection. nérmus, ethiop.; Cratomèlus, neotrop.).</li> <li>MIMNEE Front coxe simple, unarmed. (Fig. 25.). (Stenopelmàtus STENOPELM.</li> <li>4. Front tibiæ without an auditory organ</li></ul>	(Mim- MÌNÆ s, Am.). ATÌNÆ
nérmus, ethiop.; Cratomèlus, neotrop.).  MIMNEF Front coxæ simple, unarmed. (Fig. 25.). (Stenopelmàtus STENOPELM.  4. Front tibiæ without an auditory organ Front tibiæ with an auditory organ. Long-horned locusts dids. (PHASGONÙRIDÆ, LOCÜSTIDÆ).	AMÌNÆ s, Am.). ATÌNÆ
Front coxe simple, unarmed. (Fig. 25.). (Stenopelmatus STENOPELM.  4. Front tibiæ without an auditory organ. Long-horned locusts dids. (PHASGONÙRIDÆ, LOCÙSTIDÆ).	s, Am.). <b>ATÌNÆ</b>
Front coxe simple, unarmed. (Fig. 25.). (Stenopelmatus STENOPELM.  4. Front tibiæ without an auditory organ. Long-horned locusts dids. (PHASGONÙRIDÆ, LOCÙSTIDÆ).	s, Am.). <b>ATÌNÆ</b>
4. Front tibiæ without an auditory organ	ATÌNÆ
<ol> <li>Front tibiæ without an auditory organ.</li> <li>Front tibiæ with an auditory organ. Long-horned locusts dids. (PHASGONÙRIDÆ, LOCÙSTIDÆ).</li> </ol>	
Front tibiæ with an auditory organ. Long-horned locusts dids. (PHASGONÙRIDÆ, LOCÙSTIDÆ).	
dids. $(PHASGON \dot{U}RID \cancel{x}, LOC \dot{U}STID \cancel{x}).$	
	,
1211100	NÌIDÆ
This very extensive family includes a number of subfamilie	es which
may be separated as follows:	
a. Antennæ inserted between the eyes, nearer to the top of t	he occi-
put than to the clypeal suture. (Fig. 32)	b
Antennæ inserted below the eyes, or between their lower r	nargins,
nearer to the clypeal suture than to the top of the	occiput.
(Fig. 33)	0
b. First and second joints of tarsi longitudinally sulcate la	aterally.
(Fig. 35)	c
First and second tarsal joints smooth (Fig. 36); hind tibiæ	
apical spine on each side above. A large cosmopolitar	ı group,
including many leaf-like species. (Phaneróptera, w	ndesnr •
T	idespi.,
Isopsèra, ind.; Scuddèria, Am.; Tylópsis, palæarc.,	ethiop.;
Isóphya, palearc., Am.) PHANEROPT	ethiop.; ERÌNÆ
Isóphya, palæarc., Am.) PHANEROPT: c. Auditory pits on front tibiæ open. (Fig. 37)	ethiop.; ERÌNÆ d
Isóphya, palæarc., Am.)	ethiop.; ERINÆ d d exten-
Isóphya, palæarc., Am.)	ethiop.; ERINÆ d d exten- narrows
Isóphya, palæarc., Am.)	ethiop.; ERÎNÆ d d exten- narrows g
<ul> <li>Isóphya, palæarc., Am.)</li></ul>	ethiop.; ERÎNÆd d exten- narrowsge
Isóphya, palæarc., Am.)	ethiop.; ERÎNÆd d exten- narrowsgge ed, very

## **MECONEMINÆ**



Figs. 30-41. Orthoptera

- 30. **Gryllus**, fore wing (Handlirsch) Gryllidæ.
- 31. **Ecanthus,** fore wing (Handlirsch) Gryllidæ.
- 32, 33. Superior and inferior insertion of antennæ (Caudell) Tettigoniidæ.
- 34. Mecopoda, dorsal view of head and pronotum (Caudell) Tettigoniidæ.
- 35. Tarsus with sulcate segments (Caudell) Tettigoniidæ.
- 36. Tarsus with smooth segments (Caudell) Tettigoniidæ.
- 37. Open auditory pit (Caudell) Tettigoniidæ.
- 38. Linear auditory pit (Caudell) Tettigoniidæ.
- 39. Ear-like or shell-shaped auditory pit (Caudell) Tettigoniidæ.
- 40. Tarsus with free plantula (Caudell) Tettigoniidæ.
- 41. Phyllophora, hooded form of pronotum (Caudell) Tettigoniidæ.
  - f. Pronotum hood-like, strongly and acuminately produced behind; lateral carinæ of pronotum dentate or crenulate. (Fig. 41). (Phyllóphora, indoaustr.; Hyperhómala, austromal.).

### PHYLLOPHORÎNÆ

Pronotum not hood-like, nor much produced behind. (Fig. 34). (Mecópoda, austromal.; Acridóxena, Apteroscírtus, ethiop.; Tabària, Rhammatópoda, neotrop.).....MECOPODĨNÆ

- g. Front tibiæ without terminal spines above.....h
  Front tibiæ with a terminal spine above on the outer side (except
  Arytropteris).....n
- h. Antennal scrobes (i.e. the grooves in which the antennæ lie) with the margins produced. (Pseudophýllus, palæarc., indomal.; Cleándrus, Phyllomimus, indomal.; Zabàlius, ethiop.).

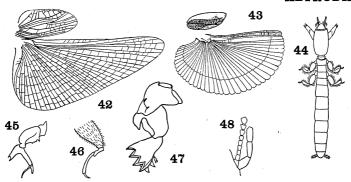
PSEUDOPHYLLÎNÆ

Antennal scrobes with the margins hardly producedi i. Hind tibiæ without apical spines above. (Såga, palæarc., Clònia, Hemiclònia, ethiop.; Hemisåga, Austr.)
Hind tibiæ with an apical spine above on one or both sidesj
j. Hind tibiæ with an apical spine only on the outer side above. ( <b>Tympanóphora</b> , Austr.; <b>Mortoniéllus</b> , malay.).
(Tympanopnora, Austr.; Mortonieilus, maiay.). TYMPANOPHORÌNÆ
Hind tibiæ with an apical spine on both sides above, or only on
the inner sidek
k. Front and middle tibiæ armed with short or medium sized spines. l
Front tibiæ, or both front and middle tibiæ, armed with long
spines decreasing in length apically. (Listroscelis, Phlùgis, neotrop.; Hexacéntrus, indomal.; Phìsis, ethiop., indoaustr.).
LISTROSCELINÆ
l. All the femora unarmed beneath, rarely the hind ones armed on
the outer or on both sides; usually smaller species. (Conocéphalus (=Xiphídion) cosmop.; Orchélimum, widespr.;
céphalus (=Xiphídion) cosmop.; Orchélimum, widespr.;
Odontoxiphídium, nearc.). (XIPHIDIÌNÆ). CONOCEPHALÌNÆ
All femora usually spined below; rarely the hind ones armed only
on the outer side, in which case the fastigium of the vertex is
either forked or extended considerably beyond the basal joint
of the antennæ; usually larger species
m. Fastigium of the vertex usually noticeably narrower than the first joint of the antennæ, sometimes dorsally sulcate. (Agrècia,
neotrop., ethiop., austromal.; Eschatócerus, neotrop.; Nícsara,
austr.: Salomòna, indoaustr.) AGRŒCIÌNÆ
Fastigium of the vertex usually distinctly broader than the first
joint of the antennæ, never sulcate. (Neoconocéphalus, Am.;
Euconocéphalus, ethiop., indoaustr.; Homorocóryphus, cosmop.; Copíphora, neotrop.). (CONOCEPHALINÆ).
COPIPHORÎNÆ
n. First joint of hind tarsi with a free plantula beneath (Fig. 40).
( <b>Ánabrus</b> , nearc.; <b>Metrióptera</b> , ĥolarc., ethiop.; <b>Décticus</b> , palæarc., ethiop.). ( <i>DECTICÌNÆ</i> ) <b>TETTIGONIÌNÆ</b>
First joint of hind tarsi without a free plantula, or with a very
short one. ( <b>Phasgonùra</b> ( $=Locústa$ ), palæarc.: <b>Onconòtus</b> .
palæarc.). $(LOC\ USTIN\cancel{E})$ PHASGONURIN\cancel{E}
o. Third joint of hind tarsi longer than the second; front tibiæ with
an apical spine on the inner side; hind tibiæ without an apical spine above on the outer sidep
Third joint of hind tarsi shorter than the second; front and hind
tibiæ with an apical spine above on both sides. (Bradýporus,
palæarc.; <b>Derállimus, Callímenus,</b> palæarc).
BRADYPORÎNÆ

p. Antennæ inserted between the lower margins of the eyes; pronotum unarmed; both sexes winged; front tibiæ with an apical spine above on the outer side; hind tibiæ with four apical spurs below. (Pycnogáster, Ephíppiger, Urómenus, Steropleùrus, palæarc.) .... EPHIPPIGERÎNÆ

Antennæ inserted distinctly below the eyes; pronotum spined; female wingless; front tibiæ without apical spines above; hind tibiæ without apical spurs below, or with only two. (Hetròdes, Acánthoplus, Eugáster, Anepiscéptus, ethiop.).

HETRODÎNÆ



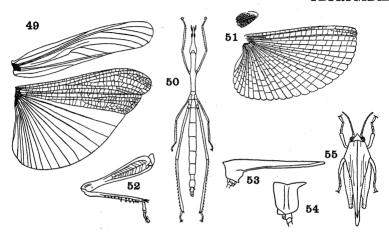
Figs. 42-48. Orthoptera

- 42. Gryllotalpa, wings (Handlirsch) Gryllotalpidæ.
- 43. **Tridactylus**, wings (Handlirsch) Tridactylidæ.
- 44. Cylindracheta (Giglio-Tos) Cylindrachetidæ.
- 45. **Tridactylus**, front leg (Saussure) Tridactylidæ.
- 46. Tridactylus, middle leg (Saussure) Tridactylidæ.
- 47. Gryllotalpa, front leg (Berlese) Gryllotalpidæ.
- 48. Cylindracheta, antenna and palpus (Giglio-Tos) Cylindrachetidæ.

	Hind tarsi one-jointed; hind legs greatly enlarged, saltatorial; three small ocelli; small species not boring in plants. ( <b>Tridácty</b> -
	lus, cosmop.; Rhipípteryx, Am.) TRIDACTÝLIDÆ
0	
8.	Front tibiæ strongly dilated, digitate; ovipositor short, not pro-
	truded; large species with very large, elongate prothorax, bur-
	rowing in the soil. (Gryllotálpa (Figs. 42, 47), cosmop.;
	Scapteríscus, Am.) GRYLLOTÁLPIDÆ
	Front tibiæ not dilated and digitate; ovipositor projecting, usu-
	ally long; antennæ always with more than thirty joints.
	(ACHÉTIDÆ) GRÝLLIDÆ
	The Gryllidæ include six subfamilies, distinguishable as follows:
a.	Tarsi compressed, the second joint minute, compressedb
	Tarsi with the second joint cordate, depressede
b.	Hind tibiæ greatly widened, furnished with a few strong movable
	spines; hind femora very broad, oval; antennæ relatively blunt
	at tips; eyes minute; small, wingless species living in ants' nests.
	(Myrmecophila, widespr.)
	Hind tibiæ slender or only slightly widened, serrulate, or serrulate
	and spinose; femora more slender; antennæ finely tapered at
	tips; usually large, free living species
$\mathbf{c}.$	Hind tibiæ spinose, but not serrulate; stout-bodied, usually dark
	colored species. (Fig. 30.). (Grýllus, Nemòbius, cosmop.;
	Gryllòdes, widespr.; Ácheta, palæarc., ethiop.). (NEMO-
	BIÌNÆ, ACHETĪNÆ)GRYLLÌNÆ
	Hind tibiæ serrulate, sometimes spinose alsod
d.	Hind tibiæ spinose, serrulate between the spines; body and legs slen-
	der; moderate or large species. Tree crickets. ( <b>Œcánthus</b> , cosmop.
	(Fig. 31); Amphiacústa, Phalangópsis, Am.; Endacústa,
	Austr., neotrop.; Phæophýllacris, ethiop.)
	Hind tibiæ with two rows of fine serrulations, but without spines
	or thorns. (Mogoplistes, palearc., ethiop., neotrop.; Ornè-
	bius, widespr.; Cyclóptilum, Am.; Ectadóderus, widespr.).
	MOGOPLISTÎNÆ
e.	Hind tibiæ not serrate, biseriately spinose and with five apical
	spurs. (Trigonidium, palearc., ethiop., indomal.; Cyrtó-
	xiphus, widespr.; Homæóxiphus, indomal.; Anáxiphus,
	neotrop.) TRIGONIDIÌNÆ
	Hind tibiæ serrate, bearing more or less numerous spines and six
	apical spurs. (Encoptera, neotrop.; Nísitra, malay.; Cardio-
	dactylus, Austromal.; Orócharis, Am.) ENEOPTERÌNÆ
9.	Tarsal claws without a pad (arolium) between them; pronotum

greatly lengthened, extending backwards to cover the entire

abdomen, fore wings vestigial, consisting of small scales at the base of the usually large hind wings; antennæ longer than the front femora. Grouse locusts. (Têtrix (Figs. 51, 53), Paratéttix, widespr.; Neotéttix, nearc.; Mazarrèdia, indoaustr.; Tettigídea (Fig. 55), Am.). (TETTÍGIDÆ, ACRYDÌIDÆ).



Figs. 49-55. Orthoptera

- 49. Dissosteira, wings (accessory veins in part omitted) (Snodgrass) Acrididæ.
- 50. Prosarthria (Brunner) Proscopiidæ.
- 51. Tetrix, wings (Handlirsch) Tetrigidæ.
- 52. Acrida, hind leg (Lugger) Acrididæ.
- 53. **Tetrix,** side view of pronotum (Packard) Tetrigidæ.
- 54. Caloptenus, side view of pronotum (Packard) Acrididæ.
- 55. **Tettigidea** (Blatchley) Tetrigidæ.

10. Body greatly lengthened and very slender, stick-like, with very long thin legs; wings absent or vestigial; head conical and greatly lengthened; prothorax tubular, not overlapping the mesonotum; antennæ eight-jointed; arolia sometimes small or indistinct. (Fig. 50). (Proscòpia, Prosárthria, Apióscelis, Corvnorhýnchus, Astròma, neotrop.).... PROSCOPÌIDÆ

11. Hind legs similar to the middle ones; their femora scarcely lengthened and not strongly thickened; body, especially the abdomen swollen or inflated; pronotum very large; green or brightly colored species. (Pneùmora, Búlla, Cystocœlia, S. Afr.). PNEUM ÓRIDÆ Hind legs very different from the middle ones, their femora large. greatly thickened at the base and lengthened; abdomen not noticeably swollen. Grasshoppers, Locusts, Short-horned locusts. (LOCÚSTIDÆ, ACRÎDÎIDÆ, ACRYDÎIDÆ). ACRÍDIDÆ This family includes the following subfamilies: a. Prosternum simple, flat.....b Prosternum with a spine, swelling, or lamellate elevation anteriorly Antennæ shorter than the front femora.....f c. Vertex and front forming together a rounded surface, the front Vertex and front meeting at an angle, the surfaces of the two forming an angle when seen in profile.....e d. Antennæ clavate, as long as the body; apterous species. (Gom-Antennæ not clavate, shorter; usually winged species. A large cosmopolitan group. (Œdípoda, widespr.; Árphia, Hippíscus, Am.; Locústa, Pachýtylus, widespr.; Dissosteira (Fig. 49), Trimerótropis, nearc.; Acrótylus, Œdàleus, old world). ŒDIPODÌNÆ e. Impressions of vertex wanting; head horizontal, the front nearly horizontal; margins of antennæ serrate; apterous species. (Psednùra, Austr.)...... PSEDNURÎNÆ Impressions of vertex present, or if rarely wanting, the head is conical, with the front more sloping, and wings are present. (Truxàlis, Am.; Stenóbothrus, widespr.; Gomphócerus, palæarc., Am.; Stauronòtus, palæarc., ind.; Mecostèthus, palæarc.). (TRYXALÌ NÆ, TRUXALÌ NÆ).... ACRIDÎNÆ f. Pronotum greatly flattened at the sides, roof-shaped and frequently with a median ridge; posterior femora dilated and compressed. (Scirtótypus, ethiop., indomal.; Chorétypus, indomal.; Brachýtypus, ethiop.) ..... CHORŒTYPÎNÆ Pronotum not greatly flattened at the sides, and not ridged above; posterior femora slender. (Eriánthus, indoaustr.; Teichophrys, Eumástax, Masýntes, neotrop.)...EUMASTACÌNÆ

g. Impressions of the vertex large and shallow, forming the anterior end of the vertex where they are separated by a very narrow

- Impressions of vertex not forming the anterior end of the vertex, placed above, at the sides or below, or obsolete.....h

# LITERATURE ON ORTHOPTERA (sens. lat.), INCLUDING PHASMATODEA, DERMAPTERA, BLATTARIÆ AND MANTODEA

BLATCHLEY, W. S. Orthoptera of Illinois. 27th Ann. Rept. Dept. Geol. Nat. Res. Indiana, 1902, pp. 123–471 (1903).

The Orthoptera of Northeastern America. Nature Pub. Co. (1920).

Bruner, L. Preliminary Catalogue of the Orthopteroid Insects of the Philippines. Univ. Nebraska Studies, **15**, pp. 195–281 (1916).

Brunner, C. v. W. Prodromus der europäischen Orthopteren. Leipzig (1882). Revision du Système des Orthoptères. Ann. Mus. Civ. Stor. Nat. Genova, **13**, pp. 1–230 (1893).

Burr, M. A Synopsis of the Orthoptera of Western Europe. London (1910). Chopard, L. Orthoptères et Dermaptères. Faune de France. (1922).

Fröhlich, C. Die Odonaten und Orthopteren Deutschlands. Jena (1903).

Fruhstorfer, H. Die Orthopteren der Schweiz. Arch. Naturg., Jahrg., 87A, Heft 5, pp. 1–262 (1921).

Hebard, M. Studies on the Dermaptera and Orthoptera of Ecuador. Proc. Acad. Nat. Sci. Philadelphia, **76**, pp. 109–248 (1924).

Studies on the Dermaptera and Orthoptera of Colombia. Trans. American Entom. Soc., **52**, pp. 275–354 (1927).

The Orthoptera of Colorado. Proc. Acad. Nat. Sci., Philadelphia, 81, pp. 303-425 (1929).

Houlebert, C. Thysanoures, Dermaptères et Orthoptères. In Encycl. Sci. (1924). Hubbell, T. H. Dermaptera and Orthoptera of Berrien Co., Michigan. Pap. Mus. Zoöl., Univ. Michigan, No. 116, 77 pp. (1922).

Innes, W. Revision des Orthoptères de l'Egypt. Mém. Soc. Entom. Egypte, 3, pp. 5-176 (1929).

KARNY, H. Zur Systematik der Orthopteroiden Insekten. Treubia, 3, p. 236 (1923).

- Kirby, W. F. A Synonymic Catalogue of the Orthoptera. Vols. 2 and 3. British Mus. Nat. Hist., London (1907–10).
- Lucas, W. J. Monograph of the British Orthoptera. London, Ray Soc., 264 pp. (1920).
- Morse, A. P. Manual of the Orthoptera of New England. Proc. Boston Soc. Nat. Hist., **35**, no. 6 (1920).

Orthoptera of Maine. Bull. Maine Agric. Sta., No. 296, 36 pp. (1921).

- Piers, H. The Orthoptera of Nova Scotia. Proc. Nova Scotia Inst., Halifax, 14, pp. 201–351 (1918).
- RAMME, W. Orthoptera, In Tierwelt Mitteleuropas, 4, Lief. 2, pp. VI 1–22 (1928).
- Saussure, H. and others. Orthoptera. In Biologia Centrali-Americana, 1, (1893–99).
- Scudder, S. H. Guide to the Genera and Classification of North American Orthoptera. Cambridge, Mass. (1897).

Catalogue of Orthoptera of the United States. Proc. Davenport Acad.

Nat. Sci., 8, pp. 1–101 (1900).

Index to North American Orthoptera. Occas. Pap. Boston Soc. Nat. Hist., No. 6, pp. 1–436 (1901).

TÜMPEL, L. Die Geradeflügler Mitteleuropas, 2d edit. Gotha. 327 pp. (1907).

Walden, B. H. Orthoptera of Connecticut. Bull. State Geol. Nat. Hist. Surv. Connecticut, No. 16, pp. 48–169 (1911).

### SUBORDER TETTIGONIODEA

- Bolivar, J. Orthoptera palæarctica. Pycnogastrinæ. Eos, **2**, pp. 423–463 (1926).
- Bruner, L. Revision of Tropical American Tettigonoidea. Ann. Carnegie Mus., 9, pp. 284–404 (1912).
- Brunner, C. v. W. Monographie der Phaneropteriden. 399 pp., Vienna (1878).

Monographie der Stenopelmatiden und Gryllacriden. Verh. zool.-bot. Ges., Wien, 38, pp. 247–394 (1888).

Additamenta zur Monographie der Phaneropteriden. Verh. zool.-bot. Ges., Wien, **41**, pp. 1–196 (1891).

Monographie der Pseudophylliden. 282 pp., Vienna (1895).

CAUDELL, A. N. Cyrtophili of the United States. Journ. New York Entom. Soc., 14, pp. 32–45 (1906).

The Decticinæ of North America. Proc. U. S. Nat. Mus., **32**, pp. 285–410 (1907).

Locustidæ, Decticinæ. Gen. Insectorum, fasc. 72, 43 pp. (1908).

Locustidæ, Prophalangopsinæ. Gen. Insectorum, fasc. 120, 7 pp. (1911).

Locustidæ, Ephippigerinæ. Gen. Insectorum, fasc. 140, 10 pp. (1913).

Locustidæ; Meconeminæ, Phyllophorinæ, Tympanophorinæ, Phasgonurinæ, Phasmodinæ, Bradyporinæ. Gen. Insectorum, fasc. 138, 25 pp. (1913).

- Locustidæ, Mecopodinæ. Gen. Insectorum, fasc. 171, 32 pp. (1916).
- Locustidæ, Hetrodinæ. Gen. Insectorum, fasc. 168, 13 pp. (1916).
- Locustidæ, Saginæ. Gen. Insectorum, fasc. 167, 8 pp. (1916).
- The Genera of Rhaphidophorinæ of America North of Mexico. Proc. U. S. Nat. Mus., 49, pp. 655-690 (1916).
- Chopard, L. Revision of the Indian Gryllidæ. Rec. Indian Mus., **30**, pp. 1–36 (1928).
- Fulton, B. B. Tree Crickets of New York. Tech. Bull. No. 42, New York Agric. Exp. Sta. (1915).
- Giglio-Tos, F. Sulla posizione systematica del genere Cylindracheta. Ann. Mus. Civ. Genoa (3), **6**, pp. 81–101 (1914).
- Hebard, M. Review of North American Species of Myrmecophila. Trans. American Entom. Soc., **46**, pp. 91–111 (1920).
  - Studies on the Gryllidæ of Panama. Trans. American Entom. Soc., **54**, pp. 233–294 (1928).
- KARNY, H. Revisio Conocephalidarum. Abh. zool.-bot. Ges., Wien, 4, Heft 3, pp. 1–114 (1907).
  - Locustidæ, Listroscelinæ. Gen. Insectorum, fasc. 131, 20 pp. (1912).
  - Locustidæ, Conocephalinæ. Gen. Insectorum, fasc. 135, 17 pp. (1913).
  - Locustidæ, Copophorinæ. Gen. Insectorum, fasc. 139, 50 pp. (1913).
  - Locustidæ, Agrœciinæ. Gen. Insectorum, fase. 141, 47 pp. (1913).
  - Fauna Buruana, Orthoptera, Fam. Gryllacridæ. Treubia, **7**, pp. 41–84 (1925).
  - Revision of the South African Gryllacridæ. Ann. South African Mus., **29**, pp. 77–151 (1929).
- Karsch, F. Über die Hetrodiden. Berliner entom. Zeits., 31, pp. 43–72 (1887).
  Über die Orthopterenfamilie der Prochiliden. Entom. Nachr., 17, pp. 97–107 (1891).
- Redtenbacher, J. Monographie der Conocephaliden. Verh. zool.-bot. Ges. Wien, 41, pp. 315–562 (1891).
- Rehn, J. A. G. and Hebard, M. Revision of the Orthopterous Group Insaræ (Phaneropterinæ). Trans. American Entom. Soc., 40, pp. 37–184 (1914).
- Saussure, H. Mélanges Orthoptérologiques. Gryllides. 2 pts., Geneva (1887–78).
  - Revision du genre Tridactylus. Rev. Suisse Zool., 4, pp. 407–419 (1897).
- Saussure, H. and Zehnter L. Monographie des Gryllotalpiens. Rev. Suisse Zool., 2, pp. 403–430 (1894).
- Schimmer, F. Monographie der Gryllodengattung Myrmecophila. Zeits. wiss. Zool., **93**, pp. 409–534 (1909).
- Scudder, S. H. The North American Ceuthophili. Proc. American Acad. Arts Sci., 30, pp. 17-113 (1894).
  - Stenopelmatinæ of the Western United States. Canadian Entom., 31, pp. 113–121 (1899).
- Shiraki, T. Monographie der Grylliden von Formosa und Japan. 129 pp. (1911).

- Tepper, J. G. O. The Phanopterinæ of Australia. Trans. Roy. Soc. South Australia, 15, pp. 77–113 (1892).
- Tindale, N. B. Australasian Mole Crickets of the Family Gryllotalpidæ. Rec. South Australian Mus., 4, pp. 1–42 (1928).
- Uvarov, B. P. Revision of the Old World Cyrtacanthacrini. Ann. Mag. Nat. Hist. (9), 11 and 12 (several parts) (1923).

### SUBORDER ACRIDODEA

Bolivar, J. Acrididæ, Pyrgomorphinæ. Gen. Insectorum, fasc. 90, 58 pp. (1909)

Acrididæ, Pamphaginæ. Gen. Insectorum, fasc. 170, 40 pp. (1916).

Bruner, L. Orthoptera, Acridiidæ. Biologia Centrali-Americana, 2 (1900–09). Synoptic List of the Paraguayan Acrididæ. Proc. U. S. Nat. Mus., 30, pp. 613–694 (1906).

South American Tetrigidæ. Ann. Carnegie Mus., 7, pp. 89–143 (1910).

Brunner, C. v. W. Monographie der Proscopiden. Verh. zool.-bot. Ges., Wien, 40, pp. 87–124 (1890).

Burr, M. Eumastacidæ. Gen. Insectorum, fasc. 15, 23 pp. (1904).

Essai sur les Eumastacides. Ann. Soc. España Hist. Nat., 28, pp. 75–112; 253–304 (1899).

Monograph of the genus Acrida (=Truxalis). Trans. Entom. Soc. London, 1902, pp. 149–187 (1902).

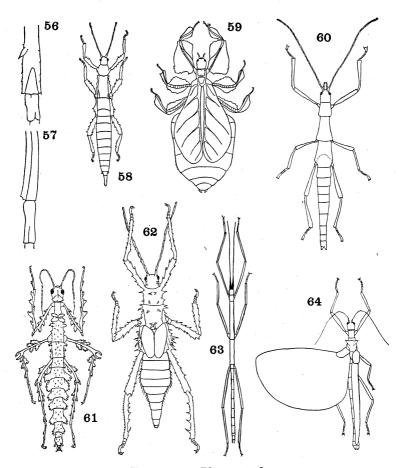
- CLAASSEN, P. W. The Grasshoppers of Kansas, Pts. 1 and 2. Bull. Dept. Entom. Univ. Kansas, No. 11, 126 pp. (1917).
- FINOT, A. Sur le genre Acridium. Ann. Soc. Entom. France, 76, pp. 247–354 (1907).
- Fletcher, T. B. Acrydidæ. Cat. Indian Ins. Govt. India, Centr. Pub. Br., Calcutta, pt. 1 (1921).
- Hancock, J. L. The Tettigidæ of North America. Chicago, 188 pp. (1902). Acridiidæ, Tetriginæ. Gen. Insectorum, fasc. 79, 4 pp. (1906).
- Hebard, M. Key to the North American genera of Acridinæ. Trans. American Entom. Soc., **52**, pp. 47–59 (1926).
  - Studies in the Tettigoniidæ of Panama. Trans. American Entom. Soc., **53**, pp. 79–156 (1927).
- Kirby, W. F. Acrididæ. Fauna British India (1914).
- MacNeill, J. Revision of the Truxalidæ of North America. Proc. Davenport Acad. Sci., 6, pp. 179–274 (1897).
- Rehn, J. A. G. North American Edipodinæ. Trans. American Entom. Soc., 45, pp. 229–255 (1919).
- Rehn, J. A. G. and Hebard, M. Study of North American Eumastacinæ. Trans. American Entom. Soc., 44, pp. 203–250 (1918).
- SAUSSURE, H. Prodrome des Œdipodiens. Mem. Soc. Phys. Hist. Nat., Geneva 28, No. 9, 254 pp. (1884).
- Scudder, S. H. Revision of the Melanopli. Proc. U. S. Nat. Mus., 20, pp. 1–421 (1897) and Proc. Davenport Acad. Sci., 7, pp. 155–205 (1899).

- Shiraki, T. Acrididen Japans. Tokio, 90 pp. (1910).
- Somes, M. P. The Acrididæ of Minnesota. Bull. Minnesota Agric. Expt. Sta., No. 141, pp. 1–100 (1914).
- Thomas, C. Synopsis of the Acrididæ of North America. Rept. U. S. Geol. Surv. Terr., Washington Govt. Printing Office, 262 pp. (1873).
- Uvarov, B. P. Revision of the Genus Locusta (= Pachytylus). Bull. Entom. Res., 12 (1921).
- WILLEMSE, C. Revision of Acrididæ of Sumatra. Tijdschr. v. Entom., 73, pp. 1-206 (1930).

# ORDER PHASMATÒDEA

# (PHÁSMIDA, PHASMÒIDEA, PHASMÒDEA, GRESSÒRIA)

Large or very large insects, generally with very slender body of cylindrical, stick- or twig-like form; rarely greatly flattened and leaflike. Head more or less freely movable, usually with long slender antennæ; eyes well developed; ocelli often absent. Prothorax short or very short, even in the very elongate forms; mesothorax short or elongate; metathorax frequently long, very closely united with the first abdominal segment (median segment), the suture between them often obscured or effaced. Abdomen usually long, cylindrical or tapering, the segments similar; cerci unjointed. Legs nearly always long or very long, similar, the tarsi five-jointed. Wings very frequently entirely absent; when present the fore wings are usually very much smaller than the hind ones, of stouter consistency; hind wings when well developed with a narrow, stouter anterior portion and a very large delicate anal expansion or fan. Metamorphosis very slight and gradual; terrestrial and vegetarian in all stages. Mainly tropical species. Walking sticks, Stick insects, Leaf insects.



Figs. 56-64. Phasmatodea

- 56. Heteropteryx, tip of hind tibia (Brunner) Phasmatidæ.
- 57. Bacteria, tip of hind tibia (Brunner) Bacteriidæ.
- 58. Heterocopus (Brunner) Bacillidæ.
- 59. Phyllium (Brunner) Phyllidæ.
- 60. Anisomorpha (Caudell) Phasmatidæ.
- 61. Trychopeplus (Hebard) Bacteriidæ.
- 62. Heteropteryx (Brunner) Phasmatidæ.
- 63. Pseudomeryle (Caudell) Phasmatidæ.
- 64. Pomposa (Brunner) Bacteriidæ.

- This family includes three groups, conveniently regarded as subfamilies, separable as follows:
- a. Prosternum with two roughened tubercles between the front coxe; antennæ often as long as or longer than the front legs, always much longer than the front femora. (Fig. 58). (Óbrimus, Heterocòpus, Tisámenus, Hoploclònia, Dátames, Dòres, malay.).
  OBRIMÎN A
- b. Antennæ much shorter than the front legs, rarely (Xylica) as long; old world species. (Pseudodátames, Círsia, Antongília, Madagascar; Xýlica, ethiop.; Bacíllus, palæarc.).

# BACILLÌNÆ

Antennæ as long as or longer than the front legs; mainly new world species. (Pygirhýnchus, Céroys, Acanthocolònia, Mirophásma, Canulèius, neotrop.; Oròbia, Madagascar).

### **PYGIRHYNCHÎNÆ**

- 3. Antennæ long in the male, many jointed, very short in the female, with few joints; mesonotum quadrate or transverse; fore wings of female covering the greater part of the abdomen; entire sides of abdomen broadly dilated into leaf-like extensions. (Fig. 59). Leaf insects. (Phýllium, ethiop., indomal.; Chitoníscus, polynes.; Nanophýllium, New Guinea)..... PHYLLĪIDÆ Antennæ long in both sexes; mesonotum longer than wide; abdo-

Four subfamilies are recognized, separable by the following key:

- - Tarsal claws pectinate; fore wings when present filiform or stalked. (Aschiphásma (= Ascepásma), Dina, malay.; Prebistus, Abrosòma, indomal.). (ASCEPASMINÆ).

# **ASCHIPHASMATINÆ**

- b. Body and legs armed with numerous spines (rarely the male not spiny); femora quadrate, not compressed, above with an apical spine; apical area of tibia produced into a spine. (Fig. 56).
  (Anisacántha, Parectatosòma, Madagascar; Leócrates, malay.; Heterópteryx, indoaustr.). HETEROPTERYGÎNÆ Body and legs armed with sparse teeth or spines; femora above
- without apical spine; apical area of tibia unarmed...........c.
  c. Sixth abdominal segment quadrate (male), or transverse (female),
- c. Sixth abdominal segment quadrate (male), or transverse (female), rarely elongate; legs unarmed; femora not compressed nor with

Sixth abdominal segment more elongate, much longer than wide (male) or quadrate (female); front femora either compressed or with leaf-like dilatations. (Donùsa, Eùcles, Strátocles, Brizòides, Phásma, Prexáspes, Prísopus, neotrop.; Phæophásma, malav.; Damasippòides. Madagascar).

PHASMATINÆ

4. Median segment short, transverse or but little longer than wide, much shorter than the metanotum; wingless species.

BACUNCULIDÆ

This family is divisible into the following subfamilies:

- a. Antennæ distinctly shorter than the front legs; old world species.

  (Clitúmnus, Cuniculìna, indomal.; Pachymórpha, indoaustr.; Gratídia, ethiop., ind.; Árphax) .... CLITUMNÌNÆ

  Antennæ as long as or longer than the front legs.............b
- b. Last dorsal segment of male more or less bilobed; operculum (eighth sternite of female) usually compressed, boat-shaped. (Menóxemus, Prómachus, Lonchòdes, Caraùsius, indo-austr.; Dixíppus, indomal.; Prosomèra, malay.).

LONCHODINÆ

- Last dorsal segment of male truncate; operculum vaulted, lanceolate. (Bacúnculus, Am.; Diapheromèra, N. Am.; Libèthra, Ocnóphila, Dỳme, neotrop.) . . . . . . . . BACUNCULÌNÆ
- Median segment as long as or longer than the metanotum, or at least much longer than wide; body frequently winged.

### BACTERÌIDÆ

This family may be divided into three subfamilies as follows:

- b. Front femora unarmed above or similarly dentate on both sides, or not three-sided. (Fig. 61). (Bactrídium, Cleonístria, Bóstra, Bactèria, neotrop.; Palòphus, ethiop.; Dimorphòdes, malay.; Eurycántha, Austr.). (PHIBALOSOMINÆ).

BACTERIÎNÆ

Front femora three-sided, spinose-dentate on the inner side above; cerci frequently large, leaf-like. (Hermárchus, Acrophýlla,

### LITERATURE ON PHASMATODEA

See also Orthoptera (sens. lat.), p. 57

Brunner, C. v. W. and Redtenbacher, J. Die Insekten-familie der Phasmiden. Parts I-III., 589 pp. Leipzig. (1906–08).

CAUDELL, A. N. The Phasmidæ or Walking Sticks of the United States. Proc. U. S. Nat. Mus., 26, pp. 863–885 (1903).

Günther, K. Die Phasmoiden Neuguineas. Mitt. zool. Mus. Berlin, 14, pp. 599–746 (1928).

Kirby, W. F. A Synonymic Catalogue of the Orthoptera, Vol. 1. British Mus. Nat. Hist., London (1904).

Shelford, R. Phasmidæ, in Biologia Centrali-Americana, 2 (1909).

Westwood, J. O. Catalogue of the Insects in the British Museum. Phasmidæ. London, British Mus., 195 pp., 48 pls. (1859).

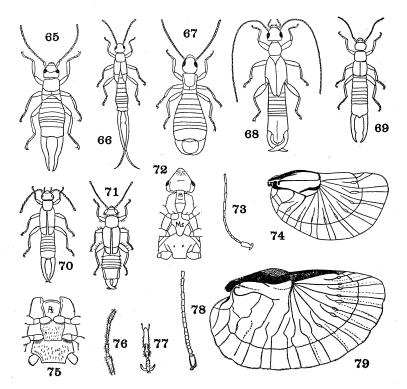
### ORDER DERMÁPTERA

Moderate-sized or small, more or less depressed insects, of elongate form, with the abdomen terminating in a pair of strong, movable forceps. Head free; antennæ filiform, with from 10 to 30, rarely more, joints; mandibles large, always (except Arixenia) fitted for biting. Prothorax free, more or less quadrate. Meso- and metathorax clearly separated; fore wings (elytra) short, leaving the abdomen exposed, heavily chitinized; hind wings (Fig. 79) large, orbicular or broadly oval, at base with two or three cells surrounded by heavy veins; anal fan very large, with a number of radiating veins; radially folded from near the center, and when at rest almost entirely covered by the elytra; sometimes one or both sexes apterous. Abdomen long, very flexible, with from eight to ten exposed segments; cerci forming the anal forceps; one-jointed, except in the nymphs of a few genera, where they are multiarticulate. Legs rather short; tarsi three-jointed, with claws. Metamorphosis slight and gradual; terrestrial in all stages. Earwigs.

1. Mandibles well developed, fitted for chewing; cerci heavily chitinized and opposable like the blades of a pair of forceps; eyes well developed; usually winged; not ectoparasitic; the body shining, rarely noticeably pubescent. (Suborder FORFICULINA) . . 2

Mandibles not fitted for chewing, toothed at tips and fringed with bristles along the inner margin; cerci weakly chitinized; eyes much reduced; apterous species, the body strongly pubescent,

	living as ectoparasites on bats. (Suborder ARIXENINA).
	(Arixènia, malayan) ARIXENÌIDÆ
2.	Metapygidium and telson present as two small plates behind the
	pygidium, or else all three are fused together with the tenth
	tergite to form a large horizontal plate (squamopygidium);
	pygidium simple, never with complex processes; ædæagus of
	male double
	Metapygidium and telson not developed or vestigial; pygidium
	well developed, often with complex processes; ædæagus of male
	a single median piece. (EUDERMÁPTERA)16
3.	Squamopygidium absent, the pygidium, metapygidium and telson
	all present as separate plates; body not strongly flattened (ex-
	cept Platylabiidæ). (PROTODERMÁPTERA)4
	Squamopygidium developed; hind wing with an oblique branch in
	the large basal cell; body very strongly flattened; a large scutel-
	lum visible between the elytra at base. (If elytra and scutellum
	are absent, see Gonólabis, couplet 13). (Apáchyus (Figs. 68, 74)
	indomal., Austr.; Dendroiketes, ind.). (Superfamily APACHY-
	ÒIDEA) (PARADERMÁPTERA)APACHŸIDÆ
4.	Metapygidium and telson not reduced, nearly as large as the rel-
	atively small pygidium; head depressed, truncate or concave
	and not emarginate behind; femora compressed and generally
	keeled. (Superfamily PYGIDICRANOIDEA) (PYGIDI-
	CRANALES)
	Metapygidium and telson much reduced in size, greatly smaller
	than the pygidium which is relatively very large; femora not
	compressed or keeled. (Superfamily LABIDURÒIDEA) (LAB-
	$IDUR\grave{A}LES$ )
5.	Femora keeled
	Femora not keeled9
6.	Antennæ with from 15 to 25 joints, the fifth and sixth joints elon-
	gate (Fig. 73)
	Antennæ with 25 to 35 joints, the fifth and sixth short, transverse $$
	or quadrate (Fig. 78)8
7.	Both elytra and hind wings absent; metasternum truncate behind;
	nymphs with forceps-like cerci. (Anatèlia (Fig. 73), Canary
	Isl.; Chállia, North China) ANATELÌIDÆ
	Elytra always, and hind wings usually perfect; metasternum sin-
	uate or excavated behind; nymphs with long, many-jointed
	cerci, not forceps-like. (Díplatys, tropicopol.).
	DIPLATŸIDÆ



Figs. 65-79. **Dermaptera** 

- 65. Burriola (Burr) Cheliduridæ.
- 66. Timomenus (Burr) Ancistrogastridæ.
- 67. Esphalmenus (Burr) Esphalmenidæ.
- 68. Apachyus (Burr) Apachyidæ.
- 69. Labidura (Burr) Labiduridæ.
- 70. **Doru** (Burr) Forficulidæ.
- 71. **Propyragra** (Burr) Pyragridæ.
- 72. Allostethus, underside of head and thorax (Burr) Allostethidæ.
- 73. Anatelia, antenna (Burr) Anateliidæ.
- 74. Apachyus, wing (Tillyard) Apachyidæ.
- 75. **Esphalmenus**, underside of thorax (Burr) Esphalmenidæ.
- 76. Ancistrogaster, base of antenna (Burr) Ancistrogastridæ.
- 77. Mesasiobia, tarsus (Burr) Anechuridæ.
- 78. Karschiella, antenna (Burr) Karschiellidæ.
- 79. Forficula, wing. Forficulidæ.

8.	Antennæ unusually thick; fourth to sixth joints transverse (Fig.
	78); nymphs with jointed cerci. (Karschiella, Bormánsia,
	ethiop.)
	Antennæ not very thick; fourth to sixth segments short, but not
	transverse; cerci of nymphs forceps-like, not jointed. (Pygidi-
	cràna, neotro.p; Kalocrània, malay.; Dicràna, ethiop., in-
	doaustr.; Cranopýgia, ind.)
9.	Prosternum convex, more or less acute anteriorly; new world
	species, the body pubescent. (Fig. 71). (Pyràgra, Pyragróp-
	sis, Echinopsàlis, Propyràgra, neotrop.)PYRÁGRIDÆ
	Prosternum not acute anteriorly; old world species, the body
	clothed with short, stiff bristles. (Echinosòma, ethiop., in-
	doaustr.) ECHINOSOMÁTIDÆ
10.	Body not very strongly flattened; forceps (cerci) not flattened
	nor sickle-shaped
	Body very strongly flattened; forceps strongly flattened and
	sickle-shaped; elytra perfectly developed, the hind wings
	short; antennæ 19-20 jointed. (Platylàbia, indomal.). (PAL-
	$ ilde{I}\!$
11.	Mesosternum strongly narrowed behind (Fig. 72); stout species,
	sometimes without elytra and wings. (Allostèthus, Gono-
	labidùra, Allostethélla, malay.) ALLOSTÉTHIDÆ
	Mesosternum not strongly narrowed behind
12.	Prosternum not narrowed behind
	Prosternum narrowed behind (Fig. 75); without wings or elytra;
	abdomen of male much widened apically (Fig. 67). (Esphál-
	menus, neotrop., ethiop.; Gonolabina, neotrop.).
	ESPHALMÉNIDÆ
13.	Mesosternum rounded behind; rather stout species, winged or
	apterous. (Anisólabis, cosmop.; Psalis, Gonólabis, Eubor-
	éllia, neotrop., ethiop., ind.). (PSÁLIDÆ, ANISOLÁBIDÆ)
	PSALÍDIDÆ
	Mesosternum truncate behind
14.	Antennæ with more than twenty-five joints; elytra always and
	wings usually present. (Fig. 69). (Labidura, cosmop.; Nala,
	old world; Forcípula, widespr.; Tomopyga, indomal.).
	LABIDÜRIDÆ
1 =	Antennæ with ten to fifteen joints; without elytra or wings 15
15.	Last dorsal segment truncate. (Idolopsàlis, neotrop.; Pseudi-
	sólabis, indoaustr.; Parisólabis, Austr.). PARISOLÁBIDÆ
	Last dorsal segment bifid; slender species with long legs. (Cten-

	isólabis, Brachýlabis, Antisólabis, ethiop., Austr.; Nanni- sólabis, ind.; Metisólabis, ethiop., ind.; Leptisólabis, wide-
	spr.) BRACHYLÁBIDÆ
16.	Second joint of tarsi simple, not lobed nor dilated. (Superfamily
	LABIÒIDEA) ( <i>LABIÀLES</i> )
	Second joint of tarsi lobed or dilated. (Superfamily FORFICU-
	LÒIDEA) (FORFIC ULÀLES)23
17.	Elytra with a sharp keel or a row of minute tubercles near the
	lateral margin
	Elytra not thus keeled
18.	Tarsi long and very slender; elytra granulose, the keel formed by
	a row of minute tubercles. (Pericòmus, neotrop.).
	PERICÓMID <i>Æ</i>
	Tarsi short and relatively thick; elytra smooth, with a sharp
	keel
19.	Antennæ with the joints cylindrical20
	Antennæ with the joints enlarged apically or clavate. (Nesogás-
	ter, malay., Austr.)
20.	
	breviated; antennæ 16-to 20-jointed; old world species of slen-
	der form. (Vándex, ethiop.)
	Abdomen of male broadened at the middle; antennæ with 12-15
	joints; elytra abbreviated; new world species of stout form.
0.1	(Strongylopsalis, neotrop.)STRONGYLOPSALÍDIDÆ
21.	Body not strongly flattened
	Body strongly flattened. (Sparátta, Parasparátta, Prospar-
	átta, neotrop.; Auchénomus, ethiop., malay.).  SPARÁTTIDÆ
99	Head transverse, the median and frontal sutures strong and deep;
22.	eyes large and prominent. (Spongiphora, Purex, neotrop.;
	Vóstox, Am.; Spongovóstox, tropicopol.; Márava, austro-
	mal.) SPONGIPHÓRIDÆ
	Head narrow, the sutures weak or obsolete; eyes small, no longer
	than the first joint of the antennæ. (Làbia, cosmop.; Prolàbia,
	tropicopol.; Làrex, neotrop.; Chætospània, ethiop., indo-
	austr.; Ándex, Austr.)LABÌIDÆ
23.	Second joint of tarsi with a narrow lobe produced beneath the third
	joint; moderately stout or robust species. (Chelisódoches,
	ethiop., indoaustr.; <b>Énkrates, Pròreus</b> , indomal.; <b>Soleno-</b>
	sòma, ind.; Kleidùchus, Austr.)CHELISODÓCHIDÆ
	Second joint of tarsi with a dilated lobe on each side (Fig. 77)24

24.	Sternal plates strongly transverse; elytra much reduced in size;
	abdomen depressed and dilated; pygidium transverse. (Fig. 65).
	(Chelidùra, Burrìola, palæarc., Mesochelidùra, palæarc.,
	ethiop.)
	Sternal plates not very decidedly transverse
25.	Antennal joints not very long and slender, the fourth often much
	shorter than the third and the first much shorter than the
	head
	Antennal joints all long and slender; fourth not shorter than the
	third; first longer and thicker than the others (Fig. 76)29
26.	Meso- and metasternum and pygidium broad; forceps remote,
	not depressed; elytra entire or abbreviated. (Fig. 77). (Ane-
	chùra, Mesasiòbia, palæarc., neotrop.; Pseudochelidùra,
	palæarc.; Pterýgida, Allodáhlia, indomal.). ANECHÜRIDÆ
	Meso- and metasternum quadrate or narrow; pygidium narrow. 27
27.	Abdomen cylindrical, not depressed
	Abdomen depressed; forceps flattened or cylindrical; mesosternum
	rounded posteriorly; antennæ with 12-15 joints; elytra rarely
	shortened. (Fig. 70). (Forficula, holarc., ethiop., indomal.
	(F. auriculària, European earwig); Chelidurélla, Apterýgida,
	palæarc., ethiop.; Dòru, Amer., Austr.; Skalistes, neotrop.;
00	Hypúrgus, indomal.) FORFICÙLIDÆ
28.	Elytra entire, not shortened; mesosternum rounded posteriorly;
	old world species. (Eudóhrnia, ind.; Kosmètor, indomal.).
	EUDOHRNÌIDÆ
	Elytra abbreviated; nusosternum truncate. (Neolobóphora,
20	neotrop.; Árchidux, ethiop.) NEOLOBOPHÓRIDÆ
49.	Sternal plates generally transverse, at least relatively broad; meta- sternum truncate; abdomen and forceps depressed; new world
	species. (Fig. 76). (Ancistrogáster, Vláx, Prãos, Tristan-
	élla, Paracósmia, neotrop.) ANCISTROGÁSTRIDÆ
	Sternal plates narrow; metasternum narrow, excavated poste-
	riorly
30.	Abdomen but little depressed, surface rather convex; legs long and
50.	slender; pygidium narrow; forceps remote, slender. (Fig. 66).
	(Dinex, neotrop.; Timómenus, Epárchus, Córdax, indomal.;
	Opisthocósmia, ethiop., malay.; Thalpèrus, ethiop.).
	OPISTHOCOSMIIDÆ
	Abdomen depressed and rather dilated; legs short; pygidium
	strongly transverse. (Diaperásticus, ethiop.).
	DIAPERASTÍCIDÆ

### LITERATURE ON DERMAPTERA

See also Orthoptera (sens. lat.), p. 57

Bormans, A. and Krauss, H. Dermaptera. Das Tierreich, Lief. 11 (1900).

Burn, M. Revision of Forficulidæ and Chelisodochidæ. Trans. Entom. Soc. London, 1907, pp. 91–134 (1907).

Dermaptera. Fauna of British India. London (1910).

Revision of the Labiduridæ. Trans. Entom. Soc. London, 1910, pp. 161–203 (1910).

Dermaptera. Gen. Insectorum, fasc. 122, 112 pp. (1911).

Revision of the Genus Diplatys. Trans. Entom. Soc. London, 1911, pp. 21–47 (1911).

Burr, M. and Jordan, K. On Arixenia, a Suborder of Dermaptera. Trans. 2d Entom. Congr. (1913).

Снораво, L. Orthoptères et Dermaptères. Faune de France (1922).

Günther, K. Die Dermapteren der deutschen Kaiserin-Augusta-Flusse-Expedition. Mitt. zool. Mus. Berlin, **15**, pp. 55–83 (1929).

Kirby, W. F. A Synonymic Catalogue of the Orthoptera, vol. 1. British Mus. Nat. Hist., London (1904).

Rehn, J. A. G. The Dermaptera of the American Museum Congo Expedition with a Catalogue of the Belgian Congo Species. Bull. American Mus. Nat. Hist., 49, pp. 349–413 (1924).

WALDEN, B. H. Dermaptera of Connecticut. Bull. State Geol. Nat. Hist. Surv. Connecticut, No. 16, pp. 44-47 (1911).

ZACHER, F. Studien über das System der Protodermapteren. Zool. Jahrb. Abth. f. Syst., **30**, pp. 303–400 (1911).

# ORDER DIPLOGLOSSÀTA

# (DERMODERMÁPTERA)

Moderate-sized, flattened, wingless species living as external parasites of rodents. Head horizontal, semicircular, with the mouth opening below. Mandibles strong, dentate; maxillæ and labium well developed, their palpi five- and three-jointed respectively. Antennæ rather short, filiform, with the basal joint greatly elongated. Thoracic segments expanded laterally into flattened plates; prothorax large, free. Legs similar, stout and short; coxæ small, separated; tarsi three-jointed, with claws. Cerci long, filiform, but unsegmented. Metamorphosis imperceptible, the females viviparous; living as external parasites of rodents.

One family. (Hemimerus, South Africa)..... HEMIMÉRIDÆ

### LITERATURE ON DIPLOGLOSSATA

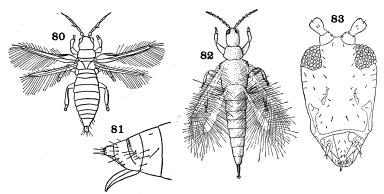
Hansen, H. J. On the Structure and Habits of *Hemimerus talpoides*. Ent. Tidskr., **15**, pp. 65–93 (1894).

JORDAN, K. Anatomy of Hemimerus. Novitat. Zool., 16, pp. 327-330 (1909). Krauss, H. Forficulidæ. Das Tierreich, Lief. 11, pp. 130-132 (1900).

## ORDER THYSANÓPTERA

(PHYSOPODA)

Small or minute, usually depressed, slender insects with the wings often reduced in size, and frequently capable of jumping, feeding generally on plant sap, or more rarely on animal juices. Head vertical, free; eyes well developed; usually three ocelli; mouthparts fitted for sucking, inferior, asymmetrical, consisting of a triangular clypeus fused



Figs. 80-83. Thysanoptera

- 80. Heliothrips (Russell) Thripidæ.
- 81. Euthrips, apex of abdomen of female, showing ovipositor (Russell).
- 82. Thrips Thripidæ.
- 83. Cephalothrips, head. (Peterson).

with the bases of the maxillæ to form a sheath that encloses the mandibular and maxillary setæ. Prothorax free; wings four, similar, narrow, with few or no veins, closely fringed with long bristles, often vestigial or absent. Legs similar; tarsi one- or two-jointed, with one or two claws, and with a bladder-like or hoof-like enlargement at tip. Abdomen with ten visible segments; terminal one frequently tubular; basal one often closely attached to the thorax. Ovipositor present or absent, when present consisting of four more or less parallel, often

strongly curved pieces. Metamorphosis gradual, the nymphs very similar to the adult; wings developing externally; penultimate instar often quiescent.

- 1. Wing surface microscopically hairy; wings usually present, fore wing with a marginal vein and at least one longitudinal vein attaining the wing tip: last abdominal segment rarely tubular. in the female usually conical and longitudinally divided beneath. in the male usually rounded at tip; ovipositor present, saw-like, composed of four pieces. (Suborder TEREBRÁNTIA).....2 Wing surface bare, without pubescence; fore wing veinless, or at most with a single, abbreviated median vein, wings often absent: last abdominal segment tubular in both sexes and not divided beneath; no ovipositor. (Suborder TUBULÍFERA)......10 2. Ovipositor curved upwards; wings usually broad, with the tips rounded; body not flattened; antennæ nine-jointed...........3 Ovipositor curved downwards; wings when present narrower and almost always pointed at tips; body more or less flattened; an-3. Labial palpi with fewer joints than the maxillary palpi; all joints of antennæ usually freely movable.....4 Labial palpi with the same number of joints as the maxillary palpi, or with one more joint; last three to five joints of antennæ 4. Maxillary palpi with seven or eight joints; labial palpi with three to five joints. (Désmothrips, Austr.; Órothrips, nearc.; Stomátothrips) ..... OROTHRÍPIDÆ Maxillary palpi with three joints; labial palpi with two joints. (Melánothrips, palæarc.; Ánkothrips, widespr.; Cránothrips, Austr.) ..... MELANOTHRÍPIDÆ 5. Antennæ extremely long and slender, the third joint at least ten times as long as thick and as long as the head (Fig. 84), joints three and four without elongated sensory areas; fore wing slender, without crossveins; anterior ocellus absent or very small. (Franklinothrips, nearc., ethiop.; Corynothripòides, ethiop.).
  - Antennæ much shorter; joints three and four with elongated sensory areas; wings broader, the fore wing with distinct crossveins; three well developed ocelli. (Æólothrips, Archæólothrips, Rhipídothrips). (COLEOPTRÁTIDÆ).

ÆOLOTHRÍPIDÆ

FRANKLINOTHRÍPIDÆ

6. Antennæ nine-jointed, sometimes apparently ten-jointed, without an apical stylus: front tarsus with a claw-like projection at the base of the second joint; third and fourth joints of antennæ enlarged, conical. (Hetérothrips, Am.). HETEROTHRÍPIDÆ Antennæ six- to eight-jointed, rarely with the second joint divided by a suture so that the antennæ appear to be nine-jointed: antennæ usually with a one- or two-jointed style at apex (Fig. 88); front tarsus simple, without appendage on second joint: third 7. Antennæ not moniliform, six- to eight-jointed, always with apical style: pronotum simple above, without longitudinal sutures: front and hind femora slender; ovipositor almost always well developed...... Antennæ moniliform (Fig. 85), eight-jointed, without apical style; pronotum with a longitudinal suture at each side; front and hind femora greatly thickened (Fig. 87); ovipositor much reduced. (Mèrothrips, nearc., neotrop.). MEROTHRÍPIDÆ 8. Sixth joint of antennæ well developed, usually as large as or Sixth or sixth and seventh joints of antennæ verv small. stylelike, very much smaller than the fifth. (Cerátothrips, palæarc.). CERATOTHRÍPIDÆ 9. Last abdominal segment in female cylindrical, very heavily chitinized, ninth and tenth segments with extremely long, stout, thorn-like bristles. (Panchætothrips, India; Dinurothrips, Macrùrothrips) ..... PANCHÆTOTHRÍPIDÆ Last abdominal segment in female conical, weakly chitinized, rarely more heavily chitinized than the preceding segments; ninth and tenth segments never with large thorn-like bristles. (Figs. 80, 88, 90). (Heliothrips, Thrips, Frankliniella, Anaphothrips. Physothrips and many other genera). (STEN-OPTÉRIDÆ) ..... THRÍPIDÆ 10. Maxillary palpi two-jointed: antennæ with seven or eight, usually eight joints; middle coxæ further apart than the other Maxillary palpi one-jointed; antennæ with four to seven joints; hind coxæ further apart than the other pairs. (Fig. 86). (Urothrips, ethiop.: Stephánothrips, ethiop., neotrop.: Brádythrips, neotrop.; Bebélothrips, palæarc.). (UROTHRIPÒI-

DEA) ...... UROTHRÍPIDÆ

Eighth abdominal segment with long, posteriorly directed, pegshaped projections along the posterior edge; last segment of abdomen extremely short and stout. (Chirothripòides).

CHIROTHRIPÒIDIDÆ

# 84 85 88 89 89

Figs. 84-90. Thysanoptera

- 84. Franklinothrips, antenna (Hood) Franklinothripidæ.
- 85. Merothrips, antenna (Hood) Merothripidæ.
- 86. Bradythrips, apex of abdomen of female (Hood) Urothripidæ.
- 87. Merothrips, leg (Hood) Merothripidæ.
- 88. Frankliniella, antenna (Karny) Thripidæ.
- 89. Pygothrips, apex of abdomen of female (Hood) Pygothripidæ.
- 90 Thrips, head and anterior part of thorax (Shaw) Thripidæ.

usually prominently overhanging the base of the antennæ and bearing the front occllus at its extremity; usually with a strong bristle in front of the eye. (Idôlothrips, Gigántothrips, Actínothrips) . . . . . . . . . . . . . . . . . IDOLOTHRÍPIDÆ

13.	Male with a stout tubular projection at each side of the sixth
	abdominal segment. (Mégathrips, Bacillothrips, Megálo-
	thrips) MEGATHRÍPIDÆ
	Sixth abdominal segment of male simple, without a tubular pro-
	jection laterally

14. Last abdominal segment greatly elongated, about as long as the remainder of the abdomen and three or four times as long as the head. (Hystrichothrips, Holùrothrips).

#### HYSTRICHOTHRÍPIDÆ

Last abdominal segment much shorter, never greatly lengthened.15

- 16. Third joint of antennæ with a strong crest-like ring of large sense cones at apex. (**Ecacánthothrips**, Indomal.; **Órmothrips**).

## ECACANTHOTHRÍPIDÆ

- 17. Sensory cones of antennæ unusually long and acute, each with an accessory cone or a long, slender bristle, the joints which bear the cones much swollen; eyes very much enlarged, contiguous. (Eupátithrips, Sedulothrips)...... EUPATITHRÍPIDÆ
  - Sensory cones not remarkably developed; eyes much smaller. (Phlæothrips, Tríchothrips, Liothrips, Acánthothrips, Zvgothrips, and many others).......PHLŒOTHRÍPIDÆ

#### LITERATURE ON THYSANOPTERA

- BAGNALL, R. S. Synopsis of Æolothripidæ. Trans. 2d Congress Ent., 2, pp. 394–397 (1913).
- The Genus Melanothrips. Entom. Monthly Mag., **60**, pp. 9–11. (1924). Hinds, W. E. Monograph of Thysanoptera of North America. Proc. U. S. Nat. Mus., **26**, pp. 79–242 (1902).
- Hood, J. D. Subfamilies of Thysanoptera. Proc. Biol. Soc. Washington, 28, pp. 53–60 (1915).
  - Keys to Genera and North American Species of Urothripidæ. Bull. Brooklyn Entom. Soc., **24**, pp. 314–321 (1929).
- Hoop, J. D. and Williams, C. B. Synopsis of Urothripidæ. Ann. Entom. Soc. America, 20, pp. 1–9 (1927).

Karny, H. Synopsis der Megathripidæ. Neue Beiträge zur systematischen Insektenkunde, Berlin, 1, pp. 105–110; 113–118 (1919).

Beiträge zur Malayischen Orthopteren-fauna, III. Thysanoptera. Treubia, 1, pp. 211–261 (1921).

Studies on Indian Thysanoptera. Mem. Dept. Agric. India, 9, pp. 187–239 (1926).

- Knechtel, W. K. Thysanoptere din Romania. Studin Monografic. Bulet. Agric., Bucarest, 2-3, pp. 1-235 (1923).
- Moulton, D. Synopsis, Catalogue and Bibliography of North American Thysanoptera. Bull. Bur. Entom. U. S. Dept. Agric., Tech. Ser. No. 21 (1911).
- PRIESNER, H. Die Thysanopteren Europas. Four parts. Vienna, Fritz Wagner (1926–28).

Thysanoptera. In Tierwelt Mitteleuropas, 4, Lief. 2, pp. VIII 1–18 1928).

Uzel, H. Monographie der Thysanopteren. 472 pp. Königgrätz (1895).

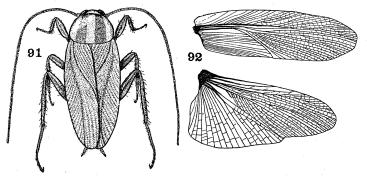
WATSON, J. R. Synopsis and Catalogue of North American Thysanoptera. Florida Agric. Expt. Sta., Tech. Bull. No. 168, 100 pp. (1923).

#### ORDER BLATTÀRIÆ

### (OOTHECÀRIA, CURSÒRIA, BLATTÒIDEA)

Moderate-sized or large, sometimes very large, rarely very small, broadly oval, flattened, quick-running insects. Head free but inflexed so as to be nearly or quite concealed beneath the pronotum, the mouth posterior or nearly so. Mandibles strong, toothed; maxillæ well developed, bilobed, with five-jointed palpi; labial palpi three-jointed; eyes usually well developed, usually two ocelli; antennæ long, filamentous, many-jointed. Prothorax large, movable, usually transverse; meso- and metathorax subequal, not freely movable. Wings often absent or much reduced in size; when present, overlapped on the abdomen; fore wings parchment-like, containing many veins; hind wings with a large anal lobe, clearly separated from the rest of the wing and radially folded. Legs strong, similar, coxæ large; tibiæ usually strongly spinose; tarsi five-jointed. Abdomen with ten tergites of nearly equal size, broadly attached at the base and not very freely movable; cerci prominent and jointed. Metamorphosis slight. Roaches.

- 3. Moderate to large, heavily pigmented species, with eyes; tenth dorsal segment of the male more or less quadrangular, often impressed, or emarginate behind. Mainly tropical. (Blåtta, cosmop. (B. orientàlis, Oriental cockroach); Eurýcotis, neotrop.; Polyzostèria, austr.; Méthana, ethiop., indoaustr.; Deropéltis, ethiop.; Periplanèta (Fig. 92) cosmop. (P. americàna, American cockroach, P. australàsiæ, Australian cockroach). (PERIPLA NETÌNÆ)



Figs. 91-92. Blattariæ

- 91. Blatella. Phyllodromiidæ.
- 92. Periplaneta, wings. Blattidæ.

Small, colorless, cave-inhabiting species, with the eyes vestigial or absent; wings reduced or absent; tenth dorsal segment of the male elongate triangular and emarginate behind. (Nocticola, Spelæoblátta, indomal.) . . . . . . . . NOCTICÓLIDÆ

- - Very small, wingless or subapterous species, 2 to 5 mm. in length, living in ant nests; entire body covered with loose hairs; legs stout, the tibiæ heavily spined; antennal joints much longer than wide. Neotropical. Ant guests. (Attáphila (Figs. 98, 99)).

**ATTAPHÍLIDÆ** 

5. Tenth dorsal segment of both sexes usually transverse and narrow; hind wings when present with an apical field; fore wings with the

branches of M and Cu strongly oblique and leading toward the

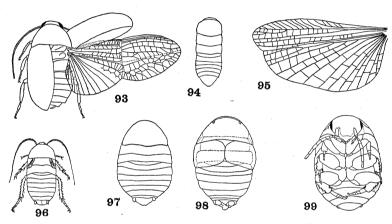
	hind margin; hind femora usually sparsely armed with spines
	beneath. (Ectòbia, cosmop.; Anaplécta, Pseudectòbia, neo-
	trop., ethiop., indomal.; Hololámpra, holarc., ethiop.).
	ECTOBÌIDÆ
	Tenth dorsal segment of both sexes more or less produced, tri-
	angular or emarginate; hind femora usually strongly spined
	beneath
6.	Tenth dorsal segment of both sexes triangular and entire, the cerci
••	distinctly projecting
	Tenth dorsal segment of the male more or less quadrate, with ob-
	tuse angles, that of the female broadly rounded or lobate, the
	cerci not projecting; tarsi with distinct pulvilli. (Calolampra,
	widespr.; Epilámpra, neotrop.; Leuroléstes, Phlebonòtus,
	Phoráspis, Hyrophicnoda (Fig. 97), neotrop.; Homalop-
	terus, indomal., neotrop.; Heterolámpra, ethiop., indoaustr.).
	(PHORASPÍDIDÆ) EPILÁMPRIDÆ
7.	Pronotum and fore wings smooth; hind wings with the radial vein
	usually emitting several parallel costal veins; pulvilli absent.
	(Caloblátta, Pseùdomops, neotrop.; Blatélla (B. germánica,
	Crotonbug) (Figs. 91, 95), Ischnoptera, Loboptera, Phyllo-
	dròmia, Temnópteryx, cosmop.; Ellipsídion, austr.).(PSEU-
	DOMÓPIDÆ)PHYLLODROMÌIDÆ
	Pronotum and fore wings covered with a silky pile; hind wings
	with the radial vein emitting irregular costal veins; pulvilli
	present; usually large species. Neotropical. (Megaloblátta,
	Nyctibora, Heminyctibora) NYCTIBÓRIDÆ
8.	Abdomen with the seventh segment normal, not enclosing the
	terminal segments, the cerci and at least the tenth dorsal seg-
	ment free9
	Abdomen with the seventh segment of both sexes broadly rounded,
	triangular and enclosing the terminal segments and also the
	cerci; tibiæ short and strongly bristly; wingless and greatly con-
	vex species. (Cryptocércus (Fig. 94), nearc.).
	CRYPTOCÉRCID <i>Æ</i>
9.	Hind wings twice as long as the fore wings, with a transverse fold
	at the middle through which the veins continue to the apex,
	folded in repose; fore wings elytra-like, with weak venation;
	insects resembling dytiscid beetles. (Diploptera (Fig. 93)

Hind wings without a middle transverse fold through which the

10. Tenth dorsal segment semicircular, broadly produced, its hind margin more or less strongly dentate; last ventral segment of the male very small, without styles; legs robust, the front pair fossorial, tibiæ strongly spinose, tarsi relatively short, without arolia; costal margin of the fore wings split, wings often reduced. (Panésthia, Salgànea, indoaustr.; Geoscápheus, austr.).

#### PANESTHÌIDÆ

Last dorsal segment without produced and dentate hind margin.11



Figs. 93-99. Blattariæ

- 93. **Diploptera** (Tillyard) Diplopteridæ.
- 94. Cryptocercus, dorsal outline (Hebard) Cryptocercidæ.
- 95. Blatella, hind wing. Phyllodromiidæ.
- 96. Cocoblatta (Saussure) Blaberidæ.
- 97. **Hyporhicnoda** (Hebard) Epilampridæ.
- 98. Attaphila, dorsal view (Wheeler) Attaphilidæ.
- 99. Attaphila, ventral view (Wheeler) Attaphilidæ.
- 12. Fore wings with the branches of M and Cu directed toward the hind margin, anal area small, with few veins extending to the

	margin, costal area short. (Chorisoneùra, Am.; Choristima,
	austr.; <b>Anáptycha, Hemiptérota,</b> neotrop.).
	CHORISONEÙRIDÆ
	Fore wings with the branches of M and Cu directed toward the
	apical margin, costal area usually long and narrow. (Areolària,
	malay.; <b>Hypnórna, Plectóptera,</b> neotrop.).
	AREOLARÌID <i>Æ</i>
13.	Very small species, 5 to 7 mm. in length, living in nests of ants14
	Larger species, not myrmecophilous
14.	3 1
	weak venation, the branches of M and Cu parallel; tibiæ with
	long bristles; cerci long and jointed. Neotropical. (Notho-
	blátta) NOTHOBLÁTTIDÆ
	Rather convex insects with abbreviated fore wings and no hind
	wings; tibial spines weak; cerci short and broad. Ant guests.
	Neotropical. (Atticola) ATTICÓLIDÆ
15.	Female with the seventh ventral segment divided behind to form
	a pair of valves
	Female with the seventh ventral segment not forming a pair of
	valves
16.	Large species; prothorax elongate-trapezoidal, not tomentose;
	legs very long, the tibial bristles weak; anal areas of the hind
	wings of the fully winged forms large and plaited several times
	in repose; fore wings with the costal cell narrow and without
	crossveins, subcosta long, reaching the middle of the wings.
	(Archiblátta, Cátara, malay.)ARCHIBLÁTTIDÆ
	Insects not conforming to the preceding description; anal area of
	the fully winged forms smaller, folded only once or twice17
17.	Small species with pubescent thorax; hind wings with a pro-
	nounced thickening surrounding the ends of the shortened sub-
	costa. Widespread, tropical. (Compsòdes, Euthýrrhapha,
	Holocómpsa) EUTHYRRHÁPHIDÆ
10	Hind wings without nodal thickening at the end of the subcosta. 18
10.	Small, delicate species; tibial bristles weak; cerci long; veins
	simple or few-branched. (Latindia, neotrop.).  LATINDÎIDÆ
	Large species; tibial bristles strong; cerci short; veins many-branched. (Homœogàmia, Am.) HOMŒOGAMÎIDÆ
10	Tarsal claws without arolia, or with a minute arolium; tenth dorsal
19.	
	segment of the male abdomen more or less deeply notched20
	Tarsal claws with a distinct arolium between them

- 21. Tenth dorsal segment of the male abdomen transverse, often constricted at the middle; large, broadly convex species. (Polýphaga, palæarc., indomal., ethiop.) . . . . . . POLYPHÁGIDÆ Tenth dorsal segment more or less produced, its hind margin notched; broad beetle-like, showy insects. (Corýdia, indomal.).
- CORYDIDÆ
  22. Dorsal segments of the abdomen with protruding lateral angles;
- - Abdominal segments without projecting lateral angles; tenth dorsal segment transverse, its hind margins straight or rounded. .23
- - Hind wings with rounded tip and no specialized apical field. (Elliptoblátta, Stenopilèma, ethiop.; Hormética, Am.; Perisphæria, indomal.; Parasphæria, neotrop.).

PERISPHÆRÌIDÆ

#### LITERATURE ON BLATTARIÆ

See also Orthoptera (sens. lat. p. 57)

Brunner, C. Nouveau Système des Blattaires. 426 pp. Vienna (1865).

Снорако, L. Orthoptères et Dermaptères. Faune de France (1922).

Blattidæ de la Nouvelle-Caledonie. Nova Caledonia, Berlin, A, Zool., **3**, pp. 301–336 (1924).

HANITSCH, R. Malayan Blattidæ. Journ. Roy. Asiatic Soc., Straits Branch, **69**, pp. 17–178 (1915).

Hebard, M. The Blattidæ of North America North of the Mexican Boundary. Mem. American Entom. Soc., No. 2 (1917).

The Blattidæ of Panama. Mem. American Entom. Soc., 4, pp. 1–148 (1919).

The Blattidæ of French Guiana. Proc. Acad. Nat. Sci. Philadelphia, 78, pp. 135–244 (1926).

Studies in Malayan Blattidæ. Proc. Acad. Nat. Sci., Philadelphia, 81, pp. 1–109 (1929).

Karny, H. H. Monographie der Phyllophorinen. Treubia, 5, suppl. 142 pp. (1924).

Kirby, W. F. A Synonymic Catalogue of the Orthoptera. Vol. 1, British Mus. Nat. Hist. London (1904).

Morse, A. P. Manual of the Orthoptera of New England. Proc. Boston Soc. Nat. Hist., **35**, No. 6 (1920).

RAMME, W. Monographie des Blattiden-Genus Ectobius. Arch. Naturg. (1923). REHN, J. G. and HEBARD, M. The Orthoptera of the West Indies: Blattidæ. Bull. American Mus. Nat. Hist., **54**, pp. 1–320 (1927).

SAUSSURE, H. Mélanges Orthoptérologiques, fasc. 2, Blattidæ. Geneva (1865).
Revision de la tribu des Hétérogamiens. Rev. Suisse Zool., 1, pp. 289–318 (1890).

Revision de la tribu des Périsphériens. Rev. Suisse Zool., 3, pp. 1–59, (1895).

Revision de la tribu des Panesthiens et Épilampriens. Rev. Suisse Zool., 3, pp. 299–361 (1895).

Shelford, R. Studies of the Blattidæ. Trans. Entom. Soc. London (1906-07) (several parts).

Blattidæ, Ectobiinæ. Gen. Insectorum, fasc. 55, 15 pp. (1907).

Blattidæ, Phyllodrominæ. Gen. Insectorum, fasc. 73, 29 pp. (1908).

Blattidæ, Nyctiborinæ. Gen. Insectorum, fasc. 74, 5 pp. (1908).

Blattidæ, Epilamprinæ, Gen. Insectorum, fasc. 101, 21 pp. (1910).

Blattidæ, Blattinæ. Gen. Insectorum, fasc. 109, 27 pp. (1910).

Tepper, J. G. O. The Blattariæ of Australia and Polynesia. Trans. Roy. Soc. South Australia, 17, pp. 25–130; 18, pp. 169–189 (1893–94).

### ORDER MANTÒDEA

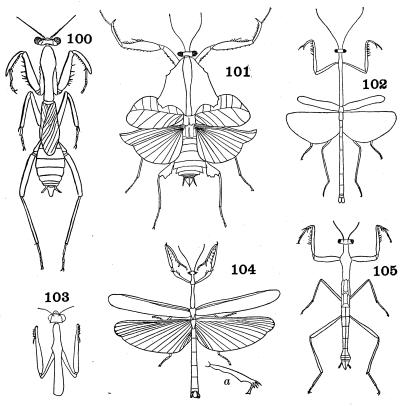
### (MANTÒIDEA)

Moderate-sized or large insects of predatory habits; the front legs very long, with the femora and tibiæ usually heavily spined and capable of being folded closely together to grasp the prey. Body elongate, sometimes very slender and usually flattened. Head freely movable, not inserted in the prothorax; eyes prominent, usually three ocelli; mandibles strong, the mouthparts inferior, rarely turned forwards; antennæ long and filamentous, many-jointed, rarely pectinate in certain males. Prothorax long or very long, freely movable, sometimes with the sides expanded; meso- and metathorax shorter, of about equal length, not freely movable. Four wings, overlapping on the abdomen, sometimes much reduced or absent, especially in the female; venation complex, fore wings usually considerably smaller than the hind pair and

of stouter consistency. Tarsi almost always five-jointed, terminating in two claws: femora and tibiæ sometimes with expanded margins. Abdomen elongate oval or long and cylindrical, the terminal segments not abbreviated, tenth tergite forming a supra-anal plate; cerci usually jointed, never very long. Metamorphosis incomplete; habits similar throughout development. Praying mantids, Soothsayers. 1. Front tibiæ without an apical hook, with two rows of spines below: cerci very long, many-jointed; small, winged species with freely movable head and simple posterior legs. (Fig. 109). (Chæ-2. Front tibiæ with one or two long teeth on the dorsal side before the apical hook, otherwise with only very few spines; cerci simple; small slender species. (Fig. 105). (Bántia, Pseudomusonia (=Mionyx), Bactromántis, Olígonyx, neotrop.; Haania, malav.) ...... OLIGONYCHÌNÆ 3. Front tibiæ without rows of strong spines below, or with only the inner row distinctly developed......4

Head of normal form, with protuberant eyes; jaws not directed forwards; stouter species with the thorax comparatively short. (Perlamántis, palearc.; Amorphóscelis, ethiop., As., Austr.). (AMORPHOSCELINÆ) ...... PERLAMANTÎNÆ

<sup>&</sup>lt;sup>1</sup>The Mantidæ include an extensive series of very diverse forms and have been grouped into a large number of subfamilies by Giglio-Tos whose divisions have been generally accepted. These together with several others of equal importance may be distinguished by the following key which follows the arrangement of Giglio-Tos. It seems probable that the Mantodea will soon be regarded as comprising several families.



Figs. 100-105. Mantodea

- 100. Stagmomantis (Rehn and Hebard) Mantidæ.
- 101. Deroplatys (Westwood) Mantidæ.
- 102. Angela (Saussure and Pictet) Mantidæ.
- 103. Hoplocorypha (Rehn) Mantidæ.
- 104. Toxodera. a, hind femur of same (Westwood) Mantidæ.
- 105. Oligonyx (Saussure and Pictet) Mantidæ.

	Hind and middle tibiæ with one or two carinæ above, except rarely in the male; prothorax long. (Fig. 111). (Oxyópsis, Pseudóxyops, Vàtes, Stagmatóptera, neotrop.; Stenovàtes, Pòpa,
	ethiop.; Æthalóchroa, Ceratocrània, indomal.)VATÌNÆ
8.	Front femora externally with a series of five to seven spines9
٥.	Front femora with four spines in the external row
9.	Front femora with the first discoidal spine longer than the second;
•	very long bodied, slender species. (Schizocéphala, indomal.;
	Euchomenélla, Ind.; Agrionópsis, ethiop.; Ángela (Fig. 102),
	neotrop.). (ANGELÎNÆ)SCHIZOCEPHALÎNÆ
	Front femora with the first discoidal spine shorter than the
	second
10.	Cerci flattened, dilated apically and more or less leaf-like; very
	elongate, slender species, with the front femora thin. (Fig. 104).
	(Toxódera, Eùthyphleps, Loxomántis, indomal.; Calamo-
	théspis, Belomántis, ethiop.; Stenophýlla, neotrop.).
	TOXODERÌNÆ
	Cerci conical or compressed, not leaf-like
11.	Front femora with three discoidal spines
	Front femora with four discoidal spines14
12.	Front tibiæ with more than eleven spines in the outer row; front
	femora with five external and three discoidal spines; small
	species. (Acontísta, Tithròne, Astóllia, Callíbia, neotrop.).
	ACONTISTINÆ
	Front tibiæ with from eight to eleven spines in the outer row13
13.	Frontal shield transverse; pronotum at least as long as the front
	coxæ, ovally dilated or sometimes with the sides more or less
	parallel. (Brunnèria, Macromántis, Photina, Orthoder-
	élla, neotrop.; Îris, widespr.) PHOTINÎNÆ
	Frontal shield subquadrate. (See couplet 23).
11	EREMIAPHILÎNÆ, part Supra-anal plate very long, lanceolate; hind femora and tibiæ
14.	with a few small spines beneath; body slender; pronotum as
	long as the front coxe. (Bolivària, Geomántis, Rivetina
	(=Fischeria), palæarc.; Deiphobe, indoaustr.; Ischnomántis,
	Omomántis, ethiop.). (FISCHERIÌNÆ) RIVETÌNÆ
	Supra-anal plate short
15.	Front tibiæ with six to eleven spines in the outer row
10.	Front tibiæ with more than eleven spines in the outer row
16.	Front femora very broad, elliptical, with the upper margin strongly
10.	arcuate; vertex conically elevated with a tubercle on each side
	next to the eye; pronotum shorter than the front coxe or barely
	as long, with conical tubercles on the disk; small species. (Oxy-
	pilus, Euoxypilus, ethiop.; Pachymántis, Ceratomántis,
	Prandavynilus indomal) OYVPILINA

	Front femora narrower, more or less triangular; pronotum with rounded lateral dilatations that give it a trifoliate appearance;
	small species. (Dystácta, Gonypetélla, Achlæna, Telo-
	mántis, ethiop.)
17.	Eyes acuminate, or the hind femora lobate; moderate-sized species
	colored like dry leaves. (Metília, Decímia, Acánthops, Epa-
	phrodita, neotrop.; Phyllocrània, ethiop.; Parablépharis,
	mal.) EPAPHRODITÎNÆ
	Eyes rounded, or the hind femora simple
18.	Frontal shield transverse (see couplet 13) PHOTININÆ, part
	Frontal shield subquadrate (see couplet 23).
	EREMIAPHILÎNÆ, part
19.	Front coxe with a minute apical lobe at the front margin20
	Front coxæ without apical lobe
20.	Front femora triangular, broad, not more than three times as long
	as wide; small species, the female often wingless. (Pseudomi-
	ópteryx, Mantíllica, Diabántia, Miobántia, neotrop.).
	PSEUDOMIOPTER Ý GÌNÆ
	Front femora slender, more than three times as long as broad; pro-
	notum long and narrow, not noticeably dilated; small slender
	species. (Fig. 103). (Musoniélla, Musònia, Diamusònia,
	Théspis, neotrop.; Hoplocórypha, ethiop.)THESPÌNÆ
21.	Front femora with the two intermediate spines of the outer row
	longer than the others; pronotum longer than the front coxæ
	fore wings long in the male, short in the female; body usually
	very slender. (See couplet 9) SCHIZOCEPHALINÆ, part
	Front femora with the two intermediate spines of the outer row
	not longer than the others
22.	Front tibiæ with four or five spines in the outer row
	Front tibiæ with more than five spines in the outer row25
23.	Front femora with the groove that receives the tibial claw remote
	from the base24
	Front femora with the claw groove close to the base. ( <b>Eremi-</b>
	áphila, palæarc.: Tarachòdes, Galépsus, Tarachódula,
	ethiop.: Parepiscòpus, Didymocórypha, indomal.). (ORTHO-
	$DERINm{\mathscr{Z}},TARACHODINm{\mathscr{Z}})\ldots\ldots$ EREMIAPHILIN $m{\mathscr{Z}}$
24.	Vertex not produced (see couplet 20) <b>THESPINÆ</b> , part
	Vertex produced into an elongate, triangular process. (Pyrgo-
	mántis, ethiop.) (see couplet 23) EREMIAPHILÌNÆ, part
25.	Lateral margins of pronotum parallel or divergent in front (see
	couplet 23). (Humbertiélla, Theopómpula, indomal.; Elæa,
	Theopómpa, ethiop.) EREMIAPHILÌNÆ, part
	Lateral margins of pronotum more or less convergent in front 26
26.	Front tibiæ with the spines in the outer row erect and remote from
	one another

I	Front tibiæ with the spines in the outer row decumbent and very close together
27. I	close together
	106
108	110
	Figs. 106-112. Mantodea
107. 108. 109. 110.	Empusa, front leg (Westwood) Mantidæ. Compsothespis, front leg (Westwood) Mantidæ. Metallyticus, front leg (Westwood) Mantidæ. Chæteessa, front leg (Westwood) Mantidæ. Mantoida, front leg (Westwood) Mantidæ. Vates, hind leg (Saussure and Pictet) Mantidæ. Vates, base of antenna of male (Saussure and Pictet) Mantidæ.
	Internal apical lobes of the front coxæ divergent, not dilated into a small lobe at the tip; small species. (Tarachìna, Bólbula, Enicophlæbia, ethiop.; Bólbe, Ciulfina, Austr.; Haplopèza, Iridópteryx, Fulciniélla, Eomántis, indomal.).  IRIDOPTERYGINÆ Internal apical lobes of the front coxæ contiguous. (See couplet 14)
	<b>RIVETINÆ</b> , part Front femora with a well marked fovea or pit between the first
	and second spines of the outer row

30.	Front femora with the discoidal spines forming a sinuous line;
	species of moderate size, with short, stout body, the pronotum
	broad and depressed. (Gonatista, Liturgusa, neotrop.; Dac-
	tylopteryx, ethiop.; Gonatistella, Austr.).
	LITURGUSÎNÆ
01	Front femora with the discoidal spines placed in a straight line31
31.	Spines of outer row on front femora very long and curved32
	Spines of outer row on front femora shorter, straight; slender-
	bodied species, with the wings well developed in both sexes. (Árria, malay.; Sibýlla, Presibýlla, ethiop.). SIBYLLÌNÆ
32.	Pronotum with two tubercles near the base of its posterior por-
θ⊿.	tion
	Pronotum simple, without tubercles; slender or very slender
	species, with well developed wings in both sexes. (Caliris, Ind.;
	Leptomántis, malay.; Deromántis, ethiop.).
	CALIRIDINÆ
33.	Hind femora with a small lobe or tooth below near tip; more or less
00.	stout and short-bodied species with strongly prominent eyes.
	(Majánga, ethiop.; Majangélla, malay.)MAJANGÌNÆ
	Hind femora simple, without such projection; body slender, flat-
	tened, the eyes moderately prominent. (Melliera, neotrop.;
	Mellieriélla, Austr.)
34.	Vertex produced into an elongate process (see couplet 6).
	OXYPILÎNÆ, part
	Vertex not elongated or produced
35.	Four posterior tibiæ or their femora minutely spinulose below36
	Four posterior tibiæ and femora with the lower edge smooth37
36.	Frontal shield transverse; discoidal portion of hind wings not
	banded. (See couplet 14) FISCHERIINÆ, part
	Frontal shield scarcely broader than high; discoidal portion of
	hind wings with black cross-bands; pronotum at least as long as
	the front coxæ. A cosmopolitan group. (Fig. 100). (Mántis,
	palæarc., Austr.; Stagmomántis, Am. (S. carolina, Carolina
	mantis); Auromántis, Uromántis, neotrop.; Calidomántis,
	Sphodromántis, ethiop.; Tenódera (T. sinénsis, Chinese
	mantis), Polyspolòta, widespr.; Sphodrópoda, Austr.).
0. <del>-</del>	MANTÎNÆ
37.	Lateral margins of pronotum strongly expanded, leaf-like 38
90	Lateral margins of pronotum not expanded
38.	
	Posterior femora lobed; large, brown, leaf-like species with con-
	spicuous foliaceous expansions at the sides of the pronotum
	and at the tips of the four posterior femora. (Fig. 101). ( <b>Deróplatys</b> , E. As.; <b>Brancsíkia</b> , Madagasc.).
	(Deropiatys, E. As.; Brancsikia, Madagasc.).  DEROPLATINÆ
	DEROFLATINA

39.	Hind metatarsi simple, not carinate; large species with the pronotum bearing leaf-like expansions at the sides. (Chœradòdis,
	neotrop., indomal.)
	Hind metatarsi carinate. (See couplet 36) MANTÎNÆ, part
40.	Eyes produced laterally to form a conical, spiniform process; body
10.	very slender, filiform. (Oxyothéspis, ethiop., As.; Hetero-
	chætula, malay.)
	Eyes rounded laterally
41.	Fore wings and antennæ of the male ciliate
41.	
42.	Fore wings and antennæ of the male not ciliate
44.	Pronotum linear or extremely slender; small species; the female
	apterous. (Miópteryx, Promiópteryx, Chloromiópteryx, neotrop.)
	Description of the second of t
49	Pronotum more or less expanded
43.	Pronotum almost trilobed, the angulations of the expansions some-
	what acuminate; fore wings of male broad (see couplet 16).
	DYSTACTÎNÆ, part
	Pronotum more or less elliptical, the angulations of the expansions
	rounded; elytra narrow. (See couplet 45) AMELÎNÆ, part
44.	Pronotum shorter than the front coxæ45
	Pronotum as long as or longer than the front coxæ46
45.	Hind wings not colored; small species. (Ameles, Pseudoyersínia
	palæarc.; Yersínia, Litaneùtria, neotrop.; Amántis, Myr-
	cinus, Gonýpeta, indomal.; Metentélla, Ligària, ethiop.).
	AMELÎNÆ
	Hind wings brightly colored, otherwise similar to the Amelinæ.
	(Compsomántis, Opsomántis, malay.).
	COMPSOMANTINÆ
46.	Front tibia with the sixth spine from the apex in the outer row
	longer than the fifth. (See couplet 32) CALIRIDINÆ, part
	Front tibia with the sixth spine not longer than the fifth47
47.	Eyes produced laterally to form a conical, spiniform process.
	Eyes produced laterally to form a conical, spiniform process. (See couplet 40)
	Eyes not thus spined48
48.	Front femora with the first spine in the discoidal row not shorter
	than the second; large species of elongate form; wings large in
	male, short in female. (Archimántis, Rheomántis, Pseudo-
	mántis, Austr.) ARCHIMANTÌNÆ
	Front femora with the first discoidal spine shorter than the
	second
49.	Supra-anal plate very long, lanceolate; large species with the pro-
	notum much longer than the front coxe. (Solýgia, ethiop.).
	SOLYGIÌNÆ
	Supra-anal plate short. (See couplet 36) MANTINÆ, part
50.	Pronotum slender, as long as the front coxæ
55.	- 10110 tolar bleffeld, as folig as the front Coate

51. Margins of pronotum broadly laminate (see couplet 17).

EPAPHRODITINÆ. part

Margins of pronotum not broadly laminate; species of moderate size, with slender pronotum which is at least as long as the front coxe. (Anaxárcha, Acromántis, Citharomántis, indomal.; Theomántis, Sigérpes, Anasigérpes, ethiop.).

ACROMANTÎNÆ

#### LITERATURE ON MANTODEA

Giglio-Tos, E. Mantidi Esotici, I-XII. Boll. Soc. Entom. Italiana (1911–17). (Various parts.)

Saggio di una nuova classificazione dei Mantidi. Boll. Soc. Entom. Italiana, 49, pp. 50–87 (1919).

Mantidæ, Perlamantinæ. Gen. Insectorum, fasc. 144, 13 pp. (1913).

Mantidæ, Eremiaphilinæ. Gen. Insectorum, fasc. 177, 36 pp. (1921). Mantidæ. Das Tierreich, Lief. 50, 707 pp. (1927).

Kirby, W. F. A Synonymic Catalogue of the Orthoptera. Vol. 1. British Mus. Nat. Hist., London (1904).

Rehn, J. A. G. Mantidæ. Vatinæ. Gen. Insectorum, fasc. 119, 28 pp. (1911). Saussure, H. Essai d'un Système des Mantides. Mittheil. schweiz. Entom.

Ges., 3, pp. 49–73 (1869).

Tindale, N. B. Review of Australian Mantidæ. Rec. South Australian Mus., 2, pp. 425-457 (1923).

Westwood, J. O. Revisio Mantidarum. London (1889).

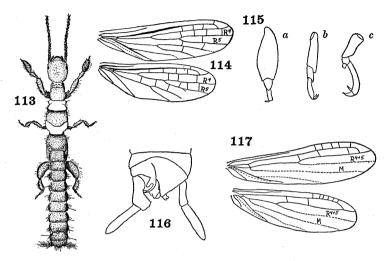
#### ORDER EMBIÒDEA

### (EMBIIDÌ NA, EMBIÒIDEA, EMBIÓPTERA, OLIGONEÙ RA, ÆTIÓPTERA)

Elongate, slender, feeble insects of small or moderate size. Head large, elongate; eyes small or of moderate size; ocelli absent; antennæ slender, many-jointed (16–32). Thorax elongate; prothorax large, free; meso- and metathorax each about as large as the prothorax; males usually winged, females always wingless. Wings similar, pubescent, elongate, the media and cubitus much reduced; no anal fan or lobe. Wings lying flat on the abdomen when at rest. Abdomen ten-segmented; long, narrow and with parallel sides; tip of abdomen and cerci usually strongly asymmetrical in the male; cerci two-jointed.

Tarsi three-jointed; first joint of front tarsi swollen, containing glands for spinning a silken web in which the insects live.

Metamorphosis gradual, incomplete. A small group, restricted to the tropics and subtropics.



Figs. 113-117. Embiodea

- 113. Oligotoma, female (Melander) Oligotomidæ.
- 114. **Donaconethis,** wings (Enderlein) Embiidæ.
- 115. **Rhagadochir**. a, front tarsus; b, middle tarsus; c, hind tarsus (Silvestri) Embiidæ.
- 116. Rhagadochir, apex of abdomen of male (Enderlein) Embiidæ.
- 117. Oligotoma, wings (Enderlein) Oligotomidæ.
- Posterior branch of the radial sector forked in both wings, or at least in the hind wing, i.e. both R<sub>4</sub> and R<sub>5</sub> developed (Figs. 114, 115, 116); sternite of first abdominal segment of female large. (Émbia, Haploémbia, widespr.; Rhagádochir, ethiop., neotrop.; Antipalùria, Clothòda, neotrop.; Donaconèthis, ethiop.). (including OLÝNTHIDÆ)......EMBÌIDÆ
  - Posterior branch of the radial sector simple in both wings, i.e. R<sub>4</sub> and R<sub>5</sub> fused (Fig. 117); sternite of first abdominal segment of female greatly reduced in size. (Fig. 113). (**Oligótoma**, cosmop.; **Teratémbia**, neotrop.). (Including *TERATEMBIIDÆ*).

**OLIGOTÓMIDÆ** 

#### LITERATURE ON EMBIODEA

ENDERLEIN, G. Embiidinen. Coll. Zool. Selys-Longchamps, fasc. 3 (1912). Hagen, H. A Monograph of the Embidina. Canadian Entom., 17, pp. 142–155 (1885).

Krauss, H. A. Monographie der Embien. Zoologica, Heft 60, 78 pp. (1911). Navás, L. Embiópteros de la America meridional. Brotéria, **16**, pp. 85–110(1919). Tillyard, R. J. The Embioptera or Web-spinners of Western Australia.

Journ. Roy. Soc. West Australia, 9, pp. 61–68 (1923).

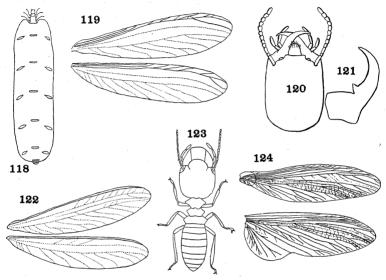
Verhoeff, K. W. Zur vergleichende Morphologie und Systematik der Embiiden. Act. Acad. Leopold. Carol., 82, pp. 145–204 (1904).

#### ORDER ISÓPTERA

Small or medium-sized, elongate, feeble insects living in large colonies and occurring as winged sexual individuals and wingless workers and soldiers; usually with weak chitinization, especially in the sterile castes which are soft-bodied and white, except for the heavily chitinized head. Head large, free, rather vertical; eyes and two ocelli usually present in the winged forms, or absent in the workers. Mandibles strong, often very large; antennæ filamentous, more or less moniliform. Prothorax free, but much smaller than the head. Legs similar, formed for running or walking; tarsi four- or rarely five-jointed, with well developed claws. Wings similar, long and narrow, deciduous soon after maturity at a preformed transverse suture near the base; long and narrow, superimposed over the abdomen, the hind pair very rarely with an anal area; venation almost always much reduced and without crossveins. Cerci short, one- to three-, rarely eight-jointed. Metamorphosis very incomplete. White ants, Termites.

#### Males and Females

- 3. Ocelli present; empodium present between the tarsal claws; pronotum large, wider than the head; wing stubs of the front wing much larger than those of the hind wing and overlapping them;



Figs. 118-124. Isoptera

- 118. **Termes,** queen (Hegh) Termitidæ.
- 119. Calotermes, wings (Calotermitidæ).
- 120. Eutermes, head of worker. Termitidæ.
- 121. Eutermes, mandible of worker. Termitidæ.
- 122. Reticulitermes, wings (Banks) Rhinotermitidæ.
- 123. Termes, worker (Hegh) Termitidæ.
- 124. Mastotermes, wings (Desneux) Mastotermitidæ.

antennæ with 13 to 23 joints; tibiæ with three apical spines. (Calotérmes (Fig. 119), Neotérmes, Cryptotérmes, cosmop.). (PROTERMÍTIDÆ, part)......CALOTERMÍTIDÆ

Ocelli absent; empodium absent; pronotum narrower than the head; wing stubs of fore wings shorter, not overlapping those of the hind pair; antennæ with 23 to 27 joints; usually more than three (3-5) apical spines on the tibiæ. (**Hodotérmes**, ethiop.;

- Anacanthotérmes, palæarc., Indo-malay.; Termópsis, nearc.). (PROTERMÍTIDÆ, part).....HODOTERMÍTIDÆ
- 4. Wings transparent, without hairs, their margins not ciliate; venation in posterior part of wing indistinct, more or less reticulate; front wing stub large. (Rhinotérmes, neotrop.; Coptotérmes, Schedorhinotérmes, widespr.; Reticulitérmes (Fig. 122), holarc., Indo-malay.). (MESOTERMÍTIDÆ).

#### RHINOTERMÍTIDÆ

### Soldiers

#### LITERATURE ON ISOPTERA

BANKS, N. Antillean Isoptera. Bull. Mus. Comp. Zoöl., Harvard, 62, pp. 475–489 (1919).

A Revision of the Nearctic Termites. Bull. U. S. Nat. Mus., No. 108 (1920).

Desneux, J. Termitidæ. Gen. Insectorum, fasc. 25, 52 pp. (1905).

EMERSON, A. E. Termites of Kartabo, British Guiana. Zoölogica, New York, **6**, pp. 291–459 (1925).

Froggatt, W. W. Australian Termitidæ. Proc. Linn. Soc. New South Wales, 10–12 (1895–97) (several parts).

Fuller, C. The Termites of South Africa. South African Journ. Nat. Hist., 3, pp. 14-52; 70-130 (1921-22).

Hegh, E. Les Termites. Brussels, 756 pp. (1922). Republished with additions in Bull. Agric. Congo Belge, Brussels, 11–14 (1920–23).

Holmgren, N. Termitenstudien. I-IV. König. Svensk. Vet. Akad. Handl., **44**, **46**, **48**, **50** (1909–13).

Hozawa, S. Revision of the Japanese Termites. Journ. Coll. Agric. Tokyo, **35**, pt. 7, 161 pp. (1915).

Silvestri, F. Termiti e termitofili dell' America meridionale. Redia, 1, pp. 1–234 (1903).

Termiti e termitofili dell' Eritrea. Redia, 3, pp. 341-359 (1906).

Termitidi e termitofili dell' Africa occidentale. Boll. Lab. Zool. Agrar. Portici, 9, 12 and 14 (1914–20).

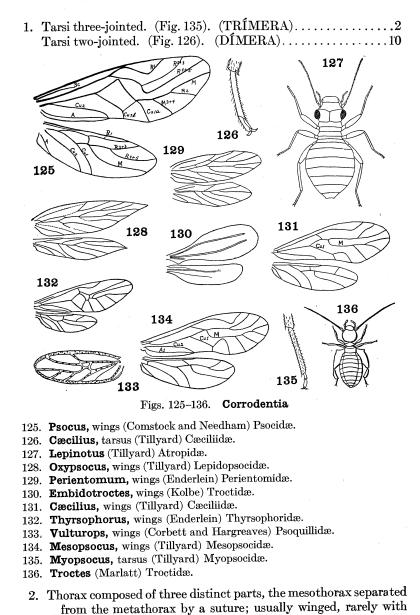
SJÖSTEDT, Y. Monographie der Termiten Afrikas. Svensk. Vet. Akad. Handl., 34 and 38 (1900–04).

Revision der Termiten Afrikas. K. Svenska Vet. Akad. Handl., (3), 3, 419 pp. (1926).

### ORDER CORRODÉNTIA

### (PSOCÓPTERA, COPEÓGNATHA)

Usually small or minute insects, rarely of moderate size, with short, soft body and usually winged. Head large, free, vertical, with a strong Y-shaped suture above; eyes large and prominent, except in a few wingless forms; three ocelli generally present; antennæ long and slender, filiform or bristle-like, many (13-50) jointed; mandibles strong, toothed and with a grinding surface. Prothorax almost always very small: mesothorax and metathorax usually separated, although rarely completely fused. Wings usually ample, sometimes much reduced or entirely absent; when at rest generally held in a sloping position, folded backwards over the body; fore pair larger than the hind pair, sometimes scaly or hairy; venation reduced, with few or no crossveins; one or several of the veins frequently strongly curved. Abdomen usually short, with nine or ten segments; cerci absent. Legs similar, fitted for running, the coxæ close together; tarsi two- or three-jointed, the first joint very long; two tarsal claws. Metamorphosis incomplete, the nymph similar to the adult form; terrestrial in all stages. Psocids, Book lice, Bark lice.



	Thorax composed of two parts, the meso- and metathorax fused
	and without suture between them; wings usually entirely absent,
	if present without forked veins; second joint of palpi without
	clubbed sense organs. (Figs. 130, 136). ( <b>Tróctes</b> (= Lipóscelis)
	(T. divinatòrius, Book louse, Cereal psocid), cosmop.; Tropùsia,
	widespr.; Pachytróctes, palæarc.; Embidopsòcus, neotrop.;
	Embidotróctes, ethiop.). (LIPOSCÉLIDÆ) TRÓCTIDÆ
3.	Wings present; prothorax much smaller than the mesothorax4
	Fore wings absent or very small and without venation; hind wings
	entirely absent; prothorax larger than the mesothorax. (Atro-
	pos (= Trògium) (A. pulsatòria, Death watch), widespr.; Lepi-
	nòtus (Fig. 127), Lepidílla, Leprólepis, Hypéretes). (Includ-
	ing LEPIDÍLLIDÆ, TROGÌIDÆ) ATRÓPIDÆ
4.	Wings fully formed, with complete venation
	Venation of wings incomplete, the fore wings oval or rounded and
	much thickened; the veins usually very broad; hind wings re-
	duced or absent; without scales. (Psoquilla, holarc.; Psoci-
_	nélla, Vúlturops, Am.) (Fig. 133) PSOQUÍLLIDÆ
5.	8
	or closely approaching each other at apex (Fig. 134)
	Second branch of cubitus and first anal vein in fore wing divergent toward apex, or at least not approaching each other; body and
	wings clothed with hairs or scales; wings more or less pointed;
	antennæ with more than thirteen joints (Fig. 128)
6.	Hind wings with a very narrow closed cell at the base between the
0.	media and cubitus; wing scales of symmetrical form, similarly
	curved on their two sides; antennæ with 20 to 25 joints. (Fig.
	129). (Periéntomum, Ind.) PERIENTÓMIDÆ
	Hind wings without a closed cell; wing scales usually asymmetri-
	cal; antennæ with 26 to 47 joints. (Fig. 128). (Lepidopsòcus,
	Echinopsòcus, Echmépteryx, Oxypsòcus). (EMPHERÌ-
	IDÆ) LEPIDOPSÓCIDÆ
7.	
	Antennæ with 22 to 25 joints; body and wings not scaled; media
	two- or three-branched; prothorax visible from above. (Phyl-
	lipsòcus, Psylloneùra, Deipnopsòcus, Rhyopsòcus).
_	PHYLLIPSÓCIDÆ
8.	
	Body and wings scaled; two anal veins in the fore wing. (Am-
	phiéntomum, ethiop., ind.; Tineomórpha, ind.; Stigmató-
	pathus, Cymatopsòcus, indomal.) AMPHIENTÓMIDÆ

- 12. Second branch of radial sector (R<sub>4+5</sub>) fused with the media or connected with it by a crossvein (Fig. 132); third and fourth antennal joints lengthened, thicker and more densely hairy than the joints beyond; large species. (Thyrsóphorus, Dictyopsòcus, Ischnópteryx, neotrop.) . . . . THYRSOPHÓRIDÆ

#### LITERATURE ON CORRODENTIA

Enderlein, G. Die Psocidenfauna Perus. Zool. Jahrb., Abth. f. Syst., 14, pp. 133–160 (1900).

Morphologie, Gruppierung und systematische Stellung der Corrodentien. Zool. Anz., **26** (1903).

Die Copeognathen des indoaustralischen Faunengebietes. Ann. Mus. Hist. Nat. Hungarici, 1, pp. 179–344 (1903).

Morphologisches System und Biologie der Atropinen und Troctinen. Jægerskiöld Zool. Exped. Egypt, No. 18, 58 pp. (1905).

Die australischen Copeognathen. Zool. Jahrb., Abth. f. Syst., 23, pp. 401–412 (1906).

Die Copeognathenfauna Japans. Zool. Jahrb., Abth. f. Syst., 23, pp. 243–256 (1906).

The Scaly-winged Copeognatha. Spolia Zeylandica,  ${\bf 4}$ , pp. 39–122 (1906).

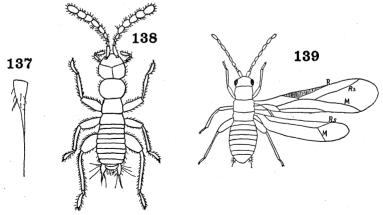
Copeognatha. In Tierwelt Mitteleuropas, 4, Lief. 2, pp. VII 1–16 (1928).

Kolbe, H. Monographie der deutschen Psociden. Jahresber. zool. Sect. Westf. Ver., 8, 1879–80, pp. 74–142 (1880).

Tillyard, R. J. Monograph of Psocoptera or Copeographa of New Zealand. Trans. New Zealand Inst., **54**, pp. 170–196 (1923).

#### ORDER ZORÁPTERA

Minute, terrestrial species of social habits, living in colonies; dimorphic, both sexes represented by winged and wingless individuals. Body depressed. Head free, somewhat inclined. Antennæ moniliform



Figs. 137-139. Zoraptera

- 137. Zorotypus, cercus (Karny) Zorotypidæ.
- 138. Zorotypus (Silvestri) Zorotypidæ.
- 139. Zorotypus, winged form (Caudell) Zorotypidæ.

or filiform, nine-jointed, the second and often also the third joint smaller than the others. Mandibles well developed, toothed and fitted for biting. Eyes and ocelli present in the winged form, absent in the wingless one. Thorax large, but no wider than the head; prothorax free, not concealing the head nor expanded laterally; meso- and metathorax distinctly separated. Abdomen elongate-oval, never much

longer than the thorax, with ten strongly transverse segments; cerci short, oval, one-jointed, with a bristle-like appendage at tip. Legs similar, formed for running; hind femora stout, armed beneath with several spiny bristles; tarsi two-jointed, the first joint short. Metamorphosis gradual, the nymph similar to the adult, especially to the wingless form. Winged form with narrow, membranous wings, the fore pair larger; venation greatly reduced; wings commonly falling off after maturity, leaving a stub attached to the body, but not separating at a preformed suture; body of winged form more heavily chit-inized than in the wingless one.

#### LITERATURE ON ZORAPTERA

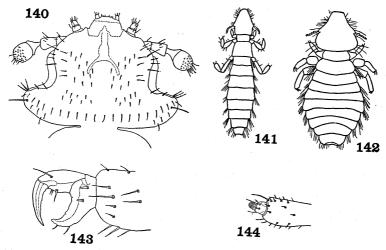
CAUDELL, A. N. Zoraptera not an Apterous Order. Proc. Entom. Soc. Washington, 22, pp. 84–97 (1920).

Silvestri, F. Descrizione di un nuova ordine di insetti. Boll. Zool. Gen. Agrar. Portici, 7, pp. 193–209 (1913).

### ORDER MALLÓPHAGA

(LIPÓPTERA)

Small wingless insects averaging two mm. and very rarely over five mm. in length. Body oval, or elongate, very strongly flattened; usually strongly chitinized and generally with a conspicuous color pattern of pale or yellowish markings contrasting with spots or bands of dark brown or black. Mouth inferior, mandibles strong; antennæ three- to five-jointed; prothorax free, rarely fused with the mesothorax; legs short, no cerci. Metamorphosis very incomplete. External parasites of birds, more rarely of mammals, during entire life, feeding on feathers, fur or skin. Bird lice, Biting lice.



Figs. 140-144. Mallophaga

- 140. Gyropus, head (Ewing) Gyropidæ.
- 141. Lipeurus (Paine) Philopteridæ.
- 142. Philopterus (Paine) Philopteridæ.
- 143. Gyropus, tip of front leg (Ewing) Gyropidæ.
- 144. **Gliricola,** tip of front leg (Ewing) Gyropidæ.

Antennæ not strongly clubbed, four-jointed......4

Prothorax not appearing like the metathorax inverted......5

DIVERS AND MEDIANDER. CHASSIFICATION OF INSECTS
5. Head evenly expanded behind, broadly triangular and strongly enlarged on the temples. (Ménopon, widespr. (M. gallinæ, Chicken louse); Colpocéphalum, Myrsídea, Trinòton, widespr.; on birds)
Head not evenly expanded and broadly triangular, not enlarged on the temples
6. Sides of head with a strong lateral swelling in front of the eye spiracles on abdominal segments three to eight. (Læmobóthrion, holarc., ethiop., neotrop.; on birds).
LÆMOBOTHRÌIDÆ
Head with the sides straight or concave; spiracles on abdominal segments two to seven. (Ricinus (=Leiótheum), widespr.
Trochilœcètes, Am.; on birds). (LEIOTHÈIDÆ).
RICĪNIDÆ

7. Tarsi with one claw; antennæ three-jointed; species infesting mammals. (Trichodéctes, Eutrichophilus, widespr.).

#### TRICHODÉCTIDÆ

- 8. Species infesting birds......9 Species infesting mammals; head heavily chitinized at the sides and armed with strong hooks; last joint of antennæ somewhat swollen or clubbed. (Trichophilópterus).

### TRICHOPHILOPTÉRIDÆ

9. Meso- and metathorax clearly separated by a suture; eyes deeply constricted. (Nesiotinus, Kerguelen Is., on penguins).

Meso- and metathorax not separated by a distinct suture. (Goniòdes, Goniocòtes, Lipeùrus (Fig. 141), Philópterus (Fig. 142), Degeeriélla, Esthiópterum, widespr.; Aptericola, N. Zeal.; on birds)..... PHILOPTÉRIDÆ

#### LITERATURE ON MALLOPHAGA

Denny, H. Monographia Anoplurorum Brittanniæ. London (1842).

Enderlein, G. Mallophaga. In Tierwelt Mitteleuropas, 4, Lief. 2, pp. VII 17-24 (1928).

EWING, H. E. Taxonomy, Biology and Distribution of Gyropidæ. Proc. U. S. Nat. Mus., **63**, art. 20, pp. 1–42 (1924).

Ferris, G. F. The Mallophagan Family Trimenoponide. Parasitology, 14, pp. 75-86 (1922).

The Mallophagan Family Menoponidæ. I. Parasitology, 16, pp. 55-66 (1924).

FULMER, L. Die Mallophagen. Mitt. naturw. Ver. Univ. Wien, 5, pp. 1-50 (1907).

GIEBEL, C. G. A. Insecta Epizoa. 308 pp. Leipzig (1874).

HARRISON, L. The Genera and Species of Mallophaga. Parasitology, 9, pp. 1–156 (1916).

Kellogg, V. L. Mallophaga. Gen. Insectorum, fasc. 66, 87 pp. (1908).

Kellogg, V. L. and Ferris, G. F. Anoplura and Mallophaga of North American Mammals. Leland Stanford Jr. Univ. Pub. (1915).

MJÖBERG, E. Studien über Mallophagen und Anopluren. Ark. f. Zool., 6, No. 13, 297 pp. (1910).

A New Family and Three New Genera of Mallophaga. Entom. Tidskr., 40, pp. 93-96 (1919).

TASCHENBERG, O. Die Mallophagen. Halle (1882).

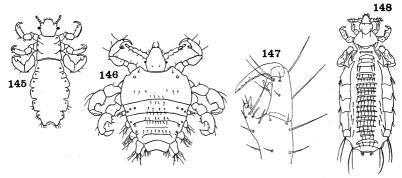
#### ORDER ANOPLÙRA

# (SIPHUNCULÀTA, PSEUDORH Y NCHÒTA, PARASÌTA, PHTHIRÁPTERA, ELLIPÓPTERA)

Small, more or less flattened, wingless ectoparasites of mammals. Head free, horizontal; eyes reduced or absent; mouth anterior, comprising an unjointed, fleshy beak; antennæ short, simple. Thoracic segments fused. Legs very stout; tarsi single-jointed, forming a claw at the end of the tibia. No cerci. Metamorphosis very slight. True lice, Sucking lice, Cooties.

- - Body thickly clothed with stout, thorn-like bristles or with spines and scales; body thick and stout; mesothorax and metathorax each with a pair of spiracles as well as abdominal segments two to eight; eyes absent; antennæ four- or five-jointed; living exclusively on marine mammals. (Echinophthírius, Lepidophthírius, Antarctophthírius) .. ECHINOPHTHIRÌIDÆ
- - Head tubularly produced anteriorly to form a beak longer than the remainder of the head; tibiæ without such a process opposing the tarsus. (**Hæmatomỳzus**, on elephants, Afr., E. Ind.).

HÆMATOM ÝZIDÆ



Figs. 145-148. Anoplura

- 145. Pediculus (Patton and Cragg) Pediculidæ.
- 146. Phthirius (Patton and Cragg) Phthiriidæ.
- 147. **Pediculus,** tip of leg (Ewing) Pediculidæ.
- 148. **Hoplopleura** (Ferris) Hæmatopinidæ.

#### LITERATURE ON ANOPLURA

Dalla Torre, K. W. Anoplura. Gen. Insectorum, fasc. 81, 22 pp. (1908).

Denny, H. Monographia Anoplurorum Britanniæ. London (1842).

Enderlein, G. Läuse-Studien. Zool. Anz., 27, pp. 220-223 (1904); 28, pp. 626-638 (1905).

EWING, H. E. Revision of the American Lice of the Genus Pediculus. Proc. U. S. Nat. Mus., 68, Art. 19, pp. 1–30 (1926).

Ferris, G. F. A Catalogue and Host List of the Anoplura. Proc. California Acad. Sci., (4) 6, pp. 129–213 (1916).

Sucking Lice, a Monograph. Pt. I., Leland Stanford Jr. Univ. Pub. Univ. Ser. (1919); Pt. II., Stanford Univ. Pub. Univ. Ser. Biol. Sci., 2, No. 2 (1921); Pt. III., No. 3 (1922); Pt. IV., No. 14 (1923).

GIEBEL, C. G. A. Insecta Epizoa. Leipzig, 308 pp. (1874).

Kellogg, V. L. and Ferris, G. F. Anoplura and Mallophaga of North American Mammals. Leland Stanford Jr. Univ. Pub. Univ. Ser. (1915).

Nuttall, G. H. F. Classification of Anoplura. Parasitology, 11, pp. 329–346 (1919).

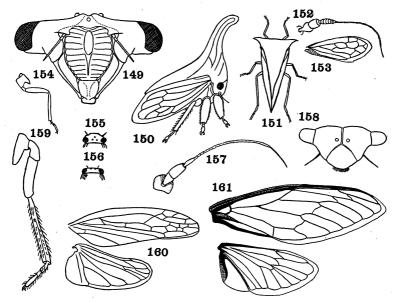
Piaget, E. Les Pédiculines. I., 714 pp. (1880); II., 162 pp. (1885) Leiden.

#### ORDER HOMÓPTERA

### (RHYNCHÒTA, part)

An assemblage of very diverse insects, difficult to define in a general way; usually of moderate or small size, rarely large; in the active forms four wings are present in both sexes; in the scale insects only the males are winged, and they have the hind wings absent; wings usually sloping over the sides of the body; fore wings never modified into a heavy basal and thinner apical portion; mouthparts forming a jointed beak, inserted at hind edge of the head and extending between the front coxe, the basal joints very short, rarely the beak is absent in the males. Beak formed of the stylet-shaped mandibles and maxillæ which are enclosed in the labium. Cerci wanting. Metamorphosis usually incomplete, sometimes complete in the male or at least with a pupal stage in the male scale insects, rarely so in the female; all the species vegetarian.

Beak appearing to arise between the front coxæ, rarely absent in male coccids and some aphids; tarsi two- or one-jointed; an-



Figs. 149-161. Homoptera

- 149. Cicada, front view of head (Berlese) Cicadidæ.
- 150. Entylia (Branch) Membracidæ.
- 151. Ceresa (Marlatt) Membracidæ.
- 152. Ceresa, antenna (Marlatt) Membracidæ.
- 153. Ceresa, fore wing (Marlatt) Membracidæ.
- 154. Cicada, hind leg (Kolbe) Cicadidæ.
- 155. Cicada, dorsal view of head (Maxwell-Lefroy) Cicadidæ.
- 156. Leaf hopper, dorsal view of head (Maxwell-Lefroy) Jassidæ.
- 157. Entylia, antenna (Branch) Membracidæ.
- 158. Entylia, front view of head (Branch) Membracidæ.
- 159. Entylia, hind leg (Branch) Membracidæ.
- 160. Gypona, wings (Metcalf) Gyponidæ.
- 161. Cicada, wings. Cicadidæ.

- Ocelli (rarely absent) placed between the eyes on the vertex, on the front margin of the head, or on the front; middle coxæ short and close together, hind coxæ movable; tegulæ absent; fore wings with the two anal veins more or less parallel, or the second absent.
  - Ocelli (rarely absent) placed beneath or near the eyes, usually in cavities of the cheeks; middle coxæ elongate, widely separate, hind coxæ immovable, fused externally with the metathorax; tegulæ present as a scale between the base of the fore wing and the side angle of the pronotum; fore wings usually with the two anal veins joining apically to form a Y-vein. (Superfamily FULGORÒIDEA. Classification from F. Muir, 1930).....24
- 3. Three ocelli, placed close together on the disk of the vertex; antennæ with short basal joint, terminated by a hair-like process which is divided into about five joints; front femora thickened and generally spined beneath, hind legs not fitted for jumping; empodia absent; male almost always with a sound-producing structure on each side at the base of the abdomen; comparatively large species with entirely membranous wings; nymphs subterranean. Cicadas, "Locusts," Harvest-flies. (Figs. 149, 154, 155, 161). (Cicàda (C. septéndecim, Periodical Cicada or Seventeen-year locust), Tibicen, Platypèdia). CICÁDIDÆ Two ocelli, rarely absent; empodia large; jumping species.....4
- 4. Pronotum not prolonged over the base of the abdomen . . . . . . 5
  Pronotum prolonged backward into a hood or process of variable form, usually much elevated and more or less concealing the scutellum and extending over the abdomen, often the prothorax is grotesquely enlarged and ornamented; head vertical, cheeks not dilated, ocelli located between the eyes, antennæ inserted between and in front of the eyes. Tree hoppers. (Membrācis, neotrop.; Cerèsa, Am. (C. bùbalis, Buffalo tree hopper, Figs. 151, 152, 153); Centrôtus, palæarc., ethiop., indomal.; Enchenôpa, Am.; Entýlia (Figs. 157, 158, 159), Am.; Górgora, ethiop., indomal.; Tricéntrus, indomal., Telamòna, Am.).

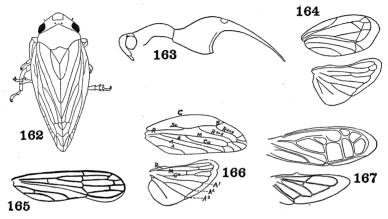
### MEMBRÁCIDÆ

5. Hind coxæ short, conical, not laterally dilated; tibiæ cylindrical, smooth, the hind pair usually armed with one or two stout solid spines and with a cluster of spinules at apex; ocelli placed on the vertex, rarely absent; flagellum composed of a large pear-shaped base and a very slender seta; nymphs usually producing a mass of froth in which they live on the stems of various plants.

- Hind coxæ transverse, reaching the side margins of the sternum; hind tibiæ ridged, with a double series of articulated spines or seriately bristly (hairy in Æthalionidæ); cheeks dilated. Leaf hoppers, Sharp shooters. (Superfamily JASSÒIDEA) . . . . . 9
- 6. Scutellum comparatively small and short (longer than pronotum only in Clastopteridæ); hind wings with outer fork of radius always present (sometimes broken at apex), thus forming a supernumerary (first) apical cell, the cubitus apically forked or simple; fore wings with claval veins when present usually distant and without connecting crossvein
- a. Scutellum not raised apically or with free apical spinous appendage; anterior margin of pronotum strongly extended between eyes; head usually obtuse-angulate; cubitus of hind wing apically forked. (Conmachæròta; Híndola, Enderleinia, malay.; Neuromachæròta, ethiop.). (ENDERLEINIÌNÆ.
- - Form very thick and stout; body of scutellum nearly flat and with dorsal furrow subobsolete; pronotum with lateral angles produced into high, thin, spreading laminæ; anterior margin of pronotum broadly, gently arcuate between eyes.

MAXUDEÌNÆ

7. Pronotal margin between eyes usually straight or slightly arcuate, pronotum commonly strongly enlarged and much broader than



Figs. 162-167. Homoptera

- 162. Spittle insect (Stearns) Cercopidæ.
- 163. Machærota, profile of head, pronotum and scutellum (Baker) Machærotidæ.
- 164. Clastoptera, wings (Metcalf) Clastopteridæ.
- 165. Oncometopia, fore wing (Ball) Cicadellidæ.
- 166. Aphrophora, wings (Metcalf) Cercopidæ.
- 167. Machærota, wings (Baker) Machærotidæ.

8. Fore wings with clavus obliquely truncate at apex; corial appendix, the apical portion of the wing, divided into two very broad subequal portions, these at rest infolded at end of the stout and broad body to overlap; fork of radius in hind wings forming a very short first apical cell considerably before apex; cubitus of hind wings not forked apically; corium with three apical cells

	and two or less subapicals; scutellum longer than pronotum. (Clastóptera (Fig. 164))
	Fore wings with the clavus narrowly acute or subacute apically;
	corial appendix either a narrow continuous membranous mar-
	gin, or wanting, never bent inward beyond the clavus to overlap
	at end of body; corial venation various but never as in the
	Clastopteridæ. (Cercòpa, Ptyelus, Aphróphora, cosmop.;
	Monécphora, widespr.; Phymatostètha, indomal.; Cosmos-
	cárta, palæarc., indomal. (Figs. 162, 166)) CERCÓPIDÆ
9.	
	concealing the head and roundly produced over the base of the
	acute scutellum. (Æthiàlion, neotrop.; Dárthula, India).
	ÆTHIALIÓNIDÆ
	Pronotum not thus modified to cover the head, although some-
	times with lateral protuberances. (JASSIDÆ, in the broad
	sense. Classification from C. F. Baker, Philippine Jour. Sci.,
	1923.)
10.	Upper part of front strongly raised and produced, its posterior
	portion forming a large part of the superior surface of the head
	(crown); the true vertex confined to basal portion of crown,
	the ocelli thus on posterior disk of crown, usually remote from
	eyes and not visible in facial view
	Upper part of front confined entirely to face, except sometimes for
	a narrow border; ocelli visible in facial view
11.	Lateral sutures of front distinctly continued over the obtuse an-
	terior margin of the crown to near the position of the ocelli, as
	in the Cercopidæ; antennæ between and near the eyes; body
	usually elongate, cylindrical, head often angulate, face large, strongly convex, the cheeks rather long and narrow. (Cica-
	délla, Dræculacéphala, Graphocéphala, Kólba, Oncome-
	tòpia (= Procònia) (Fig. 165), Tylozygus). (PROCONÌIDÆ,
	TETTIGONIÉLLIDÆ, TETTIGONÌIDÆ) CICADÉLLIDÆ
	Lateral sutures of front obsolete beyond antennæ or beyond an-
	terior border of crown
12.	Antennæ not far removed from eyes and near but never above
	level of eyes; lateral margins of front obsolete beyond scrobes. 13
	Antennæ situated entirely above and far removed from eyes; head
	anteriorly transversely thin and leaf-like, often concave be-
	neath
13.	Head acutely angled between crown and face, the face of narrow
	proportions; lateral sutures of front entering and terminating

in antennal scrobes, the face shallowly concave or weakly convex, the cheeks moderately swollen; body long, ovate, usually flattened. (**Gýpona** (Fig. 160), **Xerophlœa**).

**GYPÓNIDÆ** 

Head obtusely rounded between the strongly declivous crown and face, strongly overhanging the latter, which is deeply concave; lateral sutures of front passing mesad of antennæ; face very short, far broader than long...... PENTHIMÎIDÆ

14. Outlined lower part of front short and broad.

## THAUMASTOSCÓPIDÆ

Outlined lower portion of front long and narrow; large, brownish species. Principally indo-australian. (Lèdra, Ledrópsis).

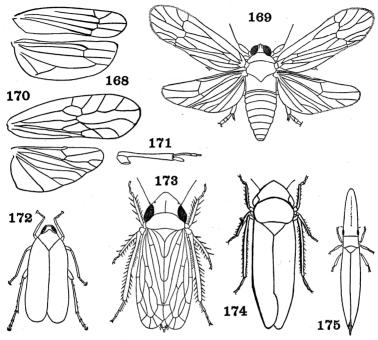
### LÉDRIDÆ

- - Basal suture of front usually obsolete, the basal lateral sutures running to and terminating at or near ocelli; vertex usually clearly connate with the front, only in highly specialized groups with a sharp edge or with transverse carinæ on anterior border; ocelli on anterior border of head or above it. (Fig. 156).

## JÁSSIDÆ

a. Fore wings with well developed veins; head variously formed but not excessively long and narrow.....b Fore wings leathery, with obliterated venation; head very long

and gradually tapering in front, body slender; tibiæ weakly spinose; Australian. (Cephalèlus (Fig. 175), Paradorýdium). CEPHALELÌNÆ



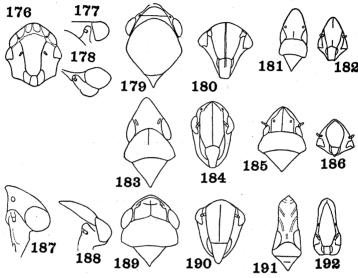
Figs. 168-175. Homoptera

- 168. Typhlocyba, wings. Jassidæ.
- 169. Sogata (Misra) Fulgoroidæ.
- 170. Liburnia, wings (Metcalf) Delphacidæ.
- 171. Liburniella, hind leg (Garman) Delphacidae.
- 172. Cixius (Metcalf) Cixiidæ.
- 173. **Deltocephalus** (DeLong) Jassidæ.
- 174. **Tettigoniella** (Ball) Cicadellidæ.
- 175. Cephalelus (Tillyard) Jassidæ.
- - Fore wings with veins, often weak at base, not branching on the disk, branching only near apex to form the apical cells; ocelli vestigial or wanting. (**Typhlócyba** (Fig. 168) (*T. austràlis*, Australian apple leaf-hopper), **Dicraneùra**, **Empòa** (*E. ròsæ*,

Rose leaf-hopper), Empoásca (E. màli, Apple leaf-hopper),

c.	Erythroneura (E. comes, Grape leaf-hopper). (EUPIERIG-IDÆ)
17.	Anterior border of vertex sharply laminately expanded, distinctly overhanging upper part of front; antennæ situated far mesad of eyes; ocelli, when distinguishable, lying between extended margin of vertex and basal margin of front in a transversely triangular (rarely linear) ocellar area and very remote from eyes
10	margin of vertex; antennæ situated close to interior line of eyes
18.	Pronotum extended between and in front of eyes; vertex very short, transverse and deeply concave
	Pronotum not abnormally extended between eyes; vertex not very short and widely transverse, the width of the vertex not more than twice the length; ocelli a little nearer to eyes than to median line, or indistinguishable
19.	Tegmina normally veined; genæ narrower than front; front strongly excavate, with high raised margins; clypeus little exserted; ocellar area very broad; hind tibiæ with very few small spines and hairs on apical half; sculpture characterized by a deep thimble pitting. (Paròpia (= Megophthálmus) (Figs. 187, 188), Mesoparòpia, malay.). (MEGOPHTHÁLMIDÆ).

Tegmina with numerous supernumerary veins; genæ wider than front; front convex; clypeus long exserted; ocellar area narrow,



Figs. 176-192. Homoptera

- 176. Koebelea, face (Baker) Koebeleidæ.
- 177. Ulopa, face in frontal view (Baker) Ulopidæ.
- 178. **Ulopa,** face in lateral view (Baker) Ulopidæ.
- 179. Signoretia, head and pronotum, dorsal view (Baker) Signoretiidæ.
- 180. Signoretia, face (Baker) Signoretiidæ.
- 181. Nirvana, head and pronotum, dorsal view (Baker) Nirvanidæ.
- 182. Nirvana, face (Baker) Nirvanidæ.
- 183. Pythamus, head and pronotum, dorsal view (Baker) Pythamidæ.
- 184. **Pythamus,** face (Baker) Pythamidæ.
- 185. Stenotortor, head and pronotum, dorsal view (Baker) Nirvanidæ.
- 186. Stenotortor, face (Baker) Nirvanidæ.
- 187. Paropia, face, lateral view (Baker) Paropiidæ.
- 188. Paropia, face, frontal view (Baker) Paropiidæ.
- 189. Euacanthus, head and pronotum, dorsal view (Baker) Euacanthidæ.
- 190. Euacanthus, face (Baker), Euacanthidæ.
- 191. Stenometopius, head and pronotum, dorsal view (Baker) Nirvanidæ.
- 192. Stenometopius, face (Baker) Nirvanidæ.

20. Genæ longer than broad, flat or concave, outwardly emarginate, normally bordering the loræ to the clypeus; scrobes very shallow and lacking strong supra-antennal ledges (as in Stenocotidæ); pronotum with very short lateral margins, converging anteriorly, ocelli distinct. (Koebèlea, nearc.). (Fig. 176).

## KOEBELÈIDÆ

- - Upper margin of face not at all extended beyond margin of vertex and not visible in dorsal view, or only a little so just in front of eyes; ocelli on anterolateral border of head or just above or below it; loræ very small and narrow; tegmina usually without anteapical cells and venation usually indistinct; antennæ situated above the eyes in facial view, rarely on upper line of or between eyes, in which case the head is long-produced... NIRVÁNIDÆ
  - a. Antennæ situated at upper angle of eyes (in facial view) or above this; lateral carinæ of vertex more or less distinct; ocelli always visible from above, on upper portion of lateral border, or on anterolateral portion of crown; eyes prominent; posterior border of pronotum more or less distinctly incurved......b
    - Antennæ situated at middle of eye margin (in facial view); lateral carinæ of vertex wanting; ocelli below anterior border of crown and not visible from above; head (from above) long spatulate, but not thin dorsoventrally; eyes not prominent, deeply set in vertex; pronotum subtruncate posteriorly; tegmina with two subapical cells. (Stenometòpius (Figs. 191, 192)).

## STENOMETOPIÌNÆ

b. Antennæ situated in deep transverse, sharp-margined scrobes; face about as broad as long or broader; eyes small; vertex short, half-ovate. (Macroceratogònia, Balbíllus, Stenotórtor (Figs. 185, 186)) . . . . . . . . . MACROCERATOGONIÌNÆ

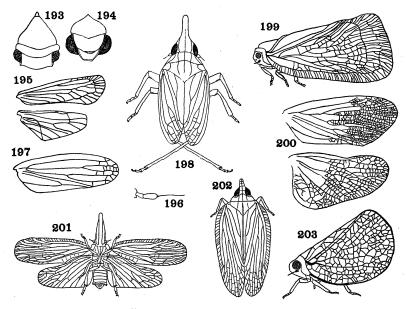
- Antennæ in shallow scrobes of ordinary type; face usually much longer than broad; vertex long; eyes large; tegmina without subapical cells, the veins of corium usually indistinguishable except by transmitted light. (Nirvàna (Figs. 181, 182), Kàna, Ophiùchus, Pseudonirvàna) NIRVANINÆ
- 23. Pronotum short, broad, broadly rounded anteriorly, the head but slightly narrower; vertex very broad, nearly twice as broad as long; width of head greater than length of head and pronotum together; ocelli situated a little within anterior margin of crown, but outside the anterolateral carina of vertex, and invisible in facial view. (Euacánthus (Figs. 189, 190); Búndera, India).

# **EUACÁNTHIDÆ**

Pronotum more or less narrowly rounded anteriorly, the head very distinctly narrower, vertex always much less than twice as broad as long; width of head always much less than length of head and pronotum together; ocelli in or very near lateral areas, and usually visible both in dorsal and facial views. (Pýthamus (Figs. 183, 184), Oniélla, Onùkia).

## PYTHÁMIDÆ

24. Flagellum of antennæ segmented; hind tibiæ without mobile spur; lateral ocelli on the front, the front reaching from eye to eye without lateral ridges dividing off a small area around the eyes; sides of face (loræ) plainly visible in front view and forming a continuous curve with the clypeus. (Tettigomètra, Egròpa, Hílda, Euphyonártex)...... TETTIGOMÉTRIDÆ Flagellum of antennæ not segmented; lateral ocelli outside the



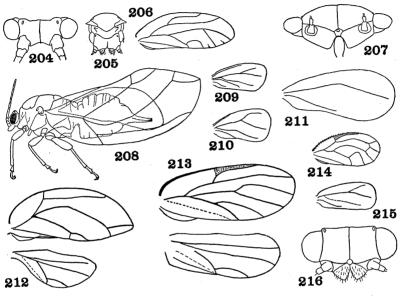
Figs. 193-203. Homoptera

- 193. Acanalonia, head (Metcalf) Acanaloniidæ.
- 194. **Aphelonema**, head (Metcalf) Issidæ.
- 195. Otiocerus, wings (Metcalf) Derbidæ.
- 196. Liburniella, antenna (Garman) Delphacidæ.
- 197. Scolops, fore wing (Metcalf) Dictyopharidæ.
- 198. Scolops (Garman) Dictyopharidæ.
- 199. **Acanalonia** (Swezey) Acanaloniidæ.
- 200. Poiocera, wings (Metcalf).
- 201. **Pyrilla** (Misra) Lophopidæ.
- 202. Tropiduchid.
- 203. Ormenis (Swezey) Flatidæ.
- 25. Second joint of hind tarsi not very small, the apex truncate or emarginate and with a row of small spines; fore wing without costal area, or with only a small one without crossveins.....26 Second joint of hind tarsus small or very small, the apex generally

	rounded or pointed and without spines or with only one at each side; costal area present or absent
26.	Claval veins not granulate; or if so, the last joint of the labium short, not longer than wide.
	One or both claval veins granulate; apical joint of labium much
	longer than wide. Abdomen compressed, the sixth to eighth
	tergites with wax secreting pores; median ocellus usually
	present. (Meénoplus, Ánigrus, Sûva, Kermèsia).
	meenopius, amgrus, suva, kermesia).  MEENÓPLIDA
27.	
21.	Sixth, seventh and eighth tergites with wax-secreting pores; ovi-
	positor reduced, incomplete. (Kínnara, Eparmène, Prosót-
	ropis, Œclídius, Atopocíxius) KINNÁRIDÆ
00	
28.	Anal area of hind wings reticulate, with many crossveins; clypeus
	with lateral carinæ; head often greatly prolonged. (Fúlgora,
	indomal.; Lanternària, neotrop.; Ámycle, Cyrpóptus, Pŷ-
	rops)FULGÓRIDÆ
00	Anal area of hind wing not reticulate
29.	Last joint of labium distinctly longer than wide30
	Last joint of labium about as long as wide. (Dérbe, Anòtia, La-
	mènia, Otiócerus (Fig. 195), Rhotàna, Venàta, Zoràida).
	DÉRBIDÆ
30.	Claval vein entering the apex of the clavus
30.	Claval vein not reaching the apex of the clavus, entering com-
	Claval vein not reaching the apex of the clavus, entering commissure before apex
30. 31.	Claval vein not reaching the apex of the clavus, entering commissure before apex
	Claval vein not reaching the apex of the clavus, entering commissure before apex
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	pha, Dichoptera, Orgamara, Orgerius, Scolops (Figs. 197,
	198)) <b>DICTYOPHÁRIDÆ</b>
	Head not prolonged in front, or only moderately so, the front
	with only a median carina, in addition to the lateral margins;
	tegulæ present; median ocellus often present. (Cíxius (Fig.
	172), Bothriócera, Koroàna, Mýndus, Oliàrus). CIXÌIDÆ
34.	Second joint of hind tarsi with a spine on each side; claval vein
	nearly always extending to and ending in the apex of the
	clavus
	Second joint of hind tarsus small, without spines
35.	
	line; fore wings with the costal area absent, or very small and
	without crossveins, or with crossveins; basal joint of hind tarsi
	usually long, rarely padded below. (Tropiduchus, Alcestis,
	Monópsis, Tambínia, Neurómeta) TROPIDÜCHIDÆ
	Mesonotum with the hind angles not marked off by a groove or
	line; first joint of hind tarsus usually short or very short36
36.	Fore wings with a crossveined costal area; without granules on
	the clavus; clypeus nearly always with lateral carinæ. (Nogo-
	dîna)
	Fore wings without a crossveined costal area, or if with such the
	clavus is granulate or the clypeus is without lateral carinæ. 37
37.	Clavus not granulate; base of costa not strongly curved 38
	Fore wings with a crossveined costal area and with the clavus
	granulate, or the base of the costa strongly curved. (Flata,
	Cerýnia, Flatòides, Nephèsa, Órmenis (Fig. 203), Phántia,
	Phỳma) FLÁTIDÆ
38.	Fore wings large, held steeply against the sides of the body; head
	about as wide as thorax; pronotum with hind edge slightly roundly emarginate, sometimes straight; mesonotum large,
	long; hind tibiæ without spines; ovipositor incomplete. (Acan-
	alònia (= Amphiscèpa) (Figs. 193, 199), Chloróchara). (AM-
	PHISCÉPIDÆ)
	Fore wings generally smaller, in Calisceline very short, or very
	narrow, parchment-like; head usually as wide as thorax or
	wider; pronotum with hind margin straight, sometimes slightly
	concave or convex; mesonotum short, not more than twice the
	length of the pronotum, with a transverse ridge parallel to the
	pronotal suture dividing it into two parts of differing sculptur-
	ing, the anterior covered by the pronotum; hind tibiæ spined;
	claval suture present (Issinæ) or absent and fore wings thick,
	ciavai suture present (issina) or absent and fore wings tinck,

	convex, and venation obscured (Hemisphæriinæ). (Brucho-
	mórpha, Aphelónema (Fig. 194), Calíscelis, Hemisphæ-
	rius, Íssus) ÍSSIDÆ
39.	Fore wings wide on apical margin, steeply held against the sides
	of the body, with a crossveined costal area; clavus long; head as
	wide or nearly as wide as the thorax; hind trochanter directed
	downward; first joint of hind tarsi at least moderately short.
	(Ricània, Armàcia, Euricània, Privèsa) RICANÌIDÆ
	Fore wings not so wide on the apical margin and not held so steeply,
	or the head is distinctly narrower than the thorax; clavus
	shorter; hind trochanter directed backward; first joint of hind
	tarsi at least moderately long40
40.	Front wider than long, the sides angulate; clypeus without lateral
	carinæ and front without longitudinal carinæ or with only a
	very obscure one. (Eurýbrachys, Messèna, Platýbrachys,
	Théssitus) EURYBRÁCHIDÆ
	Front rarely as wide as long and often without angular margins,
	nearly always with one or three longitudinal carinæ. (Lòphops,
	indomal.; Pyrilla (Fig. 201), malay.; Elasmóscelis, ethiop.,
	indomal.; Kasseròta, indoaustr.) LOPHÓPIDÆ
41.	Tarsi two-jointed, the basal joint sometimes reduced, the outer
	joint with two claws; wings, when present, four in number, with
	few veins, at rest usually held in a sloping position over the ab-
	domen; sutures between body segments distinct; mouthparts
	usually well developed in both sexes, labium usually long42
	Tarsi one-jointed (in some Monoplebidæ and in the male cochineal
	insect there is an additional minute basal joint) and with a
	single claw; females stout-bodied, always wingless, often with-
	out legs so that they rarely move after maturity, remaining
	sessile on the host plant, rarely without mouthparts; males
	delicate, usually with mesothoracic wings alone developed,
	which are gauzy and almost veinless and lie flat, overlapping
	on the abdomen when at rest; antennæ of female absent or with
	as many as eleven joints, of male with ten to twenty-five joints;
	body of female and nymphal males scale-like, gall-like, or cov-
	ered with waxy powder, tufts or scales, the sutures between the
	segments often indistinct. (Superfamily COCCÒIDEA) 47
42.	
	membranous or opaque whitish; antennæ three-tosix-jointed. 43
	Jumping insects, the femora thickened; antennæ long, five to ten-
	jointed, usually ten-jointed, the last joint with two fine apical



Figs. 204-216. Homoptera

- 204. Freysiula, head (Crawford) Chermidæ.
- 205. Trioza, head (Crawford) Chermidæ.
- 206. Carsidara, fore wing (Crawford) Chermidæ.
- 207. Paurocephala, head (Crawford) Chermidæ.
- 208. **Trioza** (Peterson) Chermidæ.
- 209. Udamoscelis, fore wing (Quaintance and Baker) Aleyrodidæ.
- 210. Aleurodicus, wing (Quaintance and Baker) Aleyrodidæ.
- 211. Aleurocanthus, wing (Quaintance and Baker) Aleyrodidæ.
- 212. Trioza, wings (Patch) Chermidæ.
- 213. Pachypsylla, wings (Patch) Chermidæ.
- 214. Ceriacremum, fore wing (Crawford) Chermidæ.
- 215. Aleyrodes, fore wing (Quaintance and Baker) Aleyrodidæ.
- 216. Pachypsylla, head (Crawford) Chermidæ.
  - a. Head deeply cleft, with the antennæ attached to the truncate anterior ends on each side of the cleft (Fig. 204); cheeks seldom produced into conical processes; media not dichotomously

- c. Vertex flat, horizontal; the front beneath it in the form of a narrow, usually elongate piece that extends from the clypeus to the anterior ocellus; wings often more or less thickened and spotted. (Aphálara, Aphalaròida, Lívia, Rhinócola) . . . . LIVIÌNÆ
- d. Fore wing with more than two marginal cells, the radial sector branched or connected to the media by a crossvein near the tip of the wing (Fig. 214). (**Ceriacrèmum**, neotrop.).

Fore wing with only two marginal cells, formed by the furcation of the media and cubitus (Fig. 212), the radial sector not branched and not connected with the media by a crossvein...e

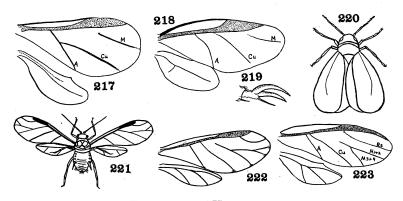
- e. First joint of hind tarsi with two black, claw-like spines at tip; radius, media and cubitus not arising at the same point from the basal vein, the media and cubitus stalked; wings rarely angulate at apex. (Arytàina, Euphálarus, widespr.; Epipsýlla, Euphyllùra, Pachypsýlla (Figs. 213, 216); Chérmes (=Psylla) cosmop. (C. pyrícola, Pear psylla). (PSYLLINÆ).
  - First joint of hind tarsi simple, without such spines at tip; radius, media and cubitus usually arising at a common point, the media and cubitus not stalked; wings usually angulate at apex. (Ceropsýlla, neotrop.; Megatriòza, widespr.; Paratriòza, neotrop.; Triòza, cosmop.). (Figs. 205, 208, 212).

TRIOZÌNÆ

43. Wings transparent, though sometimes colored, the hind wings smaller than the fore pair; tarsi with the basal joint sometimes much reduced, empodium greatly reduced or absent; body not mealy but sometimes with waxy wool; life cycle very compli-

	cated, including agamic and sexual generations, of dissimila appearance. Plant lice, Aphids. (Superfamily APHIDÒIDEA)
a.	Empodium absent; fore wings with veins R <sub>1</sub> , Rs, M, Cu and A present. ( <b>Udamóselis</b> , S. Am. (Fig. 209)). <b>UDAMOSELĪNA</b> Empodium present; fore wings with either M or Cu and A vein
b.	absent
	Fore wings with outer part of stigma bounded behind by vein R <sub>1</sub> the radial sector separate (Fig. 222); sexual females oviparous summer parthenogenetic females viviparous; new-born with anterior pronotal pleural bristles absent
a.	New-born individuals with four bristles on basal tarsal joint; head free, not fused with prothorax, adults with vertex margined labium five-jointed; cornicles broadly conical to pore-like rarely absent. (Láchnus (Pine aphids), Cínara, Euláchnus Tràma).  New-born individuals with two bristles on basal joint of tarsus labium four-jointed; cornicles pore-like to elongate cylindrical rarely absent; head of adult with vertex not margined

b. Head free; newly born with faceted eyes; hind tibiæ of oviparous female thickened. (Aphis (Fig. 221), (A. gossýpii, Cotton aphis, A. màidis, Corn aphis, A. pòmi, Green apple aphis), Rhopalosìphum, Toxóptera (T. gráminum, Green bug of wheat); Chaitóphorus; Saltusaphis; Callípterus; Pterocómma; Anuraphis (A. bàkeri, Clover aphis, A. màidi-rádicis, Corn root aphis, A. pérsicæ-nàger, Black peach aphis); Cryptosìphum; Brachýcolus (=Brevicorýne) Hyalópterus;

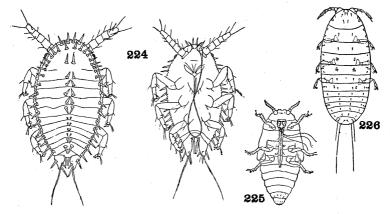


Figs. 217-223. Homoptera

- 217. Phylloxera, wings (Patch) Phylloxeridæ.
- 218. Adelges, wings (Patch) Adelgidæ.
- 219. Aleyrodes, tarsus (Quaintance) Aleyrodidæ.
- 220. Aleyrodes (Bemis) Aleyrodidæ.
- 221. Aphis (Chittenden) Aphididæ.
- 222. Macrosiphum, wings (Patch) Aphididæ.
- 223. Mindarus, wings (Patch) Aphididæ.

- Antennæ with narrow transverse sensoria; radial sector of fore wings arising from the stigma; wing veins much reduced so that the media is usually simple; cornicles usually absent or much reduced; sexual forms usually apterous and of small size; species usually producing galls; wax glands usually present. (Hormaphis, Cerataphis, Hamamelistes). HORMAPHIDINÆ

- - - Abdomen with six pairs of spiracles, the first not evident; agamic young of two kinds, either (a) delicate summer forms, the first generation usually winged, with short rostrum, moulting four times, and not overwintering, or (b) chitinized, wingless, winter form, with long rostrum, moulting three times, which rest over summer and are active in fall. (Adélges (Fig. 218) (=Cnaphalòdes), Gilletteélla, Sacchiphántes) . . . . . . . ADELGINÆ
    - Wings when at rest laid flat upon the abdomen, veins Cu and 1A fused at base forming a Y-vein; antennæ three-jointed; parthenogenetic females with beak, sucking but not defecating; sexual forms without beak; wingless agamic females not secreting a waxy flocculence, but in *Phylloxera* they secrete a waxy powder.



Figs. 224-226. Homoptera

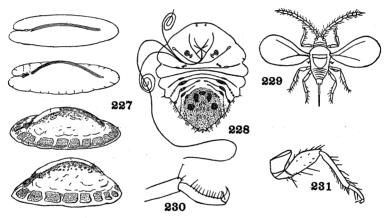
- 224. Phylloxera, dorsal and ventral views. (Doten) Phylloxeridæ.
- 225. **Phylloxera**, nymph, ventral view (Simanton) Phylloxeridæ.
- 226. Phœnicococcus, larva (Morrison) Cylindrococcidæ.
- 48. Larva and all female stages with a distinctly developed flat anal ring bearing pores and six setæ; adult male with simple nine-jointed antennæ, with a rather conspicuous seta at extreme tip of apical joint; penis sheath of adult male appearing strongly bivalved. (Orthèzia, tropicopolitan)..... ORTHEZĪIDÆ

  None of the stages with a flat anal ring bearing pores and setæ; adult male nearly always with simple ten-jointed antennæ, rarely with pectinate antennæ or with more than ten joints; penis sheath of adult male mostly entire, or merely cleft at apex, at most with short bilobate tip, in which case the compound

eves are poorly developed. ( $MARGARODID\mathcal{E}$ ).

- a. Adult female with tarsi two-jointed (Fig. 241), rarely the legs reduced to a small unsegmented protuberance; disk-like simple pores present; intermediate female legless; halteres of male with four to six long curved apical bristles (Fig. 242). Widespread. (Matsucoccus, palearc. (Figs. 242, 245); Stigmacoccus, neotrop. (Figs. 241, 248, 249); Xylococcus, nearc.).
  - XYLOCOCCINÆ
  - Adult female with tarsi one-jointed, legs, if reduced, with some segmentation; disk-like simple pores wanting.....b
- b. Adult female with six to twelve large knobbed bristles surrounding and surpassing the tarsal claw (Fig. 246); antennæ contiguous at base; intermediate females legless; male with eyes reduced to a row of facets or even to a single facet. Holarctic. (Steingèlia, palæarc.; Stomacóccus, nearc.) ... STEINGELIÌNÆ
- c. Adult female with dorsal anus, anal tube relatively well developed and provided with a simple proximal ring; intermediate female with antennæ and legs fully developed, anal tube with ring and anal opening distinctly dorsal; male tibiæ, tarsi and front femora with bifurcate setæ, middorsal area of thorax with an unchitinized area, and abdomen with one or more pairs of fleshy marginal tassels. (Drósicha, widespr.; Icèrya, tropicopol. (Figs. 230, 243); Llavèia, neotrop., indomal.; Monophlebus, indomal.; Palæocóccus (Fig. 231), widespr.; Steatocóccus).

- 50. Scale covering constructed around the first moulted skin; pygidium of the covered insect definitely formed of the fused



Figs. 227-231. Homoptera

- 227. Lecanium, growth stages of female scale. Lecaniidæ.
- 228. Diaspis, female (Howard) Diaspididæ.
- 229. Aspidiotus, male (Howard) Diaspididæ.
- 230. Icerya, tarsus of female (Riley) Monophlebidæ.
- 231. Palæococcus, hind leg. Monophlebidæ.

a. Scale of adult female or second nymphal female more or less elongate or sometimes rounded, with exuviæ at one end, if nearly circular the exuviæ near margin or when central not concentrically superposed; exuvia of first nymphal female with the remains of antennæ showing as porrect appendages . . . . b

Scale of adult and second nymphal female nearly circular, the
exuviæ central, if elongate the exuviæ concentrically superposed,
not projecting beyond margin of scale or attached at margin;
exuvia of first nymphal female never showing remains of an-
tennæ. (Aspidiòtus, widespr. (Fig. 229) (A. (Comstockáspis)
perniciòsus, San José scale); Chrysómphalus, widespr.; Tar-
giònia, widespr.)
Cools of female meanly simular with nimels control or exceptuic

b. Scale of female nearly circular, with nipple central or excentric, rarely projecting beyond margin of scale; male scale elongate, with almost parallel sides. (Aulacáspis, widespr. (Figs. 233, 234) (A. ròsæ, Rose scale); Diáspis, widespr. (D. bromèliæ, Pineapple scale; D. pirîcola, Pear scale); Howárdia, widespr.).

### DIASPIDÎNÆ

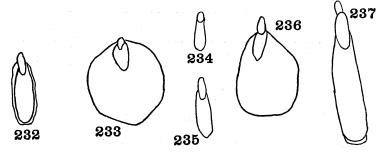
- - Female scale elongate (Fig. 232), formed in greater part by the puparium (nymphal exuvia which encloses the adult); secreted part of scale thin. (Adiscofiorinia, ethiop., indoaustr.; Fiorinia, indoaustr. (Fig. 232); Trullifiorinia, indoaustr.).

# **FIORINIÌNÆ**

- e. Pygidium usually edged with a continuous series of lobes and wide fringed processes (pectinæ), rarely with pointed narrow plates; preanal median group of wax glands often wanting, when present rarely with more than eight glands; scale white or whitish. .f
  - Pygidium of adult female or of second nymphal stage usually with pointed narrow plates, and pectinæ at most with narrow shafts; scale of adult female with second exuvia small, rarely covering half of scale; usually more than eight preanal wax glands; scale dark colored. Oyster-shell scales. (Coccomýtilus, widespr.; Lepidôsaphes (= Mytiláspis), widespr. (Fig. 240) (L. bécki, Citrus purple scale; L. ûlmi, Oyster-shell scale); Pinnáspis, Am., indomal.; Scrupuláspis, palæarc., indoaustr.).

## LEPIDOSAPHÌNÆ

f. Scale of adult female elongate, often pyriform, sometimes with parallel sides, formed mainly of the large puparium or nymphal



Figs. 232-237. Homoptera

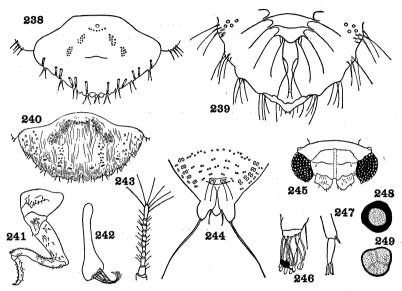
- 232. Fiorinia, female, outline of scale. Diaspididæ.
- 233. Aulacaspis, female, outline of scale. Diaspididæ.
- 234. Aulacaspis, male, outline of scale. Diaspididæ.
- 235. Chionaspis, male, outline of scale. Diaspididæ.
- 236. Chionaspis, broad female scale. Diaspididæ.
- 237. Chionaspis, narrow female scale. Diaspididæ.

Scale covering not containing the exuviæ of the early moults; pygidial segments less completely fused; legs present, even in adult female, tibio-tarsal suture obsolete; antennæ of adult female three-jointed; beak two-jointed. (Concháspis, neotrop., Ceylon (Fig. 239); Fasisûga, Chile; Scutàre, neotrop.).

### CONCHASPÍDIDÆ

51. Female with posterior end cleft; anus closed by a pair of dorsal plates; larvæ also with the anal cleft bounded on each side by a prominent seta-bearing lobe or plate; beak one-jointed; wax glands very rarely paired to resemble the figure 8; body of adult female sometimes greatly convex, bare or encased in waxy or

	cottony secretion. (Ceroplástes, cosmop. (C. ceriferus, Indian
	wax scale); Lecànium, cosmop. (Fig. 227) (L. côrni, Brown scale;
	L. hespéridum, Soft brown scale; L. pérsica, Peach scale);
	Lecaniópsis; Neolecànium, Am.; Physokérmes, nearc.;
	Pulvinària, widespr. (P. vitis, Cottony maple scale); Saissètia,
	widespr. (S. òleæ, Black scale); Toumeyélla)LECANÌIDÆ
	Anal end of abdomen not medially cleft, if apparently cleft and
	provided with lobes some of the microscopic wax glands are
	paired to resemble the figure 8
52.	End of abdomen more or less narrowed or prolonged into a tubular
	anal projection; beak two-jointed. Species inhabiting galls, or
	enclosed in wax53
	Abdomen not narrowed at tip or prolonged into an anal protuber-
	ance
53.	Insects enclosed in a mass of resinous cells, each cell with three
	adjacent openings; adult female legless, body globular or sub-
	conical, with mouthparts at one end and three tubular processes
	at the opposite end, one of the projections bearing the anus and
	the other two the mesothoracic spiracles. Lac insects. (Lácci-
	fer (= Tachárdia), ind.; Tachardiélla, widespr.; Tachardina,
	ethiop.). (TACHARDIIDÆ)LACCIFÉRIDÆ
	Insects forming galls. Usually on Eucalyptus trees; adult female
	segmented, top-shaped, with at least one pair of legs, or seg-
	mentation obsolete, head and thorax globular, abdomen re-
	duced to a tubercle, and legs and antennæ wanting. Australian.
	Peg-top Coccids. (Apiomórpha, Áscelis, Cystocóccus, Opis-
	thóscelis). $(BRACHYSCÉLIDÆ)$ APIOMÓRPHIDÆ
54	Wax glands distributed largely in pairs resembling the figure 8,
01.	generally arranged in rows; beak one-jointed; anal ring pro-
	vided with setæ; legs of adult female vestigial or wanting.
	(Asterolecànium, widespr.; Cerocóccus, widespr. (Fig. 244);
	Lecaniodiáspis, widespr.; Ollíffia, austr.).
	ASTEROLECANIDÆ
	Wax glands not seriately arranged in pairs resembling the figure 8.55
55.	Antennæ of adult female eleven-jointed; male eyes consisting of
55.	eight units arranged in a circle; anal ring distinct and provided
	with six prominent anal ring setæ, no anal lobes or anal setæ.
	(Phenacoleachia, Austr.) PHENACOLEACHÌIDÆ
	Antennæ of adult female with at most nine joints, often reduced
	or wanting; male eyes consisting of fewer parts, not arranged
	or wanting; male eyes consisting of fewer parts, not arranged



Figs. 238-249. Homoptera

- 238. **Lepidosaphes**, pygidium of adult female (Green) Diaspididæ.
- 239. Conchaspis, pygidium of adult female (Green) Conchaspididæ.
- 240. Lepidosaphes, pygidium of adult female (Quayle) Diaspididæ.
- 241. Stigmacoccus, leg of adult female (Morrison) Monophlebidæ.
- 242. **Matsucoccus**, halter of male (Morrison) Monophlebidæ.
- 243. Icerya, antenna of female (Riley) Monophlebidæ.
- 244. Cerococcus, pygidium of adult female (Green) Asterolecaniidæ.
- 245. Matsucoccus, male, dorsal view of head (Morrison) Monophlebidæ.
- 246. Steingelia, tip of tarsus of female (Morrison) Monophlebidæ.
- 247. Rhizococcus, tip of tibia and tarsus (Packard) Diaspididæ.
- 248. Stigmacoccus, simple disk pore (Morrison) Monophlebidæ.
- 249. Stigmacoccus, trilobate disk pore (Morrison) Monophlebidæ.

- Adult female with some or all legs wanting, or when all legs present the hind pair are two or three times as long as the others; antennæ, if present, with less than seven joints, often vestigial or wanting. Southern hemisphere. (Apiocóccus, neotrop.; Cylindrocóccus, austr.; Halimocóccus, ethiop.; Ourocóccus, austr.). (IDIOCÓCCIDÆ)......CYLINDROCÓCCIDÆ
- 59. Head freely articulated with the thorax; fore wings membranous, with veins and crossveins that enclose numerous cells. Terrestrial. (Pelorídium neotrop.; Xenophyes, Hemiodœcus, austr.) PELORIDÎIDÆ
  - Head more or less completely fused with the thorax, not movable; upper wings very thick, covering the whole abdomen; scutellum short and broad; aquatic. (Plèa, widespr.) .... PLÈIDÆ

### LITERATURE ON BOTH HOMOPTERA AND HEMIPTERA

- Banks, N. Catalogue of Nearctic Hemiptera-Heteroptera. Philadelphia, Am. Ent. Soc., 103 pp. (1910).
- Berg, C. Hemiptera Argentina. Anal. Soc. Cient. Argentina, **5-34** (1878–1892) (numerous parts).
- Britton, W. E. (Editor). Hemiptera and Homoptera of Connecticut. Connecticut Nat. Hist. Surv., Bull. No. 34, 783 pp. (1923). (Various authors).
- Butler, E. A. A Biology of the British Hemiptera-Heteroptera. London,  $695~\rm pp.~(1923).$
- Champion, G. C. Hemiptera-Heteroptera. In Biologia centrali-Americana, 2 (1897–1901).
- Distant, W. L. Hemiptera-Heteroptera. In Biologia centrali-Americana,  ${\bf 1}$  (1880–1893).
  - Rhynchota in Fauna of British India, 5 vols., London (1902–10).
- DOHRN, A. Catalogus Hemipterorum. Entom. Verein, Stettin, 112 pp. (1859). GUÉRIN, J. and PENEAU, J. Faune entomologique armoricaine, Hémiptères. Trav. Sci. Univ. Rennes, 13, pp. 201–301 (1915).

Horváth, G. Nomenclature des familles des Hémiptères. Ann. Mus. Nat. Hungarici, 9, pp. 1–34 (1911).

Hemiptera. Fauna Regni Hungariæ, 1, pp. 5-64 (1918).

Kirkaldy, G. W. Hemiptera. Fauna Hawaiiensis, 3, Pt. 2 (1902); 2, Pt. 6 (1902). Lethierry, L. and Severin, G. Catalogue Générale des Hémiptères, 3 vols., Brussels (1893–96).

Oshanin, B. Verzeichnis der paläarktischen Hemipteren. Ann. Mus. Zool. Acad. Imp. Sci. St. Petersbourg, **11–15** (1906–10).

Katalog der paläarktischen Hemipteren, 187 pp., Berlin (R. Friedländer und Sohn) (1912).

Vade mecum destiné a faciliter la détermination des Hémiptères. Hor. Soc. Entom. Rossicæ, **42**, No. 2, pp. 1–106 (1916). (Very complete and classified list of literature).

Peneau, M. J. Hémiptères. In Houlebert, Encyclop. Sci., 2 vols. (1922).

STÅL, C. Bidrag till Rio Janeiro-traktens Hemipter-fauna. K. Svensk. Akad. Handl., 2 and 3 (1860–62).

Hemiptera africana, 1-4, Holmiæ (1864-66).

Hemiptera Fabriciana. Svensk. Vet. Akad. Handl., 7 and 8 (1868–69). Enumeratio hemipterorum. Svensk. Vet. Akad. Handl., 9–15 (1870–76).

VAN DUZEE, E. P. Catalogue of the Hemiptera of America north of Mexico, except Aphididæ, Coccidæ and Aleurodidæ. Univ. California Pubs. Ent., 2, 902 pp. (1917).

# LITERATURE ON HOMOPTERA (GENERAL)

(See also General List, p. 134)

DISTANT, W. L. Fauna of British India, 6, 240 pp. (Homoptera appendix).

DISTANT, W. L. and FOWLER, W. W. Homoptera. In Biologia centrali-Americana, 1 (1881–1905).

EDWARDS, J. The Hemiptera Homoptera of the British Isles. 271 pp., London (1894–96).

Fowler, W. W. Homoptera. In Biologia centrali-Americana, 2, pt. 1 (1894–1909).

Jensen-Haarup, A. C. The Danish Homoptera. Copenhagen (1920).

Melichar, L. Die Homopterenfauna von Ceylon. 248 pp., Berlin (1903).

OSHANIN, B. Katalog der paläarktischen Hemipteren. 187 pp., Berlin (R. Friedländer und Sohn) (1912).

Vade mecum destiné a faciliter la determination des Hémiptères. Hor. Soc. Entom. Rossicæ, **42**, No. 2, pp. 1–106 (1916). (Very complete and classified list of literature).

Van Duzee, E. P. Catalogue of the Hemiptera of America north of Mexico, except Aphididæ, Coccidæ and Aleyrodidæ. Univ. California Publ. Tech. Bull. Entom., 2 (1917).

Walker, F. List of specimens of Homopterous Insects in the British Museum, 1–4 (1850–52); Supplement (1858) London, British Museum.

# SUBORDER AUCHENORRHYNCHA CICADOIDEA

- ASHTON, H. A Revision of the Australian Cicadidæ, Pt. 1. Proc. Roy. Soc. Victoria, 33, pp. 87–107 (1921).
- Buckton, G. B. A Monograph of the British Cicadæ. 2 vols., London (1890–91).
- Deletang, L. F. Monográfia de los Cicádidos Argentinos. An. Mus. Nac. Buenos Aires, **31**, pp. 538–649 (1923).
- DISTANT, W. L. Monograph of the Oriental Cicadidæ. London, 158 pp. (1889–92).
  - A Synonymic Catalogue of the Cicadidæ. 207 pp., London, British Museum (1906).
    - Cicadidæ, Cicadinæ. Gen. Insectorum, fasc. 142, 64 pp. (1913).
    - Cicadidæ, Gæaninæ. Gen. Insectorum, fasc. 158, 38 pp. (1914).
- HAUPT, H. Neueinteilung der Homoptera-Cicadina. Zool. Jahrb. Abth. f. Syst., 58, pp. 173–286 (1929).
- HorvAтн, G. Cicadidarum genera palæarctica. Ann. Mus. Nat. Hungarici, **10**, pp. 599-609 (1912).
- LAWSON, P. B. The Cicadidæ of Kansas. Kansas Univ. Sci. Bull., 12, pp. 307–352 (1920).
- Melichar, L. Die Cicadinen von Mitteleuropa. Berlin, 364 pp. (1896).
- Moulton, J. Cicadas of Malaysia. Journ. Fed. Malay States Mus., 11, pp. 69–182 (1923).
- MYERS, J. G. Insect Singers. A Natural History of the Cicadas. London, Geo. Routledge and Sons (1929). (Very complete bibliography).

#### MEMBRACOIDEA

- Buckton, G. B. Monograph of Membracidæ. 296 pp., London (1901-03).
- Funkhouser, W. D. Review of the Philippine Membracidæ. Philippine Journ. Sci., **10**D, pp. 365–405 (1915).
  - Membracidæ. Gen. Cat. Hemip. fasc. 1 (1927) Smith College, Northampton, Mass.
- Goding, F. W. Catalogue of Membracidæ of North America. Bull. Illinois State Lab. Nat. Hist., **3**, pp. 391–482 (1892).
  - A Monograph of the Australian Membracidæ. Proc. Linn. Soc. New South Wales, **38**, pp. 2–41 (1903).
  - Classification of the Membracidæ of America. Journ. New York Entom. Soc., 34, pp. 295–317 (1926).
  - Revision of the Membracidæ of South America and the Antilles. Journ. New York Entom. Soc., **35**, pp. 183–191 (1927).
  - The Membracidæ of South America and the Antilles. Pt. 4, Trans. American Entom. Soc., **55**, pp. 197–330 (1929).
- Van Duzee, E. P. Review of North American Membracidæ. Bull. Buffalo Soc. Nat. Sci., 9, pp. 29–129 (1908).

### CERCOPOIDEA

- Baker, C. F. Some Philippine and Malaysian Machærotidæ. Philippine Journ. Sci., **32**, pp. 529-548 (1927).
- Doering, K. Synopsis of North American Cercopidæ. Journ. Kansas Entom. Soc., 3, pp. 53-64; 81-108 (1930).
- Goding, F. W. Synopsis of Genera of North American Cercopidæ. Bull. Illinois State Lab. Nat. Hist., 3, pp. 483-501 (1895).
- LALLEMAND, V. Cercopidæ. Gen. Insectorum, fasc. 143, 167 pp. (1913).
- Matsumura, S. Monographie der Cercopiden Japans. Journ. Sapporo Agric. Coll., 2, p. 15 (1903) and Annot. Zool. Japonenses, 5, pp. 31–55 (1904).
- SCHMIDT, E. Monographie der Macherotiden. Stettiner Entom. Zeitg., 68, pp. 165–200 (1907).

### JASSOIDEA

- Buys, J. L. Cicadellidæ of Ithaca, New York. Mem. Cornell Univ. Agric. Expt. Sta., No. 80, 115 pp. (1924).
- CRUMB, S. E. Partial Key to Genera of North American Jassidæ. Trans. Kansas Acad. Sci., **25**, pp. 129–137 (1914).
- LATHROP, F. H. The Cicadellidæ of South Carolina. Bull. 199, South Carolina Agric. Expt. Sta. (1919).
- LAWSON, P. B. The Cicadellidæ of Kansas. Kansas Univ. Sci. Bull., 12, No. 1, Ent., 4, pp. 1–306 (1920).
- MATSUMURA, S. Monographie der Jassinen Japans. Termes. Füzetek, 25, pp. 353–404 (1902).
- McAtee, W. L. The Genera of the Eupterygidæ. Proc. Biol. Soc. Washington, **31**, pp. 109–124 (1918).
- NAUDÉ, T. J. Cicadellidæ of South Africa. Entom. Mem. Dept. Agric. Pretoria, No. 4, 106 pp. (1926).
- Osborn, H. The Leafhoppers of Ohio. Ohio State Univ. Bull., **32**, pp. 199–374 (1928).
  - Neotropical Homoptera, Part 6. (Typhlocybinæ). Ann. Carnegie Mus., 18, pp. 253–298 (1928).

### FULGOROIDEA

- Baker, C. F. Spolia Mentawiensia: Homoptera Fulgoroidea. Philippine Journ. Sci., **32**, pp. 391–412 (1927).
- CRAWFORD, D. L. Monograph of American Delphacidæ. Proc. U. S. Nat. Mus., 46, pp. 557–640 (1914).
- Dozier, H. L. The Fulgoridæ of Mississippi. Tech. Bull. Mississippi Agric. Expt. Sta., No. 14, 152 pp. (1928).
- GIFFARD, W. M. A Review of the Hawaiian Cixiidæ, with Descriptions of Species (Homoptera). Proc. Hawaiian Entom. Soc., 6, pp. 51-156 (1925).

Matsumura, S. Uebersicht der Fulgoriden Japans. Entom. Nachricht., 26, pp. 205–213; 257–270 (1900).

Beitrag zur Kenntnis der Fulgoriden Japans. Ann. Mus. Nat. Hungarici, 12, pp. 261–305 (1914).

Die Cixiinen Japans. Annot. Zool. Jap. Tokyo, 8, pp. 393-434 (1914).

Melichar, L. Monographie der Ricaniiden. Ann. Hofmus. Wien, 13, pp. 197–359 (1898).

Monographie der Acanaloniden und Flatiden. Ann. Hofmus. Wien, **16**, pp. 178–258 (1901); **17**, pp. 1–253 (1902).

Monographie der Issiden. Abh. zool.-bot. Ges. Wien, **3**, pp. 1–327 (1906). Monographie der Dictyopharinen. Abh. zool.-bot. Ges. Wien, **7**, pp. 1–221 (1912).

Monographie der Tropiduchinen. Verh. naturh. Ver. Brünn, **53**, pp. 82–225 (1914).

Monographie der Lophopinen. Ann. Mus. Hist. Hungarici, **13**, pp. 337–385 (1915).

METCALF, Z. P. Fulgoridæ of Eastern North America. Journ. Elisha Mitchell Sci. Soc., **38**, pp. 139–230 (1923).

Muir, F. Derbidæ of the Philippine Islands. Philippine Journ. Sci., **12**D, pp. 49–104 (1917).

Classification of the Fulgoroidea. Proc. Hawaiian Entom. Soc., **5**, pp. 205–247 (1922).

South American Fulgoroidea, Pt. 1, Delphacidæ. Bull. Hawaiian Sugar Planters' Exp. Sta., No. 18 (1926).

Classification of the Fulgoroidea. Ann. Mag. Nat. Hist. (10), 6, pp. 461–478 (1930).

Muir, F. and Giffard, W. M. Studies in North American Delphacidæ. Entom. Bull. Hawaiian Sugar Planters' Expt. Sta., No. 15, 53 pp. (1924).

Van Duzee, E. P. Review of North American Delphacidæ. Bull. Buffalo Soc. Nat. Sci., **5**, pp. 225–261 (1897).

# SUBORDER STERNORRHYNCHA CHERMOIDEA

Aulmann, G. Psyllidarum catalogus. 92 pp. W. Junk, Berlin (1913).

Blöte, A. C. Overzicht der Nederlandsche Psyllidensoorten. Tijdschr. Entom., **69**, pp. 57–84 (1926).

Crawford, D. L. Monograph of Psyllidæ of the New World. Bull. U. S. Nat. Mus., No. 85 (1914).

The Psyllidæ of the Hawaiian Islands. Proc. Hawaiian Entom. Soc.,  ${f 3},$  pp. 430–457 (1918).

#### ALEYRODOIDEA

Baker, A. C. and Moles, M. L. Aleyrodidæ of South America. Rev. Chilena Hist. Nat., 25, pp. 609–648 (1923).

- Bemis, F. E. Aleurodidæ of California. Proc. U. S. Nat. Mus., 27, pp. 471–537 (1904).
- Maskell, W. M. Contributions toward a Monograph of the Aleurodidæ. Trans. New Zealand Inst., 28, pp. 411–449 (1896).
  - Quaintance, A. L. Aleyrodidæ. Gen. Insectorum, fasc. 87, 11 pp. (1908).
  - QUAINTANCE, A. L. and BAKER, A. C. Classification of the Aleyrodidæ. Bull. Bur. Entom., U. S. Dept. Agric. Tech. Ser., No. 27, 109 pp. (1913).

A Contribution to our Knowledge of the Aleyrodinæ. Proc. U. S. Nat. Mus., **51**, pp. 335–445 (1917).

#### APHIDOIDEA

- Annand, P. N. A Contribution toward a Monograph of the Adelginæ (Phylloxeridæ) of North America. Stanford Univ. Publ. Biol. Sei., 6, pp. 1–146 (1928).
- Baker, A. C. A Generic Classification of the Hemipterous Family Aphididæ. Bull. U. S. Dept. Agric. 826, pp. 1–109 (1920).
- Börner, C. Das System der Phylloxerinen. Zool. Anz., **33**, pp. 600–612 (1908). Beiträge zu einem neuen System der Blattläuse. Arch. Klass. Phylog. Ent., **1**, pp. 115–194 (1930).
  - Monographische Studie über die Chermiden. Arb. Kais. biol. Anst. Land. Forstwirtsch., 6, Pt. 2 (1908).
- Buckton, G. B. A Monograph of the British Aphides. 4 vols. London, Ray Soc. (1875–82).
- Сногоркоwsкy, N. Die Koniferen-Läuse Chermes. 44 pp. Berlin, Friedländer (1907).
- DAVIDSON, J. A List of British Aphides. 176 pp. London, Longmans, Green & Co. (1925).
- Essig, E. O. The Aphididæ of California. Univ. California Publ. Entom., 1, pp. 301–346 (1917).
- Goot, P. van der. Zur Systematik der Aphididen. Tijdschr. Entom., **55**, pp. 69–154 (1903).
- Kirkaldy, G. W. Catalogue of the Genera of Aphidæ. Canadian Entom., **37**, **38**, **40** (1905–08) (various parts).
- Lombardi, D. Contribuzione alla conoscenza della tribu Fordina. Bull. Lab. Zool. Gen. Agrar. Portici, 7, pp. 149–188 (1913).
- Mordvilko, A. K. Aphidoidea. Faune Russie, Hemiptera, 1, pp. 237–508 (1919).
- Oestlund, O. W. Key to Aphididæ of Minnesota. 19th Rept. Minnesota State Entomologist, pp. 114–151, St. Paul (1923).
- Schouteden, H. Catalogue des Aphides de Belgique. Mém. Soc. Entom. Belgique, 12, pp. 189-246 (1906).
- Takahashi, R. Aphididæ of Formosa. Pt. 1, from Agric. Expt. Sta. Govt. Formosa (1921); Pts. 2–5, Repts. 4, 10, 16, and 22 from Govt. Res. Sta. Formosa (1923, 24, 25, and 27).

- Theobald, F. V. Aphididæ of Great Britain. I. 372 pp. London, Headley Bros. (1926).
- WILSON, H. F. A Synopsis of the Aphid tribe Pterocommini. Ann. Entom. Soc. America, 8, pp. 347–358 (1915).
- WILSON, H. F. and VICKERY, R. A. Species List of Aphididæ of World and Food Plants. Trans. Wisconsin Acad. Sci., Arts, Letters, 19, Pt. 1 (1918).

#### COCCOIDEA

- Chamberlin, J. C. Systematic Monograph of the Tachardiinæ. Bull. Entom. Research, 14, pp. 147–212 (1923).
- Cockerell, T. D. A. Coccoidea. In Biologia centrali-Americana, Homoptera. 2, Pt. 2 (1899).
- Fernald, M. E. Catalogue of the Coccidæ of the World. Bull. Massachusetts Agric. Expt. Sta., No. 88, 360 pp. (1903).
- Froggatt, W. W. A Descriptive Catalogue of the Scale Insects of Australia.

  Agric. Gazette New South Wales, 25 (1914) (six parts).
- Green, E. E. The Coccide of Ceylon. 5 vols. London (1896-1922).
- Hall, W. J. Observations on the Coccidæ of Southern Rhodesia. Bull. Entom. Res., 20, pp. 345–376 (1929).
- Hempel, A. As Coccidas do Brazil. Cat. Fauna Brasiliera. Rev. Mus. São Paulo, 3, pp. 1–77 (1912).
- Kuwana, I. The Diaspine Coccidæ of Japan. Imp. Plant Serv. Tech. Bulls., Nos. 1–3, 80 pp. (1925).
- Leonardi, G. Monografia delle Cocciniglie Italiane. 555 pp. Portici (1920).
- LINDINGER, L. Die Schildläuse (Coccidæ) Europas, Nordafrikas und Vorderasiens. Stuttgart (1912).
- MacGillivray, A. D. The Coccidæ. Scarab Co., Urbana, Ill. (1921).
- Morrison, H. The Non-diaspine Coccidæ of the Philippines. Philippine Journ. Sci., 17, pp. 147–202 (1920).
  - Classification of Ortheziinæ. Journ. Agric. Res., 30, pp. 97–154 (1925).
- Newstead, R. A Monograph of the Coccidæ of the British Isles. 2 vols. London, Ray Soc. (1901–03).
- STEINWEDEN, J. B. Bases for the Generic Classification of the Coccidæ. Ann. Entom. Soc. America, 22, pp. 197–243 (1929).

# ORDER HEMÍPTERA

# (HETERÓPTERA; RHYNCHÒTA, part)

Terrestrial or aquatic species ranging from minute to large size; usually more or less flattened or cylindrical; feeding on the juices of plants or animals. Head free, bearing a sucking, inflexed, jointed beak which is usually inserted toward the front end of the head; antennæ with few joints, those of the terrestrial species usually long; in the

aquatic forms very short. Prothorax large, free; mesothorax and metathorax firmly united; scutellum very large. Wings overlapping on the abdomen, the fore pair (hemelytra) tough at the base and membranous apically, the hind pair with large anal field, the venation much reduced and irregular, wings sometimes reduced or absent; legs of variable form, tarsi usually three-jointed, rarely reduced to two, or one joint. Abdomen with ten visible segments, frequently the sternites are larger than the tergites; no cerci. Metamorphosis incomplete. True Bugs.

250

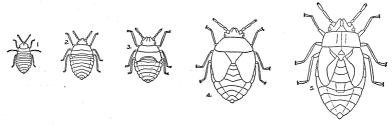


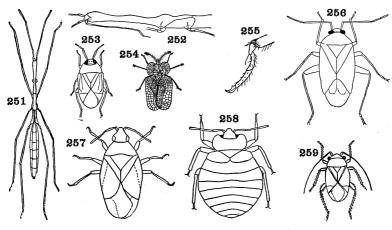
Fig. 250. Hemiptera

Piesma, developmental stages. Five larval instars, 1-5 (Barber) Piesmidæ.

	wings of uniform texture, the clavus, corium and membrane
	confluent; underside of body with silvery, velvety pubescence;
	aquatic, surface-living. Water striders, Jesus-bugs. (Super-
	family GERRÓIDEA)45
4.	Head shorter than thorax including the scutellum; body rarely
	very narrow
	Body linear; head horizontal, as long as the entire thorax and
	widened toward the apex; legs slender; upper wings with corium
	and membrane not separate; wings often absent; antennæ four-
	jointed. Marsh-treaders. ( <b>Hydrómetra</b> (=Limnóbates), cosmop.) (Figs. 251, 252). (LIMNOBÁTIDÆ) <b>HYDROMÉTRIDÆ</b>
-	Antennæ four-jointed, disregarding minute intermediate ring-
5.	
	joints or antenniferous tubercles on the head which are some-
	times present; head not shield-like, the antennæ visible from
	above. (If the antennæ are five-jointed with the basal two
	joints thickened and visible from above, see Hebridæ, couplet
	40)
	Antennæ with five principal joints39
6.	Upper wings more or less lace-like in appearance, the small reticu-
	late cells usually with membranous center; body with reticulate
	sculpturing; tarsi two-jointed; small, more or less flattened
	bugs, less than five mm. in length. Lace-bugs. (Superfamily
	TINGIDÒIDEA) 7
	Upper wings and body not so reticulate; ocelli usually present $8$
7.	Middle lobe of head (tylus) not extending forward as much as the
	side lobes (juga), the head appearing bifid in front; ocelli pres-
	ent; upper wings with the membrane not reticulate but the
	remainder reticulately punctate; pronotum not covering the
	scutellum. (Piésma (Fig. 250))
·	Side lobes of head not prominent; ocelli absent; upper wings en-
	tirely reticulate; pronotum with an angular process extending
	over the scutellum and often with an anterior hood more or
	less covering the head. (Phatnòma, Cantácader, widespr.;
	Acalýpta, Corýthucha (Fig. 254), Galeatus, palæarc., in-
	domal.; Gargáphia, Gelchóssa). (TÍNGIDÆ, TINGÍTIDÆ).
	TINGÍDIDÆ
8.	Tarsal claws devoid of basal pads (arolia), if very rarely the arolia
	are present (Miridæ, Reduvioidea) the meso- and metasternum
	are composite or the front legs are raptorial
	Tarsal claws always provided with arolia; proboscis generally
	four-jointed; meso- and metasternum simple31

9.	Antennæ whip-like, the basal two joints very short, last two joints long and very slender, pilose, the third joint thickened at the base; ocelli present; proboscis three-jointed; tarsi three-jointed; veins of upper wings forming cells; small or minute species. (Superfamily DIPSOCORÒIDEA)
	longer than the third or equal to it, rarely shorter
10.	Head more or less extended horizontally, or slightly bent down; proboscis long; eyes small; front coxal cavities not prominent. (Ceratocómbus, Crescéntius, ind.). (CERATOCÓMBIDÆ, CRYPTOSTEMMÁTIDÆ)DIPSOCÓRIDÆ
	Head transverse, inflexed between the prominent front coxæ;
	costa of fore wings not fractured. (Hypselosòma (=Glypto-
	cómbus))SCHIZOPTÉRIDÆ
11.	Meso- and metasternum composite, formed of more than one
	piece, very rarely the sutures obsolete, in which case the clypeus
	is triangulate (Cimicidæ); cuneus of the fully winged forms
	more or less distinct; hind coxæ hinged (except in a few Miridæ).
	(Superfamily CIMICÒIDEA)
	Meso- and metasternum simple, formed of a single piece; hind coxæ rotating with a ball and socket joint (except in Saldidæ)
12.	Proboscis three-jointed; upper wings, when developed, with an
	embolium; when the wings are vestigial no ocelli are present.
	(Compare also Microphysidæ, couplet 18, some of which have
	three-jointed proboscis, but no embolium)
	Proboscis four-jointed
13.	Metapleuræ without glands; body not broadly oval and thorax
	not flattened; occurring under stones along Atlantic coast of
	Europe. (Aepóphilus) AEPOPHÍLIDÆ
4.4	Metapleuræ with glands
14.	Wings vestigial; clypeus triangular, broader apically; ocelli absent; parasitic on man, bats and birds. Bedbug family. (Cimex
	(C. lectularius, Bedbug) (Figs. 255, 258), Hæmatosiphon). (ACANTHÌIDÆ of authors, CLINOCÓRIDÆ). CIMÍCIDÆ
	Wings usually well developed; sides of clypeus parallel or sub-
	parallel; ocelli present; proboscis three-jointed15
15.	
10.	and thin; proboscis long; gland opening of metathorax small;
	legs long and thin, similar; eyes large and bulging; moderately
	large species. (Velocípeda, indomal.)VELOCIPÉDIDÆ

Membrane with few veins, legs not lengthened; small species. Minute pirate-bugs. (Anthócoris, Tríphleps, cosmop. (T. insidiòsus, Predatory flower-bug)). (Fig. 257). ANTHOCÓRIDÆ



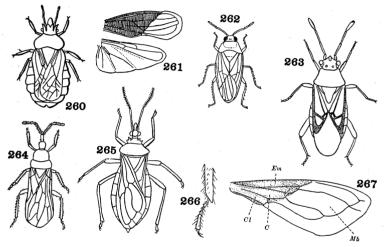
Figs. 251-259. Hemiptera

- 251. **Hydrometra** (Miall) Hydrometridæ.
- 252. **Hydrometra,** profile of anterior portion of body (Hungerford) Hydrometridæ.
- 253. **Isometopus** (Heidemann) Isometopidæ.
- 254. Corythuca Tingididæ.
- 255. Cimex, tip of tibia and tarsus (Eysell) Cimicidæ.
- 256. Lygus (MacGregor) Miridæ.
- 257. Triphleps (MacGregor) Anthocoridæ.
- 258. Cimex (Patton and Cragg) Cimicidæ.
- 259. Halticus (Distant) Miridæ.
- 17. Proboscis with basal joint scarcely longer than wide, not extending backward beyond middle of eyes; membrane of upper wings with a single large quadrangular cell. (Hesperophylum, Termatophylum) ...... TERMATOPHÝLIDÆ
  - Proboscis with basal joint longer than broad, usually reaching beyond hind margin of head; membrane with two, sometimes

	one, small cells near base, rarely with irregular free veins.
	Leaf-bugs, Plant-bugs. (Hálticus (Fig. 259), Psállus, Caló-
	coris, Pœcilocápsus, Irbísia, widespr.; Lygus (Fig. 256),
	(L. praténsis, Tarnished plant-bug), Miris, cosmop.; Cápsus,
	Camptóbrochis, palæarc., indomal.). (CÁPSIDÆ).
	MÍRIDÆ
18.	
10.	joint very small, or three-jointed. (Mallochìola, Cyrtostér-
	num, indomal.; Pachytársus, indomal.). MICROPHÝSIDÆ
	Tarsi three-jointed. (Corticoris, Diphleps, Isometòpus, pa-
10	læarc., indomal. (Fig. 253), Myiómma) ISOMETÓPIDÆ
19.	Front legs not raptorial; prosternum without medial stridulation
	groove; head rarely cylindrical20
	Front legs more or less raptorial; prosternum usually with a median
	transversely striated or granulated stridulation groove in front
	of the front coxæ; pronotum with a transverse groove; head
	cylindrical; proboscis three-jointed, fitted for piercing, rarely
	with an extra very short basal joint, the first joint stout and
	usually curved. (Superfamily REDUVIÒIDEA)26
20.	Ocelli absent; proboscis three-jointed or apparently so when the
	basal joint is minute; body flat, adapted for living under bark.
	(Superfamily ARADÒIDEA)
	Ocelli present, when rarely absent the proboscis is four-jointed and
	the head is not apically widened23
21.	Proboscis arising before the end of the head and lying in a groove
	between the cheeks
	Proboscis terminal, not lying in a ventral groove. (Isodérmus,
	austr., neotrop.; Procympiùtus, austr.)ISODÉRMIDÆ
22.	Head not wide behind the eyes, which are prominent; proboscis
	longer than the head; trochanters very short, fusing with the
	femora; abdominal spiracles placed near the base of the seg-
	ments. (Áradus, Brachyrhýnchus, cosmop.; Eumenòtes,
	indomal. (Fig. 260), Carvéntus, neotrop., indoaustr.).
	ARĀDIDÆ
	Posterior part of head wide, enclosing the eyes, often spinose;
	proboscis rarely longer than the head; trochanters distinct;
	abdominal spiracles remote from the base of the segments.
	(Aneùrus, Mezira, Neuróctenus). (MEZÍRIDÆ).
	DYSODÌIDÆ
23.	Membrane of upper wings destitute of veins, more or less con-
	fluent with the membranous clavus24

Membrane furnished with four or five long closed cells, the clavus more or less distinct. Shore-bugs. (Sálda, cosmop.; Sáldula, Pentácora (Fig. 262)). (ACANTHÌIDÆ of some authors).

SÁLDIDÆ



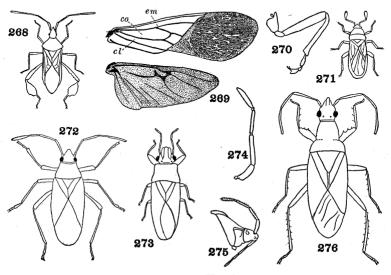
Figs. 260-267. Hemiptera

- 260. Aradus (Howard) Aradidæ.
- 261. Pentatomid bug, wings (Handlirsch) Pentatomidæ.
- 262. Pentacora (Torre Bueno) Saldidæ.
- 263. Corizus (Hambleton) Corizidæ.
- 264. **Henicocephalus** (Maxwell-Lefroy) Henicocephalidæ.
- 265. **Triatoma** (Chagas) Reduviidæ.
- 266. Reduvius, tip of tibia and tarsus (Eysell) Reduviidæ.
- 267. **Triatoma**, wings (Patton and Cragg). Em., embolium; Cl., clavus; C., corium; Mb., membranaceous area. Reduviidæ.
- - Basal two joints of antennæ thicker than the others; body robust, not over 2.5 mm. in length; tarsi two-jointed; head and thorax grooved beneath; body densely clothed with velvety pile.

	(Merragàta; Hèbrus, widespr.). (NÆOGÆIDÆ). (See couplet
0.0	39) <b>HÉBRIDÆ</b> , part
26.	Pronotum divided into three lobes; head constricted at the base
	and behind the eyes, swollen between; upper wings wholly mem-
	branous, with longitudinal veins and a few crossveins; front
	tibiæ swollen; front tarsi one-jointed, hind tarsi two-jointed; minute, delicate species. ( <b>Henicocéphalus</b> , cosmop. (Fig.
	264), Systellóderes). (ENICOCEPHÁLIDÆ).
	HENICOCEPHÁLIDÆ
	Pronotum simple, often large and broad, or long and narrow; head
	not constricted at the base behind the eyes
27.	Antennæ elbowed, slender, filiform or often very thin apically28
	Antennæ short, with the last joint swollen or enlarged; membrane
	with the veins joined, frequently forked and uniting; tarsi two-
	jointed; front legs very stout, raptorial, the front femora greatly
	thickened. (Phýmata (Ambush-bug); Macrocéphalus; Am-
	blythyreus, indomal.; Carcinócoris, indoaustr.). (MACRO-
	CEPHÁLIDÆ)PHYMÁTIDÆ
28.	Prosternum with a cross-striated median stridulation groove;
	proboscis three-jointed
	Prosternum without a stridulation groove; proboscis usually four-
	jointed, rarely three-jointed
29.	Front coxæ short; rather robust species, the body not linear;
	ocelli usually present; front legs raptorial, but not greatly modi-
	fied. Assassin bugs. (Acantháspis, ethiop., indomal.; Apio-
	$merus(A.\ crássipes,\ Bee\ assassin);\ Arilus(A.\ cristàtus,\ Wheel-$
	bug); Melanoléstes (M. picipes, Kissing-bug); Redùvius,
	palæarc., ethiop., indoaustr. (Fig. 266); Sinea, holarc.; Tri-
	átoma (=Conorhinus) widespr. (T. sanguisùga, Big bedbug)
	(Figs. 265–267); Oncocéphalus, Harpáctor, Piràtes, widespr.)
	REDUVÎIDÆ
	Front coxe greatly elongated; body greatly elongated, the middle
	and hind legs long and thin, the front legs highly raptorial;
	ocelli absent. Thread-legged bugs. (Bárce, Ploiària (=Émesa),
	widespr.; Myióphanes, palæarc., indoaustr.; Stenolæmus,
20	Gárdena, widespr.). (EMÉSIDÆ)PLOIARÌIDÆ
30.	Legs slender, the front pair strongly raptorial; tarsi three-jointed;
	membrane of upper wings with more or less distinctly branched
	veins, or with two or three longitudinal cells emitting radiating veins. Damsel-bugs. (Nabis (=Coriscus, = Reduviolus), cos-
	veins. Damsei-bugs. (Nadis (=Coriscus, = Reduviolus), cosmop., Págasa)
	mop., ragasa) NABIDA

	Legs short, tarsi two-jointed; membrane with four free veins.
	(Joppèicus, palæarc.) JOPPEÍCIDÆ
31.	Membrane of upper wings with many longitudinal veins which
	often unite; antennæ inserted well up on the sides of the head;
	ocelli present. (Superfamily COREÒIDEA)
	Membrane usually with a few veins, if many branching veins are
	present the ocelli are absent. (Superfamily LYGÆÒIDEA,
	et al.)
32.	Fourth dorsal segment of the abdomen constricted medially; gland
	openings of the metathorax usually obsolete, if rarely visible,
	placed behind the hind coxal cavities and emitting two diver-
	gent grooves. (Córizus, cosmop. (Fig. 263); Harmóstes;
	Serinètha, ethiop., indomal.)
	Basal margin of fourth and fifth dorsal segments of the abdomen
	usually sinuate in parallel manner; gland openings of meta-
	thorax almost always distinct
33.	Protessing and
	ally reaching behind the insertion of the antennæ; exterior
	margin of hind coxal cavities nearly parallel with axis of the
	body. ( <b>Ánasa</b> (Figs. 269, 270, 274, 275) (A. tristis, Squash-bug);
	Leptoglóssus, widespr. (Fig. 268) (L. phýllopus, Leaf-footed
	bug); Acanthócoris, Stenocéphalus, widespr.; Homæó-
	cerus, ethiop., palæarc., indomal.; Míctis, widespr.; Ano-
	plocnèmis, ethiop., indomal.)
	Head nearly as broad and long as the prothorax, the cheeks
	scarcely extending behind the base of the antennæ; exterior
	margin of the hind coxal cavities more or less transverse.
	(Álydus, Leptocórisa (L. trivittàta, Box-elder bug), widespr.;
	Cùru, neotrop. indomal.; Prôtenor, Stachyocnèmus). (COR-
94	ÍSCIDÆ)         ALÝDIDÆ           Ocelli present
34.	Ocem present
	Ocalli abaant
25	Ocelli absent
35.	Wings when present long and narrow, without distinct veins;
35.	Wings when present long and narrow, without distinct veins; slender bugs with long thin antennæ and abdomen narrowed
35.	Wings when present long and narrow, without distinct veins; slender bugs with long thin antennæ and abdomen narrowed at base. Indoaustral. and neotropic. (Colobathristes, Per-
35.	Wings when present long and narrow, without distinct veins; slender bugs with long thin antennæ and abdomen narrowed at base. Indoaustral. and neotropic. (Colobathristes, Per- ùda, Phænacántha, Málcus)COLOBATHRÍSTIDÆ
	Wings when present long and narrow, without distinct veins; slender bugs with long thin antennæ and abdomen narrowed at base. Indoaustral. and neotropic. (Colobathrístes, Per-ùda, Phænacántha, Málcus)COLOBATHRÍSTIDÆ Membrane with veins
<ul><li>35.</li><li>36.</li></ul>	Wings when present long and narrow, without distinct veins; slender bugs with long thin antennæ and abdomen narrowed at base. Indoaustral. and neotropic. (Colobathrístes, Per-ùda, Phænacántha, Málcus)COLOBATHRÍSTIDÆ Membrane with veins
	Wings when present long and narrow, without distinct veins; slender bugs with long thin antennæ and abdomen narrowed at base. Indoaustral. and neotropic. (Colobathrístes, Per-ùda, Phænacántha, Málcus)COLOBATHRÍSTIDÆ Membrane with veins
	Wings when present long and narrow, without distinct veins; slender bugs with long thin antennæ and abdomen narrowed at base. Indoaustral. and neotropic. (Colobathrístes, Per-ùda, Phænacántha, Málcus)COLOBATHRÍSTIDÆ Membrane with veins

37. Veins of membrane usually four or five in number and not forming anteapical cells. (**Geócoris**, **Nýsius**, cosmop. (False chinchbug); **Graptostèthus**, palæarc., ethiop., indomal.; **Oncopéltus**, widespr.; **Lygæus**, **Ligyrócoris** (Fig. 276), **Lygæosòma**, widespr.; **Blíssus** (B. leucópterus, Chinch-bug (Fig. 271))). (GEO-CÓRIDÆ, MYODÓCHIDÆ).



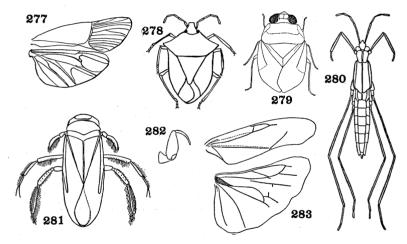
Figs. 268–276. Hemiptera

- 268. Leptoglossus (Chittenden) Coreidæ.
- 269. Anasa, wings (Tower) Coreidæ.
- 270. Anasa, leg (Tower) Coreidæ.
- 271. Blissus (Webster) Lygæidæ.
- 272. Dysdercus (Barber) Pyrrhocoridæ.
- 273. **Xylastodoris** (Barber) Thaumastocoridæ.
- 274. Anasa, antenna (Tower) Coreidæ.
- 275. Anasa, lateral view of prothorax and head (Hyatt and Arms) Coreidæ.
- 276. Ligyrocoris (Barber) Lygæidæ.

38. Membrane of upper wings with two large basal cells which emit seven or eight branching veins; stout bugs of moderate size. (**Dysdércus**, widespr. (Fig. 272) (Cotton stainers); **Physo-**

	pélta, widespr.; Euryophthálmus; Pyrrhócoris, palæarc.,
	ind.; <b>Éctatops, Antílochus,</b> ethiop., indoaustr.).
	PYRRHOCÓRIDÆ
	Membrane with few veins forming one or two basal cells. (See
	couplet 17) MÍRIDÆ, part
39.	Upper wings with the clavus membranous and confluent with the
00.	membrane which is devoid of veins; head and thorax grooved
	beneath; antennæ with the two basal joints stouter than the
	others; tarsi two-jointed; small, semiaquatic bugs. (See couplet
	25). (Hèbrus (= Neogœus), widespr.)
	Upper wings with the clavus noticeably heavier than the mem-
	brane; antennæ with the first joint thickened, the second joint
	slender; head more or less expanded, the side margins acute in
	front of the eyes and thickened above the base of the antennæ;
	ocelli present; scutellum large or very large; terrestrial. (Super-
	family SCUTELLERÒIDEA). (PENTATOMÒIDEA)40
40.	Scutellum excessively large, U-shaped and convex, covering the
	greater part of the abdomen; opaque part of the corium much
	narrowed toward the apex41
	Scutellum nearly always narrowed behind, more or less triangular;
	opaque part of the corium subtriangular, broad apically 44
41.	
	Tibiæ with two or more rows of distinct spines. Negro-bugs.
	(Corimelæna (= $Thyre \acute{o}coris$ )). ( $THYREOC\acute{O}RID\cancel{E}$ ).
er*	CORIMELÆNIDÆ
19	Fore wings about twice as long as the abdomen, folded at the
12.	middle and at rest tucked under the scutellum; tarsi two-jointed.
	(Brachýplatys, ethiop., indoaustr.; Coptosòma, old world;
	Platáspis, palæarc.). (COPTOSÒMIDÆ, PLATÁSPIDÆ).
	PLATASPÍDIDÆ
	Fore wings of normal length, not folded; tarsi nearly always three-
	jointed
43.	Sides of the prothorax without a strong tooth or lobe in front of
	the humeral angles and another on the front angles; hind wings
	with a heavy, abrupt, spur-like vein (hamus). Shield-back bugs.
	(Eurygáster, holarc.; Homæmus, widespr.; Scutéllera,
	Chrysócoris, Pœcilócoris, indomal.) SCUTELLÉRIDÆ
	Sides of the thorax with a prominent tooth or lobe in front of both
	the humeral and the front angles; eyes protuberant; hind wings
	with no hamus. (Arctócoris, palæarc.; Oncozýgia, Pòdops,
	widown (CD ADUOSOM ÁTID Æ)

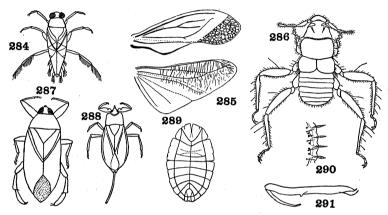
Tibiæ unarmed, or at most with weak spines, front legs not fossorial; veins of membrane extending from a vein which arises near the inner basal angle and lies nearly parallel with the edges of the corium. Stink-bugs. (Brochýmena, Cosmopépla, Euschístus (Fig. 278), Murgántia (M. histriónica, Harlequin cabbage-bug); Pentátoma, Períbalus, Podisus, Catacántha (Fig. 277)) ...... PENTATÓMIDÆ



Figs. 277-284. Hemiptera

- 277. Catacantha, wings (Kirkaldy) Pentatomidæ.
- 278. Euschistus. Pentatomidæ.
- 279. Ochterus (Garman) Ochteridæ.
- 280. Gerris (Miall) Gerridæ.
- 281. Corixa (Miall) Corixidæ.
- 282. Corixa, font leg (Kolbe) Corixidæ.
- 283. Corixa, wings (Handlirsch) Corixidæ.
- 45. Hind femora extending much beyond the apex of the abdomen, the posterior pairs of legs arising close together and very distant from the front pair; proboscis four-jointed, but the first joint short. (Gérris, cosmop. (Fig. 280); Halóbates, tropicopol.;

	Rheumatóbates; Onychótrechus, Chimarrhómetra, indo-
	mal.). (HYDROBÁTIDÆ)
	Hind femora not extending much beyond the apex of the abdomen;
	middle legs about midway between the front and hind pairs
	(except Rhagovelia); proboscis three-jointed. Broad-shouldered
	water-striders. (Microvèlia, cosmop.; Rhagovèlia, cosmop.;
	Vèlia, Am., palæarc.)VELÌIDÆ
<b>46.</b>	Head articulated with the thorax as usual or at most partly fused
	with it; tarsi with more than a single joint
	Head completely fused with the thorax, the boundary more or less
	indicated by a shallow impression; antennæ one- or two-jointed;
	eyes located rather dorsally; proboscis four-jointed; no distinct
	venation; anterior tarsi one-jointed, hind tarsi two-jointed, all
	tarsi with two claws; male genitalia strongly asymmetrical. <b>HELOTRÉPHIDÆ</b>
	Two subfamilies, probably deserving family rank, may be dis-
	tinguished as follows:
я	Antennæ two-jointed; body globose, shining; suture between head
	and pronotum visible as a fine impressed line; scutellum rounded
	on the sides, but little wider than long. (Helótrephes, in-
	domal.) HELOTREPHÎNÆ
	Antennæ composed of a single flat, disk-like joint; suture between
	head and pronotum completely obliterated, except in the
	nymph; scutellum elongate, acutely triangular toward apex. (Idiócoris, Páskia, ethiop.)
47.	Front tarsi of the usual form
	Front tarsi consisting of a single spatulate joint bearing a leaf-like
	claw; body flattened above; head overlapping the pronotum;
	proboscis very short, hidden, one- or two-jointed; middle legs
	long, hind legs formed for swimming; hind tarsi with indistinct bristle-like claws. Water boatmen. (Coríxa, cosmop.; Micro-
	nécta, palæarc., indomal.). (Figs. 281, 282, 283) CORÍXIDÆ
48.	Upper wings of leathery consistency, with the clavus, corium and
40.	membrane developed; legs often modified for swimming or
	grasping
	Upper wings transparent, the corium and membrane not sepa-
	rated, with longitudinal veins and crossveins that enclose
	numerous cells. Austr. and neotrop. ( <b>Pelorídium</b> ). (See page
	134PELORIDÌIDÆ
49.	Ocelli present; proboscis four-jointed; shore-living species50
	Ocelli absent; proboscis usually three-jointed; aquatic species51



Figs. 284–291. Hemiptera

- 284. Notonecta (Miall) Notonectidæ.
- 285. Nepa, wings (Handlirsch) Nepidæ.
- 286. Polyctenes (Westwood) Polyctenidæ.
- 287. Lethocerus (Smith) Belostomatidæ.
- 288. Nepa (Miall) Nepidæ.
- 289. Termitaradus (Myers) Termitaphididæ.
- 290. **Termitaradus,** marginal lobe (Myers) Termitaphididæ.
- 291. Ranatra, apical portion of front leg (Hungerford) Nepidæ.
- 52. Posterior tibiæ and tarsi ciliate; abdomen with a median carinate ridge below; beak four-jointed; eyes large. (Notonécta, cosmop.; Ánisops, widespr.; Buénoa, Am. (Fig. 284)).

NOTONÉCTIDÆ

- - Membrane without veins; beak without labial palpi; hind coxæ hinged; hind tibiæ slender, with small spines. Water creepers. (Heleócoris, widespr.; Cheirochèla, indomal.; Ambrysus, Naucoris, Pelócoris).

## NAUCÓRIDÆ and APHELOCHÍRIDÆ

- 54. Hind coxæ hinged, hind legs fitted for swimming, posterior tibiæ flattened and fringed, hind femora usually sulcate; tip of abdomen with two short, flat, retractile appendages. Giant waterbugs, Electric light bugs, Toe-biters. (Belóstoma, Benàcus, Ábedus, Lethócerus, widespr. (Fig. 287); Zàitha, Sphæródema) ...... BELOSTOMÁTIDÆ
  - Hind coxæ globular, rotating; hind legs formed for walking, not flattened; apical appendages of abdomen long and slender, not retractile, forming a respiratory siphon. Water-scorpions. (Laccótrephes, palæarc., indoaustr.; Cercomètus, indomal. (Figs. 285-288); Nèpa, Ránatra, cosmop. (Fig. 291)).

## NÉPIDÆ

## LITERATURE ON HEMIPTERA (GENERAL)

(See also general list on Homoptera and Hemiptera, p. 134)

- Blatchley, W. S. Heteroptera of Eastern North America. Indianapolis, Nature Pub. Co., 1116 pp. (1926).
- Gulde, J. Die Wanzen (Hemiptera-Heteroptera) der Umgebung von Frankfurt a. M. Abh. Senckenberg. naturf. Ges., **37**, pp. 329–503 (1921).
- Myers, J. G. The order Hemiptera in New Zealand with special reference to its biological and economic aspects. New Zealand Journ. Sci. Tech., 5, pp. 1-12 (1922).
- MYERS, J. G. and CHINA, W. E. A List of New Zealand Heteroptera. Ann. Mag. Nat. Hist. (10) 1, pp. 378–388 (1928).

- Parshley, H. M. List of Hemiptera of New England. Occ. Pap. Boston Soc. Nat. Hist., 7, pp. 1-125 (1920).
  - Bibliography of North American Hemiptera-Heteroptera. 252 pp. Smith College, Northampton, Mass. (1925).
- Puton, A. Synopsis des Hémiptères-Hétéroptères de France, 4 vols. (1878–81).
- Saunders, E. The Hemiptera-Heteroptera of the British Isles. 350 pp. London (1892).
- Seabra, A. F. Sinopse dos Hemípteros Heterópteros de Portugal. Mem. Estud. Mus. Zool. Univ. Coimbra (1), No. 1, pp. 1–67 (1924–25).
- STICHEL, W. Illustrierte Bestimmungstabellen der deutschen Wanzen. Berlin (1927).
- Taeuber, H. W. Beiträge zur Kenntnis der Heteropteren-Fauna der Philippinen. Konowia, 6, pp. 165–201 (1927); 8, pp. 194–233 (1929).

#### SUBORDER GYMNOCERATA

- ATKINSON, E. T. Catalogue of the Capsidæ. Suppl. Journ. Roy. Asiatic Soc., 58, pt. 2, pp. 25–200 (1889).
- BARBER, H. G. Synoptic Keys to the Lygaidæ of the United States. Psyche, 24, pp. 128-135 (1917); 25, pp. 71-88 (1918).
  - Tingitoidea of New Jersey. Circ. New Jersey Dept. Agric. Bur. Statistics, No. 54, 24 pp. (1922).
- Bergroth, E. Gerridæ, Subfamily Halobatinæ. Ohio Naturalist, 8, pp. 371–382 (1908).
  - Isometopidæ of New Zealand. Notulæ Entom., 4, pp. 3–9 (1924).
- Bruner, S. C. Sinopsis de los Redúvidos de Cuba. Mem. Soc. Cubana Hist. Nat., 7, pp. 65–82 (1926).
- China, W. E. and Myers, J. G. Classification of the Cimicoid Families. Ann. Mag. Nat. Hist. (10), 3, pp. 97–125 (1929).
- Costa Lima, A. Insectos da Familia Polyctenidæ. Arch. Esp. Agric. Med. Vet. Nictheroy, 4, pp. 61–71 (1920).
- Deay, H. O. The Coreidæ of Kansas. Kansas Univ. Sci. Bull., **18**, pp. 371–415 (1928).
- DISTANT, W. L. Revision of Pentatomidæ in the Hope collection. Proc. Zoöl. Soc. London, 1900, pp. 807–825 (1900).
- ESAKI, T. The Halobatine in the Hungarian National Museum. Ann. Mus. Nat. Hungarici, 23, pp. 117-164 (1926).
- Esaki, T. and China, W. E. A New Family (Helotrephidæ) of Hemiptera. Trans. London Entom. Soc., **75**, pp. 279–295 (1927).
  - Monograph of the Helotrephinæ. Eos, 4, pp. 129–172 (1928).
- Fracker, S. B. Reduviidæ of North America. Proc. Iowa Acad. Sci., 19, pp. 217-247 (1912).
  - Alydinæ of the United States. Ann. Entom. Soc. America, 11, pp. 255–280 (1918).

Review of North American Coreini. Ann. Entom. Soc. America, 16, pp. 165–173 (1923).

Gibson, E. H. Isometopidæ of North America. Bull. Brooklyn Entom. Soc., 12, pp. 73–77 (1917).

HANDLIRSCH, A. Monographie der Phymatiden. Ann. Hofmus. Wien, 12, pp. 127–230 (1897).

HARRIS, H. M. A Monographic Study of the Hemipterous Family Nabidæ in North America. Entom. Americana. 9, pp. 1–90 (1928).

HART, C. A. Pentatomoidea of Illinois with keys to nearctic genera. Ill. Nat. Hist. Surv. Bull., 13, pp. 157–223 (1919).

Horváth, G. Monographia Colobathristidarum. Ann. Mus. Hist. Nat. Hungarici, 2, pp. 117–172 (1904).

Synopsis Tingitidarum regionis Palæarcticæ. Ann. Mus. Nat. Hungarici, 4, pp. 1–118 (1906).

Revision des Leptopodides. Ann. Mus. Nat. Hungarici, 9, pp. 358–370 (1911).

Revision of the American Cimicidæ. Ann. Mus. Nat. Hungarici, 10, pp. 257–262 (1912).

Monographie des Mésoveliides. Ann. Mus. Nat. Hungarici, 13, pp. 535–556 (1915).

Analecta ad cognitionem Cydnidarum. Ann. Mus. Nat. Hungarici, 17, pp. 205–273 (1919).

Description of a New Bat-Bug from India. Rec. Indian Mus., 27, pp. 191–192 (1925).

Mesoveliidæ. Gen. Cat. Hemiptera, pt. 2, 24 pp., Northampton, Mass., Smith College (1929).

HUSSEY, R. F. Pyrrochoridæ. Gen. Cat. Hemiptera, 144 pp. Northampton, Mass., Smith College (1929).

JOHANNSEN, O. A. North American Henicocephalidæ. Psyche, 16, pp. 1–4 (1909).
JORDAN, K. Morphology and Systematics of the Polyctenidæ. Novit. Zool.,
18, pp. 555-579 (1912).

The American Polyctenidæ. Ectoparasites, 1, pp. 204–215 (1922).

Kiritschenko, A. Coreidæ, Coreinæ. Fauna Russie Hemiptera, 6, pt. 2, pp. 1–395 (1916) (In Russian).

Kirkaldy, G. W. Catalogue of Hemiptera. Pentatomidæ (Cimicidæ). 392 pp. Berlin (1909).

Generic Key to Phyllocephalinæ, Phlocinæ and Dinidorinæ (Pentatomidæ). Canadian Entom., **45**, pp. 81–84 (1913).

KNIGHT, H. H. A Revision of the Genus Lygus in America North of Mexico. Bull. Cornell Agr. Exp. Sta., No. 391, pp. 555-645 (1917).

Key to Subfamilies of Miridæ. Journ. New York Entom. Soc., 26, pp. 40-44 (1918).

Monograph of the North American species of Deræocoris (Miridæ). 18th Rept. St. Entom. Minnesota for 1920, pp. 76–210. Reprinted as Tech. Bull. I, Univ. of Minnesota Agr. Exp. Sta. (1921).

MCATEE, W. L. Key to Nearctic Geocorinæ (Lygæidæ). Proc. Biol. Soc. Washington, 14, p. 102 (1914).

Key to the Nearctic species of Paracalocoris (Miridæ). Ann. Entom. Soc. America. 9, pp. 366–390 (1916).

Key to Piesmidæ. Bull. Brooklyn Entom. Soc., **14**, pp. 80–93 (1919). Key to Nearctic Berytidæ. Journ. New York Entom. Soc., **27**, pp. 79–92 (1919).

MCATEE, W. L. and Malloch, J. R. Revision of Cryptostemmatidæ in the United States National Museum. Proc. U. S. Nat. Mus., 67, Art. 13, 42 pp. (1925).

Revision of the Ploiariinæ (Reduviidæ). Proc. U. S. Nat. Mus., 67, Art. 1, 135 pp. (1925).

Morrison, H. On three apparently new species of Termitaphis. Zoologica, New York, 3, pp. 403–408 (1923).

Myers, J. G. On the Systematic Position of the Family Termitaphididæ. Psyche, **31**, pp. 259–278 (1924).

MYERS, J. G. and CHINA, W. E. The systematic position of the Peloridiidæ. Ann. Mag. Nat. Hist. (10), 3, pp. 282–294 (1930).

OSBORN, H. and DRAKE, C. J. The Tingitoidea of Ohio. Ohio State Univ. Bull., 20, pp. 217–251 (1916).

Parshley, H. M. Synopsis of the Pentatomida of New England. Psyche, 22, pp. 170-177 (1915).

American species of Aradus. Trans. American Entom. Soc., 47, pp. 1–106 (1921).

Pennington, M. S. Notas sobre Coréidos Argentinos. Physis, Buenos Aires, 5, pp. 125–170 (1922).

Pinto, C. Ensaio Monogróphico dos Reduvideos hæmatóphagos o "Barbieros." 118 pp. Rio de Janeiro (1925).

Classification des genres de la famille Triatomidæ. Bol. Biol. São Paulo, fasc. 8, pp. 103–114 (1927).

Poppius, B. R. Beiträge zur Kenntnis der Anthocoriden. Acta Soc. Sci. Fennica, 37, No. 9, pp. 1–43 (1909).

Zur Kenntnis der Cylapinæ. Acta Soc. Sci. Fennica, 37, No. 4, pp. 1–45 (1909).

Beiträge zur Miridenfauna Australiens. Öfv. vet. Soc. Förh., **53**, No. 3, pp. 1–16 (1911).

Beiträge zur Kenntnis der Miriden von Ceylon. Öfvers Vet. Soc. Helsingfors, **53**, No. 2, pp. 1–36 (1911).

Miriden der æthiopischen Region. Part I., Acta Soc. Sci. Fennica, **41**, pp. 1–203 (1912); Part II., ibid., **44**, pp. 1–138 (1914).

Readio, P. A. Biology of the Reduviidæ of America North of Mexico. Sci. Bull. Univ. of Kansas, 17, pp. 1–248 (1927).

Reuter, O. M. Monographia anthocoridarum orbis terrestris. Acta Soc. Sci. Fennica, 14, pp. 555–758. Also published separately (1885).

Monographia Ceratocombidarum orbis terrestris. Acta Soc. Sci. Fennica, **19**, pp. 1–28 (1891).

Die Familie der Bettwanzen. Zeits. wiss. Insektenbiol., 9, p. 251 etc. (several parts) (1913).

Monographia Nabidarum orbis terrestris. Acta Soc. Sci. Fennica, 37, pp. 1–62 (1909).

Neue Beiträge zur Phylogenie und System der Miriden. Acta Soc. Sci. Flor. Faun. Fennica, **37**, No. 3, pp. 1–167 (1910).

REUTER, O. M. and Poppius, B. R. Zur Kenntnis der Termatophyliden. Öfv. Finska Vet. Soc. Förh., **54**A, pp. 1–17 (1912).

Schouteden, H. Pentatomidæ, Scutellarinæ. Gen. Insectorum, fasc. 24, 100 pp. (1904).

Pentatomidæ, Graphosomatinæ. Gen. Insectorum, fasc. 30, 46 pp. (1905). Pentatomidæ, Aphylinæ. Gen. Insectorum, fasc. 47, 4 pp. (1906).

Pentatomidæ, Asopinæ = Amyoteinæ. Gen. Insectorum, fasc. 52, 82 pp. (1907).

Pentatomidæ, Dinidorinæ. Gen. Insectorum, fasc. 153, 19 pp. (1913).

SIGNORET, V. Revision du Groupe des Cydnides. Ann. Soc. Entom. France, 1-3 (various parts) (1881-83).

Stoner, D. The Scutelleroidea of Iowa. Univ. Iowa Studies Nat. Hist., 8, No. 4, pp. 1–140 (1920).

Van Duzee, E. P. Keys to Genera of North America Miridæ. Univ. California Pubs., Ent., 1, pp. 199–216 (1916).

List of Pentatomidæ of America North of Mexico. Trans. American Entom. Soc., **30**, pp. 1–80 (1904).

Watson, S. A. The Miridæ of Ohio. Ohio State Univ. Bull., 33, 44 pp. (1928). White, F. B. Report on the Pelagic Hemiptera. Challenger Exped., Zoöl., 7,

pt. 19, p. 82 (1883).

ZIMMER, J. F. The Pentatomidæ of Nebraska. Univ. Nebraska, Contr. Dept. Entom., No. 4, pp. 1–33 (1912).

#### SUBORDER CRYPTOCERATA

Ferrari, E. Die Hemipterengattung Nepa. Ann. Hofmus. Wien, **3**, pp. 161–194 (1888).

Hale, H. M. Two New Hemiptera from New South Wales. Proc. Linn. Soc. New South Wales, 49, p. 462 (1924).

Studies in Australian Aquatic Hemiptera. Rec. South Australian Mus., **3**, pp. 195–217 (1926).

Hueber, T. Deutsche Wasserwanzen. Jahrb. Ver. Vaterl. Naturk. Würtemburg, **61**, pp. 91–176 (1905).

Huncerford, H. B. The Biology and Ecology of Aquatic and Semiaquatic Hemiptera. Univ. Kansas Sci. Bull., 11, pp. 3–265 (1919).

The Nepidæ of North America north of Mexico. Kansas Univ. Sci. Bull., 14, pp. 425–469 (1922).

Revision of the Notonectide and Corixide of South Africa. Ann. South African Mus., 25, pp. 375–474 (1929).

KIRKALDY, G. W. A guide to the study of British waterbugs (aquatic Rhynchota). Entom., 31, pp. 177–180 (1898); continued in vols. 32, 33, 38, 39, 41 (1899–1908).

Revision of Notonectidæ. Trans. Entom. Soc. London, 1897, pp. 393–426 (1897) and Wiener Entom. Zeitg., 23, pp. 111–135 (1904).

Notes on Corixidæ. No. I. Canadian Entom., **50**, pp. 117–120 (1908). MAYR, G. L. Die Belostomiden. Verh. zool.-bot. Ges. Wien, **21**, pp. 399–440 (1871).

MELIN, D. Neotropical Gelastocoridæ (partial revision). Zool. Bidrag., 12, pp. 151-198 (1930).

Reuter, O. M. Hemiptera Gymnocerata Europæ. 5 vols. (1878–96).

TORRE-BUENO, J. R. The Genus Notonecta in North America. Journ. New York Entom. Soc., 13, pp. 143-167 (1905).

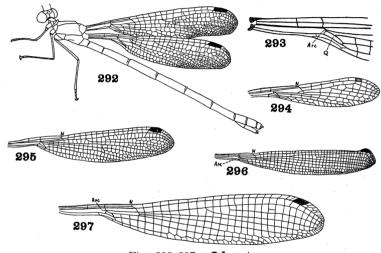
## ORDER ODONÀTA

# (LIBELLULÒIDEA, PARANEURÓPTERA)

Slender predatory insects, usually of large or very large size and usually strong fliers; head mobile, eyes large, three ocelli; antennæ minute, four- to seven-jointed; mouth inferior, mandibles strong, maxillary palpi one-jointed, labial palpi two-jointed; prothorax small but free, meso- and metathorax fused, oblique; cerci one-jointed; legs not large, similar, usually armed with spines, placed far forward, tarsi three-jointed; wings four, nearly alike, elongate, membranous, netveined, not folded, with characteristic nodus, arculus and triangle (see Figs. 298, 299). Abdomen long and narrow, cylindrical or flattened, sometimes clubbed at apex; ovipositor sometimes complete; male sexual apparatus attached to the second sternite. Metamorphosis considerable, the nymphs aquatic, no resting pupal stage. Damsel flies, Dragon flies.

## Adults

- - Discoidal cell divided into two cells, a triangle and supratriangle; wings without a distinct narrowed stalk-like portion at the base; hind wings noticeably different from the fore ones in shape and venation; nodus at or beyond the middle of the wings, at least



- Figs. 292–297. **Odonata**
- 292. Lestes (Garman) Lestidæ.
- 293. Chromagrion, base of fore wing (Garman) Cœnagriidæ.
- 294. **Hemiphlebia**, wing (Needham) Hemiphlebiidæ.
- 295. Megalestes, wing (Needham) Synlestidæ.
- $296. \ \ \textbf{Mecistogaster,} \ wing \ (Needham) \ Pseudostigmatidæ.$
- 297. Nesobasis, wing (Tillyard) Cœnagriidæ.

Eyes less noticeably projecting laterally; separated by less than their width (female) or almost contiguous above (male); mesothorax wider than long; abdomen distinctly swollen apically. (Epiophlèbia, Ind., Japan). (Suborder ANISOZYGÓPTERA).

## EPIOPHLEBÌID*Æ*

3. Only two antenodal crossveins (except *Thaumatolestes* and *Neurolestes*) (Fig. 295); arculus at least as near to the nodus as to the

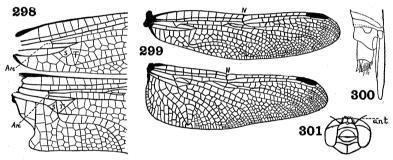
	base of the wing; wings strongly petiolate; mesopleura not
	divided by an oblique suture. (Superfamily CŒNAGRI-
	OIDEA (=CENAGRIONOIDEA))
	Five or more antenodal crossveins; arculus farther from the nodus
	than from the base of the wing (equidistant in some Libella-
	ginidæ); wings less strongly petiolate; frequently with metallic
	coloration; mesopleura with a distinct oblique suture extending
	from the root of the wing toward the middle coxa. (Superfamily
	AGRIÒIDEA (= $AGRIONÒIDEA$ ))
4.	Second branch of cubitus strongly arched upwards at its base,
	just beyond the apex of the discoidal cell; discoidal cell very
	narrow, pointed apically; two, rarely three antenodal cross-
	veins; nodus at the basal third of the wing. (Fig. 295). (Syn-
	léstes, Austr.; Chloroléstes, ethiop.; Periléstes, neotrop.;
	Megaléstes) SYNLÉSTIDÆ
	Second branch of cubitus straight, or very weakly curved up-
	wards at the base. (Fig. 294)5
<b>5</b> .	Postnodal crossveins more or less completely in line with the
	crossveins beneath them (i.e. extending as straight lines from
	the costa to the first branch of the radius)6
	Postnodal crossveins not at all in line with the crossveins beneath
	them; discoidal cell open basally. Small, slender, metallic green
	species, with short wings. (Fig. 294). (Hemiphlèbia, Austr.).
	HEMIPHLEBÌIDÆ
6.	Wing without any supplemental sectors extending inwards from
	the apical part of the hind margin7
	Wing with one or more supplemental sectors8
7.	Second branch of cubitus long, extending to beyond the middle
	of the wing (except Chlorocnemis); first anal vein usually
	present. (Fig. 297). (Comágrion (= Ágrion, auctt.), wide-
	spr.; Ischnùra, Enallágma, cosmop.; Pseudágrion, indo-
	austr.; Árgia, Am.; Nehallènia, holarc., neotrop.; Chloro-
	cnèmis, ethiop.; Platycnèmis, palæarc.). (Including PLATY-
	CNÉMIDÆ). (CŒNAGRIÓNIDÆ)CŒNAGRÌIDÆ
	Second branch of cubitus short, entering the wing margin before
	the middle of the wing; first anal vein usually absent. (Noso-
	stícta, Neostícta, Austr.; Protoneura, neotrop.).
	PROTONEŪRIDÆ
8.	Last two branches of radial sector arising nearer to the nodus
	than to the arculus; apical angle of discoidal cell obtuse, rarely
	more or less acute

	Last two branches of radial sector (R <sub>4+5</sub> and IR <sub>3</sub> ) arising near
	the base of the wing, closer to the arculus than to the nodus
	(Fig. 292); discoidal cell drawn out to a very acute angle at
	apex. (Léstes, cosmop.; Sympýcna, palæarc.; Archiléstes,
	Am.; Austroléstes, austr.) LÉSTIDÆ
9.	Nodus at the basal third or fourth of the wing; pterostigma regu-
	lar in shape, short or long. (Megapodágrion, neotrop.; Argio-
	léstes, Lestòidea, austr.; Podoléstes, austromal.).
	MEGAPODA GRÌIDÆ
	Nodus at the basal sixth or seventh of the wing; pterostigma weak,
	absent or of abnormal shape. (Fig. 296). (Pseudostígma,
	Mecistogáster, Megalóprepus, Microstígma, Thaumato-
	neura, neotrop.)PSEUDOSTIGMÁTIDÆ
10.	Wings without a distinct petiole at the base, the posterior margin
	without any angulation basally near the arculus; antenodal
•	crossveins numerous between the costa, subcosta and radius. 11
	Wings with a distinct petiole at the base, marked off by an angula-
	tion of the posterior margin basally near the arculus; antenodal
	crossveins few in number (seven or less) and sometimes absent
	between the subcosta and radius. (Amphipteryx, neotrop.;
	Diphlèbia, neotrop., austr.; Devadátta, indomal.).
	AMPHIPTER Ý GIDÆ
11.	Sectors of arculus arising at or near the middle or below the
	middle of the arculus; discoidal cell with the basal side not
	longer than the apical one
	Sectors of arculus arising at the extreme upper end of the arculus
	and thus attached to the radius; discoidal cell irregular, the
	basal side longer than the apical one. (Thore, Eùthore, Chal-
	cópteryx, neotrop.)POLYTHÓRIDÆ
12.	,
	pterostigma long and regular
	Sectors of the arculus arising from the lower third of the arculus;
	pterostigma frequently imperfect or obsolete in the male;
	whole wing, including the anal field, closely reticulate. (Agrion
	(=Calópteryx) holarc.; Hæterina, Am.; Pentaphlèbia,
	Sàpho, ethiop.; Vestàlis, indomal.; Neuróbasis, indoaustr.).
10	(CALOPTERÝGIDÆ)
13.	responding; arculus much closer to the base of the wing than
	to the nodus. ( <b>Pseudophæa</b> , indomal.; <b>Dysphæa</b> , mal.;
	Anisopleùra, ind.) EPALLÁGIDÆ

Antenodal crossveins of the first and second series not corresponding beyond the arculus; arculus often nearly as close to the base of the wing as to the nodus. (Libellàgo, ethiop.; Rhinocỳpha, indomal.; Dictèrias, Heliócharis, neotrop.).

## LIBELLAGÍNIDÆ

14. Antenodal crossveins of the first and second series (i.e. above and below the subcostal vein) not corresponding or continuous, except for two greatly thickened ones that extend straight across from the costa to the radius; triangles in fore and hind wings alike or closely similar in shape and occupying the same position with reference to the arculus in both wings; labial palpi two-jointed. (Fig. 298). (Superfamily ÆSCHNOIDEA)... 15



Figs. 298-301. Odonata

- 298. Petalura, base of wings (Needham) Petaluridæ.
- 299. Libellula, wings. Libellulidæ.
- 300. Cordulegaster, tip of abdomen of female (Tillyard) Cordulegastridæ.
- 301. Libellula, head; ant., antenna (Hyatt and Arms) Libellulidæ.

- - Ovipositor reduced to a pair of valves attached to the eighth segment; only one oblique crossvein between the third and fourth branches of the radius; smaller species. (Gómphus, Ophiogómphus, holarc.; Lindènia, palæarc.; Erpetogómphus, Am.; Ictìnus, indoaustr.; Austrogómphus, austr.).

#### GÓMPHIDÆ

- 17. Third branch of radial vein (nodal sector) strongly arched below the pterostigma; a thickened crossvein (brace vein) present just below the base of the pterostigma; eyes almost always contiguous for a long distance above. (Anax, cosmop.; Æschna, widespr.; Gynacántha, neotrop., ethiop., indoaustr.; Basiæschna, nearc.; Austrophlèbia, austr.)..... ÆSCHNIDÆ
  - Third branch of radial vein gently curved; no brace vein; eyes just touching or very slightly separated above; ovipositor long. (Fig. 300). (Cordulegáster, holarc.; Anotogáster, palæarc., indomal.; Allogáster, indomal.)....CORDULEGÁSTRIDÆ
- 18. Triangle in fore wing not shortened, its upper side much more than half as long as its basal side; anal angle of wing almost always angulated in the male; posterior margin of eye slightly lobed at middle. (Cordùlia, holarc.; Somatochlòra, holarc., austr.; Macromia, widespr.; Tetragoneùria, nearc.).

## CORDULIIDÆ

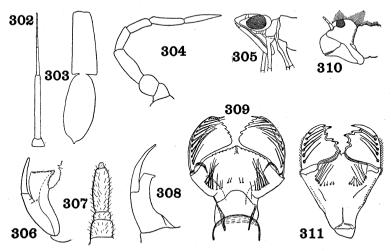
Triangle in fore wing greatly shortened, its upper side usually less than half as long as its basal side; anal angle of wing rounded in both sexes; hind margin of eye evenly rounded. (Figs. 299, 301). (Libéllula, Leucorrhínia, Sympètrum, holaic.; Pántala, cosmop.; Crocóthemis, ethiop., indoaustr.; Tràmea, Am.; Celíthemis, nearc.) . . . . . . . LIBELLÜLIDÆ

## Nymphs 1

1. Body slender, bearing three long tracheal gills attached to the tip of the abdomen; these are usually leaf-like in shape and traversed by conspicuous tracheal tubes (rarely much reduced

<sup>&</sup>lt;sup>1</sup> The following key will serve as an aid in determining the families, but must not be relied on as infallible.

	in some Cœnagriidæ); lateral abdominal gills usually absent. (Suborder ZYGÓPTERA)
	Body stout, the abdomen not narrow; without tracheal gills attached to the tip of the abdomen; the caudal gills concealed in an enlargement of the rectum; tip of abdomen bearing three spine-like or triangular processes
2.	Gizzard with four to eight dental folds. (Suborder ANISÓP—TERA)
	Gizzard with sixteen dental folds; very rare Indian and Japanese species. (Suborder ANISOZYGÓPTERA).
	EPIOPHLEBÌID <i>Æ</i>
3.	Mask bearing at least two pairs, and usually with numerous stiff bristles or setæ
	Mask destitute of setæ both on the mentum and the lateral lobes. 6
4.	Median lobe of mask incised; lateral lobes deeply cleft; caudal
	gills with the secondary tracheæ lying at right angles to the
	gill axis; legs long LÉSTIDÆ
	Median lobe of mask projecting, not incised; legs short or moder-
	ate in length5
<b>5.</b> ,	Caudal gills stalked, with pointed, leaf-like apical portion.
	PSEUDOSTIGMÁTIDÆ
	Caudal gills usually slender, lamellate, held in a vertical plane,
	not clearly stalked, often distinctly constricted.
	CŒNAGRÌIDÆ
6.	Second joint of antennæ greatly lengthened, as long as or longer than the following together. (Fig. 302)
	Second joint of antennæ not noticeably lengthened8
7.	Median caudal gill flat, much shorter than the lateral ones which
	are triangular in section
	All three caudal gills of approximately the same size and shape.
	SYNLÉSTIDÆ
8.	Caudal gills strongly constricted at the middle. (Fig. 303).
	PROTONEÙRIDÆ
	Caudal gills not constricted at the middle9
9.	Abdomen with six or seven pairs of lateral gills; caudal gills swollen, sac-like
	Abdomen without lateral gills11
10.	Caudal gills swollen, oblong, with sharply pointed tips.
	EPALLÁGIDÆ
	Caudal gills with the apex broad, not pointed.
	TO THE TOTAL TOTAL TO THE TOTAL



Figs. 302-311. Odonata, Nymphs

- 302. Synlestes, antenna of nymph (Tillyard) Synlestidæ.
- 303. Isosticta, caudal gill of nymph (Tillyard) Protoneuridæ.
- 304. Æschnid, antenna of nymph (Howe) Æschnidæ.
- 305. Æschnid, lateral view of head, with labium closed (Howe) Æschnidæ.
- 306. Tachopteryx, lateral lobe of labium (Howe) Petaluridæ.
- 307. Gomphid, antenna (Howe) Gomphidæ.
- 308. Æschna, lateral lobe of labium (Tillyard) Æschnidæ.
- 309. Plathemis, labium, open (Garman) Libellulidæ.
- 310. Libellulid, lateral view of head, with labium closed (Howe) Libellulidæ.
- 311. Cordulegaster, labium, open (Garman) Cordulegastridæ.
- 13. Labium forming a more or less spoon-shaped mask which covers the ventral and sometimes also the front surface of the head as far as the antennæ (Fig. 310); setæ usually numerous....14 Labium flat, not forming a spoon-shaped mask for the lower surface of the head (Fig. 305), almost always without setæ.....16

- - Lateral lobes of labium variable in form, smooth, serrate, regularly dentate or sometimes with a few long teeth, but in the last case these do not interlock with those of the other side; median lobe triangular, projecting, not bifid or emarginate. (Fig. 309). 15
- - Teeth on inner margin of lateral lobes of labium usually reduced to crenulations or obsolete (in a few forms with long teeth the mask is either greatly enlarged (*Pantala*) or the species are very small (Tetratheminæ)) .......................LIBELLÜLIDÆ
- - Antennæ short and stout, the segments wider than long; lateral lobes of labium with a short movable hook (Fig. 306).

#### PETAL URIDÆ

#### LITERATURE ON ODONATA

- Andres, A. The Dragonflies of Egypt. Mem. Soc. Entom. Egypte, **3**, pp. 1–43 (1928).
- Bartenev, A. N. Libellulidæ II. Faune Russ. Pseudoneuroptera, 1, pp. 353–576 (1919).
- CALVERT, P. P. Odonata. In Biologia centrali-Americana, Neuroptera, 1, pp. 17–420 (1901–08).
  - Odonata of Cuba. Trans. American Entom. Soc., 45, pp. 335-396 (1919).
- Fraser, F. C. Indian Dragonflies. Journ. Bombay Nat. Hist. Soc., 25–29 (numerous parts) (1918–25).
- Fröhlich, C. Die Odonaten und Orthopteren Deutschlands. Jena (1903).
- Garman, P. The Zygoptera of Illinois. Bull. Illinois State Lab. Nat. Hist., 12, pp. 411-587 (1917).
  - Odonata of Connecticut. Bull. State Geol. Nat. Hist. Surv., Connecticut, No. 39, 331 pp. (1927).
- Howe, R. H. Manual of Odonata of New England. Mem. Thoreau Mus. Concord, Mass., 2, 7 parts, 138 pp. and supplement (1917–23).
- Kellicott, D. S. Odonata of Ohio. Ohio Acad. Sci., Special Paper, No. 2, 114 pp. (1899).

- Kennedy, C. H. Classification of Zygoptera. Ohio Journ. Sci., 21, pp. 83–88 (1920).
- Kirby, W. F. Catalogue of Odonata. 202 pp., London (1890).
- Krüger, L. Einführung in das Studium der Libellen. Abh. Ber. pommers. naturf. Ges., 6, pp. 53–106 (1925).
- Lucas, W. J. British Dragonflies. London (1900).
- Martin, R. Corduliidæ. Collection Zoologiques, Selys-Longchamps, fasc. 17 (1909).
  - Æschnidæ. Coll. Zool. Selvs-Longchamps, fasc. 18–19 (1909).
  - Æschnidæ, Æschninæ, Gen. Insectorum, fasc. 115, 33 pp. (1911).
  - Cordulidæ, Gen. Insectorum, fasc. 155, 32 pp. (1914).
- Munz, P. A. Keys for identification of genera of Zygoptera. Mem. American Entom. Soc., No. 3, 78 pp. (1919).
- MUTTKOWSKI, R. A. Odonata of Wisconsin. Bull. Wisconsin Nat. Hist. Soc., 6, pp. 57–123 (1908).
  - Catalogue of Odonata of North America. Bull. Pub. Mus. Milwaukee, 1, Art. 1 (1910).
- Needham, J. G. Manual of Dragonflies of China. Zoologica Sinica, Series A, 344 pp. Peiping, China (1930).
- NEEDHAM, J. G. and HEYWOOD, H. B. Handbook of Dragonflies of North America. 378 pp., Springfield, Illinois (1929).
- Oguma, K. The Japanese Æschnidæ. Insecta Matsumurana, 1, pp. 78–100 (1926).
- Ris, F. Odonata. Süsswasserfauna Deutschlands, Heft 9, 67 pp. (1909).

  - Neuer Beitrag zur Kenntnis der Odonatenfauna der Neu-Guinea Region. Nova Guinea, Livr. 13, pt. 2, pp. 81–131 (1913).
  - Die Odonatenfauna von Argentina. Mem. Soc. Entom. Belgique, 22, pp. 55–102 (1913).
  - Odonata of South Africa. Ann. South African Mus., 18, pp. 247–452 (1921).
- Schmidt, E. Odonata. In Tierwelt Mitteleuropas, 4, Lief. 1b, pp. IV 1–66 (1929).
- SEEMAN, T. M. Dragonflies, Mayflies and Stoneflies of Southern California. Pomona Journ. Entom. Zool., 19, pp. 1–69 (1927).
- TILLYARD, R. J. The Biology of Dragonflies. University Press, Cambridge, England (1917).
- WALKER, E. M. North American Species of Æschna. Univ. Toronto Biolog. Stud., No. 11, pp. 1–213 (1912).
  - North American Somatochlora. Univ. Toronto Stud. Biol. Ser., No. 26, 202 pp. (1925).
- WILLIAMSON, E. B. Dragonflies of Indiana. Rept. Dept. Geol. Ind., No. 24, pp. 229–333 (1901).
  - North American Species of Macromia. Proc. U. S. Nat. Mus., 37, pp. 369–398 (1909).

## ORDER PLECTÓPTERA

# (EPHEMERÓPTERA, EPHEMÉRIDA, AGNÁTHA)

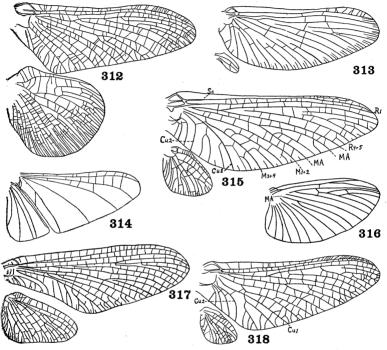
Delicate insects with short, filiform antennæ and vestigial mouthparts; abdomen slender, bearing two or three long, many-jointed caudal filaments; four wings, usually with very complex venation. Head not very freely movable, with the compound eyes and three ocelli present; antennæ with two large basal joints and a bristle-like, indistinctly iointed terminal portion. Prothorax more or less free, small or very small; mesothorax large. Fore wings much larger than the hind ones, usually with many longitudinal veins and a great number of crossveins: commonly with numerous short longitudinal veins near the margin that are not attached at the base; media more complex than in other living orders, its anterior, convex branch preserved; hind wings often very small or even absent. Abdomen slender, usually cylindrical or tapering, with ten segments. Legs weak; tarsi usually with five or four joints. Metamorphosis incomplete, the nymph aquatic and provided with abdominal gills and three caudal filaments. Aerial form emerging from the nymph as a subimago which is similar to the imago. but which molts when it transforms to the imago. Imaginal life very short. Mayflies.

#### Adults

- - Subcosta completely developed, visible for its entire length.....3
- 3. Wings translucent, subopaque in the male, entirely opaque in the female; hind margin of wings without unattached veinlets; legs weak, the front pair sometimes long in the male; hind legs almost always short and weak. (Euthyplòcia, neotrop., ethiop.;

	Exeuthyplòcia, Povílla, ethiop.; Campsùrus, neotrop.; Polymitárcys, widespr.)
	Wings transparent, shining; margin of wings, especially the hind
	pair, with numerous short unattached veinlets; legs strong,
	functional
4.	First branch of cubitus in fore wing simple, not branched, but
	connected with the wing margin by numerous crossveins; fork
٠	of R <sub>2</sub> and R <sub>4</sub> in hind wing much longer than its stalk. (Ephém-
	era, holarc., indoaustr.; Hexagènia, Am., ethiop.; Eatónica,
	ethiop.; Pentagènia, nearc.) EPHEMÉRIDÆ
	First branch of cubitus in fore wing forked; not connected to the
	wing margin by crossveins; fork of R2 and R4 in hind wing
	shorter or no longer than its stalk. (Potamánthus, holarc.;
	Rhænánthus, palæarc., indoaustr.; Potamanthòdes, in-
	domal.) POTAMÁNTHIDÆ
5.	Hind tarsi with four freely movable joints; if with the indication
	of a fifth joint, this is immovably attached to the tibia. (Super-
	family BAETÒIDEA)6
	Hind tarsus with five freely movable joints10
6.	Subcosta in fore wing fused with the radius or wanting, at most
	visible at the base; wings milky or grayish, with very simple
	venation, the fore wing with only four to seven longitudinal
	veins, with crossveins in only from two to five of the anterior
	spaces; hind wing without or with very few crossveins which
	are restricted to the anterior part; large or medium sized
	species. (Fig. 314). (Oligoneùria, palæarc., neotrop., ethiop.;
	Homoneùria, Lachlania, nearc.; Spaniophlèbia, Nòya,
	neotrop.; Elassoneùria, ethiop.)OLIGONEURÌIDÆ
	Subcosta in fore wing free, well developed and visible for its entire
	length
7.	Anterior median vein in fore wing (MA) clearly forked8
	Anterior median vein in fore wing, simple, not forked, although
	behind it are two free veins which are not attached at the base
	(Fig. 313); fore wing usually with few crossveins; hind wings
	very small and narrow, sometimes absent, with at most two or
	three longitudinal veins. (Clòeon, cosmop.; Baètis, widespr.;
	Centróptilum, holarc., ethiop.; Callibaètis, Am.). BAÉTIDÆ
8.	Wings milky or infuscated, ciliate on the hind margin; hind wings
	absent, although sometimes present in the subimago; no un-
	attached intercalated veins; frequently with only a few cross-
	vaine emall anguing (Fig. 316) (Cannie widgen: Tricory

phòdes, Leptòhyphes, Am.; Leptohyphòdes, neotrop.; Tricórythus, ethiop.). (Including *PROSOPISTOMÁTIDÆ*?). (BRACHYCÉRCIDÆ). ... CAÉNIDÆ



Figs. 312-318. Plectoptera

- 312. Baetisca, wings (Eaton) Baetiscidæ.
- 313. Baetis, wings (Eaton) Baetidæ.
- 314. Oligoneuria, wings (Eaton) Oligoneuriidæ.
- 315. **Ephemerella,** wings (Eaton) Ephemerellidæ.
- 316. Caenis, wing (Eaton) Caenidæ.
- 317. Chirotenetes, wings (Needham) Siphluridæ.
- 318. Atalophlebia, wings (Eaton) Leptophlebiidæ.
  - 9. Second branch of cubitus in the fore wing usually widely separated at base from the first branch, but lying close to the first anal vein; no unattached intercalated veins between the media and cubitus and none in front of the posterior branch of the media.

(Fig. 318). (Thraùlus, palæarc., neotrop., indoaustr.; Atalo-

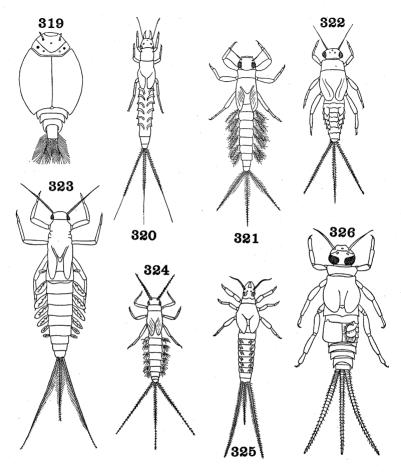
	phlèbia, neotrop., indoaustr.; Habrophlèbia, Leptophlèbia,
	holarc.; Adenophlèbia, ethiop.) LEPTOPHLEBÌIDÆ
	Second branch of cubitus in fore wing approximate at base to the
	first branch, but widely separated from the first anal vein;
	several (usually two) unattached intercalated veins between the
	media and cubitus and also in front of the posterior branch of
	the media. (Fig. 315). (Ephemerélla, Chitonóphora, hol-
	arc.; Drunélla, nearc.; Melanomerélla, neotrop.; Teloga-
	nòdes, indoaustr.) EPHEMERÉLLIDÆ
10.	First and second branches of the cubitus running more or less
	parallel to the first anal vein; Cu <sub>1</sub> and Cu <sub>2</sub> connected by cross-
	veins, but the cubital area without paired crossveins or sinuous
	veins extending to the wing margin; hind wings rounded, with
	numerous long intercalated veins extending to the posterior
	part of the wing margin; prothorax very small. (Fig. 312).
	(Baetísca, nearc.) BAETÍSCIDÆ
	First and second branches of the cubitus very close together at
	base and strongly divergent apically, the second branch much
	shorter than the first and strongly curved backwards towards
	apex, hind wings oval; pronotum well developed11
11.	Cubital area of fore wing with a number of more or less sinuous
	veins extending from the first branch of the cubitus to the wing
	margin. (Fig. 317). (Amelètus, holarc., austr.; Chirotónetes,
	Siphl $\hat{\mathbf{u}}$ rus (= $Siphlon\hat{u}$ rus), $holarc.$ ; Siphlon $\hat{\mathbf{sca}}$ , $nearc.$ ; Onis-
	cigáster, indoaustr.). $(SIPHLON \dot{U}RID\cancel{E})$ SIPHL $\dot{\mathbf{U}}$ RID\cancel{E}
	Cubital area of fore wing without oblique sinuous veins extending
	from the first branch of the cubitus to the wing margin, but
	with from two to four straight unattached veins more or less
	parallel to the branches of the cubitus12
12.	Only two unattached intercalated veins in the cubital area of the
	fore wing; if sometimes with a second pair indicated, these are
	short and lie close to the second branch of the cubitus; two or
	three caudal filaments. (Amétropus, Métropus, palæarc.).
	AMETROPÓDIDÆ
	Four unattached intercalated veins in the cubital area of the fore
	wing, the longer pair lying close to the second branch of the
	cubitus; two caudal filaments. (Heptagènia, Epeòrus, hol-
	arc.; <b>Ìron</b> , nearc., <b>Ecdyonùrus</b> , palæarc., Am.; <b>Rhithrógena</b> ,

palearc.; Atópopus, indoaustr.). (ECDYONURIDÆ).

HEPTAGENÌIDÆ

# Nymphs

l.	Abdominal gills visible at their insertion above or on the sides of the abdomen
	of the abdomen
	thorax behind which covers most of the abdomen. (Fig. 319).
	Imago unknown. ( <b>Prosopistoma</b> , palæarc., ethiop.).
	PROSOPISTOMÁTIDÆ
2.	Mandibles usually very long and extended anteriorly; six or seven
-	pairs of plumose gills, the first sometimes much reduced; legs
	stout
	Mandibles very short, not extended anteriorly; gills not plumose;
	legs slender
3.	Mandibles extremely large and projecting far beyond the head in
	front; gills extended dorsally over the abdomen4
	Mandibles much shorter, but slightly projecting in front of the
	head; gills extended laterally, away from the abdomen. (Fig.
	321)POTAMÁNTHIDÆ
4.	Front with two tubercles anteriorly; mandibles curved outwards
	at tips; antennæ with long cilia EPHEMÉRIDÆ
	Front without tubercles; mandibles curved downwards at tips;
_	antennæ without or with short cilia
5.	Body short and stout; six pairs of similar gills; caudal filaments
	short, the median one shorter than the lateral ones. (Fig. 325).
	PALINGENÎIDÆ
	Body long and slender; seven pairs of gills, the first pair much reduced in size; anal filaments long, of equal length. (Fig. 320).
	POLYMITÁRCIDÆ
6.	Body strongly flattened, the head orbicular, or more or less trans-
٠.	verse, the eyes placed on its dorsal surface
	Body more or less cylindrical, not or very slightly flattened, the
	eyes placed at the sides of the head
7.	Gills uniform, extending from the sides of the abdomen; anal
	filaments at least as long as the body HEPTAGENÎIDÆ
	First pair of gills inserted on the under side of the first abdominal
	segment, the six following pairs extended from the sides of the
	abdomenOLIGONEURÌIDÆ
8.	Lateral caudal filaments (cerci) fringed on both edges9
	Lateral caudal filaments ciliated on the inner border only 11
9.	Seven pairs of gills, inserted laterally at the sides of the abdomen;
	sometimes all are filamentous or the first is much reduced and
	the others leaf-like. (Fig. 324)LEPTOPHLEBÎIDÆ



Figs. 319-326. Plectoptera, Nymphs

- 319. Prosopistoma, nymph (Rousseau) Prosopistomatidæ.
- 320. Polymitarcys, nymph (Rousseau) Polymitarcidæ.
- 321. Potamanthus, nymph (Rousseau) Potamanthidæ.
- 322. Ephemerella, nymph (Rousseau) Ephemerellidæ.
- 323. Baetis, nymph (Rousseau) Baetidæ.
- 324. Habrophlebia, nymph (Rousseau) Leptophlebiidæ.
- 325. Palingenia, nymph (Rousseau) Palingeniidæ.
- 326. Tricorythus, nymph (Rousseau) Caenidæ.

Five pairs of gills, the last or the last two pairs not visible. (Fig. 322) ..... EPHEMERÉLLIDÆ

#### LITERATURE ON PLECTOPTERA

- Снорга, В. Indian Ephemeroptera: Ephemeroidea. Rec. Indian Mus., 29, pp. 91–138 (1927).
- EATON, A. E. Revisional monograph of recent Ephemeridæ. Trans. Linn. Soc. London, (2), 3, (1883–85).

Ephemeridæ. In Biologia centrali-Americana, pp. 1–16 (1892).

- Gros, A. J. and Lestage, J. A. Contribution a l'étude des larves des Éphéméroptères. Ann. Biol. Lacust., 15, pp. 123, 133, 136, 161 (1927).
- Klapálek, F. Ephemerida: Süswasserfauna Deutschlands, Lief. 8 (1909).
- Lestage, J. A. Notes sur les Éphémères de la "Monographical Revision" de Eaton. Ann. Soc. Entom. Belgique, **64**, pp. 33–60 (1924).

Les Éphéméroptères de la Belgique. Bull. Ann. Soc. Entom. Belgique, **68–69** (several parts).

Contribution a l'étude des larves des Éphémères paléarctiques. Ann. Biol. Lacust., 8, pp. 213–457 (1918).

- Needham, J. G. Ephemeridæ of New York. Bull. New York State Mus., No. 86, pp. 17–62 (1905).
- Rousseau, E. Les larves et nymphes aquatiques des insectes d'Europe, Vol. 1, pp. 162-273 (1921).
- Seeman, T. M. Dragonflies, Mayflies and Stoneflies of southern California. Pomona Journ. Entom. and Zool., 19, pp. 1–69 (1927).
- $\label{eq:Tumpel} \textbf{Tumpel}, R. \ Die \ Geradeflügler \ Mitteleuropas. \ (Plectoptera, pp. \ 73–108) \ (1901).$
- Ulmer, G. Übersicht über die Gattungen der Ephemeropteren. Stettiner Entom. Zeit., **81**, pp. 97–144 (1920).

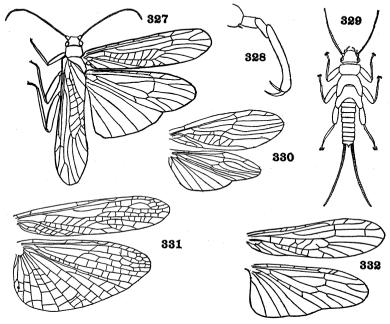
Verzeichnis der deutschen Ephemeropteren. Konowia, **6**, pp. 234–262 (1927).

Ephemeroptera. In Tierwelt Mitteleuropas, 4, Lief. 1b, pp. III 1-43(1929).

## ORDER PLECÓPTERA

(PERLÀRIA)

Body soft, of moderate or large size; four membranous wings, usually with many veins and numerous crossveins, rarely reduced in size: anal area of hind wing large and pleated, usually separated by a notch from the rest of the wing. Head broad and flattened: mandibles either well developed or much reduced; antennæ long, thread-like; three ocelli; cerci usually long and many-jointed. Prothorax large, free; legs strong, tarsi three-jointed. Nymphs aquatic, commonly with tracheal gills; antennæ long, much like those of the adults; eyes well developed; ocelli present; cerci usually long, many-jointed. Metamorphosis slight. Stoneflies, Salmon-flies.



Figs. 327-332. Plecoptera

- 327. Isopteryx Perlidæ.
- 328. Perla, tarsus. Perlidæ.
- 329. Perla, nymph (Pictet) Perlidæ.
- 330. Notonemoura, wings. Nemouridæ.
- 331. Stenoperla, wings (Tillyard) Eustheniidæ.
- 332. Perla, wings. Perlidæ.

No such transverse cord near the middle of the wing; distal portion of wing with crossveins. Australian, neotropical. (Leptopérla, Dinotopérla). (Including GRIPOPTERÝGIDÆ, part)....LEPTOPÉRLIDÆ

6. Cerci very short, sometimes reduced to a single joint, never with more than ten joints; last anal vein in fore wing forked beyond

the anal cell. (Nemoùra, Leùctra). (Fig. 330). (Including $T \not\equiv NIOPTER \not YGID \not\equiv MIDETER \not YGID \not\equiv MIDETER \not YGID \not\equiv MIDETER \not\equiv MID$
Nymphs
1. Visible gills present
No visible gills
Gills on the abdomen, but none on the thorax
3. Gills on the thorax and also on the underside of abdominal seg-
ments 1 and 2, or 1 to 3PTERONÁRCIDÆ Gills present on thorax only
4. Gills on the underside of the prothorax only <b>NEMOÙRIDÆ</b>
Gills on all three thoracic segments5
5. Three pairs of gills in the form of filamentous tufts on the pleuræ;
legs densely fringed with long hairs
A few <b>NEMOÙRID</b>
6. Gills a series of paired latero-ventral abdominal appendages on
segments 1 to 5 or 1 to 6; large species <b>EUSTHENIIDÆ</b> Gills a rosette of small filaments surrounding the anus.
Gins a rosette of sman maments surrounding the ands.  LEPTOPÉRLIDÆ
7. Palpi with the apical joints more slender than the basal ones
(Fig. 329)
Palpi with the apical joints as stout as the basal ones
third NEMOÙRIDÆ
Second joint not very short9
9. Third joint of tarsi not longer than the first and second together;
cerci glabrous, as long as the body Some <b>NEMOÙRIDÆ</b> Third joint of tarsi twice as long as the first and second together.
CAPNIDÆ
LITERATURE ON PLECOPTERA
ENDERLEIN, G. Klassifikation der Plecopteren. Zool. Anz., 34, pp. 385–419
(1911).
Frison, T. H. Fall and Winter Stoneflies or Plecoptera of Illinois. Bull. Illinois Nat. Hist. Surv., <b>18</b> , pp. 345–409 (1929).
nois Nat. flist. Surv., 10, pp. 343–409 (1929).

Klapálek, F. Die Plecoptera. Coll. Zool. Selys-Longchamps, fasc. 4 and 4A,

266 pp., Brussels (1912-23).

- LeRoi, O. Zur Plecopteren von Rheinland-Westfalen. Ver. bot. zool. Ver. Rheinl-Westf. 1912, pp. 25-51 (1912) (Extensive bibliography).
- MacLachlan, R. Monograph of the British Neuroptera Planipennia. Trans. London Entom. Soc., 1868, pp. 145–224 (1868).
- Needham, J. G. and Broughton, E. Central American Stoneffies. Journ. New York Entom. Soc., 35, pp. 109-120 (1927).
- Needham, J. G. and Claassen, P. W. A Monograph of the Plecoptera or Stoneffies of America North of Mexico. 397 pp., Lafavette, Indiana (1925).
- Schoenemund, E. Plecoptera. In Tierwelt Mitteleuropas, 4, Lief. 2, pp. V 1–18 (1928).
  - Die Larven der deutschen Perla-Arten. Entom. Mitt., **14**, pp. 113–121 1925).
- Seeman, T. M. Dragonflies, Mayflies and Stoneflies of Southern California. Pomona Journ. Fntom. and Zool., 19, pp. 1-69 (1927).
- SMITH, L. W. Studies in North American Plecoptera. (Pteronarcinæ and Perlodini). Trans. American Entom. Soc., 43, pp. 433–489 (1917).
- Tillyard, R. J. A new classification of the Order Perlaria. Canadian Entom., 53, pp. 35–43 (1921).

Revision of the Family Eustheniidæ. Proc. Linn. Soc. New South Wales, **46**, pp. 221–226 (1921).

The Stone-flies of New Zealand. Trans. New Zealand Inst., 54, pp. 197–217 (1923).

## ORDER MEGALÓPTERA

## (SIALÒIDEA)

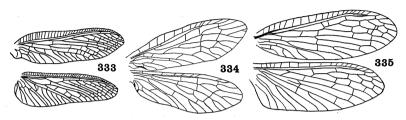
Soft-bodied species with large wings, long and sometimes pectinate antennæ and simple, similar legs. Costal cell with many transverse veins, subcosta and first radius simple, apically fused, the radial sector arising near the base; hind wings with the anal space normally large, folded fan-like when at rest. Prothorax quadrate. Metamorphosis complete; larvæ aquatic, living in freshwater streams; predatory, mandibulate, possessing lateral abdominal gill-filaments; wings appearing in the pupal stage; no cocoon.

#### Adults

Three ocelli present; fourth joint of the tarsi simple, not bilobed; venation regular, with the crossveins weakly formed, branches of the radial sector directed backward; large or medium sized species, 45 to 100 mm. in wing expanse. (Figs. 333, 335). (Archichauliòdes, Austr.; Chauliòdes, Neohérmes, Corýdalis (Dobson, Hellgrammite), Am.; Neochauliòdes, indomal.; Hérmes, Neùromus, Protohérmes, Neoneùromus, Asia).

CORYDÁLIDÆ

Ocelli absent; fourth joint of the tarsi prominently bilobed; some of the branches of the radial sector directed forward; medium sized species, 20 to 40 mm. in wing expanse. (Fig. 334). (Protosialis, Am.; Austrosialis, Stenosialis, Austr.; Sialis, holarc.).



Figs. 333-335. Megaloptera

- 333. Corydalis, wings. Corydalidæ.
- 334. Austrosialis, wings (Tillyard) Sialidæ.
- 335. Chauliodes, wings. Corydalidæ.

#### Larvae

## LITERATURE ON MEGALOPTERA

(See also general papers on Neuroptera)

Barnard, K. H. Cape Megaloptera. Trans. Roy. Soc. South Africa, 19, pp. 169–184 (1931).

DAVIS, K. C. Sialididæ of North and South America. Bull. New York State Mus., No. 68, pp. 442–487 (1903).

ESBEN-PETERSEN, E. Megaloptera. Gen. Insectorum, fasc. 154 (1913).

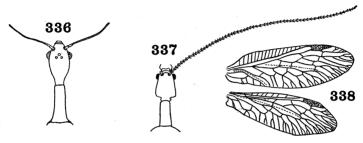
Stitz, H. Megaloptera. In Tierwelt Mitteleuropas, 6, Lief. 1, pp. XIV 1–2 (1928).

Weele, H. W. Megaloptera. Coll. Zool. Selys-Longchamps, fasc. 5, Brussels (1910).

## ORDER RAPHIDIÒDEA

(RAPHIDIÒIDEA, EMMENÓGNATHA, part.)

Moderate-sized, slender, predatory species with elongate cylindrical prothorax; head large, nearly horizontal, mandibles strong, antennæ



Figs. 336-338. Raphidiodea

- 336. Raphidia, head and prothorax of female (Albarda) Raphidiidæ.
- 337. Inocellia, head and prothorax of female (Albarda) Raphidiidæ.
- 338. Raphidia, wings (Handlirsch) Raphidiidæ.

# LITERATURE ON RAPHIDIODEA

Albarda, H. Revision des Raphidides. Tijd. Entom., **34**, pp. 65–184 (1891). Esben-Petersen, E. Raphididæ. Gen. Insectorum, fasc. 154, 13 pp. (1913).

Navás, L. Monografia de l'orde dels Rafidiopters. Arch. Inst. Catalans, Barcelona, 93 pp. (1918).

Stitz, H. Raphidiodea. In Tierwelt Mitteleuropas,  ${\bf 6},$  Lief. 1, pp. XIV 2–4 (1928).

# ORDER NEURÓPTERA

(SYNISTÀTA, part., DICTYÓPTERA, part.)

Small to rather large, slender, predaceous insects with large wings, but of slow flight. Head free, vertical, eyes prominent, mouth inferior, mandibles strong; prothorax more or less freely movable and prominent, meso- and metathorax not closely grown together; abdomen long and narrow, the first sternite reduced, no cerci; wings similar, membranous, no large anal field; when at rest the wings usually lie roof-like over the abdomen, longitudinal veins almost always very

numerous through repeated branching, costal cell almost always with crossveins. Metamorphosis complete, larvæ terrestrial, aquatic only in Sisyridæ, strongly mandibulate, the mandibles and maxillæ specialized for sucking the body juices of the insect prey; pupæ in cocoons.

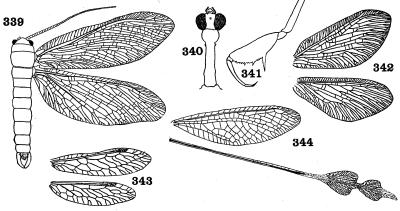
1. Veins and usually crossveins abundant, radial sector with several branches or forkings; wings not covered with a whitish powder. 2 Veins and crossveins few in number, the radial sector at most only forked; wings covered with a whitish powder; very small, slender, pale-colored rare species, of 3 to 10 mm. wing expanse. (Superfamily CONIOPTERYGOIDEA).

## CONIOPTERÝGIDÆ

The following subfamilies are distinguished:

- a. Outer lobe of maxilla three-jointed; abdomen with four to six pairs of everted ventral pouches; stem of media of fore wings usually with two thickenings which bear bristles. (Aleurópteryx, palæarc.; Coniocómpsa, Helicocònis, holarc., austr., neotrop.; Heterocònis, austr.) .... ALEUROPTERYGINÆ
- 2. Large, stout, showy, moth-like, cursorial species, of 30 to 70 mm. wing-expanse; costal area not broad, Sc, R and R<sub>s</sub> not forming a distinct triple vein; head small and closely set on the prothorax; antennæ long and filiform, and tapering in both sexes, 40- to 50-jointed; abdomen of the male terminating in large forcipate appendages; ovipositor not exserted (Fig. 339). (Superfamily ITHONOIDEA). (Ithône, Várnia, austr.; Oliárces, Calif.; Rapísma, Asia) . . . . . . . . . . . . ITHÔNIDÆ
- - Antennæ at least thickly cylindrical, variable in length, usually gradually enlarged toward the end, or filiform with clavate end; at least the discal portion of the wings densely reticulate, Sc

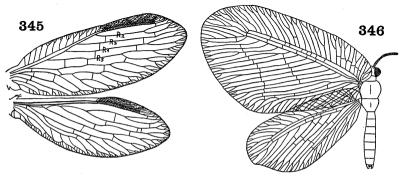
4. Hind wings not longer than the fore wings, the two pairs similar in form and venation. (Superfamily HEMEROBIOIDEA)..5



Figs. 339-344. Neuroptera

- 339. Ithone, Outline of body and wings (Tillyard) Ithonidæ.
- 340. Mantispa, head and thorax from above. Mantispidæ.
- 341. Mantispa, raptorial front leg. Mantispidæ.
- 342. **Dilar,** wings (Handlirsch) Dilaridæ.
- 343. **Mantispa,** wings (Handlirsch) Mantispidæ.
- 344. Chasmoptera, wings (Tillyard) Nemopteridæ.

Eur.: Nallàchius. Am.: Nèpal. Asia) DILÁRIDÆ

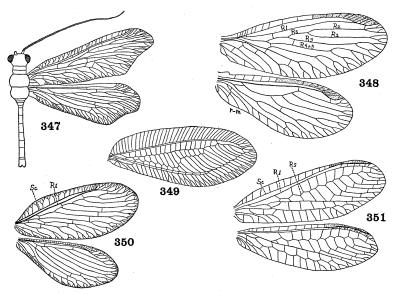


Figs. 345-346. Neuroptera

- 345. Micromus, wings (Tillvard) Hemerobiidæ.
- 346. Psychopsella, outline of body and wings (Tillyard) Psychopsidæ.
- 9. Rather large, moth-like species, with broad wings, the costal area of the fore wings very wide, Sc, R<sub>1</sub> and R<sub>s</sub> closely parallel, forming a chitinized triple midrib and fusing at the apical fourth of the wing; antennæ short; rare, nocturnal species. (Fig. 346). (Psychópsis, Megallànes, Austr.; Psychopsélla, Arteriópteryx, Psychóphasis, ethiop.; Megapsýchops).

PSYCHÓPSIDÆ

10.	Not moth-like, wings not broadly rounded, with normal costal area and without the above-described prominent triple vein 10 Ocelli present; discal area of the wings with many crossveins, marginal area with no crossveins but with many forked veinlets; moderate to large, slender species. Widespr., but not North American. (Euporismus, Œdósmylus, Ósmylus, Porismus, Spilósmylus)
11.	Ocelli absent
	forked veinlets; Sc and R <sub>1</sub> fused near wing-tip; vertex convex; antennæ moderate in length; rather large, nocturnal species, wing-expanse 40 to 75 mm. ( <b>Polystæchotes</b> , N. Am.).
	POLYSTŒCHÓTIDÆ
	No recurrent vein at the humeral angle of the wings; discal area of the wings not differentiated from the marginal area; antennæ
	longer than the head and thorax; smaller species
19	Vertex convex; wing venation relatively simple, radial sector of
14.	fore wings without definitive accessory veins, Sc and R <sub>1</sub> coal-
	esced near tip of wing, costal crossveins not forked, r-m cross-
	vein of hind wings in the axis of the wing; size small, 6 to 8
	mm. in length; larvæ aquatic, feeding on freshwater sponges.
	(Fig. 348). Spongilla-flies. (Climàcia, nearct.; Neurórthrus,
	Sisyrélla, palæarct.; Sísyra, holarct.) SISÝRIDÆ
	Vertex flattened; the single radial sector in the fore wings with
	definitive accessory veins; hind wings with the r-m crossvein
	oblique or transverse; size larger13
13.	
	away from R <sub>1</sub> , the cell R <sub>1</sub> broad and containing many cross-
	veins; wings rounded, not falcate
	Costal crossveins forked, cell R <sub>1</sub> narrow and almost devoid of
	crossveins, apical portion of the hind margin of the fore wings
	sometimes widely notched, leaving the apex more or less acute (the falcate condition); wings and body hairy, especially the
	hind margin of the wings
14	Wings of nearly equal width, a crossvein placed near base of the
<b>.</b>	subcostal cell, less than thirty crossveins in the costal cell before
	the stigma. (Fig. 351). Green lacewings, Stink-flies; the larva
	are Aphis-lions. Cosmopolitan. (Allochrysa, Chrysòpa
	Malahma Nathaahrina) CHPVSÓPIDA



Figs. 347-351. Neuroptera

- 347. Trichoma, outline of body and wings (Tillyard) Trichomatidæ.
- 348. Sisyra, wings (Tillyard) Sisyridæ.
- 349. Oligochrysa, fore wing (Tillyard) Apochrysidæ.
- 350. Protobiella, wings (Tillyard) Berothidæ.
- 351. Chrysopa, wings (Tillyard) Chrysopidæ.
- 15. Fore wings with Sc and R fused before the wing-tip; peculiar seed-like scales often present on some part of the wings. (Fig. 350). (Acroberòtha, ethiop., Ind.; Beròtha, indomal.; Isoscelípteron, palæarc.; Lomamŷia, Am.; Cycloberòtha, austr.).
  BERÓTHIDÆ

Fore wings with Sc and R not fused apically; hairs of body and wings conspicuously long. (Fig. 347). (Stenobiella, Trichòma, austr.). TRICHOMÁTIDÆ

16.	Wings about one-third as wide as long	, costal area wide, marginal
	veinlets forked, subcostal cell with	
	long, cylindrical. (Myiodáctylus,	
		MYIODACTÝLID <i>A</i>

- Wings much narrower, the marginal area at least in large part
- 17. Antennæ elongate cylindrical; subcostal area with many crossveins. (Austronýmphes, Nýmphes, Nymphídion, austr.).

## NÝMPHIDÆ

- Antennæ more or less distinctly clavate, or apically swollen or
- 18. Antennæ about as long as the head and thorax; wings usually with an elongate narrow cell immediately behind the point of fusion
  - Antennæ long, slender, strongly clavate apically; eyes usually divided into two parts by a groove; no elongate hypostigmatic cell differentiated. Widespread, mainly tropical. (Acmonòtus, austr.; Ascálaphus, palæarc.; Colobópterus, Am.; Hỳbris, indomal.; Neuróptynx, nearc.; Suhpalácsa, ethiop., indoaustr.; Ululòdes, Am.; Nephoneùra, ethiop.).

## ASCALÁPHIDÆ

- 19. Antennæ weakly clubbed, or flattened at tip; hypostigmatic cell elongate; body and wings pubescent; weak fliers. Larvæ are called ant-lions or doodle-bugs. Widespread, mainly tropical. (Brachynemurus, Creagris, Dendroleon, Formicaleo, Hesperòleon, Myrmèleon, Palpàres, Protopléctron). (MYR-MELEÓNIDÆ) ..... MYRMELEÓNTIDÆ
  - Antennæ strongly clubbed; hypostigmatic cell variable; abdomen and wings shining; crepuscular, strong fliers, superficially resembling dragonflies. (Stilbópteryx, austr.).

# STILBOPTER Ý GIDÆ

#### LITERATURE ON NEUROPTERA

- BAGNALL, R. S. Review of British Coniopterygidæ. Entom. Rec., 27, pp. 241-247 (1915).
- Banks, N. Revision of Nearctic Chrysopidæ. Trans. American Entom. Soc., **29**. pp. 137–162 (1903).
  - A Revision of the Nearctic Hemerobiidæ. Trans. American Entom. Soc., **32**, pp. 21–51 (1905).
  - Catalogue of the Neuropteroid Insects of the United States. American Entom. Soc., Philadelphia, 53 pp. (1907).

A Revision of the Nearctic Coniopterygidæ. Proc. Ent. Soc. Washington, 8, pp. 77–86 (1907).

Revision of the Nearctic Myrmeleonidæ. Bull. Mus. Comp. Zoöl. Harvard, **68**, pp. 1–84 (1927).

Enderlein, G. Monographie der Coniopterygiden. Zool. Jahrb. Abth. f. Syst., 23, pp. 173-242 (1906).

Coniopterygidæ. Gen. Insectorum, fasc. 67, 18 pp. (1908).

Klassifikation der Mantispiden. Stettiner Entom. Zeit., **71**, pp. 341–379 (1910).

Die Klassifikation der Coniopterygiden. Arch. Klass. Phylog. Entom., **1**, pp. 98–114 (1930).

HAGEN, H. A. Neuroptera of North America. Smithsonian Misc. Coll. IV., (1862).

KRÜGER, L. Revision of Osmylidæ. Stettiner Entom. Zeitg., 75, pp. 9–130 (1914); 76, pp. 60–87 (1915).

Systematische Uebersicht der Neuropteren. Settiner Entom. Zeitg., **78**, pp. 116–137 (1917).

Revision of Psychopsidæ. Stettiner Entom. Zeitg., **83**, pp. 17–48 (1922). Revision of Berothidæ. Stettiner Entom. Zeitg., **83**, pp. 49–88 (1922).

Revision of Hemerobiidæ. Stettiner Entom. Zeitg., 83, pp. 138–173 (1922).

Kuwayama, S. Studies on the Dilaridæ of Japan. Trans. Sapporo Nat. Hist. Soc., 8, pp. 51–83 (1921).

Lacroix, J. L. Faune des Plannipennes de France. Ascalaphidæ. Bull. Soc. Étude Sci. Nat. Elbeuf., **41**, pp. 65–100 (1923).

Mocsary, A. Neuroptera. Fauna Regni Hungariæ, pp. 33-44 (1918).

Nakahara, W. Osmylinæ of Japan. Annot. Zool. Japonenses, Tokyo, 8, pp. 489–518 (1914).

Navás, P. L. Neurópteros (s. lat.) de España y Portugal. Brotéria, 5, 6, 7 (three parts) (1906–08).

Monográfia de la familia de los Diláridos. Mem. Roy. Acad. Barcelona, 7, No. 17, pp. 619–671 (1909).

Monográfia de los Menoptéridos. Mem. Roy. Acad. Barcelona, 8, No. 18, pp. 1–70 (1910).

Nemopteridæ. Gen. Insectorum, fasc. 136, 23 pp. (1913).

Ascaláfidos sudamericanos. Revist. Chil. Santiago, 17, pp. 41–74 (1914). Dilaridæ. Gen. Insectorum, fasc. 156, 14 pp. (1914).

Les Myrméléonides d'Europe. Insecta, Rennes, 5, pp. 57–62 (1915).

Crisopids d'Europe. Arixius Inst. Cien. Barcelona, 3, No. 2, pp. 1–98 (1915).

Stitz, H. Neuroptera. In Tierwelt Mitteleuropas, 6, Lief. 1, pp. XIV 5–19 (1928).

Tillyard, R. J. Studies in Australian Neuroptera. Proc. Linn. Soc. New South Wales, **41**, pp. 269–332 (1916).

The Family Psychopsidæ (Australian). Proc. Linn. Soc. New South Wales, 43, pp. 750–786 (1918).

Revision of the Family Ithonidæ. Proc. Linn. Soc. New South Wales, 44, pp. 414-437 (1919).

Weele, H. W. Ascalaphiden. Coll. Zool. Selys-Longchamps, fasc. 8, Brussels (1908).

WITHYCOMBE, C. L. A Contribution towards a Monograph of the Indian Coniopterygidæ. Mem. Dept. Agric. India, 9, pp. 1–19 (1925).

## ORDER MECÓPTERA

# (PANORPÀTÆ, PANORPÌNA, MECÁPTERA)

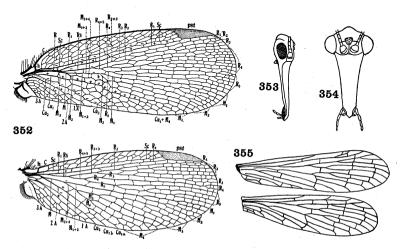
Small or moderate-sized, rather slender, insects with the head nearly always prolonged downwards to form a sort of beak. Eyes large; ocelli present or absent; mandibles small. Wings usually present, almost always long and narrow, similar; radius extensively branched; media and cubitus with few branches; crossveins only rarely numerous; anal area almost always very small, not separated; wing surface without scales. Antennæ long, many-jointed, slender. Prothorax small, free; meso- and metathorax similar. Legs long, slender, similar, fitted for running; coxæ large, pendant and approximate; tarsi five-jointed. Abdomen usually slender; cerci small; genitalia of male usually greatly swollen, forming a reflexed bulb. Metamorphosis complete, the larvæ caterpillar-like. Scorpion flies.

- Tarsi slender, filiform; the apical joint bearing two claws and not capable of folding back on the fourth.
   2 Tarsi each with a single claw; the apical joint modified for grasping, the fifth joint folding back on the fourth; legs extremely long and slender, wings usually present. (Bittacus, cosmop.; Neobittacus, Kalobittacus, Pàzius, neotrop.; Harpobittacus, Austr.) (BITTACÙSIDÆ)

BORÈIDÆ

Radial sector or media, or both with four branches or less; radial sector arising at or beyond the basal third of the wing; media

This family is divisable into three well-defined subfamilies which are accorded family rank by some workers.



Figs. 352-355. Mecoptera

- 352. Notiothauma, wings (Crampton) Notiothaumidæ.
- 353. Panorpa, head, lateral view (Packard) Panorpidæ.
- 354. Panorpa, head, frontal view (Packard) Panorpidæ.
- 355. Panorpa, wings. Panorpidæ.

- Radial sector four-branched; cubitus and main stem of media of fore wing touching only at a point or for a very short distance; larger species. (Chorista, Tæniochorista, austr.).
  - CHORISTINÆ
- 4. Ocelli present; wings with very dense and irregular venation, the anal area well developed; radius and media coalescent on the basal fifth of the wing. (Fig. 352). (Notiothauma, neotrop.) . . . . . . . . . . . . . . . . NOTIOTHAUMIDÆ
  - Ocelli absent; venation of wings more open and regular, the anal area slightly developed; radius and media not coalescent at the base of the wing. (Mérope, nearc.) ...... MERÓPIDÆ

#### LITERATURE ON MECOPTERA

- CARPENTER, F. M. Revision of Nearctic Mecoptera. Bull. Mus. Comp. Zoöl. Harvard Univ., 72, pp. 205-277 (1931).
- Enderlein, G. Ueber die Phylogenie und Klassification der Mecopteren. Zool. Anz., **35**, pp. 385–399 (1910).
- ESBEN-Petersen, P. Tables for determination of Danish Mecoptera and Megaloptera (in Danish). Kopenhagen Fauna and Flore, pp. 129–144 (1914); pp. 1–16, 41–47 (1915).
  - A Synonymic List of the Mecoptera. Entom. Meddel., **10**, pp. 216–242 (1915).
    - Mecoptera. Coll. Zool. Selys-Longchamps, fasc. 5 (1921).
- MIYAKE, T. Studies on the Mecoptera of Japan. Journ. Coll. Agric. Imp. Univ. Tokyo, 4, pp. 265–400 (1913).
- STITZ, H. Mecoptera. In Tierwelt Mitteleuropas, 6, Lief. 1, pp. XIV 19–22 (1928). TILLYARD, R. J. Nannochoristidæ. Proc. Linn. Soc. New South Wales, 17, pp. 284–301 (1917).

# ORDER TRICHÓPTERA

# (PHRYGANÒIDEA)

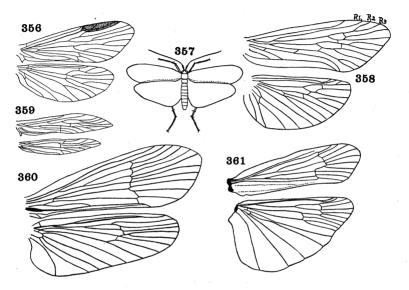
Small to medium-sized, slender, flying insects; head freely movable, vertical, eyes prominent, ocelli three or none, mandibles vestigial or absent, palpi prominent, antennæ thread-like, often very long; prothorax small, free; meso- and metathorax similar; wings more or less clothed with hairs, with many veins and a few crossveins; the hind wings often with a folded anal area; wings rarely reduced in size; legs similar, coxæ pendant and approximate, tibiæ always with spurs, tarsi five-jointed. Metamorphosis complete. Larvæ aquatic; usually with tufted tracheal gills; more or less caterpillar-like and usually living in cases constructed of small objects spun together with silk. Caddis (or Caddice) flies; larvæ called Caddis worms.

# Adults

1.	the front wings closely covered with projecting, clubbed hairs;
	marginal fringe of wings very long, that of hind wings longer;
	discal cell of hind wings open or wanting; wings usually very
	long and narrow, more or less pointed; antennæ at most as long
-	as the fore wings, usually much shorter and thickened; maxil-
	lary palpi five-jointed, strongly hairy, their last joint neither
	bowed nor ringed; ocelli usually present. (Fig. 359). (Hydróp-
	tila, Allotríchia, holarc.; Oxyethira, widespr.; Mortoniélla,
	neotrop.) HYDROPTÍLIDÆ
	Rarely minute species; fore wings without or with solitary, thick-
	ened, projecting hairs; marginal fringe shorter than width of
	wing; antennæ almost always longer than the fore wings2
2.	Ocelli present
	Ocelli absent
3.	Maxillary palpi strongly hairy or scaly; tibial spurs 1-3-4 or
	2-4-4 a few SERICOSTOMÁTIDÆ
	Maxillary palpi with only weak hairs, not scaly 4
4.	Last joint of maxillary palpi divided into false ring-joints,
	curved and as long as the third and fourth joints together;
	front tibiæ without or with two, three, or four spurs
	Last joint of maxillary palpi not ringed, rarely curved, subequal
	to the other joints (palpi absent in some Hydropsychidæ)6
5	Hind wings not dilated, in shape similar to the fore wings. (Chi-
٥.	márrha, cosmop.; Philopótamus, holarc.).
	PHILOPOTÁMIDÆ
	Hing wings with expanded anal angle, much wider than the fore
	wings. (Stenopsyche, indomal.; Parastenopsyche, palæarc.;
	Pseudostenopsýche, neotrop.) STENOPSÝCHIDÆ, part
6	Front tibiæ with one or no spur; middle tibiæ with three or two
0.	spurs
	Front tibiæ with two or three spurs, middle tibiæ with four spurs;
	maxillary palpi four- or five-jointed8
7.	Middle tibiæ widened, their tarsi broadened and armed with stiff
١.	hairs (see Couplet 8). (Agrypnètes, holarc.).
	PHRYGANÉIDÆ, part
	Middle tibiæ simple, normal; maxillary palpi of male three-jointed,
	of female five-jointed, but of similar structure in the two sexes.
	(Anabòlia, Stenóphylax, Limnéphilus, Hálesus, Apatània,
	holarc.) (Figs. 357, 361)LIMNEPHÍLIDÆ

8. Maxillary palpi five-jointed, the basal two joints very short and thick (Fig. 356). (Rhyacóphila, Glossosòma, Agapètus, holarc.; Hydrobiòsis, Psilochorèma, Austr.).

## RHYACOPHÍLIDÆ



Figs. 356-361. Trichoptera

- 356. Hydrobiosis, wings (Tillyard) Rhyacophilidæ.
- 357. Limnephilus, outline. Limnephilidæ.
- 358. Hydropsyche, wings. Hydropsychidæ.
- 359. Allotrichia, wings. Hydroptilidæ.
- 360. Phyllorheithrus, wings (Tillyard) Calamoceratidæ.
- 361. Limnephilus, wings. Limnephilidæ.

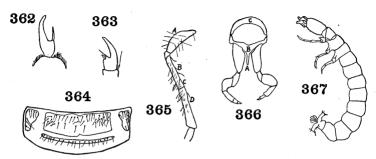
Maxillary palpi of male four-jointed, of female five-jointed, the joints cylindrical, the second joint not short, the palpi of the two sexes similar. (Neurònia, Phrygànea, Agrýpnia, holarc.)

## **PHRYGANÈIDÆ**

10.	Mandibles robust; maxillary palpi with the second joint short. (Stenopsychòdes, austr.) (see couplet 5).
	STENOPS ÝCHIDÆ, part
	Mandibles slender and sinuous; maxillary palpi with the second
	joint bulbously enlarged internally. (Polycéntropus, Plec-
	trocnèmia, Neureclípsis, holarc.; Polypléctropus, neotrop.,
	austr.) POLYCENTROPÓDIDÆ
11.	Last joint of the five-jointed, scarcely hairy, maxillary palpi an-
	nulate and arcuate
	Last joint of the usually strongly hairy maxillary palpi neither
	ringed nor curved
12.	First vein from the discal cell of the fore wing (anterior branch
	of the radial sector) forked; maxillary palpi long and thin.
	(Fig. 358). (Hydropsyche, Macronèma, cosmop.; Hydro-
	psychòdes, widespr.; Dipléctrona, holarc.; Smícrides, Am.).
	HYDROPS ÝCHIDÆ
	First fork (R <sub>2</sub> ) wanting in both fore and hind wings; first joint
	of the maxillary palpi small. (Psychomyia, Tinòdes, holarc.;
10	Lŷpe, Metalŷpe, palæarc.) PSYCHOMYÌIDÆ
13.	Both median and discal cells of fore wings present and closed; maxillary palpi five-jointed (Fig. 360). ( <b>Heteroplectron</b> , Am.;
	Ganonèma, Am., Ind.; Anisocéntropus, holarc., indoaustr.;
	Calamóceras, palæarc.; Phyllòicus, neotrop.; Phyllorhei-
	thrus, austr.)
	Median cell of fore wings absent
14.	Maxillary palpi of the male three-jointed, of the female five-
17.	jointed, of different structure in the two sexes; antennæ usu-
	ally thick, hairy and with large basal joint; wings thickly hairy,
	discal cell present. (Brachycentrus) SERICOSTOMÁTIDÆ
	Maxillary palpi of both sexes five-jointed
15.	Discal cell of both wings absent, neuration of the two sexes usually
10.	different, apical veins few. (Molánna, holarc., indomal.;
	Beræa, holarc.; Molannòdes, Beræòdes, palæarc.).
	MOLÁNNIDÆ
	Discal cell of fore wings present
16.	Middle tibiæ with two spurs; discal cell of hind wings almost
	always open or absent, only the upper branch of the radial
	sector forked, only the first apical fork present; joints of maxil-
	lary palpi uniform; antennæ long and slender. (Leptócerus,
	Œcetis, Setòdes, widespr.; Mystácides, holarc.; Leptocélla,
	Am.; Notanatólica, indoaustr.)LEPTOCÉRIDÆ

	Middle tibiæ usually with four spurs; discal cell of hind wings closed, both branches of radial sector of fore wings forked, at least the first and second apical forks present; basal joint of antennæ large. (Neróphilus, nearc.; Psilotrèta, holarc.; Odontócerum, palæarc.; Marília, Am.; Barypénthus, neotrop.) ODONTOCÉRIDÆ
	Larv x
1.	Body campodeiform; head held straight forward, forming a continuation of the long axis of the body; abdomen depressed, the sutures between the segments deeply impressed; tracheal gills usually absent; larvæ generally not constructing a movable case (Fig. 367)
	sutures between the segments feebly impressed; tracheal gills usually present; larvæ always living in a movable, tubular
2.	case
	Abdomen not much wider than the thorax, much larger species
	usually with only the prothorax heavily chitinized
3.	Last abdominal segment with a chitinized shield above.
	RHYACOPHÍLIDÆ
	Last abdominal segment without a chitinized shield 4
4.	Mandible with numerous bristles on the outer edge; tracheal gills
	present along the sides of the abdomen; legs very unequal.
	HYDROPSÝCHIDÆ
	Mandible with only two bristles on the outer edge; tracheal gills absent
5	Labrum soft, whitish, retractile under the edge of the clypeus.
υ.	(Fig. 367) PHILOPOTÁMIDÆ
	Labrum chitinized, yellowish or brownish
6.	Claws of legs long and slender (Fig. 362), nearly straight, with only one basal spur
	Claws of legs short, stout, curved
7.	Claws with a single basal spur STENOPS ÝCHIDÆ
	Claws of middle and hind legs each with two basal spurs (Fig.

8. Labrum with a transverse row of many stout bristles before the middle; tracheal gills filamentous .... CALAMOCERÁTIDÆ Labrum not thus armed, usually with scattered bristles on the disc and several pairs of spines on the front margin .................9



Figs. 362-367. Trichoptera, Larvæ

- 362. Phylocentropus, tarsal claw of front leg (Krafka) Polycentropodidæ.
- 363. Timeodes, tarsal claw (Rousseau) Psychomyiidæ.
- 364. **Psilotreta**, dorsal view of metathorax showing chitinous plates. (Krafka)
  Odontoceridæ.
- 365. Leptocella, front leg. a, coxa; b, trochanter; c and d, femur. (Krafka) Leptoceridæ.
- 366. **Arctæcia**, anterior view of prothorax in section showing prosternal horn (Krafka) Sericostomatidæ.
- 367. Philopotamus, larva (Rousseau) Philopotamidæ.
- 9. Labrum much longer than broad. Pronotum and mesonotum chitinized; metanotum with four chitinized plates (Fig. 364).

# ODONTOCÉRIDÆ

- 10. Body suberuciform; mesonotum generally soft like the metanotum, rarely with two small chitinous plates; abdominal constrictions well marked; lateral gills on segments 2 to 7 covered with black hairs. Larval case tubular, open at both ends, straight or but little bent; living in tranquil water.

# **PHRYGANÈIDÆ**

11. Mesonotum entirely corneous; metanotum with three pairs of chitinous plates; middle legs stouter and longer than the hind ones; gills never with black hairs..... LIMNEPHÍLIDÆ

Mesonotum not entirely corneous; metanotum generally soft
middle legs not longer than the hind ones
12. Femur of hind leg divided into a short basal and long apical piece
(Fig. 365); right mandible without inner bristles; no accessory
bristles on back of mandibles LEPTOCÉRIDÆ
Femur not divided
13. Prosternum with a horn or spine that projects downward between
the front coxe (Fig. 366) Some <b>SERICOSTOMÁTID</b>
Prosternum unarmed14
14. First abdominal segment with three nipple-shaped protuberances, one above and one at each side
First abdominal segment narrower than the following ones and without such protuberances Some <b>SERICOSTOMÁTID</b>
15. Body slender; claws of anal prolegs small, simple.
MOLÁNNIDA
MOLÁNNIDÆ Body robust; claws of anal prolegs stout, formed of two or three
MOLÁNNIDÆ
MOLÁNNIDÆ Body robust; claws of anal prolegs stout, formed of two or three large superimposed hooksSome SERICOSTOMÁTIDÆ
MOLÁNNIDÆ Body robust; claws of anal prolegs stout, formed of two or three
MOLÁNNIDÆ Body robust; claws of anal prolegs stout, formed of two or three large superimposed hooksSome SERICOSTOMÁTIDÆ
MOLÁNNIDÆ Body robust; claws of anal prolegs stout, formed of two or three large superimposed hooks Some SERICOSTOMÁTIDÆ  LITERATURE ON TRICHOPTERA  Banks, N. Genera of Nearctic Leptoceridæ. Trans. American Entom. Soc. 25, pp. 199–218 (1898).
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Krafka, J. Key to the families of Trichopterous larvæ. Canadian Entom., 47, pp. 217–225 (1915).

IWATA, M. Trichopterous larvæ from Japan. Annot. Zool. Japonenses, 11,

HAGEN, H. A. Phryganidarum synopsis. Verh. k. k. zool.-bot. Ges. Wien,

Beiträge zur Kenntnis der Phryganiden. Verh. k. k. zool.-bot. Ges.

pp. 125–151 (1873).

pp. 202–233 (1927).

**14**, pp. 799–890 (1864).

Wien, 23, pp. 377-452 (1873).

- Kuwayama, S. The Stenopsychidæ of Japan. Ins. Matsumarana., 4, pp. 109–119 (1930).
- Lestage, J. A. Trichoptera, in Rousseau, Larves et Nymphes aquatiques des Insectes d'Europe, pp. 343–964 (1921).

Catalogue des Trichoptères d'Afrique. Rev. Zool. Africaine, **6**, pp. 251–335 (1919) and Ann. Soc. Entom. Belgique, **59**, pp. 130–135 (1919).

Note Trichoptérologique. Bull. and Ann. Entom. Soc. Belgique, 65, pp. 363–386 (1926).

- LLOYD, J. T. Larvæ of North American Trichoptera. Bull. Lloyd Libr., Cincinnati, Entom. Ser., No. 1 (1921).
- Martynov, A. V. Preliminary revision of Phryganeidæ. Ann. Mag. Nat. Hist. (9), **14**, pp. 209–224 (1924).

On the family Stenopsychidæ. Eos, 2, pp. 281–308 (1926).

- McLachlan, R. A monographic revision and synopsis of the Trichoptera of the European fauna. London. Van Voorst (1874–80).
- Morton, K. J. North American Hydroptilidæ. Bull. New York State Mus., No. 86, pp. 63–75 (1905).
- Ulmer, G. Trichoptera. Gen. Insectorum, fasc. 60, 289 pp. (1907).

Trichoptera. Süsswasserfauna Deutschlands, Lief. 5 and 6, 326 pp. (1909). Literature on Trichoptera (1910–14). Zeits. wissensch. Insektenbiol., 15 (various parts) (1919–21).

Trichoptera. In Tierwelt Mitteleuropas, 6, Lief. 1, pp. XV 1-46 (1928).

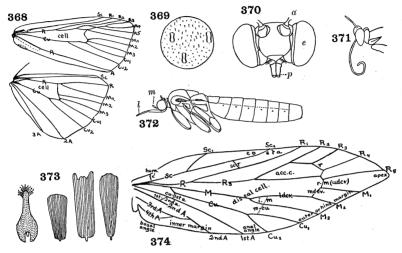
# ORDER LEPIDÓPTERA

# $(GLOSS \stackrel{?}{A}TA)$

Rather large, sometimes small, or very large insects; wings and body thickly clothed with scales that form a color pattern, the wings opaque, with the venation obscured by the scales; scaly covering rarely restricted to certain portions of the wings or absent in a very few unusual forms; wings very rarely absent. Antennæ long, many-jointed, variously modified, filiform, pectinate or clubbed; ocelli sometimes present. Mouthparts suctorial, frequently vestigial or absent, when not in use coiled under the head; the maxillæ incorporated into an unjointed tongue; mandibles absent, except in a few primitive forms; palpi usually well developed, the labial ones generally larger than the maxillary. Prothorax small; wings large, membranous, similar, the fore pair somewhat longer; venation complete, but not complex, few crossveins. Legs similar, tarsi ordinarily five-jointed. No cerci. Metamorphosis very great: larvæ with biting mouthparts, usually caterpillar-like, and with paired false-legs on some of the abdominal segments in addition to three pairs of thoracic legs; larvæ almost always plant-feeders. Moths, Butterflies and Skippers.

#### Adults

1. Fore and hind wings (if wings are absent or greatly reduced in size, see couplet 169) with four or five radial veins, rarely with three, Sc and R<sub>1</sub> separate (Fig. 374); wings of similar shape,



Figs. 368-374. Lepidoptera

- 368. **Noctua,** wings (Forbes): Sc, subcosta, R, radius, M, media, Cu, cubitus (their branches indicated by numbers), A, 2A, 3A, anal veins; cell, discal cell. Noctuidæ.
- 369. Portion of bleached wing membrane, showing attachments of three scales and numerous aculeæ.
- 370. **Danais**, head in frontal view: a, base of antenna; e, eye; p, base of proboscis. Danaidæ.
- 371. **Hamadryas**, head, prothorax and part of mesothorax (Scudder) Nymphalidæ.
- 372. **Crambus,** lateral outline of body (Fernald): m, maxillary palpus; l, labial palpus.
- 373. Scales from the wings of various Lepidoptera (Scudder).
- 374. **Micropteryx** (Forbes): Sc, subcosta; R, radius; M, media; Cu, cubitus; A, anal (their branches indicated by numbers); hum, humeral crossvein; udcv, upper discocellular vein (radio-medial crossvein); ldcv, lower discocellular vein; i, intercalated cell; ac. c, accessory cell. Micropterygidæ.

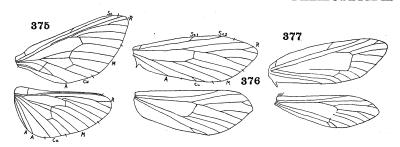
more or less pointed at tip, with ten or more veins, the membrane with minute spines (Fig. 369); fore wings with a thumb-

	like lobe at basal angle (jugum); a spiral proboscis never developed. (Suborder JUGATÆ) (MICROPTER YGÌNA, HOMONEÙRA, ARCHILEPIDÓPTERA, ZEUGLÓPTERA)2
	Fore and hind wings dissimilar in shape and venation, hind wings with Sc and $R_1$ fused at least at tip (not to be confused with $R_s$
	which is usually the only free radial vein (Fig. 368)), rarely two free radial veins present, at most six veins arising from the cell;
	jugum and mandibles not developed. (Suborder FRENATÆ) (HETERONEÙRA)8
2.	margin; small species, wing-expanse about one-half inch; tibial spurs usually present; jugum usually minute, frenulum present. (Superfamily MICROPTERYGÒIDEA). (MICROJUGÀTÆ,
	JUGOFRENÀTÆ)
	one to nine inches; tibial spurs usually wanting; jugum usually underlapping the hind wing, no frenulum. (Superfamily (HEPIALÒIDEA). (MACROJUGÀTÆ)
3.	T,
	functional mandibles present; Sc forked near middle. ( <b>Epi-martýria</b> , No. Am.; <b>Micrópteryx</b> (=Eriocéphala) (Fig. 374), holarc.; <b>Sabatínca</b> , Austr.). (ERIOCEPHÁLIDÆ).
	MICROPTERÝGIDÆ
	Middle tibiæ with one or two spurs; mandibles vestigial or wanting; Sc simple or forked near tip4
4.	wing; jugum large, overlapping the hind wing; scales not scal-
	loped nor coarsely striated. (Mnemónica, holarc.; Eriocrània, holarc.; Neopseùstis, indomal.)
	Middle tibiæ with two spurs; Sc simple; jugum much reduced, underlapping the hind wing; scales with scalloped border and coarse striation. (Mnesarchæa, Austr.).
	Middle tibiæ with two spurs; Sc simple; jugum much reduced, underlapping the hind wing; scales with scalloped border and coarse striation. (Mnesarchæa, Austr.).  MNESARCHÆIDÆ
5.	Middle tibiæ with two spurs; Sc simple; jugum much reduced, underlapping the hind wing; scales with scalloped border and coarse striation. (Mnesarchæa, Austr.).  MNESARCHÆIDÆ  R <sub>1</sub> arising near base of wings, discal cell containing a large intercalated cell formed by the basal petiolate branch of the media,
5.	Middle tibiæ with two spurs; Sc simple; jugum much reduced, underlapping the hind wing; scales with scalloped border and coarse striation. (Mnesarchæa, Austr.).  MNESARCHÆIDÆ  R <sub>1</sub> arising near base of wings, discal cell containing a large intercalated cell formed by the basal petiolate branch of the media, i.e. three cells present (Fig. 375)
	Middle tibiæ with two spurs; Sc simple; jugum much reduced, underlapping the hind wing; scales with scalloped border and coarse striation. (Mnesarchæa, Austr.).  MNESARCHÆIDÆ  R <sub>1</sub> arising near base of wings, discal cell containing a large intercalated cell formed by the basal petiolate branch of the media, i.e. three cells present (Fig. 375)

ANOMOSÉTIDÆ

Tibial spurs absent (Fig. 376). (Palæoses, austr.).

PALÆOSÉTIDÆ



Figs. 375-377. Lepidoptera

- 375. Phassus, wings (Hampson) Hepialidæ.
- 376. Palæoses, wings (Turner) Palæosetidæ.
- 377. Anomoses, wings (Turner) Anomosetidæ.

Wings entire, not cleft nor divided into finger-like divisions, rarely

(Gelechiidæ) the fore wings moderately cleft	12
10. Wings divided into two to four divisions	
Each wing divided into six plumes; small, silve	ry white moths.
(Orneòdes, widespr.) (Fig. 384)	
380 381 38	
383 388	
379	
₩ 384	
Figs. 378–385. Lepidoptera	
378. Plumose antenna of moth (Duncan).	
379. Pectinate antenna of moth (Duncan).	
380. Melittia, head, in lateral view (Beutenmüller) Ægeri	idæ.
381. Bembecia, middle leg (Beutenmüller) Ægeriidæ.	
382. Melittia, wings (Beutenmüller) Ægeriidæ.	
383. Pterophorus, wings (Berlese) Pterophoridæ.	
384. Orneodes, wings (Berlese) Orneodidæ.	
385. Platyptilia, wings (Fernald) Pterophoridæ.	
11. Fore wings divided into two plumes, rarely four, three; small, delicate moths, usually prettily tilus, cosmop.; Platyptília, cosmop.; Pter	colored. (Oxýp-
spr.). (Figs. 381, 385, 404). (ALUCÍTIDÆ).	EROPHÓRID <i>Æ</i>
Each wing divided into two plumes. (Cenóloba, (See couplet 43d) OXYCHI	

12. Underside of hind wing with a double series of enlarged and divergent scales along the cubital vein; tibiæ exceptionally long, thin and with long spurs. (Agdístis, palæarc., ethiop.).

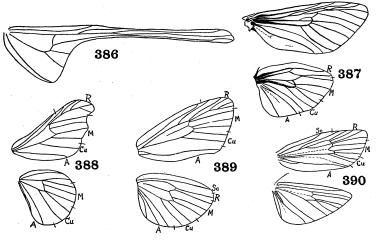
13. Wings in large part transparent and usually devoid of scales, ex-

	cept on margins and veins; fore wing narrow, at least four times as long as wide; inner margin of fore wing and costal margin of hind wing with a row of recurved, interlocking spines; Sc of hind wings close to cell and to next vein, hidden in a fold and apparently absent; frenulum well developed; ocelli present; wasp-like, day-flying moths (Figs. 380, 381, 382). Clear-wing moths. (Paranthrène, widespr.; Trochílium, holarc., ethiop.; Ægèria, cosmop.). (SESÌIDÆ).
	ÆGERÌIDÆ
	Wings scaled throughout, or if clear, the fore wings are triangular;
	wings not interlocking by series of spines; Sc of hind wing pres-
14.	ent, though sometimes close to or in part fused with R14 Antennæ thin, swollen at tip to form a more or less distinct club,
17.	as in the butterflies, or even recurved at tip, as in the skippers. 15
	Antennæ variously modified, if swollen subapically or toward
	the middle they gradually taper on distal portion18
15.	Fore wing with some branches of R stalked
	Fore wing with all veins arising from discal cell, medial stem reduced; both wings with first anal vein lacking; eyes not ciliated; male alone with frenulum (see couplet 155). ( <b>Euschèmon</b> ,
10	austr.)
16.	Fore wings with $M_2$ arising nearer $M_3$ than $M_1$
	Fore wings with $M_2$ arising midway between $M_1$ and $M_3$ , or closer to $M_1$ ; hind wings with first A wanting. African. ( <b>Apopró</b> -
	genes, Pemphegóstola)APOPROGÉNIDÆ
17.	Both wings with first A present, discal cell small and closed; pro-
	boscis developed. (Cástnia, neotrop.; Synèmon, Austr.).
	CASTNIIDÆ
	Both wings with first A reduced; discal cell open; proboscis vesti-
	gial. ( <b>Táscina</b> (= Neocástnia, India)). (NEOCASTN <b>Ì</b> IDÆ).
	TASCÍNIDÆ
18.	Hind wings with three anal veins (if less than three anals, small
	species with narrow wings, the hind pair with reduced venation
	and bearing a long fringe of hairs on the hind margin almost
	as wide as, or wider than, the wing, and the tibial spurs more than twice the width of the tibiæ); fore wings usually with first
	anal vein complete, <i>i.e.</i> usually two anal veins reaching the
	margin. (MICROFRENATÆ)
	Hind wings with two anal veins, rarely with one, in addition to
	11 11 1011 1 1 1 1 1

a possible unthickened fold in the membrane, or in the Aus-

	tralian genus Oxychirota both wings lack all anals; wings
	almost never very narrow, the hind wing not more than half
	longer than wide, except rarely in large species when a tail-like
	projection is developed, and not long-fringed; fore wings usu-
	ally with but one complete anal vein, when accessory cell is
	present it is not completely contiguous with the discal cell (if
	accessory cell is completely contiguous refer to couplet 109,
	Cecidosidæ). (MACROFRENATÆ)
19.	Hind wings with veins Sc + R <sub>1</sub> and R <sub>s</sub> widely separate beyond
	discal cell
	Hind wings with veins Sc+R <sub>1</sub> and R <sub>s</sub> fused or very closely parallel
	for a greater or less distance between the end of the discal cell
	and the tip of the wing, the base of R sometimes evanescent.
	(If R <sub>1</sub> of fore wing is as long as cell, see Drepanidæ, couplets
	65, 81, 95). (PYRALÍDIDÆ, s. lat.)
20.	Hind wings ribbon-like, with a long apical tail (Fig. 386). (Hi-
	mantópterus, Semióptila African). HIMANTOPTÉRIDÆ
	Wings normal
21.	Hind wings with veins Sc + R <sub>1</sub> and R <sub>s</sub> fusing to near the end of
	the discal cell, or fusing beyond the middle of the cell, or these
	veins coincident throughout
	Hind wings with veins $Sc + R_1$ and $R_s$ separate from the base, or
	fusing only a short distance along the discal cell, the fusion
	located at the base or before the middle of the cell, or some-
	times connected by a bar
22.	Proboscis well developed; wings thinly scaled, translucent; hind
	wings with basal part of R represented as a spur in the cell,
	or entirely lost. (Acolòithus, Pyromórpha, Am.) (see coup-
	let 45). (ZYGÆNIDÆ, of authors) PYROMÓRPHIDÆ, part
	Proboscis obsolete; hind wings with Sc + R <sub>1</sub> and R <sub>8</sub> separate in part;
	wings heavily and loosely clothed with soft scales, mixed with
	curly hair in the northern species; hind wings (in American
	species) with R free at base. Flannel moths. (Norape, Mega-
	$ extbf{lopŷge}$ ). $(LAG\grave{O}ID\pounds)$ MEGALOP ÝGIDÆ
	Proboscis and palpi absent; veins Sc + R <sub>1</sub> and R <sub>8</sub> coincident through-
	out. (Engyóphlebus, ethiop.)ENGYOPHLÉBIDÆ
23.	The fringe on the anal angle of the hind wings not or but slightly
	longer than elsewhere; tibial spurs at most about as long as the
	width of the tibiæ24
	The fringe on the anal angle of the hind wings distinctly longer
	than elsewhere; tibial spurs more than twice the width of the

	tibiæ; when the accessory cell is present its longest side is con-
	tiguous with the discal cell. (TINEÒIDEA)
24.	Fore wings with accessory (radial) cell
	Radial cell not formed
25.	Mouthparts vestigial
	Mouthparts usually developed, with scaled proboscis; tibial spurs
	long; small to minute moths. (TINEOIDEA)



Figs. 386-390. Lepidoptera

- 386. Himantopterus, hind wing (Westwood) Himantopteridæ.
- 387. Prionoxystus, wings (Comstock and Needham) Cossidæ.
- 388. Bombyx, wings. Bombycidæ.
- 389. Metarbela, wings (Hampson) Metarbelidæ.
- 390. Simaethis, wings (Spuler) Glyphipterygidæ.

28.	Wings strong and more or less lanceolate; body heavy and surpassing the hind wings; median cell and strong medial stem nearly always present in discal cell of each wing; larvæ borers. Carpenter moths
	DALCERIDÆ
29.	Fore wings with first and second anal veins free, or first anal
	wanting30
	Fore wings with first and second anal veins connected by a cross-
00	vein near the margin. (Gívira) HYPÓPTIDÆ
30.	Fore wings with first anal vein present
01	Fore wings with first anal vein absent
31.	Frenulum present
32.	Frenulum absent. (Ratárda, indoaustr.) RATÁRDIDÆ
o∡.	Hind wings with R <sub>s</sub> and M <sub>1</sub> widely separate; palpi very short. ( <b>Zeùzera</b> (Z. pyrìna, Leopard moth, holarc., ind.), <b>Phragma</b> -
	tècia, palæarc.)
	Hind wings with R <sub>s</sub> and M <sub>2</sub> stalked beyond apex of discal cell or
	close together; palpi upturned to middle of front (Fig. 387).
	(Cóssus (= $Trýpanus$ ), cosmop., <b>Prionoxýstus</b> ).
	CÓSSIDÆ
33.	Frenulum present. (Argyrótypus, Chrysótypus, Madagascar).
	(CHRYSOTÝPIDÆ) ARGYROTÝPIDÆ
	Frenulum absent; tropical distribution (Fig. 389). (Metarbèla;
	Teràgra; Salágena). (ARBÉLIDÆ, HOLLÁNDIDÆ, TER-
	ÁGRIDÆ) METARBÉLIDÆ
34.	Frenulum absent or vestigial, humeral angle of hind wing more or
	less expanded; M2 of fore wings arising midway between M1
	and M <sub>3</sub> or closer to M <sub>1</sub> ; chætosema absent; moderately large
	moths with broad hind wings
	Frenulum present (if absent in moths of small size with narrow
	and more or less oblong wings, refer to couplet 90) 37
35.	Fore wings with radial branches $R_{2+3+4+5}$ united on a common
	stalk; hind wings with Sc and R connected by a bar (Fig. 388).
	(See couplet 64) BOMBÝCIDÆ
	Fore wing with radial branches $R_{2+3}$ , $R_{4+5}$ stalked independently
	of $R_{1+2}$ ; hind wings with Sc and R free beyond base36

36.	The five branches of R very long, occupying more than one-third
	the apical edge of the wing, only R <sub>3</sub> and R <sub>4</sub> on a short stalk,
	the other radial branches free; wings large. (Chrysopóloma,
	Éctropa, ethiop.). (ECTRÓPIDÆ). CHRYSOPOLÓMIDÆ
	Radial branches occupying one-fourth of the apical edge of the
	fore wings, $R_{2+3}$ , or $R_{4+5}$ , or $R_{3+4+5}$ stalked, tip of fore wings
	extended and rather pointed, the apical margin sinuate. Ameri-
	can, mostly neotropical. (See couplet 67) MIMALLÓNIDÆ
37.	
	Cu <sub>1</sub> united for a considerable distance beyond the discal cell,
	R <sub>2,3,4,5</sub> united, and first A absent; hind wings with Sc free from
	R <sub>s</sub> from base, and first A evanescent on basal half. (See coup-
	let 56). (Phryganídia, Cal.)
	Fore wings with $M_2$ arising closer to $M_3$ than to $M_1$ , thus causing
	the cubitus to appear four-branched
38	Hind wings very small, with Sc and R separate beyond the base;
00.	hind tibiæ with strong middle and apical spurs; antennæ di-
	lated apically; chætosema absent. African. (Charídea, Toòsa).
	CHARIDÈIDÆ
	Hind wings with Sc and R connected by a bar, or by fusion before
	the middle of the discal cell
39	Fore wings with all five branches of the radius arising from the
	discal cell. (Hyblæa, nearc.)
	Fore wings with some of the radial branches absent or coalesced
	beyond the end of the discal cell
40.	Fore wings with first and second anals connected by a crossvein,
	or fusing before the tip. (Fig. 391). Bagworm moths. (See
	couplet 46). (Pachythèlia, palæarc.; Fùmea, palæarc.).
	PSÝCHIDÆ, part
	Fore wings with first and second anals not connected, nor apically
	fusing, or 1A absent
41.	Proboscis well developed; chætosema present; antennæ dilated
	or pectinate in the male. (Cyclòsia; Gíngla; Campylòdes;
	Erásmia; Zygæna, palæarc.; Pròcris, palæarc., indoaustr.;
	Chalcòsia; Aglàope). (Including CHALCOSÌIDÆ).
	ZYGÆNIDÆ
	Proboscis and palpi much reduced
42.	Fore wings with R <sub>3</sub> , R <sub>4</sub> and R <sub>5</sub> stalked or united. (Sabine; Pho-
-	bètron; Eùclea; Adonèta; Prolimacòdes; Heterogènea, pa-
	læarc.; Ápoda). (COCHLIDÌIDÆ, HETEROGENÈIDÆ, LIM-
	ACÓDIDÆ) EUCLÈIDÆ

Fore wings with only three simple branches of the radius, all arising from the cell. (**Heterógynis** &, **Somábrachys** &, S. Eur.). (See couplet 170). (EPICNOPTER ÝGIDÆ).

#### HETEROGÝNIDÆ

43. Antennæ thickened, spindle-shaped, sometimes hooked or recurved at the tip, the joints usually carinated beneath, sometimes pectinated; hind wings with Sc and R<sub>s</sub> connected by a strong crossvein near the middle of the discal cell, then extending closely parallel to the end of the cell or beyond; proboscis and palpi present; chætosema absent; stout, often large moths, with rather narrow wings, the hind pair much shorter than the fore wings. Hawk moths, Humming bird moths. (Phlegethóntius (= Protopárce); Sphínx (= Hylòicus), cosmop.; Hémaris



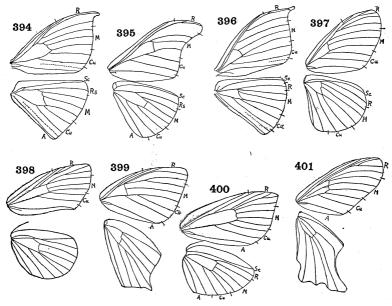
Figs. 391-393. Lepidoptera

- 391. Solenobia, wings (Spuler) Psychidæ.
- 392. Harrisina, wings (Jones) Pyromorphidæ.
- 393. Hemiceras, wings (Hampson) Notodontidæ.

46.	Fore wings with the anal veins more or less fused or connected
	by a crossvein so as to end as a single vein. (Eurycýttarus,
	Thyridópteryx). (See couplet 40)PSÝCHIDÆ, part
	Fore wings with a single complete anal vein, i.e. 2A, the first
	anal always reduced, at most represented by a fold, and the
	third anal short or wanting, generally connecting with the
	second anal so that the latter appears to have a basal fork47
47.	Fore wings with M <sub>2</sub> arising from the middle of the end of the
	discal cell, or in front of the middle, i.e. the cubitus apparently
	three-branched
	Fore wings with M <sub>2</sub> arising behind the middle of the discal cell,
	<i>i.e.</i> the cubitus apparently four-branched73
48.	Hind wings with Sc strongly angled, or rarely swollen and sinuous,
	at the base, usually with a strong humeral brace-vein at the
	bend, thence very close to or fusing with R <sub>s</sub> for a greater or
	less distance along the cell; palpi and proboscis well developed;
	chætosema present
	Hind wings with Sc and R <sub>s</sub> fused from base to beyond the middle
	of the wing, swollen at base, then rapidly diverging; rather
	small, very slender species, with finely scaled wings, the fore
	pair narrow, the hind pair broad; the apparent three-branching
	of the cubitus of the fore wings due to the absence of $M_2$ and $M_3$ .
	(See couplet 90) LITHOSÌIDÆ, part
	Hind wings with Sc straight or gently curving at base, separate
	from R <sub>s</sub> , no connecting bar present; proboscis often weak or
	undeveloped; chætosema weak or absent
49.	Antennæ dilated toward tip; eyes hairy and ciliated. (Sematura,
	Madagasc.; Anurápteryx, Am.). (MA NIDÌIDÆ).
	SEMATŪRIDÆ
	Antennæ slender or feathered, if dilated toward tip the eyes are
50	bare. (GEOMÉTRIDÆ, s. lat.)
50.	Eyes small and oval. (Bréphos, holarc.; Eránnis, holarc.). (MONOCTENÌIDÆ) BRÉPHIDÆ
	Eyes round and usually large
51	Hind wings with $M_2$ reduced or absent, the cubitus apparently
51.	three-branched. (Macària (=Semiothisa), widespr.; Boármia,
	cosmop.; Selidosèma, cosmop.; Geómetra, holarc.; Abráxas,
	holarc., ind.; Campæa (= Metrocámpa), holarc.; Énnomos
	(=Eugònia), holarc.; Paleácrita (P. vernàta, (Spring canker-
	worm)). (BOARMIIDÆ, SELIDOSEMÁTIDÆ).
	GEOMÉTRIDÆ, s. str.
	CHOTTELLULDIE, S. St.

	Hind wings with M <sub>2</sub> well developed, the cubitus apparently four-
	branched52
52.	$M_2$ of hind wings and often of both pairs arising very close to $M_1$ .
	(Chlorissa (= Nemòria), $widespr.$ , Euchlòris, $widespr.$ ;
	Hemíthea, Mesòthea). (GEOMÉTRIDÆ Meyrick, Tillyard).
	HEMITHEIDÆ
	$M_2$ of hind wings arising nearly midway between $M_1$ and $M_3$ 53
53.	Hind wings with Sc and Rs fused to or beyond the middle of the
	cell, or connected by a bar beyond the middle, or fused for a
	short distance toward the end of the cell. (Hydriómena,
	widespr.; <b>Tephroclýstis</b> (=Eupithècia), widespr.; <b>Trichóp</b> -
	teryx, holarc.; Xanthorhòe, cosmop.; Laréntia, Eucymá-
	toge, Eucéstia, palæarc., ind.). (HYDRIOMÉNIDÆ).
	LARENTÌIDÆ
	Hind wings with Sc and R <sub>s</sub> free, or with a connecting bar, or fused
	for a short distance before the middle of the discal cell $\dots 54$
54.	Hind wings with Sc free from R <sub>s</sub> , though close to it along the
	second fourth of the discal cell. (Dýspteris, nearc.).
	ŒNOCHROMÁTIDÆ
	Hind wings with Sc and R <sub>s</sub> fused for a short distance. (Ptychó-
	poda, Leptómeris, cosmop.; Leucophthálmia, holarc.;
	Cinglis $(=Acidàlia)$ ). $(STÉRRHIDÆ)$ ACIDALÌIDÆ
55.	Frenulum well developed, more than one-fifteenth the length of
	the wing
	Frenulum vestigial or absent; hind wings with Sc never fusing
	with R <sub>s</sub> , but sometimes connected by a weak bar
56.	• • • • • • • • • • • • • • • • • • •
	tance beyond the cell, stem of M indicated through the cell;
	fore wing with R <sub>2-5</sub> or R <sub>3-5</sub> stalked; proboscis present; tympana
	small and subdorsal, or absent; hind tibiæ with both middle
	and apical spurs; slender, butterfly-like species. Neotropical.
	(Myònia, Orícia, Tithraùstes, Phryganídia). (See couplet
	37)
E 77	Of other conformation
<b>57</b> .	
	fore wings with R <sub>5</sub> stalked with M <sub>1</sub> and well separated from
	R <sub>4</sub> ; proboscis present; no tympanum. Tropical. ( <b>Epiplèma</b> ,
	Medùsia, Melèaba) EPIPLÉMIDÆ Hind wings with Sc close to $R_s$ ; fore wings with $M_1$ not stalked
	with R <sub>s</sub>

58.	Hind wings with R <sub>s</sub> and M <sub>1</sub> stalked beyond the cell; fore wings
	commonly with a radial areole. (If $R_1$ of fore wings is much
	shorter than discal cell, see couplet 79, Pyralididæ) 59
	Hind wings with M <sub>1</sub> free, R <sub>s</sub> arising before or at the end of the
7 g	discal cell
59.	Proboscis present, often weak, sometimes absent; tympanum
	present; fore wings fully scaled, usually with R <sub>2-5</sub> stalked, often
	with a radial areole. (Fig. 393). Principally neotropical.
	(Cerùra, palæarc., Am., Indo-austr.; Notodónta, Drymònia,
	holarc.; Stauròpus, palæarc., indomal.; Phalèra, palæarc.,
	indomal.; <b>Datàna</b> (D. ministra, Yellow necked apple worm);
	Schizura (S. concinna, Red humped apple worm); Hetero-
	cámpa). (CERÙRIDÆ)NOTODÓNTIDÆ
	Proboscis wanting; tympanum absent; fore wings with $R_{2,3}$ and
	R <sub>4</sub> , 5 stalked together; wings of northern species with clear
	spots. (See couplet 63). (Apatelòdes, Thaumetopœa (=Cne-
	thocámpa), Euptérote)). (THAUMETOPŒIDÆ).
	EUPTERÓTIDÆ
60.	
	middle of the wing; hind wings with cubitus apparently three-
	branched. (Áxia, Epicimèlia, S. Eur.) AXÌIDÆ
	Discal cell not or scarcely extending beyond the middle of the
	wing; hind wings with cubitus apparently four-branched; fren-
	ulum of male knobbed. (Fig. 398). (See couplet 76). (Polýploca,
	Palimpséstia, palæarc.; Habrósyne, holarc.; Cymatóphora.)
	CYMATOPHÓRIDÆ, POLYPLÓCIDÆ THYATÍRIDÆ
61	Fore wings with $M_1$ stalked with $R_5$ and hind wings with two
. 01.	complete anal veins; if M <sub>1</sub> and R <sub>5</sub> are short-stalked, the second
	anal diverges widely from Cu <sub>2</sub> and a vestige of the first anal is
	present
	Fore wings with $M_1$ arising from R at or before the end of the
	discal cell, or $M_1$ is free from $R_5$ ; if $M_1$ and $R$ are stalked beyond
	the cell, the hind wings have but one complete anal vein65
62.	
04.	Frendum vestigiai
	(E. imperiàlis, Imperial moth); Citherònia (=Ceratocámpa);
	Sýssphinx; Anisòta; Adelocéphala). (CERATOCÁMPIDÆ,
	SYSSPHÍNGIDÆ)CITHERONÌIDÆ
63.	Hind wings with Sc and R <sub>s</sub> close together but not joined by a bar;
00.	fore wings with $R_2$ , 3 and $R_4$ , 5 joined on a common stalk. (See
	couplet 59) EUPTERÓTIDÆ, part



Figs. 394-401. Lepidoptera

- 394. Janiodes, wings (Jordan) Cercophanidæ.
- 395. Antheræa, wings (Hampson) Saturniidæ.
- 396. Oxytenia, wings (Jordan) Oxyteniae.
- 397. Brahmæa, wings (Hampson) Brahmæidæ.
- 398. **Thyatira**, wings (Hampson) Thyatiridæ.
- 399. **Urapteroides,** wings (Hampson) Uraniidæ.
- 400. Addæa, wings (Hampson) Thyrididæ.
- 401. **Epicopeia**, wings (Hampson).
- 65. Hind wings with the cubitus apparently three-branched. . . . . . . 66
  Hind wings with the cubitus apparently four-branched; one anal
  vein; outer margin of fore wings strongly sinuous; tympanum
  present. (Fig. 405). (See couplets 81, 95) . **DEPÁNIDÆ**, part

66.	Fore wings with R <sub>4</sub> , 5 stalked
	Fore wings with $R_5$ free, arising from the discal cell71
67.	Fore wings with $R_2$ , 3 long-stalked, separated from $R_4$ , 5. (Mimállo;
	Cicinnus (= $Per\acute{o}phora$ ); Lacosòma). (See couplet 37). ( $PRO$ –
	TOPSÝCHIDÆ, PEROPHÓRIDÆ, LACOSÓMIDÆ, LA-
	$COSM \acute{A}TID \rlap{E})$
	Fore wings with R <sub>3,4</sub> stalked
68.	
	and R <sub>4,5</sub> which groups are again stalked; M <sub>1</sub> arising from the
	upper angle of the cell; hind wings with Sc close to R <sub>s</sub> but free;
	proboscis present. (Fig. 397). ( <b>Brahmæa</b> , Asia, Afr.).
	BRAHMÆIDÆ
	Fore wings with only three or four branches of the radius devel-
	oped, of which two or three are on a common stalk; hind wings
	with Sc more or less distant from $R_s$ . (Fig. 396)69
69.	Proboscis bearing carinate papillæ; antennæ bipectinate ventrally
	in both sexes. (Fig. 396). (Asthénida; Oxýtenis, neotrop.).
	OXYTÉNIDÆ
	Proboscis thin and reduced
70.	Hind wings with Sc and R connected by a bar. (Fig. 394). (Cer-
	cóphana, Janiòdes, neotrop.) CERCOPHÁNIDÆ
	Hind wings with Sc and R <sub>s</sub> not connected by a bar. (Fig. 395).
	Principally neotropical and ethiopian. (Antheræa; Áttacus
	(A. átlas, Atlas moth); Autómeris; Callosamia (C. pro-
	mèthea, Promethea moth); Hemileùca; Philosàmia (=Sàmia
	auctt.) (P. walkeri (cýnthia), Ailanthus moth); Pseudohàzis;
	Sàmia (= Platysàmia) (S. cecròpia, Cecropia moth); Satúrnia;
	Tèlea (T. polyphèmus, Polyphemus moth); Tropæa (= Ác-
	tias) (T. lùna, Luna moth). (Including ATTÁCIDÆ, AR-
	SENURIDÆ, HEMILEÙCIDÆ)) SATURNÌIDÆ
71.	Fore wings with $M_1$ free, not stalked with $R_5$
	Fore wings with M <sub>1</sub> stalked with R <sub>5</sub> , or arising very close to R <sub>5</sub>
	at the apex of the discal cell (Fig. 399). (Urània, Alcidis,
=0	Nyctálemon, Urapteròides) URANÌIDÆ
<i>7</i> 2.	
	Fore wings with R <sub>2</sub> , 3, 4 stalked. (Fig. 401). ( <b>Epicopèia</b> , indomal.).
79	EPICOPEÌIDÆ
73.	the state of t
	or closed discal cell, rarely $R_3$ , 4 or $R_4$ , 5 short-stalked; maxillary

palpi minute; wings often with unscaled spots. (Fig. 400).

	Mainly tropical. (Thỳris, Striglìna, Adæa, Rhodoneùra, Dysòdia, Risàma, Méskea). (See couplet 124).  THYRÍDIDÆ
74.	Veins variously fusing beyond the discal cell
<b>7</b> 5.	Hind wings with Sc and R <sub>s</sub> separate from near the base, or fused beyond the cell, when joined near the base the fusion not ex-
	tending to the middle of the cell
76.	Shaft of antennæ tapering evenly from base to tip
77.	approaching it, or briefly touching near the apex of the cell, sometimes fused for a greater or less distance beyond the cell. 78 Hind wing with Sc fused with $R_{\rm s}$ for a short distance before the
78.	middle of the cell
	Hind wings with Sc and $R_s$ connected by a bar before the discal cell. (Fig. 402)
79.	Fore wings with $R_1$ about as long as the cell, or longer80 Fore wings with $R_1$ much shorter than the cell. (Fig. 406). (See couplets 19, 58). A very large, widespread group.  PYRALIDIDÆ, part
a.	Fore wing with A1 present; proboscis weak or absent; hind wings with the fringe on Cu weak or absent; larvæ bore in aquatic or marsh plants. (Schænòbius, Acéntropus, Ràmila). (SIGÌNÆ)SCHŒNOBIÌNÆ
b.	Fore wings with A1 absent
1 T	his family is sometimes divided into a series of families, but its numerous well-marked

<sup>&</sup>lt;sup>1</sup> This family is sometimes divided into a series of families, but its numerous well-marked groups are generally regarded as of only subfamily rank.

<b>c.</b>	Proboscis and ocelli absent; male usually with third joint of the palpi vestigial
	Proboscis and ocelli present; labial palpi normale
d.	Front with a conical tuft; general vestiture deep and mixed;
	larvæ living as scavengers in nests of social Hymenoptera or
	in dried foods. (Gallèria (G. melonélla, Bee moth), Paralíspa,
	Achròia). (TINEÌNÆ, Hampson)
	Front and thorax smooth-scaled; larvæ feeding on scale insects.
	(Macrothèca) MACROTHECÌNÆ
e.	Maxillary palpi rather well developed; hind wings with Sc free.
•	(Pýralis (P. farinàlis, Meal moth), Aglóssa, Hercùlia, Cle-
	deòbia, Omphalócera, Hypsopygia (H. costàlis, Clover-
	hav worm)  PVRALIDÎNÆ
	hay worm)
	(Arta, Chrysauge, Clydonópteron, Gephyra, Náchaba).
	(SEMNIÌNÆ) CHRVSAIIGÌNÆ
f	(SEMNIÌNÆ)
	separated from Sc, cell closed by a weak but distinct vein, R
	more or less weakened; labial palpi beak-like, maxillary palpi
	triangular; mainly Old World species. (Ancylolòmia, Prion-
	ápteryx, Eufernáldia) ANCYLOLOMIÌNÆ
	Hind wings with $M_1$ closely approximate to $R$ g
g.	Fore wings with $R_5$ stalked with $R_{3+4}$ ; only one free vein below
۶.	the forked vein which runs to the apex from the radial stemh
	Fore wings with R <sub>5</sub> free; two free veins from the radial stem below
	the forked one; hind wings with Sc and R almost always fus-
	ing m
h.	Fore wings with R <sub>3</sub> and R <sub>4</sub> completely fused; hind wing with
	strong fringe on Cui
	Fore wings with R <sub>3</sub> and R <sub>4</sub> stalked, separating apicallyk
i.	Cell in hind wing closed by a delicate but nearly complete vein;
	frenulum of female consisting of a single spinej
	Cell in hind wing widely open; frenulum of female multiple. (See
	couplets k, m). (Raphiptera) CRAMBINÆ, part
i.	Proboscis strong, separating the palpi toward the base: larvæ
J.	usually leaf-rollers, a few feeding in stored foods. (Phỳcita,
	<b>Ephéstia</b> (Fig. 406) (E. kuchniélla, Mediterranean flour moth),
	Plòdia (P. interpunctélla, Indian-meal moth, world wide pest
	of dried fruits, nuts, corn and other cereals), Acróbasis, Diorýc-
	tria, Nephrópteryx). (ANERASTIÌNÆ, Hampson).
	PHYCITÎNÆ
	Proboscis weak or vestigial, not separating the palpi and concealed
	by them when coiled. (Peòria, Ponjàdia, Anerástia, Hypsó-
	tropa). $(HYPSOTROPINÆ)$ ANERASTIÌNÆ
k.	Hind wings with a heavy fringe on the base of Cu; labial palpi
	parpi

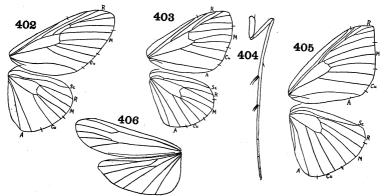
	beak-like, the maxillary palpi triangular. (Argýria, Crámbus,
	Diatræa (corn and sugar cane borers), Thauamtópsis). (See
	couplets i, m) CRAMBINÆ, part
	Fringe on base of Cu very light or wanting; maxillary palpi plu-
1	mose, or small and concealed
1.	Fore wings with raised scale-tufts. (Epipáschia, Tetrálopha, Oneida, Pocócera). (POCOCERINÆ) EPIPASCHIÌNÆ
	Fore wings smooth, without raised scale-tufts. (See couplet e).
	PYRALIDÎNÆ, part
m	Hind wings with fringe on Cu heavy; labial palpi beak-like, the
111.	maxillary palpi triangular. (See couplets i, k).
	CRAMBINÆ, part
	Fringe on Cu light or absent; labial palpi rarely beak-like, the
	maxillary palpi usually moderate or small, not triangularn
n.	TT' 1 ' '1 ' ' A
11.	Some loose hair near the inner margin of the hind wing, part of it
	forming a weak fringe on Cu which runs into a group of spatu-
	late hairs or scales below Cu. (Glaphýria, Dicymolòmia,
	Lipocósma) GIAPHVRIÌNÆ
٥.	Lipocósma)
	phila, Nýmphula (= Hydrocámpa), Eurrhýpara, Muso-
	tima). (HYDROCAMPINÆ)NYMPHULINÆ
	Fore wings with $R_2$ freep
p.	Labial palpi beak-like; maxillary palpi large and triangular;
	fore wing usually slightly rough-scaled, with M <sub>1</sub> well separated
	from $R_5$ at origin, about as far from base of $R_{3+4}$ as from $M_2$ .
	(Scopària, Xeróscopa) SCOPARIÌNÆ
	Labial palpi often upturned; maxillary palpi very rarely large and
	triangular ( $Loxosteg\acute{o}psis$ ) and in that case with $R_{3+4}$ and $M_1$
	closely approximate. (Loxóstege (Web-worms), Désmia,
	Phlyctænia, Agrótera, Sylépta, Margarònia, Héllula,
	Pantógrapha, Diaphània (Melon and Pickle-worms), Py-
	rausta (P. nubilàlis, European corn borer)). (AGROTER-
	$(NE) \dots PYRAUSTÎNE$
80.	Tympanum developed; chætosema reduced or wanting; dip in Sc
	of hind wings located beyond the discal cell and sometimes
	resulting in a fusion with $R_s$
	Tympanum not developed; chætosema present; dip in Sc of hind
	wings opposite the middle of the discal cell, humeral vein
	present; day flying, butterfly-like moths. (See couplet 96).
	CALLIDULIDÆ
Ω1	Ocelli well developed; frenulum of male knobbed. (Thyatira, pa-
01.	læarc.). (See couplet 60)
	Ocelli vestigial; fore wings commonly with recurved tip. (Fig.
	Oceni vesugiai; fore wings commonly with recurved tip. (Fig.

405). (See couplets 65, 95). (**Euchèra, Edápteryx, Drépana, Cilix,** palæarc.; **Falcària,** holarc.). (*DREPAN ÙLIDÆ*).

### DREPÁNIDÆ

82. Proboscis present, palpi recurved above the vertex, with bare third joint; ocelli present, eyes bare; thorax and abdomen clothed with smooth scales; day-flying, often colored. (Fig. 402). (Peridròmia, Callimórpha, palæarc.; Hýpsa (= Asòta), Agánais). (CALLIMÓRPHIDÆ, AGANÀIDÆ, ASÓTIDÆ).

HÝPSIDÆ



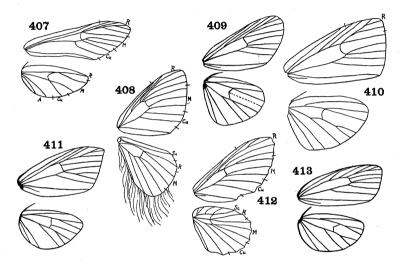
Figs. 402-406. Lepidoptera

- 402. **Hypsa**, wings (Hampson) Hypsidæ.
- 403. Lymantria, wings (Hampson) Lymantriidæ.
- 404. Pterophorus, hind leg of male (Fernald) Pterophoridæ.
- 405. Euchera, wings (Hampson) Drepanidæ.
- 406. Ephestia, wings (Chittenden) Pyralididæ.
- - Fore wings with two radial areoles, the basal one very long, apex of wing acute; female without frenulum. (See couplet 93). Australasian. (Anthèla, Chelépteryx, Munichryia).

# ANTHÉLIDÆ, males

	Tympanal hoods less conspicuous and more lateral; fore wings with radial areole usually present
85.	Ocelli present; hind wings with $M_2$ sometimes weak, or rarely absent, $M_1$ independent or very short-stalked with $R_s$ , basal areole formed by Sc and R very small, less than one-sixth the
	length of the discal cell
	Ocelli absent; hind wings with M <sub>2</sub> as strong as the other veins,
	$M_1$ stalked with $R_s$ , basal areole more than one-sixth the length of the discal cell. (Fig. 403). (See couplet 83). Principally
	indomalayan and ethiopian, but containing the following im-
	portant species of holarctic distribution: ( <b>Lymántria</b> (= <i>Liparis</i> ) ( <i>L. mónacha</i> , Nun moth); <b>Porthètria</b> (= <i>Ocnèria</i> ) ( <i>P.</i>
	dispar, Gipsy moth); <b>Nýgmia</b> (=Eupróctis) (N. phæorrhæa,
	= chrysorrhæa, Browntail moth); Stilpnòtia (S. sálicis, Satin
	moth); Orgyia (= Notólophus); Hemerocámpa (Tussock
	moths). (LIPARIDIDÆ, LIPÁRIDÆ, OCNERIIDÆ).
	T.VMANTRÌIDÆ
86.	Hind wings with vein M <sub>2</sub> imperfect or obsolete. (CARADRÍN-
	<i>IDÆ</i> ) <b>NOCTÙIDÆ</b> a
	Hind wings with vein $M_2$ well developed <b>PLUSÌIDÆ</b> d
a.	Eyes hairy. (Alètia, Círphis (=Leucània), cosmop. (C. unipúncta, Army worm))
	Army worm))
	Army worm)). MELANCHRĪNÆ Eyes bare
b.	Army worm)). MELANCHRĪNÆ  Eyes bare
b.	Army worm)).  Eyes bare
b.	Army worm)).  Eyes bare
b.	Army worm)).  Eyes bare
b.	Army worm)) MELANCHRĪNÆ  Eyes bare b  Eyes not ciliated c  Eyes furnished with a marginal row of long cilia curving over them. (Pòlia, Conístra, Orthòsia, holarc.; Cucúllia, holarc., ethiop.) POLIĪNÆ  The four posterior tibiæ spinose. (Heliothis (H. obsolèta, Corn earworm, Cotton boll-worm), Eùxoa, Agròtis, widespr.; Triphæna, holarc.; Nóctua). (AGROTĪNÆ). NOCTUĪNÆ  Tibiæ not spinose. (Acronýcta. (= Anátela), Luperīna, holarc.;
b.	Army worm)) MELANCHRĪNÆ  Eyes bare b  Eyes not ciliated c  Eyes furnished with a marginal row of long cilia curving over them. (Pòlia, Conístra, Orthòsia, holarc.; Cucúllia, holarc., ethiop.) POLIĪNÆ  The four posterior tibiæ spinose. (Heliothis (H. obsolèta, Corn earworm, Cotton boll-worm), Eùxoa, Agròtis, widespr.; Triphæna, holarc.; Nóctua). (AGROTĪNÆ). NOCTUĪNÆ  Tibiæ not spinose. (Acronýcta. (= Anátela), Luperīna, holarc.;
b. с.	Army worm))  Eyes bare  Eyes not ciliated  Eyes furnished with a marginal row of long cilia curving over them. (Pòlia, Conístra, Orthòsia, holarc.; Cucúllia, holarc., ethiop.)  POLIÌNÆ  The four posterior tibiæ spinose. (Heliothis (H. obsolèta, Corn earworm, Cotton boll-worm), Eùxoa, Agròtis, widespr.; Triphæna, holarc.; Nóctua). (AGROTÌNÆ).  NOCTUÌNÆ  Tibiæ not spinose. (Acronýcta (= Apátela), Luperìna, holarc.; Nonàgria, widespr.; Monòdes, mainly Amer.; Hadèna, cosmop.; Carádrina)  CARADRINÌNÆ  Hind wings with M2 and M3 parallel. (Hermínia, Bomólocha,
b. с.	Army worm)).  Eyes bare
b. с.	Army worm))  Eyes bare  Eyes not ciliated  Eyes furnished with a marginal row of long cilia curving over them. (Pòlia, Conístra, Orthòsia, holarc.; Cucúllia, holarc., ethiop.)  POLIÎNÆ  The four posterior tibiæ spinose. (Heliothis (H. obsolèta, Corn earworm, Cotton boll-worm), Eùxoa, Agròtis, widespr.; Triphæna, holarc.; Nóctua). (AGROTÎNÆ).  NOCTUÎNÆ  Tibiæ not spinose. (Acronýcta (= Apátela), Luperìna, holarc.; Nonàgria, widespr.; Monòdes, mainly Amer.; Hadèna, cosmop.; Carádrina)  CARADRINÎNÆ  Hind wings with M2 and M3 parallel. (Hermínia, Bomólocha, holarc.; Hypèna, cosmop.)  HYPENÎNÆ  Hind wings with M2 and M3 close together at base, diverging
b. c. d.	Army worm))  Eyes bare  Eyes not ciliated  Eyes furnished with a marginal row of long cilia curving over them. (Pòlia, Conístra, Orthòsia, holarc.; Cucúllia, holarc., ethiop.)  POLIÌNÆ  The four posterior tibiæ spinose. (Heliothis (H. obsolèta, Corn earworm, Cotton boll-worm), Eùxoa, Agròtis, widespr.; Triphæna, holarc.; Nóctua). (AGROTÌNÆ).  NOCTUÌNÆ  Tibiæ not spinose. (Acronýcta (= Apátela), Luperìna, holarc.; Nonàgria, widespr.; Monòdes, mainly Amer.; Hadèna, cosmop.; Carádrina)  CARADRINÌNÆ  Hind wings with M2 and M3 parallel. (Hermínia, Bomólocha, holarc.; Hypèna, cosmop.)  HYPENÌNÆ  Hind wings with M2 and M3 close together at base, diverging apically
b. c. d.	Army worm))  Eyes bare  Eyes not ciliated  Eyes furnished with a marginal row of long cilia curving over them. (Pòlia, Conístra, Orthòsia, holarc.; Cucúllia, holarc., ethiop.)  POLIÌNÆ  The four posterior tibiæ spinose. (Heliothis (H. obsolèta, Corn earworm, Cotton boll-worm), Eùxoa, Agròtis, widespr.; Triphæna, holarc.; Nóctua). (AGROTÌNÆ).  NOCTUÌNÆ  Tibiæ not spinose. (Acronýcta (= Apátela), Luperìna, holarc.; Nonàgria, widespr.; Monòdes, mainly Amer.; Hadèna, cosmop.; Carádrina)  CARADRINÌNÆ  Hind wings with M2 and M3 parallel. (Hermínia, Bomólocha, holarc.; Hypèna, cosmop.)  HYPENÌNÆ  Hind wings with M2 and M3 close together at base, diverging apically  e Eyes glabrous  f
b. с. d.	Army worm)).  Eyes bare
b. с. d.	Army worm)).  Eyes bare

87.	Hind wings small, with Sc apparently wanting; tympanal hoods
	very large, the abdomen sometimes constricted behind them;
	day-flying, usually brightly colored moths. (Fig. 407). Mainly
	indoaustr. (Euchròmia, Ctenùcha, Amàta (=Sýntomis),
	Scépsis, Cosmosòma). (SYNTÓMIDÆ, AMÁTIDÆ, SYN-
	TOMÍDIDÆ) EUCHROMÌIDÆ
	Hind wings with Sc well developed; tympanal hoods not excep-
	tionally large. Widespread
88.	Hind wing with Sc and R fused from near the base to the middle
	or near the middle of the cell, leaving a small basal areola.
	(Fig. 410). Mainly tropical. (Hylóphila, palæarc.; Eàrias,
	Old World; Sarrothripus, palæarc., indoaustr.).
	HYLOPHÍLIDÆ
	Hind wings with Sc and R fused from extreme base to near or
	beyond the middle of the cell
89.	Ocelli present. Large, widespread group. (Phægóptera (=
	Ophàrus), Parasèmia (= $Hyphoràia$ ), Nemeóphila (= $Para$ -
	sèn ia, pt.), Árctia, Háploa, Utetheisa, Apántesis, Hyphán-
	tria (Webworms), Euprèpia, Autómolis, Halisidòta, Estíg-
	mene, Diacrísia) ARCTÌIDÆ
	Ocelli absent
90.	Fore wings smoothly scaled, rarely M <sub>2</sub> or M <sub>3</sub> wanting. (See
	couplet 48). (Eudésmia (=Cisthene, auct.), Cisthene (=
	Íllice), Lithòsia, Hypoprèpia, Chionèma, widespr.).
	LITHOSÌIDÆ
	Fore wings with tufts of raised scales. (Fig. 411). (Nola, Ræsèlia,
	Ùraba, Célama) NÓLIDÆ
91.	Hind wings with Sc entirely free from R. Indomalayan. (Cocýtia).
	(EUCOCYTÌIDÆ) COCYTÌIDÆ
	Hind wings with Sc and R connected for a short distance at base,
	sometimes forming a very small basal areolet; rather small
	or medium-sized day-flying moths of brilliant colors, often
	dark, ornamented with large pale spots. (Fig. 409). Forester
	moths. Principally ethiopian and indoaustralian. (Agarista,
	Alýpia, Androlòma, Eusèmia, Phalænòides). (PHALÆ-
	$N\grave{O}IDID\pounds)$ AGARÍSTIDÆ
92.	Hind wings with the straight Sc connected with the cell by a bar
	near or before the middle of the discal cell93
	Hind wings with Sc free from R or fusing for a greater or less
	distance, but not joined to the cell by a bar near the base of the
	cell



Figs. 407-413. Lepidoptera

- 407. Euchromia, wings (Hampson) Euchromiidæ.
- 408. Pterothysanus, wings (Hampson) Pterothysanidæ.
- 409. Alypiodes, wings (Hampson) Agaristidæ.
- 410. Earias, wings (Hampson) Hylophilidæ.
- 411. Nigetia, wings (Holland) Nolidæ.
- 412. Epicnaptera, wings (Hampson) Lasiocampidæ.
- 413. Hypoprepia, wings (Hampson) Lithosiidæ.
- 94. Fore wings with R<sub>4</sub> long and free, or arising from base of the stalked R<sub>2</sub>, 3; R<sub>5</sub> and M<sub>1</sub> short-stalked; hind wings with Sc and R<sub>s</sub> fused for a greater or less extent near the middle of the discal cell, humeral angle wide and strengthened by one or more humeral veins. (Fig. 412). Widespread, mainly tropical.

	(Tólype, Epicnáptera (= $Gastrópacha$ ), Lasiocámpa, Eriogáster, Malacosòma (= $Clisiocámpa$ ). Tent caterpillars). ( $LACHNÈID\pounds$ ) LASIOCÁMPIDÆ
	Fore wings with R <sub>4</sub> short and arising apically from R <sub>3</sub> or from
	$ m R_{2+3}$
95.	Fore wings with R <sub>5</sub> free, R <sub>4</sub> remote from R <sub>3</sub> and ending beyond
	tip of the wing, outer margin not sinuate; hind wings with Sc
	dipping toward R <sub>s</sub> in front of cell96
	Fore wings with R <sub>2-5</sub> stalked, R <sub>4</sub> very close to R <sub>3</sub> and entering the
	costal margin; hind wings with Sc dipping toward or fusing with
	R <sub>s</sub> beyond the cell. (See couplets 65, 81). (Orèta). <b>DREPÁNIDÆ</b> , part
96.	Fore wings with $R_2$ free, only $R_{3+4}$ stalked; hind wings with lower
٠.	margin bearing a double fringe of long woolly hairs. (Fig. 408).
	Indian and African. (Pterothýsanus, Hibríldes).
	PTEROTH YSÁNIDÆ
	Fore wings with R <sub>2-4</sub> stalked; hind wings without long fringe. Indo-
O=	malayan. (See couplet 80). (Callidula, Clèis). CALLIDÜLIDÆ
97.	Maxillary palpi conspicuous, folded in resting position98
	Maxillary palpi straight and porrect, or vestigial (if of folded type,
98.	inconspicuous or invisible)
00.	an eye-cap, larger than the eye; wing-membrane aculeate99
	Eye-cap small or undeveloped; fore wings usually with large cell
	and branched veins100
99.	Fore wings with branched veins, normally with short trapezoidal
	aculeæ distributed over entire membrane; cell sometimes absent;
	media usually dipping deeply into wing; extremely small moths,
	the wing expanse as little as 3 mm. (Fig. 414). Leaf miners.
	(Neptícula, cosmop.; Scoliaula, Eur.; Glaucólepis; Ectœdèmia; Obrússa; Trifúrcula). (STIGMÉLLIDÆ).
	NEPTICÙLIDÆ
	Fore wings with three or four simple veins only, aculeæ pointing
	forward, in rows and confined to a small area at the base of
	the wing; hind wings linear; mostly oriental, three North
	American species. Larvæ legless miners in bark or rind. (Opó-
100	stega) OPOSTÉGIDÆ
100	The state of the s
	not aculeate; head with a few erect hairs behind. (Acrolèpia, cosmop.)
	Fore wings with $R_5$ extending to the costa, sometimes absent .101

101. Head usually entirely smooth; strongly flattened species, coxæ

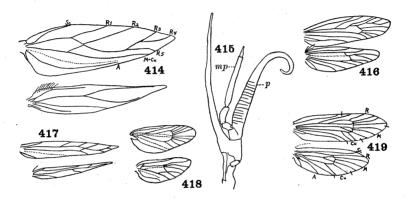
	flat and appressed, fore wings curved down at apex, hind wings
	narrow-lanceolate; venation sometimes reduced; mostly orien-
	tal. (Opógona, ethiop.; Phèoses, nearc.; Œnóphila, ethiop.,
	indomal.). (OINOPHÍLIDÆ)ŒNOPHÍLIDÆ
	Head tufted, at least on the vertex, sometimes with a naked area
	above the eyes behind the antennæ, in which case the hind wings
	are ample; venation complete, accessory cell commonly present,
	four or five branches of R extending to the costal margin102
109	Wing-membrane aculeate (Fig. 369); hind wings with Sc having
102.	a strong basal fork (the lower branch being R <sub>1</sub> ) or considerably
	swollen at the base, R and Sc usually sharply divergent from
	the base; vertex roughly hairy
	strong as the other veins and when distinct placed well out from
	the base of the wing, connecting Sc and R which are closely
	parallel toward the base. Widespread. (See couplet 112).
100	(Isocórypha, Diachorísia, Leucómele, Enoe). TINÈIDÆ
103.	Folded part of maxillary palpi half as long as the width of the
	head. (Eudárcia, Incurvària, holarc.; Phyllopòria, palæarc.;
	Lamprònia, holarc., ethiop.; Nemóphora, Paracleménsia).
	(LAMPRONIIDÆ) INCURVARÌIDÆ
	Folded part of maxillary palpi two-thirds as long as the width of
	the head. (Fig. 415). Yucca moths; larvæ boring in seed pods
	of Yucca. (Prodóxus, Tegetícula (= Pronùba), Amer.).
	PRODÓXIDÆ
104.	Basal joint of the antennæ enlarged and concave beneath, form-
	ing an eye-cap
	Basal joint of the antennæ not forming an eye-cap, sometimes
	provided with tufts of scales or a pecten of bristles
105.	Venation complete and nearly parallel, hind wings with Sc end-
	ing at apical fourth and discal cell reaching the middle of the
	wing. (Calósima, holarc.). (see couplet 146).
	BLASTOBÁSIDÆ
	Venation reduced, wings linear-lanceolate with sharply pointed
	apex, fore wings with the veins at the apex of the discal cell
	radially diverging; hind wings without discal cell, the principal
	vein axial. (Fig. 417)
106.	Labial palpi minute and drooping, or absent; fore wings typically
	with a closed anal cell, or the vertex rough. (Fig. 417). (Buc-

culàtrix (Ribbed cocoon makers), Lyonètia, cosmop.; Be-

déllia, Leucóptera (= Cemióstoma), widespr.; Phyllobróstis). (BEDELLÌIDÆ, CEMIOSTOMÁTIDÆ, ERECHTHÌIDÆ, HIEROXÉSTIDÆ). (Including BUCCULATRÍGIDÆ).

#### LYONETÌIDÆ

Labial palpi moderate, upcurved; no anal cell; vertex smooth; eye-cap small, no pecten. (**Phyllocnístis**, part). (See couplet 139)



Figs. 414-419. Lepidoptera

- 414. Ectœdemia, wings (Braun) Nepticulidæ.
- 415. **Tegeticula**, mouthparts of female (Packard): m p, maxillary palpus; p, palpifer.
- 416. Setomorpha, wings (Busck) Setomorphidæ.
- 417. Bedellia, wings (Clemens) Lyonetiidæ.
- 418. Cecidoses, wings (Brethes) Cecidosidæ.
- 419. Tinea, wings (Spuler) Tineidæ.
- 108. Labial palpi with the first joint short, and the second joint bristly, on the outer side and equal to the fusiform third joint.....110 Mouthparts wholly vestigial. South American gall-making species, with very indistinct venation. (Ridiáschina).

RIDIASCHÍNIDÆ

109.	Labial palpi scaled or short-hairy, the third joint usually long and pointed, or very short in the species with roughest vestiture
	Mouthparts wholly undeveloped; front and hind wings each with a single anal vein, fringe short; larvæ gall-makers. (Fig. 418). South American. (Cecidòses, Olièra)
110.	Wing-membrane aculeate over all; female with piercing ovi-
110.	positor; antennæ often longer than the wings. (Adèla, palæ-
	arc., neotrop.; Nemotòis, Old World; Chalcèopla, nearc.).
	ADÉLIDÆ
	Wing-membrane not generally aculeate; ovipositor membranous,
	retractile
111.	Fore wings with R <sub>3</sub> free, arising from the accessory cell. (Fig.
111.	419)
	Fore wings with a hyaline aculeate fovea in the accessory cell
	distorting the venation, R <sub>3</sub> , 4, 5 seriately stalked. (Fig. 416).
	(Setomórpha, widespr.) SETOMÓRPHIDÆ
112.	Vertex with high, rough, bristling vestiture. (Fig. 419). ( <b>Tinea</b>
	(Clothes moths), Monòpis, Scárdia, cosmop.; Tinèola
	(T. bisseliélla, Clothes moth), Trichophaga (T. tapetzélla, Case-
	bearing clothes moth), Ténaga, widespr.; Elatôbia, holarc.).
	(See couplet 102). (Including MONÓPIDÆ) TINÈIDÆ
	Vertex with short, flattened hair. (Amýdria) AMYDRÌIDÆ
113.	Labial palpi elongate, deeply scaled, but not bristly, with the
	first joint at least subequal to the second joint, or much longer,
	of female typically porrect, of male upturned; eyes sometimes
	hairy or scaly114
	Labial palpi with the first joint small; eyes bare
114.	
	sized species; no ocelli
	Proboscis well developed
115.	, 8 66
	or tufted; antennæ exteriorly ciliated on apical half; third
	joint of labial palpi minute and bare. (Ashinàga, Formosa).  ASHINÁGIDÆ
	Vestiture deep and spatulate on the thorax, with well marked an-
	terior and posterior tufts; eyes more or less distinctly hairy; stout, Noctuid-like moths; American, mostly tropical. (Acró-
	lophus, Burrowing webworms) ACROLÓPHIDÆ
116.	
110.	sinuate, veins $R_s$ , $M_1$ and $M_2$ nearly parallel; fore wings with

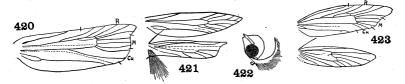
	$R_4$ and $R_5$ stalked or coincident; find tible thinly clothed with
	long hairs above, densely rough-haired between spurs beneath.
	Indomalayan. (Amphithèra, Agriothèra, Telethèra).
	AMPHITHÉRIDÆ
	Hind wings with greatly reduced venation, frenulum of female
	simple; ocelli posterior; larvæ on cocoanut. Austromal. (Aga-
	nóxena, Hæmolytis). (See couplet 153). AGONOXÉNIDÆ
117.	Hind wings relatively ample, with well developed anal region,
	rounded at apex or trapezoidal, or more or less deeply notched
	below the apex, often wider than their fringe; venation more
	or less complete
	Hind wings narrow-lanceolate and pointed, or linear, never much
	wider than their fringe; sometimes without closed cell, venation
	often much reduced
118.	Hind wings with $M_1$ and sometimes also $M_2$ absent
110.	Hind wings with $M_1$ present, associated with $R_s$ (if $M_1$ of the hind
	wings and A1 of the fore wings are rarely absent (Gelechiidæ)
	the outer margin of the hind wings is more or less emargin-
110	ate)
119.	Hind wings with the cell closed, M <sub>1</sub> and M <sub>2</sub> both wanting, R <sub>8</sub>
	extending alone, the outer margin not concave; fore wings
	heavily tufted, all the branches of R free, R <sub>5</sub> ending beyond the
	wing tip, Cu <sub>2</sub> from near end of cell, A1 absent; mostly Australian
	and Hawaiian. (Carposina (C. fernaldàna, American currant
	fruit worm), Bóndia)
	Hind wings trapezoidal-ovate, with cell open and M <sub>1</sub> and some-
	times also M <sub>2</sub> absent; fore wings with R <sub>5</sub> absent. (Metachánda,
	Ancylomètis, Chanýstis, ethiop., ind.) . METACHÁNDIDÆ
120.	Hind wings with 3A distally forked; venation complete, acces-
	sory cell present, all veins separate; front wings with two medial
	stems in cell. (Larvæ at first parasitic in Homoptera, later
	myrmecophilous). (Cyclotórna, austr.). CYCLOTÓRNIDÆ
	Anal vein not distally forked
121.	Hind wings with radial vein shortly furcate at apex; labial palpi
	with second joint thickened and with rather appressed scales,
*	last joint very short, filiform, obtuse; front tarsi much longer
	than their tibiæ. (Anomóloga, ethiop.) ANOMOLÓGIDÆ
	Hind wings with radial vein not furcate at apex
122.	Fore wings with Cu2 arising before the last fourth of the cell;
	labial palpi beak-like, with the third joint short and blunt and
	the second joint closely or roughly scaled and usually porrect

- 123. Upper side of the hind wings with a fringe of long hairs on the basal part of Cu (if rarely absent in somes pecies of Laspeyresia the fore wings have M<sub>1</sub> and M<sub>2</sub> rather close together at the tip); fore wings with R<sub>4</sub> and R<sub>5</sub> separate, or with veins M<sub>2</sub>, M<sub>3</sub> and Cu<sub>1</sub> converging strongly toward the margin. (Fig. 420). A cosmopolitan group. (Áncylis (A. comptàna, Strawberry leafroller), Evêtria (Pine twig moths), Carpocápsa (=Cýdia) C. pomonélla, Codling moth; C. sáltitans, Mexican jumping bean moth); Rhyaciònia (=Evètria, = Retínia), Anarmònia (=Epinòtia), Laspeyrèsia (L. prunívora, Lesser apple worm; L. interstinctàna, Clover-seed worm), Spilonòta (=Tmetócera) (S. ocellàna, Budmoth), Olethreùtes, Melíssopus, Polychròsis (P. viteàna Grape-berry moth), Gypsónoma, Eucósma, Argyróploce, Hemímene, Pámmene). (EUCÓS-MIDÆ, EPIBLÉMIDÆ, GRAPTOLÍTHIDÆ).

## OLETHREÙTIDÆ

ANÓTHIDÆ) ..... TORTRÍCIDÆ

125. Labial palpi beak-like, with the second joint rough-scaled on the upper side and smooth within, and the third joint porrect and inconspicuous; fore wings with A1 lost, all veins independently arising from the discal cell, rarely R<sub>4</sub> and R<sub>5</sub> stalked; hind wings with R and M₁ approximate or stalked and widely distant from M₂. (If the second joint of the palpus is bristly on the outer side and third joint is well set off refer to couplet 101; if R<sub>4+5</sub> are stalked and both terminate in the costa and M₂ of the hind wings is not widely separate from M₁ see Anarsia ♂ of the Gelechiidæ, couplet 126). Mainly holarctic. (Commóphila, Hysteròsia, Phalònia (=Cónchylis), Phármacis, Chlidònia, Phtheóchroa, Euxánthis). (COMMOPHÍLIDÆ, CONCHÝLIDÆ).



Figs. 420-423. Lepidoptera

- 420. **Hystricophora**, fore wing (Heinrich) Olethreutidæ.
- 421. **Agnippe,** wings (Busck) Gelechiidæ.
- 422. Gnormoschema, head from side (Busck) Gelechiidæ.
- 423. Glyphipteryx, wings (Spuler) Glyphipterygidæ.

126. Both wings with A1 lost (extreme tip present in Symmoca, spp.); outer margin of hind wing usually concave, sometimes quite emarginate and the apex produced; fore wings with R₅ running into the costa, stalked with R₄. (Figs. 421, 422). The largest family of Microfrenatæ, with about 400 genera and 3,700 species including many destructive species. (Anacámpsis (Leafrollers), Anársia (A. lineatélla, Peach twig-borer), Aristotélia (A. fragàriæ, Strawberry crown-miner), Dûvita, Dichómeris, Glyphidócera, Recurvària, Sitotròga (S. cerealélla, Angoumois grain-moth), Pectinóphora (P. gossypiélla, Pink boll-

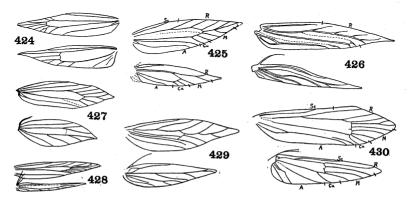
	worm), Phthorimæa (P. operculélla, Potato tuber moth), Telphùsa, Thiótricha, Gnorimoschèma (Goldenrod gall
	moth), Geléchia, Sophrònia, Sýmmoca, Trichótaphe). (See couplet 149). (DICHOMÉRIDÆ). Most GELECHÌIDÆ
	A1 preserved, at least at margin
127.	Hind wings with R and $M_1$ close together, connate, or stalked 128
	Hind wings with R and M <sub>1</sub> well separated at origin, at least half
	as far apart as at the margin
128.	
	where R <sub>5</sub> terminates, Sc, R <sub>1</sub> and R <sub>2</sub> short, ending before middle
	of wing, $M_1$ absent, $M_2$ and $M_3$ both ending in costal margin;
	maxillary palpi vestigial. (Strepsímana, India). STREPSIMÁNIDÆ
	Fore wings with anterior veins longer, M <sub>3</sub> at least ending beyond
	wing tip
1289	Wings relatively ample, fore wings blunt; maxillary palpi of
120a	the folded type
	Wings rather narrow, fore wings sometimes falcate; maxillary
	palpi porrect. (Ceróstoma, holarc., neotrop.). (See couplet
	133) PLUTÉLLIDÆ, part
129.	Fore wings with R <sub>5</sub> long-stalked and extending to the outer
	margin, Cu <sub>1</sub> and Cu <sub>2</sub> widely separated; mainly Old World.
	(Ptochorýctis, Cryptóphasa, Xyloríctes). (CRYPTOPHÁS-
	IDÆ, UZÜCHIDÆ) XYLORÍCTIDÆ
	Fore wings with R <sub>5</sub> usually free and usually extending to the
	costa, Cu <sub>1</sub> and Cu <sub>2</sub> usually connate or stalked; mostly New
	World species. (Stenòma, Menésta, Setióstoma). ( $STENO-M\acute{A}TID\pounds$ ) STENÓMIDÆ
130.	
190.	Ocelli usually very large and conspicuous; fore wings with R <sub>5</sub>
	ending beyond the wing-tip (Fig. 423). Largely oriental.
	(CHOREÙTIDÆ, HEMEROPHÍLIDÆ, SIMAÉTHIDÆ).
	GLYPHIPTERÝGIDÆ
	Two well-marked subfamilies may be distinguished:
a.	Wings relatively narrow, the apex of the fore wings more or less
	extended as a lobe-like prolongation. (Glyphipteryx, cosmop.,
	mainly austr.)
	widespr., Simaèthis (=Allononyma), cosmop., mainly tropi-
	cal) CHOREUTINÆ

131.	Fore wings with $R_4$ and $R_5$ stalked
	Fore wings with R <sub>4</sub> and R <sub>5</sub> separate, R <sub>5</sub> ending beyond the wing-
132.	tip
104.	cosmop.) ETHMIDÆ
	Hind wings with $M_2$ arising nearer $M_3$ than to $M_1$ ; palpi long,
	reaching or surpassing the vertex. ( <b>Ecóphora</b> , palæarc.;
	Depressària, widespr.; Agonópteryx, Dasýcera, Pleùrota).
	(See couplet 149). (DEPRESSARIIDÆ).
	Most <b>ŒCOPHÓRIDÆ</b>
133.	Hind wings with M <sub>1</sub> and M <sub>2</sub> stalked; antennæ extending forward
	in repose. (Plutélla (P. maculipénnis (=cruciferàrum), Dia-
	mond-back cabbage moth), cosmop.). (See couplet 128).
	PLUTÉLLIDÆ
	Hind wings with M <sub>1</sub> and M <sub>2</sub> separate. (Yponomeùta, widespr.;
	Atteva, tropicopol.; Orthotælia, palæarc.; Uròdus). (AT-
	TÉVIDÆ, based on pupal characters only, HYPONÓTIDÆ,
	misprint, HYPSELÓPHIDÆ, HYPONOMEÙTIDÆ, OR-
104	THOTALIIDA: YPONOMEŪTIDA
134.	Fore wings without closed cell
135.	Fore wings with discal cell formed
199.	and hind wings linear, with three or four unbranched veins only;
	middle or hind legs commonly displayed when at rest. (See
	couplet 141)
	Hind tibiæ hairy; fore wings lanceolate, with seven veins reach-
	ing the margin. (Coptodisca, "Shield-bearers"). (See coup-
	let 139)
136.	Hind wings lanceolate, sometimes very small, at least one-sixth
	as wide as long, with the R-stem axial, widely separated from
	Se137
	Hind wings with the R-stem closely associated with Sc at the
	base, or lost; or wing linear and the veins crowded or much re-
	duced, the R-stem not prominently axial
137.	Hind wings without discal cell, Cu-stem often simple; palpi
	drooping
	Hind wings usually lanceolate, with a discal cell, the Cu-stem at
٠.	least two-branched; fore wings with R <sub>1</sub> arising before the middle
	of the cell; palpi usually upturned beyond the middle of the
	front, often strongly divergent. (Fig. 425). (Elachísta, cosmop.; Cycnòdia, Aphelosètia, Chrysopelèia, nearc.; Per-

ittia, palæarc.). (APHELOSETÌIDÆ, CYCNODÌIDÆ, CHRYSOPELEÌIDÆ) ..... ELACHÍSTIDÆ

138. Hind wings with an oblique branch from the R-stem to the costa near the middle of the wing, and sometimes another nearer the tip. (**Tinágma**, nearc.; **Douglasia**, palæarc.).

**DOUGLASIIDÆ** 



Figs. 424-430. Lepidoptera

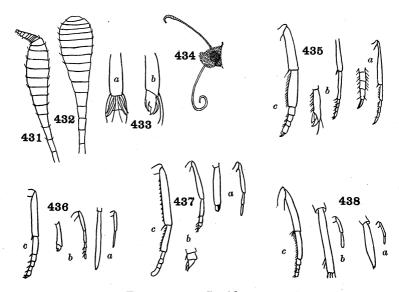
- 424. Scythris, wings (Walsingham) Scythrididæ.
- 425. Elachista, wings (Spuler) Elachistidæ.
- 426. Coleophora, wings (Forbes) Coleophoridæ.
- 427. Coptotriche, wings (Walsingham) Tischeriidæ.
- 428. Mompha, wings (Busck) Cosmopterygidæ.
- 429. Antispila, wings (Spuler) Heliozelidæ.
- 430. Holcocera, wings (Forbes) Blastobasidæ.
- 140. Accessory cell unusually large, extending halfway to the base of the wing and hind wing without closed cell and with reduced

venation; hind tibiæ very hairy; antennæ long, those of the male

	heavily ciliate; vertex often with a large loose semi-erectile tuft. (Fig. 427). ( <b>Tischèria</b> , widespr. ( <i>T. malifoliélla</i> , Apple
	leaf-miner)
	Accessory cell smaller, or more often absent (if long, hind wing
	with closed cell and complete venation); otherwise of different conformation
1/1	Hind tarsi with more or less distinct groups of bristles near the
141.	
	ends of the several joints, the hind tibiæ smooth-scaled or with
	stiff bristles; middle or hind legs displayed when in resting
	position, either raised or held out sideways. (Heliodines,
	holarc., austr.; Schreckensteinia, holarc.; Pancàlia, Au-
	gásma, palæarc.; Strathmópoda, indoaustr., ethiop., palæarc.;
	Idioglóssa, Eucleménsia). $(TIN \cancel{E}GER \dot{I}ID \cancel{E})$ .
	Most <b>HELIODÍNIDÆ</b>
	Hind tarsi without evident groups of bristles
142.	Fore wings with four veins or less, either free or stalked, extend-
	ing from the cell to the costa, and five or six from the cell to
	the inner margin, the last branch of R ending beyond the tip of
	the wing; hind wings with R and M usually widely separated
	at the margin
	Fore wings with five veins extending from the cell to the costa,
	or with only three or four to the inner margin, the last branch of
	R ending before the tip of the wing (except Epimarptidæ)145
143.	Fore wings with $R_1$ arising beyond the middle of the cell, about
110.	as long as $R_0$ (Fig. 424) (Scythris (= $Ruthis$ ) cosmon:
	as long as $R_2$ . (Fig. 424). (Scỳthris (=Butàlis), cosmop.; <b>Paralogístis</b> , ethiop.). (SCÝTHRIDÆ, BUTÁLIDÆ).
	SCYTHRÍDIDÆ
	Fore wings with $R_1$ arising before the middle of the cell, longer
	than $R_2$
144.	Hind tibiæ stiffly bristled, normally in tufts at the spurs; hind
144.	wings with $M_1$ and $M_2$ separate. ( <b>Epermènia</b> (=Chauliòdus),
	widespr.; Acanthèdra, nearc.; Catapléctica, palæarc.).
	EPERMENÎIDÆ
	Hind tibiæ with long loose hair; palpi small and drooping; hind
	wings with M <sub>1</sub> and M <sub>2</sub> united or long-stalked. (Argyrésthia
	(A. conjugélla, Apple fruit-miner), widespr.; Zellèria, cosmop.;
	Hofmánnia) ARGYRESTHÌIDÆ
145.	Fore wings with the discal cell set obliquely, the end distinctly
	closer to the hind margin than to the costa, Cu <sub>2</sub> very short and
	usually extending directly back to the margin146

	Fore wings with the discalcell axial and central, Cu <sub>2</sub> normally longer
1.10	and continuing parallel with the medial veins, rarely obsolete, 148
146.	Fore wings with blunt discal cell, R <sub>1</sub> arising from the middle of
	the wing, veins R <sub>2</sub> to Cu arising from the end of the discal cell,
	with a long stigmal thickening between the costa and R <sub>1</sub> ; hind
	wings with Sc and R normally fused for a short distance near
	the base; antennæ with a heavy pecten on the basal joint.
	(Fig. 430). Mainly tropical species. (Blastóbasis, widespr.;
	Auximóbasis, Am.; Dryopèria, Holcócera, Pigrítia, Valen-
	tínia). (See couplet 105)
	Fore wings without stigma, R <sub>2</sub> arising distinctly before the end
	of the discal cell; hind wings with R not fused with Sc, some-
	times vestigial. (Fig. 426)
147.	Front tibiæ slender, with the epiphysis small and apical or want-
111.	ing, the hind tibiæ with the upper spurs above the middle;
	antennæ porrect in repose. (Fig. 426). (Coleóphora (= $Hap$ -
	loptilia) cosmop. (C. fletcherélla, Cigar case-bearer; C. malivor-
	élla, Pistol case-bearer); Goniodòma, Metriòtes, palæarc.).
	(EUPÍSTIDÆ, HAPLOPTILIIDÆ) COLEOPHÓRIDÆ
	Front tibiæ stouter, with the epiphysis at the middle; antennæ
	turned back in repose. (Batráchedra, Blastodácna, Pyro-
	dérces). (See couplet 153) COSMOPTER ÝGIDÆ, part
148.	Labial palpi with the third joint normally blunt, fusiform and
	more or less angulate with the second, the joints not curving;
	maxillary palpi porrect but not folded across the proboscis, or
	reduced, sometimes absent; A2 not forked at the base. (Acro-
	cércops, Gracilària (G. syringélla, Lilac leaf-miner); Litho-
	collètis (= $Phyllonor\acute{y}cter$ ) (Leaf blotch-miners); <b>Mármara</b> ( $M$ .
	pomonélla, Apple-skin miner); Paréctopa; Parórnix, Órnix).
	(EUCÉSTIDÆ, LITHOCOLLÉTIDÆ, PHYLLORYCTÉR-
	IDÆ)GRACILARÌIDÆ
	Labial palpi with the third joint long, tapering, pointed, the
	second joint curving up; maxillary palpi small, but of folded
	type, curving over the base of the proboscis; A2 commonly
	formed at the base
149.	Both wings with A1 preserved at the margin and A2 forked at the
110.	base; basal joint of the antennæ with a strong pecten of bristles.
	(Borkhausènia, Éndrosis). (See couplet 132).
	(Borkhausema, Endrosis). (See couplet 152).  • CCOPHÓRIDÆ, part
	Hind wings and usually also the fore wings lacking A1; antennæ
	often without a pecten

150,	Fore wings with no veins emerging from the end of the discal cell
	between the continuations of the R and Cu + M stems. (Hèlice,
	The isoa). (See couplet 126) GELECHIDE, part The oblique end of the discal cell of the fore wings emitting
	several veins between the R and Cu stems
151.	Labial palpi with appressed scales, the second joint with a pro-
	jecting pencil of scales above; antennæ with the basal joint
	elongate, a notch near the base of the stalk covered by an
	oblique tooth beneath it; hind tibiæ rough-haired above; fore
	wings with R <sub>5</sub> ending beyond the wing-tip. (Epimárptis,
	India) EPIMÁRPTIDÆ
	Labial palpi without dorsal tuft; base of antennæ not toothed;
	$R_5$ ending before wing-tip
152.	Hind wings elongate-ovate or ovate-lanceolate with the costal
	margin simple, regularly arched. Hawaiian. (Hyposmócoma, Diplósara, Aphthonètus). (DIPLOSÁRIDÆ).
	HYPOSMOCÓMIDÆ
	Hind wings lanceolate or linear, at most with a slightly devel-
	oped costal lobe toward the base, accentuated by a tuft of stiff
	scales beyond which the costa is straight or slightly concave,
	the apex always acute
153.	Hind wings with reduced neuration, R, M and Cu apparently
	simple veins, no crossveins, subcosta very short, frenulum
	simple; labial palpi strongly flattened and rather rough-scaled
	on anterior edge throughout. Austromalayan. (Agonóxena,
	Hæmólytis). (See couplet 116)AGONOXÉNIDÆ
	Hind wings, except when linear, with several branches of Cu
	and M and the crossvein preserved, frenulum usually multiple; palpi with the second joint smooth or tufted below.
	(Fig. 428). A large cosmopolitan group. (Cosmópteryx, Chry-
	soclista, Homalèdra, Synallágma, Wálshia, Stagmató-
	phora, Lavérna, Lophóptilus, Mómpha, Perimède, Psacá-
	phora, Limnècia (L. fragmitélla, Cat-tail moth)). (Compare
	couplet 147). (LAVÈRNIDÆ). Most COSMOPTERÝGIDÆ
154.	
	forked, and arising from the discal cell; eyes strongly lashed
	in front; antennæ separated at the base by a distance greater
	than half the width of the eye, usually hooked at the tip; stout
	bodied butterflies of rapid, erratic flight; hind tibiæ usually
	with a middle spur. Skippers. (HESPERIOIDEA)155
	Fore wings with some of the branches of R stalked or absent;



Figs. 431-438. Lepidoptera

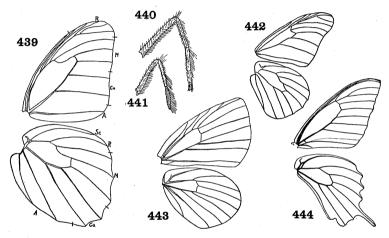
- 431. Antenna of skipper, apical portion (Duncan) Hesperiidæ.
- 432. Antenna of butterfly, apical portion (Duncan).
- 433. **Epargyreus,** last joint of tarsus of male (Scudder): a, dorsal view; b, lateral view. Hesperiidæ.
- 434. Epargyreus, side view of head (Scudder) Hesperiidæ.
- 435. **Lycæna**, details of legs (Scudder): a, front leg of male with tarsal joints at left more enlarged; b, front leg of male with last tarsal joint at left more enlarged; c, middle leg of male. Lycænidæ.
- 436. **Calephelis**, details of legs (Scudder): a, tibia and tarsus of front legs of male, with tarsus at left more enlarged; b, tibia and tarsus of front leg of female, with last joint at left more enlarged; c, tibia and tarsus of middle leg of male.
- 437. **Euphydryas**, details of legs (Scudder): a, tibia and tarsus of front leg of male, with last joints of tarsus on left more enlarged; b, tibia and tarsus of front leg of female with last joints of tarsus below more enlarged; c, tibia and tarsus of middle leg of male. Nymphalidæ.
- 438. **Megisto**, details of legs (Scudder): a, tibia and tarsus of front leg of male, with tarsus at left more enlarged; b, tibia and tarsus of front leg of female, with tarsus at left more enlarged; c, tibia and tarsus of middle leg of male. Satyridæ.

eves rarely lashed; antennæ closer together, the antennal club never pointed and recurved at the tip (Fig. 432); hind tibiæ never with a middle spur. Butterflies. (PAPILIONOI-155. Hind wings with the discal cell divided by the forked base of the media; M<sub>2</sub> fully developed; male with frenulum. Australian. (Euschèmon). (See couplet 15). Females of EUSCHEMÓNIDÆ Hind wings with the base of the media wanting and M2 more or 156. Head much narrower than the metathorax; antennal club large, not drawn out at the tip nor recurved; wing expanse 40 mm. or more; fore wings with  $M_2$  nearer to  $M_3$  than to  $M_1$ ; palpi porrect. Giant skippers. (Megathymus. N. Am.). MEGATHÝMIDÆ Head large; antennal club usually drawn out at the tip and with a distinct recurved apical crook (rarely the crook is not formed in some species whose small size distinguishes them from the a. Antennal club large, and cylindrical or cylindrical-conical, usually reflexed before the swollen part. Mostly South and Central (Pyrrhopýge, Mysôria, Mýscelus, Jemàdia, Apýrrothrix). (THAMYRÍDIDÆ)....PYRRHOPYGÌNÆ Antennal club not wholly reflexed.....b b. Antennal club bent near the middle, ending in a long hooked or bent point, often as long as the swollen part; palpi stout, the second joint raised against the face, the third joint long, slender, bare and porrect; male without costal fold; discal cell of fore wing two-thirds as long as the wing-length, M<sub>2</sub> ending somewhat closer to  $M_1$  than to  $M_3$ . (Ismene, indomal.; Hasòra, indoaustr.; Rhopalocámpa, ethiop.) ..... ISMENÎNÆ Antennal club without a long hooked terminal portion, and fore wings with discal cell less than two-thirds the wing-length. If the antennal club has a long hooked terminal part and the discal cell is more than two-thirds the wing-length, the palpi c. Fore wings with discal cell at least two-thirds as long as the wing, when shorter  $M_2$  arises closer to  $M_1$  than to  $M_3$ . (Figs. 433, 434). (Achlyòdes, neotrop.; Telégonus, Thỳmele, neotrop.;

**Eùdamus, Epargýreus,** Am. mostly trop.; **Urbànus** (= Hespèria, = Pýrgus, auct.) widespr.; **Erýnnis** (= Thánaos) palæarc. Am.; **Tagìades**, Old World tropic.; **Pholisòra**, nearct.; **Celœn**-

	orrhinus, tropicopol.). (ACHLYÓDIDÆ, EUDÁMIDÆ,
	PÝRGIDÆ, TELEGÓŃIDÆ, THYMÉLIDÆ, URBANÌNÆ, HESPERIÌNÆ) ERYNNÌNÆ
	Fore wings with discal cell less than two-thirds as long as wing,
	$M_2$ arising closer to $M_1$ than to $M_3$ ; and usually with an oblique
	brownish brand across the disk of the fore wings. Cosmo-
	politan. (Cènides, Cyclópides, ethiop.; Ancylóxipha,
	Atrytène, Thymélicus, Am.; Ochlèdes, nearc.; Hespèria
	Atrytène, Thymélicus, Am.; Ochlèdes, nearc.; Hespèria (= Pámphila) holarc.; Hálpe, indomal.; Dálla, Thràcides,
	neotrop.; <b>Párnara</b> , ethiop., indomal.). (CYCLOPIDIDÆ,
	PAMPHILÌNÆ, auct.) HESPERIÌNÆ
157.	Front legs normal, or if slightly reduced in size and structure,
	with the claws toothed or bifid
	Front legs, at least in the male, more or less strikingly different
	from the other pairs, usually not used for walking, the claws of
	their tarsi, when present, never toothed or split160
158.	
100.	fore wings with Cu apparently four-branched and with two or
	three anal veins; hind wings with the anal area reduced, one
	anal vein present
	Tarsal claws bifid; anterior tibiæ without pads; fore wings with
	Cu apparently three-branched and one anal vein present;
	hind wings with two anal veins: medium sized or rather small
	butterflies with broad wings, typically yellowish or white with
	blackish marginal markings. Cosmopolitan. ( <b>Áscia</b> (= Póntia,
	Pieris) (A. ràpæ, Cabbage butterfly), Eùrymus (=Còlias),
	Eurèma, Nathàlis, Catopsília, Zerène, Anthócharis,
150	Synchloe (Orange-tips)). (PIÉRIDÆ)ASCÌIDÆ
159.	,
	hind wings usually with a wavy margin and a tail-like prolon-
	gation; ground color of wings yellow or black; large showy
	butterflies with contrasting color pattern. (Fig. 444). Swallow-
	tails. (Iphiclides, Tròides (=Ornithóptera), Laértias, Pa-
	pílio). (EQUÍTIDÆ) PAPILIÓNIDÆ
	Fore wings with radius four-branched, no crossvein between the
	base of the discal cell and the anal vein; medium sized butter-
	flies, ground color of wings cream-white, marked with dusky
	and usually with a red eye-spot on the hind wing; mainly alpine
	species. Parnassians. (Parnássius, holarc.).

160. Front legs much reduced in size in both sexes and without tarsal



Figs. 439-444. Lepidoptera

- 439. Caligo, wings (Stichel) Brassolidæ.
- 440. Caligo, tibia and tarsus of front leg of female (Stichel) Brassolidæ.
- 441. Caligo, tibia and tarsus of front leg of male (Stichel) Brassolidæ.
- 442. Danais, wings (Scudder) Danaidæ.
- 443. Calephelis, wings (Stichel) Riodinidæ.
- 444. Papilio, wings (Comstock) Papilionidæ.
- 162. Front feet of female ending in a corrugated knob; fore wings with the subcostal vein forked at the extreme base, A3 preserved; antennæ not scaled above; generally large butterflies with bold contrasting coloration; mainly tropical. (Figs. 370, 442). (See couplet 166). (**Dánais** (= Anòsia) (D. plexippus, Milkweed

	butterny), Heliconius, Acræa). (ACRÆIDÆ, EUPLŒ-
	$ID\mathscr{E}$ , $LYMN\acute{A}DID\mathscr{E}$ , $MANI\acute{O}LID\mathscr{E}$ ) <b>DANÀID</b> $\mathscr{E}$
	Front tarsi of female present, though more or less abbreviated;
	fore wings with A3 lacking. (Fig. 438) 163
163.	Fore wings elongate oval, twice as long as broad; mainly tropi-
	cal
	Fore wings much less than twice as long as broad165
164.	Antennæ clothed with scales, at least above; front tarsi of the
	female four-jointed; wings opaque; medium sized, brightly
	colored butterflies. Almost entirely neotropical. (Apostrà-
	phia, Colànis, Diòne (= $Agraulis$ ), Migonitis (= $Helicònius$ ,
	auct.); Cethòsia, indomal.). (PALÆOTRÓPIDÆ, HELI-
	CONÌIDÆ)EUEÍDIDÆ
	Antennæ naked; wings often in great part translucent and desti-
	tute of scales. (Dircénna, Ithòmia)ITHOMÌIDÆ
165.	Fore wings with some of the veins greatly swollen at the base:
	usually small butterflies, rarely rather large, feeble fliers,
	frequently of brownish colors, with yellowish eye-spots or
	ringed marks. (Fig. 438). (Megísto (=Cissia, Neonýmpha),
	Cœnonýmpha, Sátyrus (=Cercyonis), Árgus (=Satyròdes),
	Œneis, Enòdia). (AGAPÉTIDÆ)SATÝRIDÆ
	Fore wings with the veins not swollen at the base: large, tropical
	species with very broad wings, above with deep rich colors,
	below with eye-spots and intricate lines. (Figs. 439, 440, 441).
	Neotropical. (Caligo, Brássolis, Opsíphanes). (CALIGÓN-
100	IDÆ) BRASSÓLIDÆ
166.	Hind wings with a large, cradle-like depression along the anal
	edge in which the abdomen rests: large species, usually with
	brilliant metallic blue color; tropical. (Mórpho, neotrop.;
	Amathùsia, Amathuxídea, indoaustr.). (ÁRGIDÆ).  MORPHÒIDÆ
	Hind wings without a structure of this sort: usually moderate-
	sized species without brilliant blue coloration; many common
	brightly colored butterflies. (Figs. 437, 371). (Dryas (= Ar-
	gýnnis), Brénthis, Euptoièta (Fritillaries); Euphýdryas
	(=Lemònias, Melitæa), Phyciòdes (=Crescent-spots); Ha-
	màdryas (= Aglais, Euvanéssa, Vanéssa), Eugònia, Junònia,
	Polygònia (= $Grápta$ ), Vanéssa (= $Pyrámeis$ ), (Angle-wings)
	Basilárchia (=Limenitis), Heteróchroa (Sovereigns),
	Chlorippe (= Apatùra) (Emperors); Anà (Goatweed
	butterflies). $(ARGYRÈID\pounds)$ NYMPHÁLIDÆ

167. Palpi very long, porrect, from one-fourth to one-half as long as the body and thickly hairy. Snout butterflies. Cosmopolitan. (Libýthea, Hýpatus) LIBYTHÈIDÆ
Palpi not elongated, of ordinary size
a. Hind wings with base of costa developed (Fig. 443); almost wholly neotropical. (Mesosèmia, Riodina (=Erycina, auct.), S. Am.; Eurýbia, Cària, Bàotis, Caléphelis, Lýmnas). RIODININÆ
Hind wings without basal vein. ( <b>Dodòna</b> , <b>Zémeros</b> , indomal.; <b>Abísara</b> , indomal., ethiop.; <b>Euselàsia</b> , nearct.; <b>Hàdes</b> , <b>Helicòpis</b> (= <i>Plebèius</i> ), neotrop.; <b>Nemeòbius</b> , Eur.; <b>Dicallaneùra</b> , malay.)
Hind wings with the costa not thickened at the base and without the humeral vein; fore wings with $M_1$ almost always arising from the anterior angle of the discal cell: generally small, delicate species, the antennæ ringed with white; often brightly colored and with very slender tail-like appendages on the hind wings. Widespread. (Strymon (= $Th\`{e}cla$ ), Atlides, Inci-
sàlia, Mitoùra (Hair-streaks); Lycèna (= Heòdes, Chrysó-phanus), Tharsàlea (Coppers); Evères, Lycenópsis, Glaucopsyche (Blues). (CUPIDÍNIDÆ, RURÁLIDÆ).
LYCÆNIDÆ  169. Head of female moth of the usual form and structure170  Head of female of abnormal form and structure, resembling that of the caterpillar. (See couplet 42).  Females of some HETEROGÝNIDÆ
170. Moth developing in and frequently never leaving a sack or case constructed by the larva and carried about by the latter during growth. (See couplets 26, 40, 42). Females of <b>PSÝCHIDÆ</b> TALÆPORÎIDÆ and some HETEROGÝNIDÆ
Moth not developing in such a sack constructed by the larva, the latter entirely free-living
171. Body scaly or hairy; or if woolly, without ocelli
couplet 89) Females of some ARCTIDA  172. Palpi usually long and extended forward
Females of some TORTRÍCIDA

173. Body stout, densely woolly, with short legs. (See couplet 85).

Females of some **LYMANTRÎIDÆ**Body slender, hairy or scaly, with relatively long legs. (See couplet 50) . . . . . . Females of a few **GEOMETRÒIDEA** 

# Lepidoptera, Larvlpha

The key given below follows closely that used by Forbes in his Lepidoptera of New York state, which is based on the system elaborated by Fracker. It includes the more important families as they may be distinguished on the basis of those genera whose larvæ have been carefully and systematically studied. Wherever possible other families have been added, but at present a knowledge of even the European and North American forms is so incomplete that the present key must be used only as a guide in identification, and must in no way be relied upon as presenting characters of definite and final accuracy. Many families are necessarily omitted.

1.	Thoracic legs present, formed of distinctly chitinized segments;
	abdominal prolegs usually present or indicated by crochets2
	Thoracic legs wanting or reduced to fleshy swellings without chit-
	inized segments; prolegs frequently vestigial or absent7
2.	Body segments bearing setæ, stiff hairs, or spines, arising from

# MICROPTER ÝGIDÆ

- - Head concealed within the prothorax which bears a slit below through which the mouthparts are protruded for feeding; body almost always with spines or secondary hair, the primary setæ

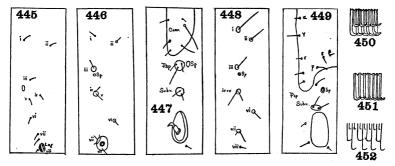
obsolete; body with obscure incisures, but usually with pits. Slug-caterpillars ..... EUCLÈIDÆ

6. Setæ iv. and v. distant on the abdominal segments (Fig. 51); prolegs present, but without hooks; living in the pods of Yucca.

Tegeticula of the PRODÓXIDÆ

Setæ iv. and v. adjacent (Fig. 446); prolegs absent.

A few GELECHIIDÆ



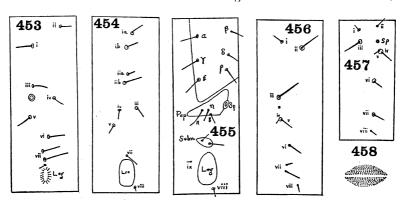
Figs. 445-452. Lepidoptera, Larvæ

- 445. Adela, setal map of third abdominal segment (Forbes) Adelidæ.
- 446. **Dichomerus**, setal map of sixth abdominal segment (Forbes) Gelechidæ.
- 447. Dichomerus, setal map of prothorax (Forbes) Gelechiidæ.
- 448. **Dicymolomia**, setal map of third abdominal segment (Forbes).
- 449. Sthenopis, setal map of prothorax (Fracker).
- 450. Biordinal or double-rowed arrangement of crotchets on abdominal proleg (Forbes).
- 451. Uniordinal or single-rowed arrangement of crochets on abdominal proleg (Forbes).
- 452. Triordinal or triple-rowed arrangement of crochets of abdominal proleg (Forbes).

Head with six small ocelli on each side......13

9.	Two ocelli on each side of the head; front not extending to the ver-
	tex; larvæ leaf-miners, forming blotch mines <b>HELIOZÉLIDÆ</b>
	One large ocellus on each side of head or ocelli obsolete10
10.	Front triangular, the ocellus frontal; larva making a large blotch
	mine from which it drops to spin a cocoon in the ground.
	ERIOCRANÎID <i>Æ</i>
	Front quadrangular; ocellus lateral in position
11.	Front narrowed behind, wider in front; body cylindrical; prolegs,
	if present, vestigial and borne on the second to seventh ab-
	dominal segments
	Front widest behind; body usually depressed; prolegs, if present,
	borne on the third to fifth segments of abdomen. See couplet
	13) GRACILARÌIDÆ
10	Body not greatly lengthened, about five times as long as thick;
14.	prolegs usually present; larvæ mining in leaves, bark or fruits,
	or forming galls in twigs or petioles NEPTICULIDÆ
	Body very slender, about ten times as long as thick; prolegs ab-
	sent; larvæ mining near the surface of the stems of herbaceous
	or other plants OPOSTÉGIDÆ
13.	Vestigial prolegs, bearing hooks, present on the third to sixth
	abdominal segments; larvæ mining in leaves, usually making
	a blotch mine
	Abdomen without prolegs on the sixth segment; larvæ leaf miners,
	at least in the early stages, the older larvæ sometimes spinning
	a shelter on the surface of the leafGRACILARÌIDÆ
14.	Body without secondary or tufted setæ; tubercle vi. with a single
	seta; vii. with at most three setæ, unless the proleg has a multi-
	serial circle of hooks when it may bear four setæ15
	Body bearing tufted or secondary hairs; at least two setæ on
	tubercle vi. on the sixth abdominal segment, or with addi-
	tional setæ on the proleg52
15.	Sixth abdominal segment bearing prolegs, although prolegs may
	be absent on the more anterior segments
	Sixth abdominal segment without prolegs. (See couplet 13).
	GRACILARÎIDÆ
16.	Hooks (crochets) of the prolegs arranged in a circle or ellipse,
	which may be incomplete, or in transverse band
	Crochets forming a single band, sometimes with a few vestigial
	ones in addition
17	Prespiracular wart of prothorax with two setæ
± •, •	Prespiracular wart of prothorax bearing three setæ
	Enopie dual wait of productax bearing diffee serge

18.	Hooks on prolegs uniordinal, i.e. with their tips forming a single
	line (Fig. 25); body cylindrical; tubercle vii. of mesothorax
	bearing a single seta; surface of body rough and granular.
	ORNEÓDIDÆ
	Hooks on prolegs biordinal or triordinal, <i>i.e.</i> with their tips form-
	ing two or three parallel lines (Figs. 26, 27), or uniordinal in
	larvæ with stout body and bisetose tubercle vii. of mesothorax.
	Chrysauginæ of the <b>PYRALÍDIDÆ</b>
19.	Hooks on prolegs forming two transverse bands, rarely reduced
	to one
	Hooks on prolegs forming a circle or ellipse which is sometimes
	broadly interrupted26
20.	Prolegs with the hooks either in a single transverse row, or in two
	multiserial bands21
	Prolegs with the hooks arranged in two simple (uniordinal)
	series
21.	Prolegs represented by one uniserial band of very small hooks,
	the prolegs practically absent; leaf-miners or case-bearers.
	INCURVARÌID <i>Æ</i>
	Prolegs with many, short, vestigial hooks arranged in two trans-
	verse multiserial bands (Fig. 458); case-bearers living in a
	portable lenticular case made of a piece of leaf ADÉLIDÆ
22.	Setæ iv. and v. of abdomen remote; young larvæ living in ser-
	pentine mines in leaves, later feeding externally.
	Bucculatrix of the LYONETIIDÆ
	Setæ iv. and v. of abdomen adjacent (Fig. 448)23
23.	Hooks on the anal prolegs disposed in two groups; habits various.
	GELECHÌIDÆ
	Hooks on the anal prolegs in a single series24
24.	5,
	the vertex
	Front short extending about one-third the way to the vertex.
	$Cossula  ext{ of the } \mathbf{C} \mathbf{\acute{O}SSID} oldsymbol{\mathcal{E}}$
25.	Spiracles elliptical, normal in size, those of the eighth abdominal
	segment placed higher up than the others; body white, without
	markings; boring in woody, or more rarely in the stems of
	herbaceous plants ÆGERÌIDÆ
	Spiracles very small, circular, the last pair about in line with the
	others; larvæ usually living in portable cases and feeding ex-
	ternally or sometimes mining, but never boring into the stems
	of plants



Figs. 453-458. Lepidoptera, Larvæ

- 453. Scardia, setal map of third abdominal segment (Forbes) Tineidæ.
- 454. Acrolophus, setal map of mesothorax (Fracker) Acrolophidæ.
- 455. Acrolophus, setal map of prothorax (Fracker) Acrolophidæ.
- 456. Carposina, setal map of first abdominal segment (Forbes) Carposinidæ.
- 457. **Carposina**, setal map of eighth abdominal segment (Forbes) Carposinidæ.
- 458. Adela, arrangement of crochets on proleg (Forbes) Adelidæ.
- 29. Seta i. of abdomen much lower than ii. (Fig. 453); larvæ generally case-bearers; the case usually ending in a triangular valve, more rarely lenticular; often living on animal matter, fungi, etc.

TINÈIDÆ

Seta i. of abdomen not lower than ii.; habits varied.

**HELIODÍNIDÆ** 

30. Meso- and metathorax with setæ i.a and i.b close together (Fig.

	454); abdomen with seta iv. below the level of the spiracle31
	Meso- and metathorax with seta i.a in front of i.b and well sepa-
	rated from it; larvæ boring in woody plants, commonly in the
	roots
31.	Prothorax with seta beta at a higher level than alpha (Fig. 455).
01.	ACROLÓPHIDÆ
	Prothorax with seta beta at a lower level than alpha. The Plu-
	tellidæ, Argyresthesiidæ and Acrolepiidæ will also run out here.
	Families related to the <b>YPONOMEÙTIDÆ</b>
00	
32.	Last pair of spiracles placed very high up, nearer to the mid-
	dorsal line than setæ i. of the anterior abdominal segments;
	larvæ internal feeders in the fruits of various plants. (Figs.
	456, 457) CARPOSÍNIDÆ
	Last pair of spiracles placed lower down, in the normal position. 33
33.	Mesothorax with two setæ on tubercle vii., above base of leg. 34
	Mesothorax with only a single seta on tubercle vii. (Fig. 549);
	seta ii. on ninth abdominal segment placed higher up than i.
	(Fig. 463)
34.	Prothoracic spiracle higher than wide, i.e. with its longer axis
	vertical; seta i. of ninth abdominal segment placed higher up
	than ii.; larvæ boring in the tissues of plants, or leaf-rollers.
	THYRÍDIDÆ
	Prothoracic spiracle with its longer axis horizontal; larvæ living
	in a portable case, from the anterior end of which they protrude
	the anterior end of the body when feeding
25	Thoracic legs with the last two segments stout; seta i. on abdomi-
55.	nal segments below the level of seta ii. (Fig. 460). PS ÝCHIDÆ
	Thoracic legs with the last two segments very slender; seta i.
	on abdominal segments above the level of seta ii. (as in Fig.
0.0	461) TALÆPORÌIDÆ
36.	Setæ ii. of ninth segment nearer together across the dorsum than
	these setæ are on the preceding segments, frequently on the
	same plate
	These setæ as far apart across the dorsum on the ninth segment
	as on any of the preceding segments, very rarely on the same
	plate
37.	
	461); prolegs with the hooks forming a uniordinal row (Fig.
	451); larvæ boring in herbaceous plants or feeding in the seeds.
	PHALONÌIDÆ

	Setæ iv. and v. on abdomen not at the same level, a line connect-
	ing them diagonal or vertical; hooks on prolegs usually multi-
	ordinal (Fig. 452). (Including the Olethreutidæ and several
	other related families) TORTRÍCIDÆ
38.	Coxæ of the metathoracic legs separated by less than twice their
	own width39
	Coxæ of the metathoracic legs separated by twice their own width;
	prolegs small; small species; larvæ with varied habits, mining
	in leaves, boring in stems, rolling leaves or feeding in seeds.
	COSMOPTER Ý GIDÆ
39.	Setæ i. and ii. on abdomen widely separated40
	Setæ i. and ii. on abdomen close together. (See couplet 29).
	HELIODÍNIDÆ, part
40.	Front short, not extending more than half way to the vertex,
	usually much less than half way41
	Front longer, extending almost or fully two-thirds of the distance
	to the vertex, narrowly pointed above; small species with uni-
	ordinal or biordinal hooks on the prolegs
41.	Species boring in plants, usually in wood; setæ iv. and v. placed
	on separate tubercles on the ninth abdominal segment; body
	white
	Leaf feeders; setæ iv. and v. together on the same tubercle on the
	ninth abdominal segment; body brightly colored. STENÓMIDÆ
42.	Setæ ii. of the ninth abdominal segment placed on a common
	dorsal plate; adfrontal plates short, not attaining the vertex.
	ZEUZÉRIDÆ
	Setæ ii. of the ninth abdominal segment on separate tubercles on
	each side of the mid-dorsal line adfrontal plates attaining
	the slightly cleft vertex CÓSSIDÆ
43.	Hooks of prolegs biordinal, i.e. of two lengths, their tips forming
	two parallel lines (Fig. 450)
	Hooks of prolegs uniordinal, all of the same length with their tips
	forming one continuous line
44.	Ocelli irregularly arranged, three of them closer together in a
	group (Fig. 464); larvæ usually webbing or rolling leaves.
	ŒCOPHÓRIDÆ
	Ocelli evenly spaced in a single group on each side of the head;
	habits various
45.	Seta iii. on the eighth segment of the abdomen usually placed above
	and behind the spiracle; habits varied, often scavengers, feeding
	in puts, or produced on scale insects RIASTORÁSIDÆ

	Seta iii. on eighth segment of abdomen usually placed just above or slightly before the spiracle; larvæ leaf-rollers or feeding
10	generally on foliage GLYPHIPTERÝGIDÆ
46.	Prespiracular wart of prothorax bearing three setæ
417	Prespiracular wart of prothorax bearing two setæ
47.	Setæ iv. and v. of abdomen remote, or if approximate the setæ
	beta of the two sides of the body much closer together than the
	setæ alpha of the two sides of the body on the prothorax (Fig.
	462); prolegs long and slender. (See couplet 31).  YPONOMEŪTIDÆ
	TPONOMEUIIDA
	int =   460
	** 1
	"\ 461   "F 461   "F 461   "F 461   "T
Ī	459
	VII \
	Owii
	Figs. 459-464. Lepidoptera, Larvæ
459. 460.	Phalonia, setal map of mesothorax (Forbes) Phaloniidæ.  Thyridopteryx, setal map of fourth abdominal segment (Forbes)
	Psychidæ.
461.	
462.	
463.	
464.	Depressaria, arrangement of larval ocelli (Forbes) Œcophoridæ.
	Setæ iv. and v. of abdomen placed close together; beta about as far apart as alpha; prolegs usually short ETHMÎIDÆ
48.	Tubercle vii. on meso- and metathorax with two setæ
40	Tubercle vii. on meso- and metathorax with a single seta51
49.	Body setæ minute, the tubercles placed in obscure rings; head
	unusually wide; prolegs reduced; larvæ forming nests in loosely
	rolled leaves
	Body setæ heavy, almost always spinulose, on conspicuous tu-

<b>50</b> .	Tubercle iii. of abdomen bearing two setæ; larvæ usually feeding
	on lichens LITHOSÌIDÆ
	Tubercle iii. of abdomen with a single seta. (See couplet 69).
	Utethersa of the ARCTIIDÆ
51.	Body usually with enlarged contrasting tubercles; eighth ab-
	dominal segment with a conspicuous hump; banded or spotted
	with black; larvæ feeding externally on foliage. AGARÍSTIDÆ
	Body not as above; usually dully colored and not with conspicu-
	ous transverse bands of black; usually external feeders on the
	leaves of plants, sometimes boring into fruits, or cutworms.
	NOCTŪIDÆ
52.	Less than four pairs of ventral prolegs or with the first pair greatly reduced
	Four pairs of ventral prolegs bearing hooks, and sometimes with
	additional ones not bearing hooks
53.	Body hairs tufted; hooks on prolegs uniordinal (Fig. 25); three
	pairs of ventral prolegs; larvæ feeding externally on foliage.
	NÓLIDÆ
	With only a few accessory hairs or sometimes with fine secondary
	hair; usually only one pair of ventral prolegs (on the sixth ab-
	dominal segment) in addition to the anal pair (on the eighth
	segment). Including also most of the families of Geometroidea.
	GEOMÉTRIDÆ
54.	Four pairs abdominal prolegs; anal prolegs sometimes reduced or absent
	Four pairs of abdominal prolegs on segments three to six and in
	addition a pair without hooks on segments two and seven;
	body bearing stinging hairs mixed with tufts of dense, soft hair.
	MEGALOPÝGIDÆ
55.	Anal prolegs entirely absent; body with secondary hair below, but
	none above except on the few enlarged tubercles. <b>DREPÁNIDÆ</b>
	Anal prolegs present as a pair of large tubercles, or flagella at least
	normally fully developed
56.	Hooks on the prolegs uniordinal, all of equal length, their tips
	forming a single continuous line (Fig. 451)57
	Hooks on the prolegs biordinal or triordinal, of two or three
	lengths, their tips forming two or three parallel lines71
57.	Setigerous tubercles vestigial or absent, or obscured by secondary
	hair
	At least tubercle vi. many haired and distinct; secondary hairs
	sparse or absent above the prolegs65

58.	Anal plate bifurcated; head roughly papillose; third ocellus very
	$large \dots SAT $ ÝRIDÆ
	Anal plate simple; head smoother; third ocellus rarely much en-
	larged
59.	Body caterpillar-like in form; vegetarian species, not parasitic on
	other insects
	Body hemispherical, with a complete circle of uniordinal hooks;
	living as external parasites of Homoptera (Jassidæ and Ful-
	goroidea) EPIPYRÓPIDÆ
60.	Spiracles small, circular; ventral prolegs slender, more or less
	petiolate, with expanded walking surface; usually leaf-rollers,
	more rarely boring in the stems of plants PTEROPHÓRIDÆ
	Spiracles elliptical, larger; ventral prolegs short61
61.	
	Secondary setæ very sparse or absent above the prolegs; with
	simple setæ or a few accessory ones
<b>62.</b>	1, 1
	oped as the ventral ones. (See couplet 51). A few <b>NOCTUIDÆ</b>
	Notch of labrum acute, with convergent sides; anal prolegs much
	reduced, not functional; body often with spines, long fleshy
	tubercles or humps, frequently brightly colored.
	NOTOD ÓNTIDÆ
63.	Tubercle iv. much lower on the seventh abdominal segment than
	on the other segments; anal prolegs more or less reduced or
	modified
	Tubercle iv. at about the same level on abdominal segments six,
	seven and eight. (See couplet 65). A few LYMANTRIDÆ
64.	
	Skin smooth, not shagreened. (See couplet 62). NOTODÓNTIDÆ
65.	Sixth and seventh abdominal segments bearing eversible glands
	in the middle above; body hairy, usually with conspicuous
	tufts of brightly colored hairs; feeding externally on foliage.
	LYMANTRÌIDÆ
	No dorsal eversible glands
66.	
0.7	Spiracles of the usual size, elliptical
67.	Ventral prolegs short, with a straight band of heavy hooks.  PYROMÓRPHIDÆ
	Ventral prolegs slender, more or less petiolate, with expanded
	walking surface sometimes bearing a circle of hooks. (See
	walking surface sometimes bearing a circle of nooks. (See couplet 60)
	couplet ou)

68.	Mesothorax with two or three setigerous tubercles above the level of the spiracles
	Mesothorax with only one such tubercle above the level of the spiracles; externally feeding species, commonly on grasses and low plants, also some on lichens EUCHROMÎIDÆ
69.	Tubercle, or seta iv. placed much lower on the seventh abdominal
	segment than on the sixth or eighth segment, or absent70 Tubercle, or seta iv. placed at the same level on the seventh as
	on the adjacent abdominal segments; body clothed with dense
	clusters of hairs and often with long, brightly colored tufts;
	feeders on a great variety of plants ARCTIDÆ
70.	Hooks on prolegs of even length, or gradually decreasing in size
• • •	toward the ends of the row. (See couplet 51).
	A few <b>NOCTŪIDÆ</b>
	Hooks on prolegs abruptly decreasing in size toward each end of
	the row; feeding externally on leaves, often brightly colored.
	PERICÓPIDÆ
71.	Body without noticeable accessory or secondary hair; with not
	more than eight hairs on the prolegs72
	Body with numerous secondary setæ, at least on the prolegs; anal
	prolegs well developed77
72.	Prolegs with the hooks arranged to form a complete circle73
	Prolegs bearing a band of hooks on the inner side, and sometimes
	also a much weaker band on the outer side
<b>7</b> 3.	Subdorsal setæ of abdomen simple
	Subdorsal setæ of abdomen represented by warts; body with
	tufted hair from small warts; often webbing leaves.
	SCYTHRÍDIDÆ
74.	
	the prolegs; larva spinning leaves together for a nest when
	young, later making a portable case of silk and pieces of leaf.
	LACOSOMÁTIDÆ
	Surface of head smooth; body widest on the first segment of the
	abdomen; larva constructing a portable case. Some <b>XYLORÝCTIDÆ</b>
75.	Prespiracular wart of prothorax bearing two setæ; setæ iv. and v.
10.	of abdomen usually distant from each other
	Prespiracular wart of prothorax bearing three setæ; setæ iv. and
	v. of abdomen approximate; anal prolegs well developed, with
	hooks; larva spinning a light web ETHMIDÆ
76.	Spiracles subequal in size. (See couplet 49) THYATÎRIDÆ
	1 ( (

	First and last spiracles twice as large as the others; young cater- pillars living in a communal web, later feeding more or less exposed EPIPLÉMIDÆ
77	Setæ very irregular in length, some ten times as long as the others;
11.	with obscure warts, at least in the earlier stages; sometimes
	provided with scale-like hairs
	Setæ subequal or sometimes with setæ and prominent warts and
	·
<del>7</del> 0	spines
78.	
	or with the notch somewhat shallower and continued as a groove
	to the base of the labrum; body hairy, strongly depressed, often
	with slender dorsal hair tufts.
	EUPTERÓTIDÆ, APATELODÌNÆ
	Labrum with a shallower notch which is not continued as a groove;
	no dorsal hair-pencils; larva very hairy, with dense secondary
	hair; usually feeding on the foliage of trees, sometimes in a
=0	communal web LASIOCÁMPIDÆ
79.	6
	or tubercle
	Eighth abdominal segment not thus armed on the mid-dorsal
00	line
80.	Body bearing numerous branching spines or enlarged tubercles. 81
	Body not thus ornamented, at most with two pairs of small spines
01	on the thorax
81.	
	Head angulated or spined above, or the abdomen with several
	mid-dorsal spines; hooks of prolegs usually triordinal. (Fig.
00	27)NYMPHÁLIDÆ
82.	Ninth segment of abdomen with a median dorsal spine; spines on
	body strongly unequal, armed with short nodules or short
	spinules; large and brightly colored CITHERONIDÆ
	Ninth segment of abdomen without a median dorsal spine,
	or the body spines subequal and armed with dense, long
വ	spinules
83.	Tubercles ii. of ninth abdominal segment from the two sides of
	the body fused into one on the dorsal line; body with bushy,
	branching spines, not brightly colored.
	SATURNÎIDÆ, HEMILEUCÎNÆ
	Tubercles ii. of ninth abdominal segment separate on each side
	of the median line; very large, brightly colored caterpillars.  SATURNÎIDÆ, SATURNIÎNÆ
	SATUKNIIDÆ, SATUKNIINÆ

84.	Abdominal segments each divided into six or eight annulets; prolegs normal, not widely separated; body more or less cylindrical, usually with oblique stripes or bands of color. Sphinx caterpillars
	Abdominal segments with two or three obscure annulets; prolegs unusually widely separated; body without oblique color markings. Silkworms
05	Head rounded, of the usual form
85.	Head strongly elevated, triangular in outline.
	Lapara of the SPHÍNGIDÆ
86.	Ninth abdominal segment without a median dorsal spine87
00.	Ninth abdominal segment bearing a small median dorsal spine.
	(See couplet 80)
87.	Hooks on prolegs forming an ellipse, at most narrowly inter-
	rupted
	Hooks on prolegs arranged in one band which is occasionally in-
	terrupted; or rarely forming two widely separated bands89
88.	Head much larger than the prothorax which forms a narrow
	"neck"; body widest at the middle, distinctly tapering toward
	both ends; larvæ commonly forming a nest in a folded leaf or
	in several leaves webbed togetherHESPERÎIDÆ
	Head smaller than the prothorax, partially retractile; body cylin-
00	drical; larvæ boring in the stems of Yucca MEGATHÝMIDÆ
89.	Band of hooks on the prolegs reduced or interrupted at the middle and with a narrow, spatulate, fleshy lobe arising near the inter-
	ruption; head small90
	Prolegs without a fleshy lobe near the middle of the band of
	hooks
90.	
	amount of secondary hair
	Head smaller, rarely more than one-third as wide as the body;
	secondary hair less prominent; body short and broad, more or
	less slug-like, with the legs and prolegs very short.
	LYCÆNIDÆ
91.	Prothorax above bearing an eversible, forked scent gland; gland
	when retracted showing as a groove; body not hairy or spiny,
	but sometimes bearing fleshy filaments
92.	Prothorax without a scent gland93 Setæ minute, not borne on tubercles or warts (except in the early
94.	stages)
	Setæ well developed: some warts present PARNASÌIDÆ

93.	Head and body entirely without spines, high tubercles or fleshy
	filaments
	Spines, high tubercles or fleshy filaments well developed on the
	body; when reduced, large spines or tubercles are present on
	the head
94.	Anal plate rounded, entire95
	Anal plate bifurcate at tip, bearing two distinct processes.
	SATÝRIDÆ
95.	Prolegs with only a single row of hooks, forming a curved band 96
	Prolegs with reduced hooks on the outer side in addition to the
	well developed band; head small; setæ never borne on promi-
	nent warts LIBYTHÈIDÆ
96.	Head noticeably larger than the prothorax NYMPHÁLIDÆ
	Head smaller than the prothorax; setæ usually borne on promi-
	nent warts ASCÎIDÆ
97.	Mesothorax and sometimes several other segments bearing fleshy
	filaments; secondary setæ short and confined to the prolegs.
	DANAIDÆ
	Body without fleshy filaments98
98.	
	on the abdomen as long as the width of the mesothorax; each
	abdominal segment with three lateral spines but no median
	ones above EUEÌDIDÆ
	Body spines, if present, not so slender; those on the abdomen
	shorter than the width of the metathorax; median spines
	usually present on the dorsum
	v r

# LITERATURE ON LEPIDOPTERA, GENERAL

- Aurivillius, C. and Wagner, H. Lepidoptorum Catalogus. Parts 1–28. W. Junk, Berlin (1911–23). (The parts so far as they have appeared are listed separately.)
- Bang-Haas, O. Katalog der im Seitz nicht enthaltenen paläarktische Macrolepidopteren. Novitates macrolep. Dresden, 1, pp. 1–238 (1926).
- Barnes, W. and McDunnough, J. H. Check list of the Lepidoptera of Boreal America. Decatur, Ill. (1917).
- Barrett, C. G. The Lepidoptera of the British Isles. 11 vols. London (1893–1907).
- Campos, F. Catálogo preliminar de los Lepidópteros del Ecuador. Rev. Col. Rocafuerte, Guayaquil, 9, pp. 3–106 (1927).
- Candèze, L. Lepidoptères Hétérocères de l'Indochine Française. (Catalogue) Encycl. Entom. Lepidop. II, fasc. 2, pp. 73–133 (1926–27).

- Cotes, E. C. and Swinhoe, C. Catalogue of the moths of India. 7 parts. Calcutta (1887–89).
- Druce, H. H. J. C. Heterocera, in Biologia centrali-Americana, **1-3** (1881–1900).
- Dyar, H. G. A List of North American Lepidoptera and key to the literature. Bull. U. S. Nat. Mus., No. 52, 723 pp. (1902).
- EECKE, R. VAN. De Heterocera van Sumatra. I. Zool. Meded., 8, pp. 153–217 (1925).
- FORBES, W. T. M. Field Tables of Lepidoptera. 141 pp. Worcester, Mass. (1906). Lepidoptera of New York and Neighboring States (Microlepidoptera, Pyraloids, Bombyces). Mem. Cornell Univ. Agric. Expt. Sta., No. 68, 729 pp. (1923).
- Fracker, S. B. The Classification of Lepidopterous Larvæ. Illinois Biol. Monog., 2, No. 1 (1915).
- GAEDE, M. Lepidoptera Heterocera. Tierwelt Deutschlands, pt. 14, pp. 1–334 (1929).
- Hampson, G. F. The Moths of India. Fauna of British India, 4 vols. (1892–96).
  The Moths of South Africa. Ann. South African Mus., 2, pp. 33–255;
  3, p. 389 (1900–05). (In three parts.)
  - Catalogue of the Lepidoptera Phalænæ in the British Museum, 12 vols. London (1898–1913). Supplement to Vol. 3 (1920).
- Hampson, G. F. and Durrant, J. H. List of the Families and Subfamilies of Lepidoptera. Novitat. Zool., 25, pp. 366–394 (1918).
- Heinrich, Carl. Revision of North American Moths of Subfamily Eucosminæ of the Family Olethreutidæ. Bull. U. S. Nat. Mus., No. 123, 298 pp. (1923).
  - Revision of North American Laspeyresiinæ and Olethreutinæ. Bull. U. S. Nat. Mus., No. 132, 216 pp. (1926).
- Hering, M. Lepidoptera. In Tierwelt Mitteleuropas, 6, Lief. 3, pp. XVIII 1–94 (1928).
- HOLLAND, W. J. The Moth Book. Doubleday, Page & Co., New York (1903).HUDSON, G. V. The Butterflies and Moths of New Zealand. 386 pp. Wellington, Ferguson and Osborn (1928).
- Janse, A. J. T. Check-List of South African Lepidoptera Heterocera. Transvaal Mus. Pretoria, 219 pp. (1918).
  - A Key to the Families, Subfamilies and Tribes of the Order Lepidoptera with special reference to the South African Species. South African Journ. Sci., **22**, pp. 318–345 (1926).
- Kirby, W. F. A Synonymic Catalogue of the Lepidoptera Heterocera. 951 pp. London (1892).
- Kusnezov, N. J. Faune de l'U. R. S. S. et des pays limitrophes. (Pub. by Mus. Zool. Acad. Sci., Leningrad) (1915). (Some parts on Lepidoptera have so far appeared).
- LEDERER, G. Handbuch für den praktischen Entomologen., 2, Rhopalocera, 172 pp., 3, Heterocera, 172 pp. Frankfurt a. M. (1921–23).

- MEYRICK, E. A Revised Handbook of British Lepidoptera. 914 pp. London, Watkins and Doncaster (1928).
- MEYRICK, E. and Walsingham, T. de G. Lepidoptera. In Fauna Hawaiiensis, 2 vols. (1899–1907).
- Moore, F. Lepidoptera of Ceylon. 3 vols. London (1880-87).
- Mosher, Edna. A Classification of Lepidoptera based on characters of the pupa. Bull. Illinois State Lab. Nat. Hist., 12, pp. 12–159 (1916).
- Rebel, H. Schmetterlingsbuch. Stuttgart. (1910).
- Seitz, A. Macro-Lepidoptera of the World (1906———). (Issued in a large number of parts containing many illustrations and much valuable material, but not yet indexed. Many families have so far been very completely treated. See also Closs, A. The Systematic Arrangement of Families in Seitz's "Macrolepidoptera." Entom. Mitteil., 9, pp. 22–30 (1920).)
- Spuler, A. Die Schmetterlinge Europas. 3 vols. and vol. 4 on Caterpillars. Stuttgart (1901–10).
- Standfuss, M. Handbuch der palaärktischen Grossschmetterlinge. Jena (1896). Standinger, O. and Rebel, H. Catalog der Lepidopteren des paläarktischen Faunengebietes. Berlin (1901).
- Swinhoe, C. Catalogue of Eastern and Australian Lepidoptera Heterocera. 2 vols. Oxford (1895–1900).
- TURNER, A. J. Revision of the Lepidoptera of Tasmania. Papers and Proc. Roy. Soc. Tasmania, 1925, pp. 81–151 (1925); Part 2, ibid., 1928, pp. 29–65 (1928).

### SUBORDER JUGATÆ

- МЕУRICK, E. Micropterygidæ. Gen. Insectorum, fasc. 132, 9 pp. (1912).
  Micropterygidæ. Lepid. Cat., pt. 6 (1912).
- Turner, A. J. Australian Lepidoptera Homoneura. Trans. Entom. Soc. London, 1921, pp. 592–603 (1922).
- Wagner, H. and Pfitzner, R. Hepialidæ. Catal. Lepid., pt. 4 (1911).

# SUBORDER FRENATÆ COSSOIDEA

- Barnes, W. and McDunnough, J. H. Revision of the Cossidæ of North America. Contrib. Nat. Hist. Lepidoptera N. Am., 1, No. 1, Decatur, Ill. (1911).
- Dalla Torre, K. W. Cossidæ. Lepid. Catal., pt. 29 (1923).
- Dalla Torre, K. W. and Strand, E. Metarbelidæ. Lepid. Catal., pt. 28 (1923).
- Janse, A. J. T. Revision of the South African Metarbelinæ. South African Journ. Nat. Hist., 5, pp. 61–100 (1925).
- STRAND, E. Catalogus Ratardidarum. Arch. f. Naturg., Jahrg. 82A, Heft. 4, pp. 53-54 (1917).
- Turner, A. J. Observations on Cossidæ and the classification of the Lepidoptera. Trans. Entom. Soc. London, 1918, pp. 155–190 (1918).

### CASTNIOIDEA

Dalla Torre, K. W. Castniidæ. Lepid. Catal., pt. 15 (1913).

HOULEBERT, C. Revision monographique des Castniinæ. In Oberthür, Études Lepidop. Comp., fasc. 15, 736 pp. (1918).

ROTSCHILD, W. Revision of the Castniidæ. Novitat. Zool., **26**, pp. 1–27 (1919). Talbot, G. Monograph of Castniinæ. Novitat. Zool., **26**, pp. 28–35 (1919).

### **PSYCHOIDEA**

Aurivillius, C. Chrysopolomidæ. Lepid. Cat., pt. 1 (1911).

Burgeff, H. Zygænidæ, part. Lepid. Cat., pt. 33 (1926).

Dalla Torre, K. W. and Strand, E. Psychidæ. Lepid. Cat., pt. 34 (1927).

DYAR, H. G. and STRAND, E. Dalceridæ, Epipyropidæ. Lepid. Cat., pt. 16 (1913).

EECKE, R. VAN. Eucleidæ. Lepid. Cat., pt. 32 (1925).

Fletcher, T. B. Zygænidæ. Cat. Indian Ins. Govt. India, Centr. Pub. Br., Calcutta, pt. 9 (1925).

Hering, M. Revision der orientalischen Chalcosiinen. Arch. f. Naturg., Jahrg. 88A, Heft. 11, pp. 1-93 (1922).

Heylaerts, F. J. M. Monographie des Psychides paléarctiques. Ann. Soc. Entom. Belgique, **25**, pp. 29–73 (1881).

Strand, E. Catalogus Heterogynidarum. Arch. Naturg., Jahrg. 88A, Heft. 3, pp. 47–50 (1922).

Turner, A. J. Revision of Australian Limacodidæ and Zygænidæ. Proc. Linn. Soc. New South Wales, **51**, pp. 411–445 (1926).

### TORTRICOIDEA

Busck, A. Review of Tortricid subfamily Phaloniinæ. Journ. New York Ent. Soc., 14, pp. 173-182 (1906).

FERNALD, C. H. The genera of Tortricidæ. Mass. Agric. Expt. Sta. Special Bull., 69 pp. (1908).

Kennel, J. Die paläarktischen Tortriciden. Zoologica, Heft, 54, 5 parts., 742 pp. (1908–21).

MEYRICK, E. Tortricidæ. Gen. Insectorum, fasc. 149, 81 pp. (1913). Tortricidæ. Lepid. Cat., pt. 10 (1912).

### PTEROPHOROIDEA

Barnes, W. and Lindsey, A. W. The Pterophoridæ of North America. Contrib. Nat. Hist. Lepid. N. Am., Decatur, Illinois, 4, pp. 281–452 (1921).

FERNALD, C. H. Pterophoridæ of North America. Mass. Agric. Sta. Spec. Bull. 84 pp. (1898). MEYRICK, E. Pterophoridæ, Orneodidæ. Lepidop. Cat., pt. 17 (1913). Orneodidæ. Gen. Insectorum, fasc. 108, 4 pp. (1910).

Pterophoridæ. Gen. Insectorum, fasc. 100, 21 pp. (1910).

### PYRALIDOIDEA

Dalla Torre, K. W. Thyrididæ. Lepidop. Cat., pt. 20 (1914).

Dyar, H. G. North American Nymphulinæ and Scopariinæ. Journ. New York Entom. Soc., 14, pp. 77–107 (1906).

North American Chrysauginæ. Proc. Entom. Soc. Washington, 10, pp. 92–96 (1908).

North American Pyralinæ. Proc. Entom. Soc. Washington, 10, pp. 96–102 (1908).

Fernald, C. H. Crambidæ of North America. Bull. Massachusetts Agric. Coll., Amherst, Mass., 93 pp. (1896).

HAMPSON, G. F. Classification of Schoenobiidæ and Crambidæ. Proc. Zool. Soc. London, 1895, pp. 897–974 (1895).

Classification of the Thyrididæ. Proc. Zool. Soc. London, 1897, pp. 603–632 (1897).

Classification of the Chrysauginæ. Proc. Zool. Soc. London, 1897, pp. 633–692 (1897).

Classification of some subfamilies of Pyralididæ, Epipaschiinæ, Endotrichinæ and Pyralidinæ. Trans. Entom. Soc. London, 1896, pp. 451–550. Hydrocampinæ, Scopariinæ, ibid., 1897, pp. 127–240 (1897).

Revision of the Pyraustinæ. Proc. Zool. Soc. London, 1898, pp. 590–761 (1898) and 1899, pp. 172–291 (1899).

Classification of the Pyralidæ, subfamily Galleriinæ. Novitat. Zool., 24, pp. 17–58 (1917).

Classification of Pyralidæ, subfamily Hypsotropinæ. Proc. Zool. Soc. London, 1918, pp. 55–131 (1918).

Hering, E. Uebersicht der Sumatra-Pyraliden. Stettiner Entom. Zeitg., 62–64 (1901–03). (Three parts.)

Holland, W. J. and Schaus, W. The Epipaschiinæ of the Western Hemisphere. Ann. Carnegie Mus., Pittsburgh, 16, pp. 49–130 (1925).

HULST, G. D. Epipaschiinæ of North America. Entomologia Americana, 1889, pp. 41–52; 61–76 (1889).

The Phycitidæ of North America. Trans. American Entom. Soc., 17, pp. 93–228 (1890).

Shibuya J. The Japanese Crambinæ and Pyralidinæ. Journ. Fac. Agric. Sapporo Univ., 21, pp. 121–176 (1928).

Systematic Study of Formosan Pyralididæ. Journ. Fac. Agric., Sapporo Univ., **22**, pp. 1–300 (1928).

Turner A. J. Studies in Australian Tineodidæ. Proc. Roy. Soc. Victoria, 35, pp. 26-62 (1922).

A Revision of the Australian Anerastriinæ. Proc. Linn. Soc. New South Wales,  $\bf 48$ , pp. 451–461 (1923).

## URANIOIDEA

Dalla Torre, K. W. Epiplemidæ, Uraniidæ. Lepid. Cat., pt. 30 (1924). Janet, A. and Wytsman, P. Epicopeiidæ. Gen. Insectorum, fasc. 16, 5 pp. (1904).

### BOMBYCOIDEA

- Bouvier, E. L. Les Saturnioides de l'Afrique tropicale française. Faune Colon, franç. fasc. 2, pp. 449–708 (1928).
- Fletcher, T. B. Lasiocampidæ. Cat. Indian Ins. Govt. India, Centr. Pub. Br., Calcutta, pt. 7 (1925).
- JORDAN, K. On the Saturnoidian families Oxytenidæ and Cercophanidæ. Nov. Zool., 31, pp. 135–193 (1924).
- Matsumura, S. Generic revision of palæarctic Notodontidæ. Insecta. Matsum., Sapporo, 4, pp. 78–93 (1929).
- PACKARD, A. S. Monograph of the Bombycine moths of America north of Mexico. Mem. Nat. Acad. Sci., Washington, 7 (1895).

Monograph of the Bombycine moths of America. Family Ceratocampidæ, subfamily Ceratocampinæ. Pt. 2, Mem. Nat. Acad. Sci., Washington, 9 (1905).

Monograph of the Bombycine moths of North America. Pt. 3, Mem. Nat. Acad. Sci. Washington, 12, 516 pp. (1914). (Edited by T. D. A. Cockerell).

- Prout, L. B. Provisional arrangement of the Dioptidæ. Nov. Zool., 25, pp. 395–429 (1918).
- Schaus, W. Revision of American Notodontidæ. Trans. Entom. Soc. London, 1901, pp. 257–344 (1901).
- STRAND, E. Brahmæidæ. Lepidop. Cat., pt. 16 (1913).
- Turner, A. J. Revision of Australian Saturniidæ, Bombycidæ, Eupterotidæ, Notodontidæ. Proc. Linn. Soc. New South Wales, 47, pp. 348–390 (1922). Revision of Australian Lasiocampidæ. Proc. Linn. Soc. New South Wales, 49, pp. 397–428 (1924).

### GEOMETROIDEA

- Culot, J. Geometridæ of Europe. Vol. 3, 269 pp., Geneva (1919).
- Forbes, W. T. M. The genera of Hydriomeninæ of the United States. Journ. New York Entom. Soc., 25, pp. 44-67 (1917).
- Goldfinch, G. M. Revision of Australian Geometridæ. Proc. Linn. Soc. New South Wales, **54**, pp. 378–407 (1929).
- Hulst, G. D. Classification of Geometrina of North America. Trans. American Entom. Soc., 23, pp. 245–386 (1898).
- McDunnough, J. H. Studies on North American Cleorini. Bull. Canada Dept. Agric. Entom., No. 18, 64 pp. (1920).

MEYRICK, E. Classification of the Geometridæ of Europe. Trans. Entom. Soc. London, 1892, pp. 53–140 (1892).

Packard, A. S. Monograph of the Geometrid moths or Phalænidæ of the United States. Rept. U. S. Geol. Survey, 10 (1876).

Prout, L. B. Geometridæ, Brephinæ. Gen. Insectorum, fasc. 103, 16 pp. (1910).

Œnochromatidæ. Gen. Insectorum, fasc. 104, 120 pp. (1910).

Hemitheidæ. Gen. Insectorum, fasc. 129, 274 pp. (1912).

Hemitheidæ. Lepidop. Cat., pt. 14 (1913).

Turner, A. J. Revision of Australian Geometridæ. Proc. Linn. Soc. New South Wales, 44, pp. 258-310; 383-413 (1919).

Australian Larentiidæ. Proc. Roy. Soc. South Australia, 46, pp. 225–294 (1922).

Revision of Australian Œnochromatidæ. Proc. Linn. Soc. New South Wales, **54**, pp. 463–504 (1929); **55**, pp. 1–40 (1930); **55**, pp. 199–220 (1930).

### SPHINGOIDEA

JORDAN, K. Monograph of the Ludiinæ. Nov. Zool., 29, pp. 249-326 (1922).

ROTHSCHILD, W. and JORDAN, K. Sphingidæ. Gen. Insectorum, fasc. 57, 157 pp. (1907).

Revision of the Lepidopterous family Sphingidæ. 2 vols., Suppl. to Nov. Zool., 9, London (1903).

SMITH, J. B. Monograph of Sphingidæ of North America. Trans. American Entom. Soc., 15, pp. 49–242 (1888).

WAGNER, H. Sphingidæ, Subfamilies Philampelinæ, Acherontiinæ, Chœrocampinæ, pts. 12 (1913), 21 (1915) and 23 (1919).

### TINEOIDEA

Beutenmuller, W. The Sesiidæ of North America. Mem. American Mus. Nat. Hist., 1 (1901).

Braun, A. F. Revision of North American species of Lithocolletis. Trans. American Entom. Soc., 34, pp. 269–357 (1908).

The Nepticulidæ of North America. Trans. American Entom. Soc., 43, pp. 155-209 (1917).

Brèthes, J. Lepidópteros argentinos productores de agallas. An. Soc. Sci., Argentina, 82, pp. 113–140 (1916). (Cecidosidæ and Ridiaschinidæ).

Busck, A. Revision of American Gelechiidæ. Proc. U. S. Nat. Mus., 25, pp. 767–938 (1903).

Synopsis of North American genera of Ægeriidæ. Proc. Entom. Soc. Washington, 11, pp. 115–118 (1909).

Generic Revision of American Œcophoridæ. Proc. U. S. Nat. Mus., **35**, pp. 187–207 (1909).

CLEMENS, B. Tineina (Tineoidea) of North America. 282 pp., London (1872).

Dalla Torre, K. W. and Strand, E. Ægeriidæ, part. Lepidop. Cat., pt. 31 (1925)

Dietz, W. G. Revision of Amydriinæ and Tineinæ of North America. Trans. American Entom. Soc., **31**, pp. 1–96 (1905).

Revision of Blastobasidæ of North America. Trans. American Entom. Soc., **36**, pp. 1–72 (1910).

Dyar, H. G. Revision of North American Prodoxidæ. Journ. New York Entom. Soc., 11, pp. 103–105 (1903).

DYAR, H. G. and STRAND, E. Epipyropidæ. Lepidop. Cat., pt. 16 (1913).

ELY, C. R. Revision of North American Gracilariidæ. Proc. Entom. Soc. Washington, 19, pp. 29-77 (1917).

Fletcher, T. B. Cosmopterygidæ. Cat. Indian Ins. Govt. India, Centr. Pub. Br., Calcutta, pt. 16 (1929).

Yponomeutidæ. Cat. Indian Ins. Govt. India, Centr. Pub. Br., Calcutta, pt. 17 (1929).

Meyrick, E. Gracilariidæ. Gen. Insectorum, fasc. 128, 36 pp. (1912).

Adelidæ. Gen. Insectorum, fasc. 133, 12 pp. (1912).

Gracilariidæ and Adelidæ. Lepidop. Cat., pt. 6 (1912).

Carposinidæ, Heliodinidæ, Glyphipterygidæ. Lepidop. Cat., pt. 13 (1913).

Yponomeutidæ, Plutellidæ, and Amphitheridæ. Lepidop. Cat., pt. 19 (1914).

Exotic Microlepidoptera. 4 vols., London (1912-1929).

Heliodinidæ. Gen. Insectorum, fasc. 165, 29 pp. (1914).

Revision of New Zealand Tineina. Trans. New Zealand Inst., 47, pp. 205–244 (1915).

STAINTON, H. The Natural History of the Tineina. Vols. 1–13, London (1855–1873).

Turner, A. J. Studies in Australian Ægeriidæ. Proc. Roy. Soc. Victoria, 35, pp. 26-62 (1922).

Wagner, H. Ægeriidæ, part. Lepidop. Cat., pt. 18 (1914).

Walsingham, T. de G. Heterocera (Tineoidea), in Biologia centrali-Americana, 4, 168 pp. (1911–12).

### NOCTUOIDEA

Barnes, W. and McDunnough, J. Illustrations of North American species of the Genus Catocala. Mem. American Mus. Nat. Hist., 3, 47 pp. (1918).

Beutenmüller, W. Descriptive Catalogue of Noctuidæ within 50 miles of New York City. Bull. American Mus. Nat. Hist., 14, pp. 229-312; 16, pp. 413-458 (1902).

Culot, J. Noctuidæ of Europe. Vol. 1 and 2 (1910-12).

Dalla Torre, K. W. Cymatophoridæ. Lepidop. Cat., pt. 25 (1921).

FLETCHER, T. B. Syntomidæ. Cat. Indian. Ins. Govt. India Centr. Pub. Br., Calcutta, pt. 8 (1925). Forbes, W. T. M. Genera of Noctuide of Northeastern North America. Journ. New York Entom. Soc., 22, pp. 1–33 (1914).

Grote, A. R. Die Apateliden. Mitt. Römer. Mus. Hildesheim, No. 3, 18 pp. (1896).

Houlebert, C. Revision monographique de la famille des Cymatophoridæ. Études Lepidop. Comp., **18**, pt. 2, pp. 23–252 (1921).

Hulstaert, P. G. Anthelidæ. Gen. Insectorum, fasc. 191, 13 pp. (1929).

Köhler, P. Agaristidæ de la república Argentina. Rev. Soc. Entom. Argentina, **2**, pp. 235–245 (1929).

Matsumura, S. Catalogue of Japanese Arctiidæ. Ins. Matsumura., Sapporo, Japan, 5, pp. 58–94 (1930).

McDunnouch, J. H. Generic revision of North American Agrotid moths. Bull. Canada Dept. Mines, No. 55, 78 pp. (1929).

MEYRICK, E. Monograph of the Noctuidæ of New Zealand. Trans. New Zealand Inst., 19 (1886); 20 (1887).

Pagenstecker, A. Callidulidæ. Das Tierreich, Lief. 17, 27 pp. (1901). Callidulidæ. Lepidop. Cat., pt. 2 (1911).

SMITH, J. B. Revision of the Deltoid moths. Bull. U. S. Nat. Mus., No. 48, 120 pp. (1895).

STRAND, E. Lithosiidæ. Lepidop. Cat., pt. 26 (1922).

Arctiidæ. Lepidop. Cat., pt. 22 (1919).

Agaristidæ. Lepidop. Cat., pt. 5 (1912).

Nolidæ. Lepidop. Cat., pt. 24 (1920).

Swinhoe, C. A Revision of the Genera of the Family Liparidæ. Ann. Mag. Nat. Hist. (9), **10**, pp. 449–484; **11**, pp. 47–97, 289–304, 400–442 (1922–23).

Turner, A. J. A New Family of Lepidoptera (Anthelidæ). Trans. Entom. Soc. London, 1919, p. 415(1919).

Revision of the Australian Noctuidæ. Trans. Roy. Soc. South Australia, 44, pp. 120–189 (1920).

Revision of Anthelidæ. Proc. Linn. Soc. New South Wales, 46, pp. 164–191 (1921).

Revision of Australian Drepanidæ. Proc. Linn. Soc. New South Wales, **51**, pp. 411–445 (1926).

ZERNY, H. Euchromiidæ. Lepidop. Cat., pt. 7 (1912).

## SUBORDER RHOPALOCERA, GENERAL

Aurivillius, C. Rhopalocera Æthiopica. 561 pp. Stockholm (1898).

Barnes, W. and Benjamin, F. H. Check-list of the diurnal Lepidoptera of Boreal America. Bull. Southern California Acad. Sci., **25**, pp. 3–27; pp. 88–98 (1926).

BINGHAM, C. T. Rhopalocera. Fauna of British India, 1 and 2, London (1905–07).

Buckler, W. (edited by G. T. Porritt). Larvæ of British Butterflies and Moths 9 vols., London (1886–1901).

- DISTANT, W. L. Rhopalocera malayana, 481 pp., London (1882–86).
- EDWARDS, W. H. Butterflies of North America, Boston, and New York. 3 vols. (1880–84–97).
- Evans, W. H. Identification of Indian Butterflies. Journ. Bombay Nat. Hist. Soc., **29**, pp. 230–260; 519–537; 780–797 (1923).
- GODMAN, F. D. and SALVIN, O. Rhopalocera, In Biologia centrali-Americana, 3 vols. (1879–1901).
- Holland, W. J. The Butterfly Book (nearctic). Doubleday and McClure Co., New York (1898). Second edition (1931)
- KAYE, W. J. The Butterflies of Jamaica. Trans. Entom. Soc. London, 1925, pp. 455-504 (1925).
- Kirby, W. F. Synonymical catalogue of Diurnal Lepidoptera. John van Voorst, London (1871–77). (Various parts.)
- Lang, H. C. Rhopalocera Europæ. 2 vols., London (1884).
- Mabille, P. Lepidoptères Rhopalocères de Madagascar. Grandidier, Hist. Nat. Madagascar, 18, 364 pp. (1885–88).
- MARSHALL, G. F. L. and NICEVILLE, L. The Butterflies of India. 3 vols., Central Press, Calcutta (1882–1890).
- MOORE, F. Lepidoptera indica (Rhopalocera). 8 vols., London (1890-1911).
- Moulton, J. C. Butterflies of Borneo. Sarawak Mus. Journ., 2, pp. 197–266 (1915).
- OBERTHÜR, C. and HOULEBERT, C. Rhopalocères. Faune Entom. Amoricaine 3, pp. 47–260 (1922).
- Schatz, E. and Röber, J. Die Familien und Gattungen der Tagfalter. 284 pp. Furth (1885–92).
- Scudder, S. H. Butterflies of Eastern United States and Canada. 3 vols., Cambridge (1889).
- Trimen, R. and Bowker, H. South African Butterflies. 3 vols., London (1887–1889).
- Tutt, J. W. British Butterflies. 10 vols., London (incomplete) (1899-1909).
- WATERHOUSE, G. A. and LYELL, G. The Butterflies of Australia, 239 pp. (1914).
- WRIGHT, W. G. Butterflies of the West Coast of the United States. San Bernardino, California, 257 pp. (1906).

### HESPERIOIDEA

- Barnes, W. and McDunnough, J. H. Revision of the Megathymidæ. Contrib. Nat. Hist. Lepidoptera N. Am., 1, No. 3, Decatur, Ill. (1912).
- Dyar, H. G. Review of Hesperiidæ of the United States. Journ. New York Entom. Soc., 13, pp. 111-141 (1905).
- ELWES, H. J. and EDWARDS, J. Revision of oriental Hesperiidæ. Trans. Zool. Soc. London, 14, pp. 101–324 (1897).
- HOLLAND, W. J. Revision and catalogue of African Hesperiidæ. Proc. Zool. Soc. London, 1896, pp. 2–107 (1896).

- LINDSEY, A. W. The Hesperioidea of America North of Mexico. Univ. of Iowa Studies, 9, No. 4, Iowa City (1921).
- Mabille, P. Hesperiidæ. Lepidop. Cat., pt. 9 (1904). Hesperiidæ. Gen. Insectorum, fasc. 17, 210 pp. (1904).
- MEYRICK, E. and LOWER, O. B. Revision of Australian Hesperiidæ. Trans. Roy. Soc. South Australia, **26**, pp. 38–129 (1902) and **31**, pp. 192–208 (1907).
- Warren, B. C. S. Revision of palæarctic Hesperiinæ. Trans. Entom. Soc. London, 74, pp. 1–170 (1926).
- WATSON, E. Y. Classification of Hesperiidæ. Proc. Zool. Soc. London, 1893, pp. 3–132 (1893).
- WILLIAMS, R. C. Studies in neotropical Hesperoidea, II. Trans. American Entom. Soc., **53**, pp. 261–292 (1927).

### PAPILIONOIDEA

- Bethune-Baker, G. T. Revision of the Amblypodia group of Lycenide. Trans. London Zool. Soc., 17, pp. 1–164 (1903).
- Bryk, F. Papilionidæ, part. Lepidop. Cat., pt. 27 (1923); pt. 35 (1928). Parnasiidæ. Lepidop. Cat., pt. 27 (1923).
- DRUCE, H. H. Monograph of Bornean Lycænidæ. Proc. Zool. Soc. London, 1895, pp. 556–627 (1895) and 1896, pp. 650–683 (1896).
- ELTRINGHAM, H. and JORDAN, K. Acræidæ. Lepidop. Cat., pt. 11 (1913).
- JORDAN, K. and ELTRINGHAM, H. Nymphalidæ, Acræinæ. Gen. Insectorum, fasc. 169, 81 pp. (1916).
- Neustetter, H. Eueididæ (Heliconiidæ). Lepidop. Cat., pt. 36 (1929).
- PAGENSTECKER, A. Libytheidæ. Das Tierreich, Lief. 14, 18 pp. (1901). Libytheidæ. Gen. Insectorum, fasc. 5, 4 pp. (1902).

Libytheidæ. Lepidop. Cat., pt. 3 (1911).

- RIPPON, R. H. F. Monograph of Ornithoptera. 2 vols., London (18 98–1907).
  Papilionidæ, Papilioninæ. Gen. Insectorum, fasc. 6, 15 pp. (1902).
- ROTHSCHILD, W. A Revision of the Papilios of the eastern Hemisphere. Novitat. Zool., 2, pp. 167–463 (1895).
- ROTHSCHILD, W. and JORDAN, K. A Revision of the American Papilios. Novitat. Zool., 13, pp. 411-752 (1906).
- Seitz, A. Das System der Schmetterlinge; III. Die Danaiden. Entom. Rundshau Jahrg. 44, p. 32 and various parts following (1927).
- STICHEL, H. Brassolidæ. Das Tierreich, Lief. 25, 258 pp. (1908).

Nymphalidæ, Dioninæ. Gen. Insectorum, fasc. 63, 38 pp. (1908).

Parnasiidæ. Gen. Insectorum, fasc. 58, 60 pp. (1907).

Riodinidæ. Gen. Insectorum, fasc. 112, 452 pp. (1911).

Papilionidæ, Zerynthiinæ. Gen. Insectorum, fasc. 59, 27 pp. (1907).

Brassolidæ. Gen. Insectorum, fasc. 20, 48 pp. (1904).

Nymphalidæ, Discophorinæ. Gen. Insectorum, fasc. 31, 16 pp. (1905). Nymphalidæ, Hyanthinæ. Gen. Insectorum, fasc. 39, 7 pp. (1906).

Morphoidæ. Gen. Insectorum, fasc. 36, 67 pp. (1906).

Eueididæ. Gen. Insectorum, fasc. 37, 74 pp. (1906).

Amathusiidæ, Morphoidæ. Das Tierreich, Lief. 34, 263 pp. (1912).

Beiträge zur Kenntnis der Riodinidenfauna Südamerikas. Deuts. Entom. Zeits., pp. 81–101 (1926).

Vorarbeiten zu einer Revision der Riodinidæ. Deuts. Entom. Zeits., 1926, pp. 385–396 (1927).

Nemeobiinæ. Das Tierreich, Lief. 51, 329 pp. (1928).

STICHEL, H. and EIFFARTH, H. Heliconiidæ (Eueididæ). Das Tierreich, Lief. 22, 305 pp. (1905).

Wytsman, P. Papilionidæ, Leptocircinæ. Gen. Insectorum, fasc. 4, 3 pp. (1902).

## ORDER DÍPTERA

# (ANTLIÀTA, HALTERÀTA, HALTERÍPTERA, HAUSTELLÀTA)

Minute to moderate-sized, rarely large (over one inch) insects, usually with good powers of flight: head usually vertical, freely movable; antennæ variable, comprising either many similar joints or frequently only three, the last joint sometimes annulated, sometimes provided with a sensory style or arista as a phyletic adaptation of the original terminal joints; mouthparts suctorial, incapable of mastication, usually constructed for lapping, sometimes for piercing; both prothorax and metathorax small and fused with the prominent mesothorax: only the mesothoracic pair of wings developed, the veins and crossveins not numerous, hind wings replaced by small knobbed structures (halteres), rarely wings vestigial or even absent; legs usually alike, the tarsi regularly five-jointed. Metamorphosis complete, the larvæ wholly unlike the adults; larvæ almost always legless maggots or grubs, never with true jointed legs, frequently with indistinct head and retracted mouthparts; pupæ with the appendages more or less adherent, the body either free or entirely encased in a seed-like capsule (puparium) formed of the indurated last larval moult. Food habits highly variable. Flies, Mosquitoes, Gnats, Midges.

## Adults

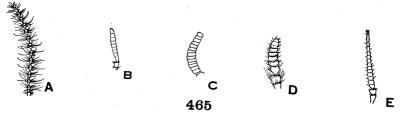


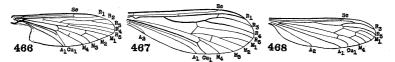
Fig. 465. Nematocerous Antennæ

- A, Perrisia (Verrall) Cecidomyiidæ; B, Simulium (Lugger) Simuliidæ; C, Ceroplatus (Johannsen) Ceroplatidæ; D, Bibio (Verrall) Bibionidæ; E, Anisopus (Verrall) Anisopodidæ.
  - Antennæ shorter, usually three-jointed, the third joint occasionally complex, with more or less distinct annulations (Fig. 518b-e), or bearing a differentiated style (Fig. 518 f, g) or arista (Fig. 556), in Rhachicerinæ the third joint divided into about 25 segments (Fig. 518a); anal cell (Cu) distally narrowed or closed, sometimes retracted and very short, or even absent, discal cell usually present, "second" vein (R<sub>2+3</sub>) never furcate; palpi short, one- or two-jointed, projecting forward; pleural suture between root of wing and middle coxæ twice bent at sharp angles. (Suborder BRACHÝCERA)......32
  - 3. Mesonotum with a more or less distinct V-shaped transverse suture beginning on each side in front of root of wings, the pointed middle part close to the scutellum (Fig. 472); post-pronotum well developed; female with a conical, generally protruding, chitinized ovipositor; male genitalia usually enlarged;

males dichoptic, eyes rounded, not excised at antennæ; legs very long and slender, easily breaking from the body at the trochanters; costa encompassing wing, nine or more veins terminating in wing-margin, subcosta long, ending beyond middle of wing. (Superfamily TIPULÒIDEA)......4

4. Radius with five branches all ending in the wing-margin, subcosta furcate at its extremity, Sc<sub>2</sub> appearing like a crossvein ending in R<sub>1</sub>; first basal cell closed at middle of wing, second basal cell distinctly shorter; one anal vein. (**Tanydèrus**, neotrop.; **Peringueyomyìna**, ethiop.; **Protóplasa**, nearc. (Fig. 466)).

TANYDÉRIDÆ



Figs. 466-468. Tanyderidæ, Trichoceratidæ, Cylindrotomidæ

- 466. Protoplasa, wing (Alexander) Tanyderidæ.
- 467. Paracladura, wing (Edwards) Trichoceratidæ.
- 468. Cylindrotoma, wing (Alexander) Cylindrotomidæ.

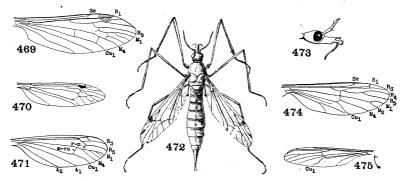
Radius with only three or four branches ending in wing-margin; basal cells long, nearly always ending distinctly beyond middle of wing and coextensive or nearly so, or the second longer than the first; an accessory or marginal cell commonly present in front of the end of the first basal cell, formed by the erect or reflexed shortened  $R_2$  or  $R_{2+3}$  located near the end of  $R_1, \ldots, 5$ 

R<sub>4</sub> and R<sub>5</sub> stalked together (Fig. 474); one distinct anal vein reaching hind margin; no discal cell formed between the branches of media; a longitudinal fold in the wing-membrane crossing anterior crossvein; ocelli absent; mesonotal suture not deep; empodium minute, pulvilli present. (LIRIOPÈIDAE).

PTYCHOPTÉRIDÆ

a. Antennæ 16-jointed; fourth vein forked as M<sub>1</sub> and M<sub>2</sub>; legs not banded. (**Ptychóptera** (=Liriòpe) (Fig. 474), widespr.).

PTYCHOPTERINÆ

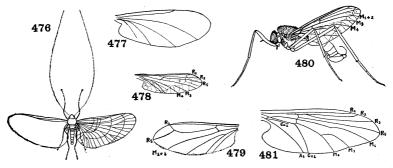


Figs. 469-475. Tipulidæ, Limoniidæ, Ptychopteridæ

- 469. Limnophila, wing (Alexander) Limoniidæ.
- 470. Tipula, wing. Tipulidæ.
- 471. Dolichopeza, wing (Alexander) Limoniidæ.
- 472. Tipula, female. Tipulidæ.
- 473. Tipula, profile of head, showing palpus (Alexander) Tipulidæ.
- 474. Ptychoptera, wing (Alexander) Ptychopteridæ.
- 475. Bittacomorpha, wing. Ptychopteridæ.
  - 6. Two or three ocelli present; last anal vein typically short, abruptly curving into the anal angle; antennal segmentation indistinct except at base; one pair of male claspers. (Trichôcera (= Petaurista) Winter-gnats, Diazósma, Nothotrichôcera, Paracladùra (Fig. 476), mostly holarc.; Íschnothrix, Cape Horn). (PETAURÍSTIDÆ)......TRICHOCERÁTIDÆ
  - 7. Last joint of maxillary palpi lash-like, much longer than the three preceding together; Sc ending in R<sub>1</sub> by an abrupt curvature of the tip but almost never also in the costa (Fig. 470); antennæ usually with 12 or 13 joints, rarely more; nasus usually distinct. Many species, widespread, but principally holarctic.

TIPÙLIDÆ

a.,	Vein R <sub>2</sub> absent, or else the second anal vein not more than one- third as long as the first anal vein; legs excessively long and
	slender. (Dolichopèza, Brachyprémna, Megistócera, Tany-
	prémna) DOLICHOPEZÌNÆ
	Vein R <sub>2</sub> present; second anal vein one-half as long as first anal
	vein; legs relatively shorter and strongerb
b.	Antennæ verticillate, i.e. with whorls of hairs, flagellum of male
	not pectinate. (Típula (Fig. 472), Holorùsia, Longùrio,
	Nephrótoma (= Pachyrrhina))
	Antennæ not verticillate; flagellum of male antennæ pectinate. (Ctenóphora, Xiphosùra) CTENOPHORÌNÆ
	Last joint of palpi shorter or not much longer than the two preced-
	ing together; Sc ending in costa and usually furcate at tip, the
	lower branch connecting as Sc <sub>2</sub> with R <sub>1</sub> (Fig. 468); antennæ
	6- to 16-jointed, rarely more, usually with 14 to 16 joints8
8.	Tibiæ spurred; two branches of radius reaching margin, due to
	the apparent fusion of $R_1$ with $R_{2+3}$ , rarely $R_2$ and $R_3$ separate,
	in which case three branches of radius reach margin, R <sub>s</sub> long,
	arising near middle of wing; larvæ eruciform. Principally hol-
	arctic. (Cylindrótoma (Fig. 468), Liógma, Triógma, Phal-
	acrócera; Stibadócera, oriental) CYLINDROT ÓMIDÆ
	Four branches of radius reaching margin, if but three branches
	reach margin R <sub>1</sub> ends in costa and R <sub>8</sub> usually arises beyond
	middle of wing. A large, cosmopolitan family. (LIMNOBIIDÆ).
	LIMONÌIDÆ
a.	Tibiæ spurred at tipb
	Tibiæ not spurred at tip
b.	Antennæ six- to ten-jointed. (Hexátoma, mainly European;
	Eriócera, Penthóptera)
	Antennæ with more than ten joints
c.	Sc <sub>2</sub> located beyond origin of R <sub>s</sub> (if before, <i>Ula</i> , the wings are
	pubescent). (Limnóphila (Fig. 469), Adelphomỳia, Epi- phrágma, Ūla, Ūlomórpha, widespr.) . LIMNOPHILÌNÆ
	Sc <sub>2</sub> located before origin of R <sub>s</sub> ; wings glabrous. (Pedicia, Di-
	cranota, Rhaphidolabis, Tricyphona, mainly nearc.).
	PEDICIÌNÆ
d.	Radius with four branches reaching margin. (Erióptera, Cladura,
	Gnophomyia, Helòbia, Molóphilus, Trimicra).
	ERIOPTERÎNÆ
	Radius with three branches reaching margin
e.	Antennæ 14-jointed; claws with teeth on lower side. (Limònia,
	1803 (=Limnòbia 1818), Dicranomỳia, Discóbola, Gera- nomỳia, Rhipídia) LIMONIÌNÆ
	mumvia. Amuliua/



Figs. 476-481. Blepharoceratidæ, Deuterophlebiidæ

- 476. **Deuterophlebia** (Edwards) Deuterophlebiidæ.
- 477. **Hammatorrhina**, wing (Bezzi) Blepharoceratidæ.
- 478. Blepharocera, wing (Comstock) Blepharoceratidæ.
- 479. Paltostoma, wing (Williston) Blepharoceratidæ.
- 480. Bibiocephala (Cole) Blepharoceratidæ.
- 481. Edwardsina, wing (Alexander) Blepharoceratidæ.
- 10. Wings large, densely covered with fine hairs, true veins almost absent but an elaborate fan-like development of secondary folds present; antennæ excessively long, six-jointed; ocelli and mouthparts absent. (**Deuterophlèbia**, India (Fig. 476)).

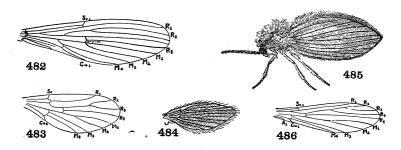
### DEUTEROPHLEBÌIDÆ

Secondary venation forming a delicate network like spider webbing, in addition to the primary veins; mesonotum with a V-shaped suture; ocelli and mouthparts present; eyes usually divided horizontally into two parts by an unfaceted stripe; hind coxæ broadly attached to thorax. Widespread, mainly holarctic and neotropical. (ASTHÉNIDÆ, LIPONEÙRIDÆ).

BLEPHAROCERÁTIDÆ

a.	Wings with M <sub>3</sub> arising from middle of upper branch of cubital fork (M <sub>4</sub> ) (Fig. 481), m-cu crossvein present and almost in transverse alignment with r-m crossvein and the short angulate bases of $R_{4+5}$
	and R <sub>s</sub> , a long spur from R <sub>s</sub> projecting basal to the angulation, radius four-branched; front trochanters scarcely half as long as coxæ. (Edwardsina, neantarc. (Fig. 481)). EDWARDSINÆ
	Wing with $M_3$ free or absent, when present disconnected from the other veins; if m-cu crossvein is present it is not in alignment with the angulations of the branches of the radius, $R_s$ without basal spur; front trochanters nearly as long as $coxeb$
b.	
	the upper $(M_{1+2})$ ; second and third veins subequal in length, usually arising separately from the cell; eight veins reaching
	margin. (Blepharócera (Fig. 478), Bibiocéphala (Fig. 480), Liponeura, Philòrus, holarc.) BLEPHAROCERATÌNÆ
	Fourth vein $(M_{1+3})$ simple; $R_2$ short or absent
c.	Labial palpi small, usually oval and pubescent, much shorter than
	basal part of labium, if somewhat longer the palpi are rigid
	and the whole labium is reduced; third vein forked, <i>i.e.</i> the second vein $(R_{2+3})$ short, arising from the third vein $(R_{4+5})$
	near its tip; seven veins reaching margin
	Labial palpi very long, slender, bare, usually curled outwardly;
	basal part of labium also long; maxillary palpi one-jointed; no macrotrichia on $R_1$ ; $R_s$ forked near tip, or simple, or absent;
	only five or six veins reaching margin. (Apistomyia, widespr.;
	Hammatorrhina (Fig. 477), Ind.; Neocurùpira, Peritheàtes,
.1	Austr.)
a.	Hind tibiæ spurred; claws of both sexes similar; female with strong mandibles. (Paltóstoma (Fig. 479), Curùpira, Kelloggìna, Limonícola, neotrop.) PALTOSTOMATÎNÆ
	Hind tibiæ without spurs; male holoptic, claws wanting; female
	dichoptic, claws dentate, mouthparts atrophied. (Hapálothrix,
	Eur.) ĤAPALOTRICHÎNÆ
11.	Costa continuing around the wing-margin, although often weaker
	along hind margin
12.	Costa disappearing beyond tip of wing
14.	At least nine veins reaching wing-margin
	holoptic
13.	Wing-veins, including hind margin, very hairy or scaly (Figs.
	484, 490); body and legs hairy or scaly; ocelli absent14
	Veins not fringed with flat scales; body and legs not scaly; sub-
	costa ending in costa at or beyond middle of wing; dichoptic; legs long and slender.

14. Wings short and broadly ovate or pointed (Fig. 484), held sloping roof-like against the body when at rest, no crossveins except sometimes near base, Sc very short, weak, ending free, radius usually five-branched; tibiæ without apical spurs; second antennal joint not enlarged; small, apparently robust species with densely hairy body, legs and wings; widespread, mostly in warm or temperate regions. Moth-flies.... PSYCHÓDIDÆ



Figs. 482-486. Psychodidæ

- 482. Bruchomyia, wing (Alexander) Psychodidæ.
- 483. Sycorax, wing (Eaton) Psychodidæ.
- 484. Pericoma, wing. Psychodidæ.
- 485. Psychoda (Cole) Psychodidæ.
- 486. Phlebotomus, wing (Alexander) Psychodidæ.
  - a. Radial sector with four branches (Figs. 482, 486).....b Radial sector with three branches, Sc short and apically erect. (Trichomyia, Sycorax (Fig. 483)).....TRICHOMYIINÆ
  - b. Distal section of Cu<sub>1</sub> elongate, extending generally parallel to M<sub>4</sub>, cell M<sub>4</sub> about equal to cell M<sub>3</sub> along the wing-margin, Sc reduced. (Psychòda (Fig. 485), Maruìna, Perícoma (Fig. 484), Telmatóscopus, Termitadélphus). PSYCHODÌNÆ
  - - Radial sector dichotomously four-branched; mouthparts not formed for sucking blood. (Bruchomyia (Fig. 482), Nemopálpus, neotrop.). (NEMOPALPINÆ).

BRUCHOMYIÌNÆ

- Wings longer and narrow (Fig. 490), not held sloping against the sides of the body, wing-margin and veins scaly, Sc ending in costa beyond middle of wing, radius four-branched; antennæ of male usually feathered with long hairs; second antennal joint enlarged: slender species, usually with long, moderately hairy or scaly legs. In all regions, many species. CULÎCIDÆ
- - Proboscis much longer than head, firm, of female adapted for piercing; wings always fully scaled; mesosternum ridged; sternopleura not divided by transverse suture (except Uranotæniini, Fig. 487); lateral sclerite of metasternum forming a triangular piece between the bases of middle and hind coxe.....b
- b. Palpi of female more than one-third as long as proboscis; abdomen sometimes without scales; scutellum crescent-shaped, with marginal bristles evenly distributed: larva without respiratory siphon, resting horizontally at surface of water; eggs provided with lateral floats. (Anópheles (Malaria mosquitoes)).

### ANOPHELÎNÆ

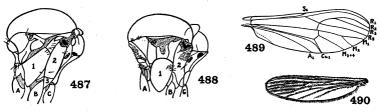
- c. Scutellum evenly rounded; clypeus much broader than long; calypteres not ciliated; bright-scaled, day-flying, not blood-sucking. (Megarhinus) . . . . . . . . . . . MEGARHINÎNÆ
- Scutellum trilobed, with marginal bristles only on the lobes....d
  d. Base of hind coxa in line with upper margin of lateral metasternal sclerite which is a small triangular piece located between bases of middle and hind coxæ: day-fliers. (Sabèthes, Goeldia, Joblotia, Limàtus, Menólepis, Wyeomyia). SABETHÎNÆ
  - Base of hind coxa distinctly below upper margin of lateral metasternal sclerite; body scales usually sparse and rarely with metallic colors: disease-bearing or obnoxious mosquitoes. . . . . e
- e. Anal vein extending well beyond fork of cubitus; wings villose; upper calypter usually ciliated. (Cùlex (C. quinquefasciàtus, Filaria mosquito), Aèdes (A. ægypti (cálopus) (=Stegomyia fasciàta), Yellow-fever and Dengue-fever mosquito), Lùtzia, Ochlerótatus, Orthopodomyia, Psoróphora, Tæniorhýnchus (=Mansonia), Theobáldia (=Culisèta)).

**CULICÌNÆ** 

Anal vein ending opposite or before fork of cubitus; wings not villose; calypteres not ciliate. (**Uranotænia** (Fig. 487)).

### URANOTÆNIÌNÆ

15. Radius with four nearly parallel curving branches (Fig. 489), the second vein (R<sub>3+4</sub>) strongly arched, R₅ ending beyond apex of wing; basal cells reaching distinctly beyond middle of wing, coextensive; ocelli absent; joints of flagellum of antennæ indistinctly separated. Few species, widespread, but mostly holarctic. (Díxa (Fig. 489); Microdíxa, Eur.; Neodíxa, N. Zeal.) . . . . . . . . . . . . DÍXIDÆ



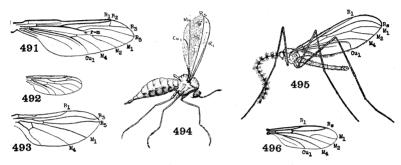
Figs. 487-490. Culicidæ, Dixidæ

- 487. Uranotænia, side view of thorax (Dyar and Shannon) Culicidæ. 1, sternopleura; 2, mesepimeron; 3, merite of middle coxa; A, front coxa; B, middle coxa; C, hind coxa.
- 488. **Eucorethra**, side view of thorax (Dyar and Shannon) Culicidæ. Explanation of parts same as for Fig. 487.
- 489. Dixa, wing. Dixidæ.
- 490. Culex, wing. Culicidæ.

Radius with three to five branches (Fig. 492), the second vein ending before apex of wing; basal cells usually not extending beyond middle of wing, the second basal cell always shorter than first; ocelli well developed; antennæ 8- to 18-jointed, the joints distinctly separated. Rare. (Hesperinus (Fig. 492); Cramptonomyia, western nearc.)..... HESPERÍNIDÆ

16. Antennæ composed apparently of two thick basal joints and a terminal nine- or ten-jointed arista; wings with seven longitudinal veins (Fig. 493), media unbranched, both basal cells closed; both sexes holoptic, ocelli absent. Small, rare, woodland species, occurring in Europe, North America and the Canary Islands. (Thaumàlea (=Orphnéphila) (Fig. 493), Androprosòpa). (ORPHNEPHÍLIDÆ). THAUMALÈIDÆ

- Antennæ long, composed of 10 to 36 cylindrical or bead-like joints; wings with greatly reduced venation (Figs. 494–496). (ITO-NÎDIDÆ) ...... CECIDOMYÎIDÆ
- a. Wings broad, with three or at most six longitudinal veins, sometimes media and cubitus branches stalked, crossveins apparently wanting; abdomen not swollen; eyes round or reniform, sometimes with confluent projections above antennæ: delicate, often minute species. Gall gnats; widespread, principally belarctic



Figs. 491–496. Pachyneuridæ, Hesperinidæ, Thaumaleidæ, Cecidomyiidæ

- 491. Axymyia, wing (Edwards) Pachyneuridæ.
- 492. Hesperinus, wing (Johannsen) Hesperinidæ.
- 493. Thaumalea, wing (Williston) Thaumaleidæ.
- 494. Mayetiola. Cecidomyiidæ.
- 495. Hormosomyia (Cole) Cecidomyiidæ.
- 496. Lestremia, wing (Kieffer) Cecidomyiidæ.

Wings much atrophied (Fig. 670) crumpled, with two longitudinal and two crossveins; first five segments of abdomen enormously swollen, the apical four segments small and slender, forming a post-abdomen; eyes confluent above, separated below antennæ: adults found exclusively in nests of termites. (**Termitomástus** (Fig. 670), South America). (See couplet 147).

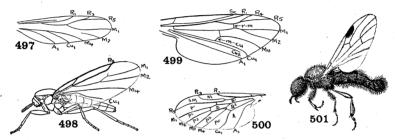
## TERMITOMASTÎNÆ

b. Wing-membrane peculiarly pubescent, the hairs (macrotrichia) directed toward base of wing; tarsi five-jointed...............
 Wing-membrane with simple microscopic pubescence; metatarsus longer than following joint, or the tarsi with less than five joints; ocelli absent; media wanting or represented by a fold.

BIBIONÎNÆ

	gárces) HETEROPEZĨNÆ
	Media $(M_{1+2})$ forked or simple, radial sector present but some-
С.	times crowded close to costa; ocelli present; metatarsus longer
	than following joint. (Lestrèmia (Fig. 496), Campylomỳza,
	Hormosomyia (Fig. 495), Micromyia, Monárdia, Prio-
	néllus Strobliélla) I.ESTREMIÌNÆ
	néllus, Strobliélla) LESTREMIIN $\rlap/E$ Media with anterior branch $(M_{1+2})$ wanting, $M_{3+4}$ absent or repre-
	sented by a fold; ocelli absent; metatarsus much shorter than
	following joint; antennal joints with whorls of looped threads,
	or sometimes with horseshoe-like appendages. (Cecidomyia
	(=Itónida), Asphondylia, Asteromylia, Colpòdia, Con-
	tarínia (C. jóhnsoni, Grape blossom-midge; C. pyrívora, Pear
	midge), Dasyneùra (D. rhodóphaga, Rose midge; D. trifolii,
	Clover leaf midge), Diarthronomyia (D. hypogwa, Chrys-
	anthemum gall-midge), Diplòsis, Lasióptera, Phytóphaga
	(P. (Mayetiola) destructor, Hessian fly, Fig. 494), Rhabdó-
	phaga) CECIDOMYIÌNÆ
	75' 1 11
17.	Discal cell present in middle of wing contiguous to end of basal
	cells (Fig. 500), media four-branched, eight veins reaching wing-
	margin; ocelli present; dichoptic; antennæ 12- to 16-jointed;
	pulvilli wanting, but empodium pulvilliform. Widespread, not
	many species. (Anisopus (= $Rhyphus$ , = $Phryne$ ) (Figs. 465e,
	$500$ ), Lobogáster, Olbiogáster). $(PHRYN\grave{E}ID\cancel{E},\ RH\acute{Y}PH-$
	$ID\mathscr{E})$ Anisopódidæ
	Wings without a discal cell formed between branches of the usu-
	ally petiolate media18
18.	Ocelli present, sometimes the lateral ocelli next to the eyes and the
	middle one vestigial or absent
	Ocelli absent or at most vestigial: coxæ not lengthened30
19.	
10.	basal and attaining middle of wing (shorter than first in <i>Plecia</i>
	(Fig. 499) which has third vein furcate), anterior veins strong;
	pulvilli present; antennæ usually shorter than thorax, rather
	stout, without constrictions between joints; male holoptic, eyes
	large and divided into upper and lower parts; palpi four-jointed.
	Widespread, mostly holarctic. March flies BIBIÓNIDÆ
a.	Third vein furcate. (Plècia (Fig. 499), Penthètria).
a.	PLECINE
	Third vein simple. (Bíbio (Fig. 501), Bibiòdes, Dílophus).
	zina com omipio ( zivio ( tig. 001), ziviouos, ziiopiius).

Second basal cell imperfectly separated from first (i.e. base of media weak or undeveloped), or apically open, or very short, never longer than first basal cell; pulvilli absent or very minute.



Figs. 497-501. Scatopsidæ, Bibionidæ, Anisopodidæ

- 497. Canthyloscelis, wing (Edwards) Scatopsidæ.
- 498. Scatopse (Cole) Scatopsidæ.
- 499. Plecia, wing. Bibionidæ.
- 500. Anisopus, wing. Anisopodidæ.
- 501. Bibio, male. Bibionidæ.

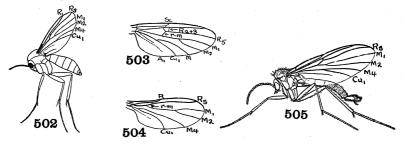
  - - Dichoptic, face rather broad; front tibiæ ending in a spine; mesonotum elevated in front. (Aspístes, Árthria). ASPISTĪNÆ

21. Radial sector with three branches, i.e. second longitudinal vein

	forked, the second vein (R <sub>2+3</sub> ) arising from the third (R <sub>5</sub> ) at
	or before the anterior crossvein (r-m) (Fig. 491); antennæ 15-
	jointed, the joints longer than wide; male dichoptic; coxæ not
	lengthened. (Pachyneura, palæarc.; Axymyia (Fig. 491),
	nearc.)
	Radial sector with two branches
22.	Second basal cell apically widely open, the posterior branch of the
	media when complete arising from the cubitus usually near the
	base, anal vein incomplete, not reaching margin of wing (see
	Fig. 504)
	Second basal cell closed (see Fig. 506), the apparent crossvein (i.e.
	the angular origin of M <sub>4</sub> ) sometimes located close to base of
	wing (Fig. 507), or the media and cubitus coalescent where the
	crossvein usually is located, or when the basal section of M
	is wanting, the media appears to arise from Cu <sub>1</sub> ; anal vein
	reaching margin at least as a fold
23.	Anterior branch $(R_{2+3})$ of radial sector abruptly extending to $R_1$ ,
	appearing like an extra crossvein closing the small rectangular
	or trapezoidal cell R <sub>1</sub> (Fig. 503); Sc usually reaching cell R <sub>1</sub> ;
	ocelli usually remote from eye-margin; microscopic hairs of
	wing-membrane typically irregularly scattered or the wings
	pubescent. Many genera; Europe, America, Australia. (Sció-
	phila (Fig. 503), Diómonus, Dziedzíckia, Monoclòna, My-
	comỳia, Neoemphèria, Polylépta, Stenophrágma).
	SCIOPHÍLIDÆ
	Radial sector not branched, the cell R <sub>1</sub> open to the wing-margin;
	Sc usually vestigial; microscopic hairs of wing-membrane seri-
	ately arranged
24.	Coxæ much elongated, fully half the length of femora; r-m cross-
	vein usually distinctly angulated from the second section of
	radial sector (Fig. 505); cubitus usually formed and long petio-
	late, rarely simple; eyes oval or reniform but without bridging
	projections above antennæ; palpi 4- or 5-jointed; prothorax
	with bristles. The dominant family of Fungus-gnats. Wide-
	spread. (FUNGIVÓRIDÆ)
a.	R <sub>1</sub> and R <sub>s</sub> running separately to base of wing, traces of base of
	$R_{2+3}$ present. (Lygistorrhina, Austr.). LYGISTORRHINÎNÆ
	$R_s$ arising from $R_1$ well beyond base of wing, or base of $R_s$ wanting;
	$R_s$ arising from $R_1$ wen beyond base of wing, or base of $R_s$ wanting; $R_{2+3}$ not present
	162+3 not present

b. Antennæ inserted plainly above middle of head; pronotum without bristles; occiput flattened, orbital bristles seriate; media with only apica disconnected parts present. (Manòta, holarc.).

MANOTÌNÆ



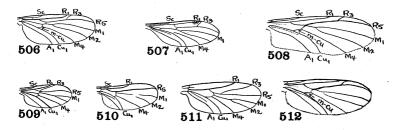
Figs. 502-505. Mycetophilidæ, Sciophilidæ, Sciaridæ

- 502. Mycetophila (Johannsen) Mycetophilidæ.
- 503. Sciophila, wing (Johannsen) Sciophilidæ.
- $504.\,$  Sciara, wing (Johannsen) Sciaridæ.
- 505. Boletina, male (Cole) Mycetophilidæ.

- b. Cubitus and  $M_4$  forking beyond origin of  $M_{1+2}$ . (Megalósphys, Fungivórides, Phorodónta) . . . . . . MEGALOSPHYĨNÆ Cubitus and  $M_4$  forking before or opposite origin of  $M_{1+2}$ . . . . . . c
- c. Medial cell (between M<sub>1</sub> and M<sub>2</sub>) not wider than adjacent cells.

  (Sciara (=Lycòria) (Fig. 504), Epídapus, Plastosciara,
  Psilosciara) SCIARINÆ

25. Second basal cell minute, much shorter than first basal cell due to the proximal location of the apparent m-cu crossvein (Fig. 507), radius three-branched, the middle branch short and commonly ending in R<sub>1</sub> near its tip; Sc complete; basal and middle sections of media continuous. (Bolitóphila (Fig. 507), Bolitophilélla).
BOLITOPHÍLIDÆ



Figs. 506-512. Mycetobiidæ, Bolitophilidæ, Diadocidiidæ, Ditomyiidæ, Ceroplatidæ, Macroceratidæ

- 506. Palæoplatyura, wing (Johannsen) Mycetobiidæ.
- 507. Bolitophila, wing (Johannsen) Bolitophilidæ.
- 508. Macrocera, wing. Macroceratidæ.
- 509. Mycetobia, wing (Johannsen) Mycetobiidæ.
- 510. Diadocidia, wing (Johannsen) Diadocidiidæ.
- 511. Ditomyia, wing (Johannsen) Ditomyiidæ.
- 512. Ceroplatus, wing, Ceroplatidæ.

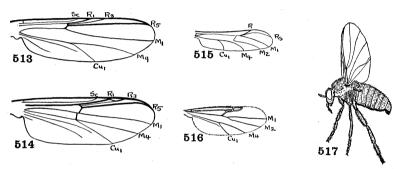
- 29. Antennæ short, usually thick-set and often flattened; tibial bristles present though usually small, posterior tibiæ with unequal apical spurs; forks of cubitus evenly diverging from the beginning. Europe, N. America, N. Africa, Australia. (Ceróplatus (Fig. 512), Asíndulum, Apèmon, Cerotèlion, Nervijûncta, Platyùra (=Zelmìra). (PLATYÙRIDÆ).

## CEROPLÁTIDÆ

Antennæ usually very slender, nearly as long as or even much longer than the whole body; tibiæ without bristles, posterior tibiæ with subequal apical spurs; forks of apparent cubitus (M<sub>4</sub> and Cu<sub>1</sub>) parallel for a short distance and then diverging. (Macrócera (Fig. 508), widespr.; Chiasmoneùra, indo-mal.).

# MACROCERÁTIDÆ

- - Cubitus forked at base, not petiolate, second basal cell open; anterior veins thick, others very weak (Fig. 517); antennæ about as long as head, ten-jointed, the flagellar joints closely united (Fig. 465b); male holoptic; first abdominal tergite with a conspicuous fringed flap-like scale; male metatarsi usually dilated. Widespread, mostly holarctic and neotropical. Blackflies, Buffalo-gnats. (MELŪSÍNIDÆ)......SIMULĪIDÆ
  - a. Radial sector forked, the two branches close together; second joint of hind tarsus without dorsal incision, the first joint apically



Figs. 513-517. Chironomidæ, Ceratopogonidæ, Simuliidæ

- 513. Chironomus, wing (Kieffer) Chironomidæ.
- 514. Anatopynia, wing (Kieffer) Chironomidæ.
- 515. Stenoxenus, wing (Kieffer) Chironomidæ.
- 516. Hartomyia, wing (Cole) Ceratopogonidæ.
- 517. Simulium (Lugger) Simuliidæ.

  - e. Front metatarsi normal. (Nevermánnia, holarc.; Frièsia, Wilhélmia, widespr.; Cnètha, Schoenbaueria, palæarc.).

### **NEVERMANNIÌNÆ**

Front metatarsi of both sexes flattened and broadened. (Simù-lium (Fig. 517), Odágmia, widespr.; Bcóphthora, holarc.; Býssodon, nearc.; Edwardséllum, ethiop.) . SIMULIÌNÆ

31. Anterior branch of media not forked; mouthparts not chitinized, not fitted for piercing; front legs lengthened, commonly raised

up and vibrated when at rest; postnotum generally with median longitudinal furrow or keel. (TENDIPÉDIDÆ).

### **CHIRONÓMIDÆ**

- a. Crossvein between M<sub>1</sub> and cubitus present, i.e. second basal cell complete (Fig. 514)
   b. Crossvein between media and cubitus absent, no second basal cell (Fig. 513)
- b. R<sub>2+3</sub> either present and forked, i.e. connected with R<sub>1</sub> by a crossvein, or else absent altogether. (Tánypus (= Pelòpia, = Protenthes), Anatopỳnia (Fig. 514), Pentaneura (= Ablabesmyia), Proclàdius). (PELOPIINÆ).....TANYPODÌNÆ
- d. Pronotum scarcely divided, an episternal suture well developed (i.e. an oblique slash-like cleft extending from base of wings nearly to front coxe, as in fig. 487); male antennæ normally plumose. (Orthoclàdius, Corynoneùra, Cricótopus, Metriocnèmis, Spaniótoma (=Camptoclàdius)).

  - Media forked (Fig. 516); mouthparts chitinized, fitted for piercing; postnotum gently rounded, without median furrow; front legs not lengthened. Punkies, No-see-ums, Sand-flies. Mainly holarctic. (HELÈIDÆ)......CERATOPOGÓNIDÆ
- a. R and M fusing to beyond middle of wing. (Stenóxenus (Fig. 515), nearc.). (STENOXÉNIDÆ) ..... STENOXENÎNÆ
  - R and M separate. (Ceratopògon, Culicòides, Bézzia, Forcipomỳia, Hartomỳia (Fig. 516), Johannsenomỳia, Palpomỳia). (Incl. JOHANNSENOMYIIDÆ).

# **CERATOPOGONÌNÆ**

# Suborder Brachycera, Section Orthorrhapha

32. Last tarsal joint furnished with three nearly equal pads under the tarsal claws, *i.e.* empodium developed pulvilliform (Fig. 534);

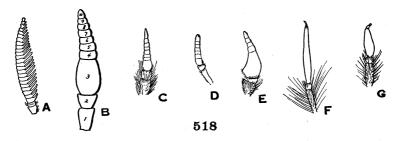
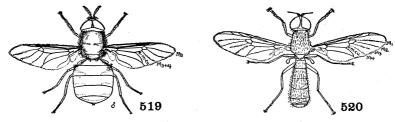


Fig. 518. Brachycerous Antennæ (Orthorrhapha)

- A, Rhachicerus (Vollenhoven) Xylomyiidæ; B, Subulonia (Enderlein) Xylomyiidæ; C, Cœnomyia (Verrall) Cœnomyiidæ; D, Xylophagus (Verrall) Xylophagidæ; E, Tabanus (Verrall) Tabanidæ; F, Bombylius (Verrall) Bombyliidæ; G, Thereva (Verrall) Therevidæ.
- - Prefurca longer, i.e. R<sub>s</sub> arises distinctly before base of discal cell (Fig. 526); at least middle tibiæ with distinct spurs; costa continuing around hind margin of wing as the ambient vein.....37
- 35. Second vein (R<sub>2+3</sub>) arising at or beyond anterior crossvein (r-m), discal cell small, usually pentagonal and located closer to the costa than usual; submarginal cell or cells very small and narrow



Figs. 519, 520. Stratiomyiidæ

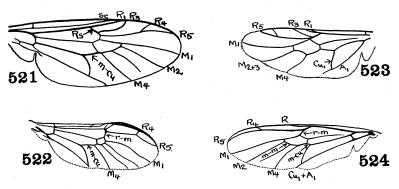
- 519. Odontomyia, male (Cole) Stratiomyiidæ.
- 520. Geosargus, male (Verrall) Stratiomyiidæ.

  - d. Media three-branched (Fig. 523), i.e. discal cell emitting two veins from its apex in addition to the vein forming its under side....e Media four-branched (Fig. 524), i.e. discal cell emitting three veins from its apex or the third just below.....g
  - e. Anterior branch of cubital fork (M<sub>3+4</sub>) joined to the discal cell by an apparent crossvein, *i.e.* discal cell emitting only two veins; apical antennal segment bristle-like, as long as remainder of antenna. (Prosopochrysa, Java) ... PROSOPOCHRYSINÆ

M<sub>3+4</sub> forming the lower side of the discal cell for a greater or less distance, the discal cell emitting therefore three veins in all. f

f. Antennæ with last, terminal or subterminal, segment (tenth) bristle-like and tipped with an extra hair, segments three to nine usually short and forming an oval or spherical complex third antennal joint. (Pachygáster, holarc.; Cynipimórpha (Fig. 523), Zabráchia, nearc.; Panàcris, Psephiócera, neotrop.; Evàsa, malay.; Plátyna, ethiop.).

PACHYGASTRÌNÆ



Figs. 521-524. Stratiomyiidæ

- 521. Archistratiomys, wing (Enderlein) Stratiomyiidæ.
- 522. Analcocerus, wing (Williston) Stratiomyiidæ.
- 523. Cynipimorpha, wing (Williston) Stratiomyiidæ.
- 524. Chrysochlora, wing. Stratiomyiidæ.

Antennæ with tenth segment not bristle-like, usually flattened, long, strap-shaped and fringed on edges, sometimes all ten antennal segments are similar, suggesting the Nematocera. (Lophóteles, Artemìta, Psegmómma, neotrop.; Isomerócera, Ptilócera, Tínda, ethiop., indomal.).

## LOPHOTELINÆ

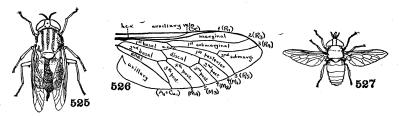
- g. All four medial branches arising from discal cell (Fig. 524).....h
  M<sub>4</sub> apparently connected with discal cell by a crossvein (Fig. 520).....k
- - Last antennal part styliform or undifferentiated, not ribbon-like, segments six to eight without furrow.....i

i.	Scutellum with marginal spines
	Anacanthélla, Porpócera, Rùba) CHRYSOCHLORÌNÆ
j.	Scutellum with four to twelve spines. (Antissa, Parantissa,
	neotrop.; Tetracanthina, Java) ANTISSÌNÆ
	Scutellum with two spines. (Clitellaria (=Ephippium), Eupary-phus, Nemótelus, Oxýcera, holarc.; Negritomyia, ethiop.,
	malay.; Euryneùra, Nothomŷia)CLITELLARIÌNÆ
k.	Antennæ with elongate terminal or dorsal arista
	Antennæ without a distinct arista
l.	Scutellum with two spines, or with vestiges of two spines. (Rha-
	phiócera, Hoplístes, Lysòzum, neotrop.).
	RHAPHIOCERATINÆ
	Scutellum without spines; metanotum prominently convex and
	with upturned hairs. (Geosárgus (= $\hat{S}$ árgus) (Fig. 520), Chry-
	sochrôma, Microchrysa, Ptécticus, widespr.; Gongròzus,
122	malay.). (SARGÎNÆ) GEOSARGÎNÆ Antennæ ending in an elongate and broadly flattened ribbon-like
111.	segment; scutellum with two spines. (Analcocerus (Fig.
	522), neotrop.)
	Antennæ with last segment of third joint usually short, never
	ribbon-liken
n.	ribbon-like
	tomyia (=Eulàlia) (Fig. 519), widespr. mostly in temperate
	zone; Cyphomyia, widespr., mostly tropical; Myxosárgus,
	Rhingiópsis, neotrop.; Hírtea, Hoplodónta).
	STRATIOMYIÌNÆ
	Scutellum unarmed. (Lasiòpa, widespr.; Chordonòta, neotrop.; Udamacántha) LASIOPÌNÆ
	Odamacantina) LASIOFINA
	Second vein arising before anterior crossvein (r-m), veins not
	crowded anteriorly; mostly neotropical species36
36.	Third vein simple, ending before wing-tip; all posterior cells open;
	third antennal joint usually three-segmented; abdomen slender,
	comprising seven segments; species under one inch in length.
	(Chiromỳza, Clavimỳia (Fig. 528), Mesomỳza, Nonàcris,
	Xenomórpha, neotrop.; Archimýza, austr.).
	CHIROM ÝZIDÆ
	Third vein forked, its branches (R <sub>4</sub> and R <sub>5</sub> ) widely divergent and
	enclosing the tip of the wing (Fig. 525), fourth posterior cell
	(M <sub>3</sub> ) closed; abdomen broadly rounded; gigantic species, not
	common. (Pantophthálmus, Acanthomèra, Rhaphior-
	rhýnchus, neotrop.). $(ACANTHOM \acute{E}RID \rlap{E}E)$ .
	DANTODUTUÁT MID Æ

- - a. Hind tibiæ without apical spurs.....b Hind tibiæ with two apical spurs which sometimes are minute. h
  - b. Third antennal joint divided into four, rarely three, segments. (Hæmatópota, widespr.; Heptátoma, Eur.).

    HÆMATOPOTÌNÆ

Third antennal joint divided into five segments, antennæ therefore apparently seven-jointed, rarely indistinctly ten-jointed. .c

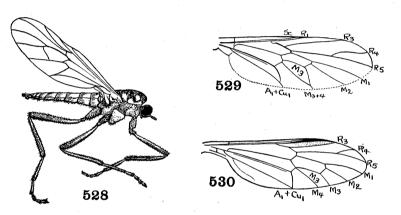


Figs. 525-527. Pantophthalmidæ, Tabanidæ

- 525. **Pantophthalmus,** photograph of specimen in O. S. Westcott collection. Pantophthalmidæ.
- 526. Tabanus, wing (Williston) Tabanidæ.
- 527. Tabanus, photographed specimen. Tabanidæ.

  - e. First antennal joint longer than wide, usually much longer; ocelli absent; first posterior cell open; relatively slender species. (Diachlorus, Acanthócera, neotrop.) . . . . DIACHLORÌNÆ
  - First antennal joint about as long as wide.....f.
    f. Antennæ with basal segment of third joint not excavated above,
  - without angle or tooth, but instead with a few short black spines; small, delicate species. (Stenotabanus, neotrop.).

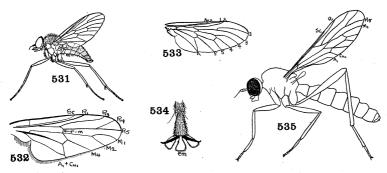
STENOTABANÎNÆ



Figs. 528-530. Chiromyzidæ, Xylomyiidæ, Xylophagidæ

- 528. Clavimyia (Enderlein) Chiromyzidæ.
- 529. **Xylomyia,** wing. Xylomyiidæ.
- 530. Xylophagus, wing. Xylophagidæ.

	First posterior cell closed. (Pangònia, palæarc.; Phàra, ethiop.; . Esenbéckia, Fidèna, Sciòne, neotrop.; Lilæa, austral.). PANGONIÌNÆ
	Calypteres small or vestigial; head not hemispherical, the occiput convex; abdomen oblong; second submarginal cell (R <sub>4</sub> ) not wide
38.	Fourth posterior cell $(M_3)$ almost or quite closed (Fig. 529); dichoptic
a.	Third joint of antennæ divided into eight annulations. (Xylomyia (Fig. 529), Eurasia, Afr., Am.; Nematocerópsis, Manchuria; Prísta, malay,; Sólva, palæarc., malay.; Subulònia, nearc.)
39.	Fourth posterior cell (M <sub>3</sub> ) open (Fig. 530)
	Facial orbits and cheeks separated from the central part; hind margin of wing veined throughout; male holoptic; scutellum of $C\alpha nomyia$ spined. Mostly robust, yellowish or blackish flies; widespread, but rare
a.	Proboscis short
b.	All tibiæ with apical spurs
c.	Abdomen broader than thorax; scutellum spined or not. (Cœnomyia (Fig. 518c), Anacantháspis, holarc.).
	Abdomen elongate, narrower than thorax; scutellum armed with spines. (Stratioléptis, Siberia, Japan).  STRATIOLEPTINÆ
40.	Costa continuing around wing-margin, venation normal (Fig. 531), anterior crossvein (r-m) distinct, five posterior cells ( <i>Hilari-morpha</i> with four); at least posterior tibiæ with spurs; calyp-



Figs. 531-535. Rhagionidæ

- 531. Rhagio, male (Cole) Rhagionidæ.
- 532. Hilarimorpha, wing. Rhagionidæ.
- 533. Chrysopila, wing. Rhagionidæ.
- 534. Rhagio, end of tarsus, showing broad empodium. Rhagionidæ.
- 535. Vermileo (Wheeler) Rhagionidæ.

Face socketed and not projecting, separated by a groove from the rather wide cheeks, antennæ inserted below middle of eyes; alula present, calypteres well developed.........................c

c. Front tibiæ with one or two spurs, hind tibiæ with two spurs.

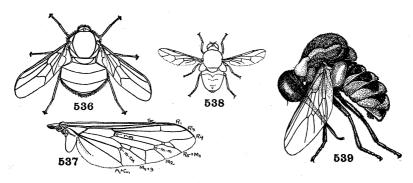
(Bicálcar, palæarc.; Bolbomŷia, Diálysis, Triptótricha, nearc.) ...... BICALCARÎNÆ

(Fig. 531), Athèrix, Átrichops). (LEPTÌNÆ).

RHAGIONINÆ

Hind tibiæ with one spur which is sometimes reduced; eyes bisected, the lower facets smaller than the upper; second vein

41. Head as wide as the depressed thorax; calypteres vestigial; posterior veins parallel with hind margin (Fig. 551), sometimes



Figs. 536-539. Acroceratidæ

- 536. Pterodontia, female (Cole) Acroceratidæ.
- 537. Ocnæa, wing. Acroceratidæ.
- 538. Acrocera (Verrall) Acroceratidæ.
- 539. Thyllis (Cole) Acroceratidæ.

forming a secondary network of small cells, first basal cell very long, its forward border continued obliquely across the wing as a "diagonal vein." Rare species, inhabiting arid regions, principally Asia Minor, S. Africa, Chile and Australia.

#### NEMESTRÍNIDÆ

a. Proboscis elongate, slender and hard, the labella usually narrow, palpi short. (Nemestrinus, Fallénia, Megistorhýnchus, Neorhynchocéphalus, Rhynchocéphalus (Fig. 551).

#### NEMESTRINÎNÆ

Proboscis short and broad, the labella fleshy, palpi long and upturned; ovipositor telescopic; alula broad. (Hirmoneùra, Hyrmophlæba, Symmíctus,) . . . . . HIRMONEURÎNÆ

Head placed low, very small as compared with the greatly hump-backed body, abdomen rounded, often inflated (Fig. 536); calypteres inflated, hiding the halteres; costa discontinued at wing-tip, posterior veins not parallel with hind margin of wing and not forming accessory cells; eyes of both sexes broadly contiguous. Spider parasites, rare species; widespread, but poorly represented in Indo-australia and tropical Africa. (CYRTIDÆ, HENÓPIDÆ, ONCÓDIDÆ).

#### ACROCERÁTIDÆ

- a. Third antennal joint small, with terminal style or hair-like rays; proboscis short.....b

  Third antennal joint large, more or less compressed, without
- b. Prothoracic lobes separated; abdomen usually inflated. (Acrócera (Fig. 538), Cýrtus, Nôthra, Oncódes, Opsèbius, Pterodóntia (Fig. 536)). (CYRTÌNÆ, ONCODÌNÆ).
- 42. Anal cell distinctly longer than second basal cell, either open, or acutely closed in or near margin of wing, basal cells usually relatively large (see Fig. 545); head bristles rarely evident . . . . 43

<sup>&</sup>lt;sup>1</sup> The following forms having short acute anal cell slightly longer than the second basal cell will cause confusion at this point of the key. The strength of the head bristles is then the best guide.

Opelia, Platypezidæ, couplet 58, Fig. 581: head bristles weak, no discal cell, the third vein not forked.

Sciadoceratidæ, couplet 52, Fig. 584: head bristles strong, subcosta ending in R1.

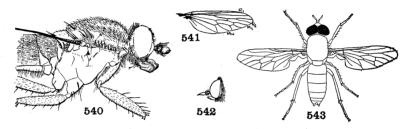
Empididæ, Brachystomatinæ, couplet 48, Fig. 559; and Hybotinæ, couplet 57, Fig. 563: head bristles weak, anal and basal cells rather long. Remaining Empididæ, couplet 53, have the anal cell small and either perpendicularly or obtusely closed.

Lonchopteridæ, couplet 54, Fig. 572: head bristles strong, wings lancet-shaped, no discal cell. The female has a false and cell.

Trypetidæ, couplet 126, Fig. 656, and the Ortalid series, couplets 94 to 97: anal crossvein (Cu1) angulately broken, the anal cell therefore with a pointed apical lobe.

Tachiniscidæ, couplet 99, Fig. 633: body bristly.

wing-veins numerous; often large species with strong legs...49



Figs. 540-543. Apioceratidæ, Therevidæ

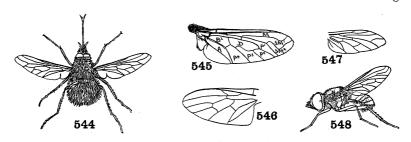
- 540. Ripidosyrma, head and thorax (Melander) Apioceratidæ.
- 541. Thereva, wing. Therevidæ.
- 542. Psilocephala, profile of head (Cole) Therevidæ.
- 543. Psilocephala, male (Cole) Therevidæ.
- 46. Apical veins curving forward, the third vein (R<sub>5</sub>) and nearly always also the fourth (M<sub>1</sub>) ending before apex of wing; at least the scutellum bristly; antennæ with a very short style; eyes

	separated; palpi broadened at tip. A small family; occurring in
	arid places; South Africa, America, Australia, Borneo. (Apió-
	cera, Apomidas, Rhaphiomydas, Ripidosýrma (Fig. 540)).
	APIOCERÁTIDÆ
	Fourth vein ending beyond apex of wing (Fig. 541); body usually
	furry rather than bristly, sometimes nearly bare; palpi not
	broadened apically. Widespread, principally holarctic. (Ther-
	eva (Fig. 541), Anaborrhýnchus, Epomỳia, Dialineùra,
	Phỳcus, Psilocéphala (Figs. 542, 543), Tabùda, Xestomỳia).
	THERÉVIDÆ
47.	Costa not continuing beyond apex of wing, fourth vein $(M_1)$ end-
	ing at or before wing-tip, three posterior cells (Fig. 552); pro-
	boscis hidden; antennæ without a style; body bare. Holarctic,
	neotropical and oriental; about thirty species; some are found
	on windows. (Scenópinus (=Omphrdle) (Fig. 552), Pseuda-
	tríchia). $(OMPHRALIDE)SCENOPÍNIDE$
	Costa continuing around entire wing, fourth vein (M1) ending
	beyond wing-tip, usually four posterior cells. (If the discal cell
	is open and the fourth vein is forked and long-petiolate (Fig.
	532), see Hilarimorphinæ, couplet 40, a)
48.	
	circlet of bristly hairs; tibiæ usually with spicules; proboscis
	usually long, thin and porrect; body usually furry and stout,
	rarely (Systropodinæ) extremely slender, bare and wasp-like;
	anal vein complete, anal cell (Cu) usually reaching margin,
	often open, alula usually distinct. Mostly occurring in sunny
	dry localities; alert, quick-flying species; many genera and species BOMBYLÎIDÆ
	-
a.	Second vein (R <sub>3</sub> ) arising almost perpendicularly from R <sub>s</sub> very
	close to the anterior crossvein (r-m) and forming a knee at its
	origin, only the third vein $(R_{4+5})$ continuous with the prefurca
	(Fig. 545); eyes with an indentation in the middle of the hind
	margin b Second and third veins forking acutely or in an arch and at a
	greater distance before the anterior crossvein than the length of that crossvein (see Fig. 548)
b	of that crossvein (see Fig. 548)
υ.	pencil of hairs at its tip; metapleuræ bare. (Anthrax (Fig. 545),
	Argyramèba, Chionamèba, Coquilléttia, Spongostylum).
	ANTHRACÌNÆ

Calypteres margined with scales; style without apical crown of hairs; metapleuræ hairy. (Exoprosòpa, Dipálta, Hemi-

pénthes, Hyperalònia, Lepidánthrax, Stónyx, Thyridánthrax, Vílla (= Hyalánthrax)) .... EXOPROSOPÌNÆ

with a rounded indentation.....g



Figs. 544–548. **Bombyliidæ** 

- 544. Bombylius (Verrall) Bombyliidæ.
- 545. Anthrax, wing. Bombyliidæ.
- 546. Mythicomyia, wing (Williston) Bombyliidæ.
- 547. Geron, wing (Williston) Bombyliidæ.
- 548. **Epacmus.** male (Cole) Bombyliidæ.

Face convex, not projecting; second vein arising acutely.....f

f. Head no broader than thorax; abdomen at least as broad as thorax, flattened; anterior crossvein much beyond middle of discal cell. (Lomàtia, Anisotàmia, Canària, Comptòsia, Oncodócera). LOMATIÌNÆ

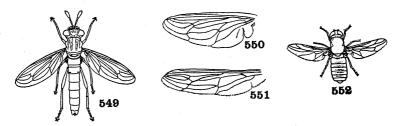
Head broader, but the hind edge narrower than the thorax; body more cylindrical than depressed; anterior crossvein near middle of discal cell. (Aphæbántus, Desmatoneùra, Epácmus (Fig. 548), Eucéssia, Petroróssia). . . . . . APHŒBANTĪNÆ

	Face at most as long as front; clypeus not separated from cheeks by a deep groove; second vein arising at an acute angleh
h.	
	or nearly so; body bare; wings narrowed at base, without alula and calypteres. (Sýstropus, Dolichomyia). SYSTROPÌNÆ
	Abdomen not remarkably slender; wings with alula and calypter.i
1.	Third vein simple, therefore only one submarginal cell; small bare flies with humped thoraxj
	flies with humped thoraxj  Third vein forked, two or three submarginal cells, second vein long, never ending in the first veink
j.	Second vein very short, ending in the first vein, or entirely absent.
	(Mythicomyia (Fig. $5\mathring{4}6$ ), Empidideicus, Glabéllula (= $Pachyneres$ )). ( $GLABELLULINÆ$ ) . MYTHICOMYIINÆ
	Second vein normal, ending independently in costa. (Cyrtòsia,
_	Cyrtomórpha, Platypỳgus) CYRTOSIÌNÆ
k.	Body more or less hunched, narrow, or at least not broad, thorax
	prominent, abdomen cylindrical or sometimes flattened; sometimes bare, or scaly, or with bristles; wings usually relatively
	short
	usually hairy and without bristles
1.	Prothorax in shape of an anterior ring beset with strong curved bristles. (Toxóphora, Heniconeùra, Lepidóphora).
	TOXOPHORÎNÆ
	Prothorax smaller and not beset with curved bristles. (Cyllènia,
	Amíctus, Éclimus, Epíbates, Hénica, Thevenetimyia).
	CYLLENIÎNÆ
m.	First antennal joint thickened and long-hairy; wings short, with four open posterior cells. (Conóphorus, Aldríchia, Codiònus,
	Platamodes) CONOPHORINÆ
	First antennal joint not thickened; wings not shortn
n.	Face protruding as a very short muzzle, proboscis short, porrect,
	with fleshy tip; eyes of male bisected; discal cell broadened at
	end, much broader than second posterior cell; nearly bare
	species. (Heterótropus (= Malthacótricha), Cænòtus, Pro- ràtes)
	Face, when developed, convex or somewhat conically projecting,
	but not beak-like, proboscis long, with small labella; rarely wholly bare
o.	Second vein and fork of third vein in line with wing-axis; body not
	broad but rather humped, vestiture fine and not abundant; leg
	bristles weak or absent. (Phthíria, Apólysis, Crocídium,
	Gèron (Fig. 547), Rhabdopsélaphus, Semiràmis).
	PHTHIRIÌN <i>Æ</i>

Second vein and fork of third vein curving forward, ending distinctly before tip of wing; body usually broad.....p
p. Vestiture short or undeveloped; no bristles on legs; head small.

(Ùsia, Corsomyza, Legnotomyia, Psiathalássius).

USIÌNÆ



Figs. 549-552. Mydaidæ, Nemestrinidæ, Scenopinidæ

- 549. Leptomydas, male (Cole) Mydaidæ.
- 550. **Mydas**, wing. Mydaidæ.
- 551. Rhynchocephalus, wing (Williston) Nemestrinidæ.
- 552. Scenopinus (Verrall) Scenopinidæ.

49. Body without bristles; fourth vein (M<sub>1</sub>) curving forward to end at or before wing-tip, neuration complex (Fig. 550), prefurca (i.e. basal section of R<sub>s</sub>) very short; antennæ with a clubbed style; proboscis with fleshy expanded tip, palpi vestigial. About 130 species; widespread, but not common; often flies of large size. (Mỳdas (Fig. 550), widespr.; Leptomỳdas (Fig. 549) nearc.; Cephalócera, ethiop.; Miltìnus, Triclònus, austr.). (MY-DÁSIDÆ)

Body usually with bristles, face bearded; fourth vein not curving forward, neuration not abnormal, prefurca long (Fig. 553); proboscis adapted for piercing, not fleshy, palpi usually prominent. A large family of nearly 4000 species, widespread, es-

- Marginal cell open; very slender species with few hairs and bristles; claws long, pulvilli absent; ovipositor without whorl of spines.
   (Leptogaster, Euscelidia, Psilonyx) ... LEPTOGASTRÎNÆ



Figs. 553-555. Asilidæ

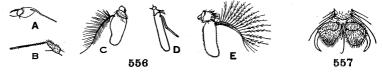
- 553. Erax, wing (Hine) Asilidæ.
- 554. Stichopogon, wing. Asilidæ
- 555. Atomosia, wing. Asilidæ.
- e. Front tibiæ without a claw-like apical projection. (Anisopògon, Cyrtopògon, Dióctria, Habropògon, Heteropògon, Holopògon, Lasiopògon, Microstỳlum, Myiélaphus, Pycnopògon, Rhádinus, Stenopògon, Stichopògon (Fig. 554)).

**EREMOCNEMINÆ** 

Front tibiæ with a claw-like apical projection. (Cenopògon, Cophùra, Dasypògon, Deromỳia, Isopògon, Nícocles,

<sup>&</sup>lt;sup>1</sup> The divisions of the Dasypogoninæ and Laphriinæ have less rank than the other two subfamilies of the Asilidæ, but are given because of the dominance of this family. The divisions of the Dasypogoninæ have no corresponding type genera.

Moderate to large, rather heavily pubescent species; third antennal joint without subapical thorn; side pieces of metanotum never bristly; crossveins closing discal and fourth posterior cells not in the same line; genitalia free and usually large. (Làphria, Andrenosòma, Ctenòta, Dasýllis, Dásythrix, Lámpria, Lámyra, Nùsa, Pogonosòma) . . . . . . LAPHRIÌNÆ, s. str.



Figs. 556, 557. Cyclorrhaphous Antennæ and Tarsus

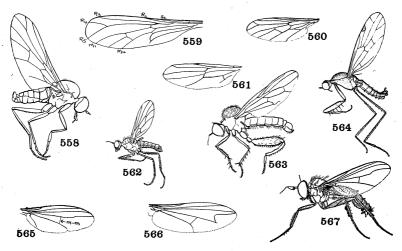
- 556. Antennæ: A, **Dolichopus**, Dolichopodidæ; B, **Drapetis** (Williston) Empididæ; C, **Volucella** (Williston) Syrphidæ; D, **Gonia** (Williston) Tachinidæ; E, **Glossina** (Hegh) Glossinidæ.
- 557. Musca, end of tarsus, showing hair-like empodium (Kellogg) Muscidæ
- - Frontal suture well developed as a horseshoe-shaped groove over the antennæ continuing down so as to separate the center of the face from the sides, frontal lunule present as a crescentic sclerite between the antennæ and the frontal suture (Fig. 594),<sup>1</sup> middle part of front nearly always differentiated from the orbits; calypteres and alula usually pronounced; arista almost

<sup>&</sup>lt;sup>1</sup> The frontal suture is forced open by the protrusion of the ptilinum, an eversible sac, for prying off the lid of the puparium at the time of emergence of the adult. Newly emerged flies of this group sometimes show the ptilinum; in older flies the ptilinum is withdrawn and the frontal suture closes to form the characteristic seam over the antennæ. The lid of the puparium opens by a circular cleft (see Fig. 763), hence the significance of the group-name Cyclorrhapha, meaning circular seam. In the coördinate group Orthorrhapha (including couplets 32 to 49) the pupa case opens by a dorsal straight longitudinal seam (see Fig. 707) and accordingly the frontal suture and ptilinum are not developed.

51.	always dorsal; costa not extended to hind margin of wing; first two dorsal segments of abdomen more or less fused. $MUSCOIDEA$ )
	obliquely across field of wing (as in Fig. 570), nor of lanceolate shape with long second vein and ambient costa (as in Fig. 571); antennæ evidently two- or three-jointed, the apical joint not spherical; male sometimes holoptic
	Venation peculiar, of the type represented by Fig. 570 or Fig. 571; no discal cell; eyes of both sexes widely separated
52.	Alular edge of wing without hairy bristles; posterior crossvein (m-m) usually present and then always located much beyond
	the anterior crossvein (r-m); anal cell acute (Fig. 563), rectangular (Fig. 561), rounded (Fig. 565) or obtuse (Fig. 560), when the anal cell is acute, the basal cells are relatively long and the subcosta is either vestigial or terminates in the costa; hind
	tarsi with first joint longer than second
	anal cell short and acute, the anal vein reaching margin, medial branches disjoined basally, Sc ending in R <sub>1</sub> ; postocular cilia present; hind metatarsus shorter than next joint. (Sciadócera, (Fig. 584), neotrop., austr.) SCIADOCERÁTIDÆ
53.	Anterior crossvein (r-m) located beyond basal fourth of wing, discal cell usually separate from second basal, third vein often forked, often four posterior cells, i.e. M <sub>2</sub> separate from M <sub>3</sub> , Sc vestigial or ending in costa; calypteres minute; eyes usually with a small incision at the antennæ; seriate postocular cilia wanting; antennal style usually terminal; proboscis usually rigid; male genitalia not inflexed; dull colored species, almost never metallic. A large family of over 2000 species, widespread, but principally holarctic and neotropical
a.	Anal and discal cells complete, or if either is incomplete the front coxæ are very long and the front legs are raptorial, or else the anal angle of the wing is rectangularb  Discal cell united with second basal (Fig. 561), anal cell and anal

vein wanting or incomplete, three posterior cells, Sc vestigial or wanting, third vein always simple. (Tachydròmia, Drapetis

	(Fig. 556b), Coloboneùra, Micrémpis, Platypálpus (Fig. 561),
	Stílpon, Tachypèza, Tachyémpis). TACHYDROMIÌNÆ
b.	Anal angle of wing not projecting, costa weakly but visibly con-
	tinuing around hind margin of wing, anal crossvein (Cu <sub>1</sub> ) acute,
	perpendicular, or rounded, rarely obtusely closing anal cell;
	front coxæ longer than posterior pairs; proboscis short; eyes
	broadly separated on the front; mesopleuræ obliquec
	Anal angle more or less distinct, if the wings taper uniformly
	toward base, the mesopleuræ are vertical or the hind margin of
	the wing is thin; front coxæ not elongate; male often holoptice
c.	
	femora in length; radial sector arising closer to anterior cross-
	vein than to humeral crossvein. (Hemerodròmia, Chelifera
	(Fig. 564), Chelípoda, Colàbris, Drymodròmia, Mono-
	dròmia) HEMERODROMIÌNÆ
	Legs slender, the front pair not distant from the others, the coxæ
	not long and the femora not thick; radial sector arising nearer
	base of wingd
d.	Second antennal joint connected with third by a finger-like pro-
	jection on inner side; no anal cell: restricted to southern hemi-
	sphere. (Ceratómerus, Icásma) CERATOMERÌNÆ
	Second antennal joint normal; anal cell present: mostly found
	about swiftly running brooks and waterfalls. (Clinócera
	(=Atalánta), Boreodròmia, Dolichocéphala, Heleodròmia,
	(Oreothàlia (Fig. 562), Synamphótera, Trichopèza, Wiede-
	mánnia). $(ATALANTINÆ)$ CLINOCERATINÆ
e.	Anal crossvein forming a distinct angle with basal part of anal vein;
	proboscis rarely longer than head; thorax often highly archedf
	Anal crossvein recurved and confluent with underside of anal cell,
	the anal vein usually an independent fold; proboscis often long,
	rarely porrect; antennæ usually three-jointed. (Émpis (Fig.
	560), Glòma, Hesperémpis, Hílara, Hilarémpis (Fig. 558),
	Hormopèza, Iteaphila, Micróphorus, Rhamphomyia).
•	EMPIDÎNÆ
f.	
	Sc distinctg
	Anal cell shorter than or about as long as second basal cell, its
	outer angle obtuse or right; Sc weak; proboscis short. (Ocy-
	dròmia, Anthàlia, Bicellària, Euthyneùra, Hoplocýrtoma,
	Leptopèza, Œdàlea, Trichina) OCYDROMIÎNÆ
g.	Discal cell emitting three veins, costa visibly continuing on hind
	margin; proboscis short and incurved; antennæ three-jointed;
	thorax not highly arched. (Brachýstoma (Fig. 559), Anomal-
	émpis, Homalocnèmis). (Including HOMALOCNEMINÆ).
	BRACHYSTOMATÌNÆ



Figs. 558-567. Empididæ, Dolichopodidæ

- 558. Hilarempis, male. Empididæ.
- 559. Brachystoma, wing. Empididæ.
- 560. Empis, wing. Empididæ.
- 561. Platypalpus, wing. Empididæ.
- 562. Oreothalia, male. Empididæ.
- 563. Euhybos, male (Melander) Empididæ.
- 564. Chelifera, male (Melander) Empididæ.
- 565. Dolichopus, wing. Dolichopodidæ.
- 566. Psilopodinus, wing (Aldrich) Dolichopodidæ.
- 567. Argyra, male (Cole) Dolichopodidæ.

Anterior crossvein located within basal fifth of wing (Fig. 566), discal cell always confluent with second basal, third vein never forked, three posterior cells, i.e. M<sub>1</sub> and M<sub>2</sub> fused, Sc when complete ending in R<sub>1</sub>; calypteres rather large and fringed; a row of postocular cilia present; proboscis almost always soft; male genitalia more or less inflexed under abdomen; color usually metallic green. A large family of over 2000 species, widespread, principally holarctic and neotropical. DOLICHOPÓDIDÆ

a.	Fourth vein typically broken, the front fork widely diverging and angulately approaching the third vein (Fig. 566); head short and broad, occiput concave, vertex sunken, ocellar triangle promi-
	nent; hypopygium free, its appendages visible; slender species
	with short and broad thorax, long narrow abdomen and long slender legs. (Sciopus, Chrysosòma, Leptorèthrum, Mes-
	órhaga, Psilopodinus (Fig. 566), Tenùopus). (AGONOSO-MATINÆ, LEPTOPODINÆ, PSILOPODINÆ, SCIAPOD-
	MATINÆ, LEPTOPODÍNÆ, PSILOPODÍNÆ, SCIAPOD-
	ÎNÆ) CHRYSOSOMATÎNÆ
	Fourth vein not angulately fractured though sometimes bowed; vertex not sunken; thorax longer than broadb
b.	
υ.	is concave and fitting against the thorax, or the palpi are broad,
	or the hypopygium is not free
	First antennal joint pubescent; occiput convex; face of male
	usually narrow and with only a weak indication of a transverse
	impression; palpi small; alar callus present; hind crossvein dis-
	tant from margin; middle tibiæ with an apical set of five bristles;
	hypopygium large, rather free, with evident and often large
	lamellæ; robust and bristly species. (Dolíchopus (Fig. 556a,
	565), Hercostomus, Orthochile, Paraclius, Pelastoneurus,
	Tachýtrechus) DOLICHOPODÌNÆ
$\mathbf{c}.$	Proboscis stout, with an incurved hook; coxæ spined; front femora
	basally with two divergent spine-like bristles; maritime species.
	(Aphrosylus, Teneriffa) APHROSYLINÆ
	Proboscis not furnished with a hook-like piercing organ; coxæ and
J	front femora not so spined
d.	Face usually broad, with evident transverse impressione Face usually narrow, with an incomplete transverse impression,
	which is sometimes entirely wanting, at least in maleh
e.	Arista dorsal; postvertical bristles evident; palpi usually very
•	broad, applied against proboscis; occiput usually convex; hy-
	popygium small, not free, with small to large appendagesf
	Arista apical or subapical; postverticals minute or wanting; occi-
	put concave; thorax with a prescutellar bare, flattened area;
	hypopygium long, without long evident appendages; alar callus
	not distinct. (Medètera, Oligochètus, Thrýpticus; Sacco-
	pherónta, ethiop.) MEDETERÌNÆ
f.	Hind crossvein nearly parallel with hind margin of wing, fourth
	vein bent forward and ending before wing-tip; upper occi-
	put concave; no acrostichal bristles; hypopygium sunk into
	sixth segment. (Plagioneùrus, America).
	PLAGIONEURÎNÆ
	Hind crossvein nearly transverse, located close to hind margin of
	wing, the distal segment of fifth vein shortg

- - Third antennal joint triangular or spherical, short, rarely somewhat lengthened with dorsal arista.....i
- i. Hypopygium large and free, appendages more or less conspicuous; thorax short, scarcely longer than broad, with prescutellar area; abdomen long; legs slender and without set of apical bristles on middle tibiæ. (Neurigòna, Oncopygius)... NEURIGONINÆ
  - Hypopygium usually small, rarely free, often hidden, the appendages never large though visible from beneath.....j
- j. Abdomen and legs elongate; antennæ located very high; ocellar triangle prominent; no pulvilli; hind margin of first abdominal segment raised. (Stolidosòma, neotrop.).
  STOLIDOSOMATÌNÆ

Abdomen short and robust; thorax longer than broad; middle tibiæ tipped with set of bristles................................. k

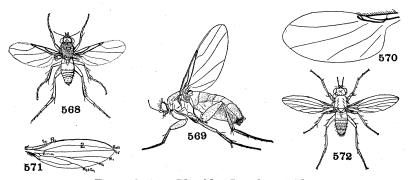
- k. Occiput rather concave; arista dorsal or subapical; hypopygium cap-shaped or hemispherical, usually with four or more strong bristles and only rarely with distinct appendages; body bristly, usually apex of abdomen furnished with bristles. (Diáphorus, Árgyra (Fig. 567), Asýndetus, Chrysotus, Leucóstola).
- Acrostichals wanting. (Xanthochlòrus, Chrysotìmus, Lamprochròmus; Micromórphus, ethiop.).

Acrostichals present, in one or two more or less evident rows. (Campsicnèmus, Sympýcnus, Syntormoneùra).

CAMPSICNEMINÆ

## Suborder Brachycera, Section Cyclorrhapha

54. Radial veins stout, running into the costa near middle of wing, medial veins weak and extending obliquely across wing, no crossveins and therefore no basal cells (Fig. 570); antennæ placed



Figs. 568-572. Phoridæ, Lonchopteridæ

- 568. Paraspiniphora (Verrall) Phoridæ.
- 569. Chætoneurophora (Cole) Phoridæ.
- 570. Megaselia, wing. Phoridæ.
- 571. Lonchoptera, wing of female. Lonchopteridæ.
- 572. Lonchoptera, male (Verrall) Lonchopteridæ.
  - a. Propleuræ lateral in position, visible from the sides, prothoracic spiracle not visible from above.....b Propleuræ small, anterior in position, the humeri formed by the
    - Propleuræ small, anterior in position, the humeri formed by the mesonotum, prothoracic spiracle visible from above; female usually wingless; ant-guests. (Platýphora (= Ænigmàtias), Ænigmatístes, Psyllomyia). (ÆNIGMATIINÆ).

### PLATYP HORÌNÆ

b. Tibiæ usually with one or several long preapical bristles; lower frontal bristles when present curving obliquely upward; mesopleuræ usually undivided; both sexes winged. (Phòra (= Trineùra), Chætoneuróphora (Fig. 569), Conícera, Diploneùra, Hypócera, Paraspiníphora (Fig. 568). PHORÎNÆ Tibiæ without any long preapical bristles, bare or ciliate on the

edge; two or four supra-antennal proclinate bristles; female often wingless or with aborted wings. (Metopina, Apocéphalus, Chonocéphalus, Gymnóphora, Ecitomyia, Megasèlia (= Aphiochæta) (Fig. 570), Pulicíphora (= Stethópathus), Rhyncophoromyia, Syneùra). (PULICIPHÓRIDÆ, STETHOPÁTHIDÆ). METOPINĨNÆ

Wings rather pointed at tip, lanceolate (Fig. 571,) costa encompassing entire wing, basal cells very small, second vein (R<sub>3</sub>) ending almost at wing-tip, anterior crossvein not obvious, no discal cell, the three branches of media arising from a common stalk from the apex of the second basal cell, Cu<sub>1</sub> of female curving forward and ending in M<sub>4</sub> at middle of wing-length, thus forming an apparent anal cell (Fig. 571), of male short and reaching hind margin (Fig. 572), veins largely setulose above; oral margin bristly; third antennal joint rounded, with a long subterminal bristle; thorax with bristles but no hairs. Principally palæarctic, few species, females rare. (Lonchóptera (Figs. 571, 572) (= Musidòra)). (MUSIDÓRIDÆ).

#### LONCHOPTÉRIDÆ

- - Proboscis distinctly longer than head, slender, stiff and often folding (Fig. 578); head wider than thorax, front broad in both sexes; face with a groove or grooves under the porrect antennæ, buccal cavity large; no body bristles; abdomen clavate, deflexed at tip; first posterior cell pointed, anterior crossvein near middle of discal cell. Widespread, about 500 species; parasitic on wasps, bees and Orthoptera. (See couplet 80).

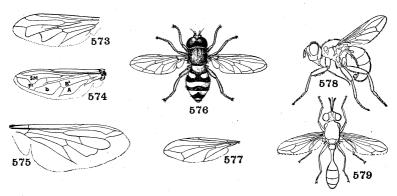
CONÓPIDÆ

a. Vertex and tibiæ without bristles; anal cell rather long and pointed; ovipositor not excessively long.....b Vertex with bristles, tibiæ spurred; anal cell small; ovipositor very long; proboscis long and geniculate; third antennal joint with subdorsal arista. (Stylogåster, mainly neotrop.).

STYLOGASTRÎNÆ

forward under abdomen. (**Dalmánnia** (Fig. 578)).

DALMANNIÌNÆ



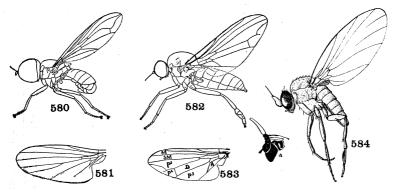
Figs. 573-579. Syrphidæ, Conopidæ

- 573. Microdon, wing (Williston) Syrphidæ.
- 574. Eristalis, wing. Syrphidæ.
- 575. Copestylum, wing. Syrphidæ.
- 576. Syrphus, male (Metcalf) Syrphidæ.
- 577. Conops, wing. Conopidæ.
- 578. Dalmannia, male (Cole) Conopidæ.
- 579. Physocephala (Lugger) Conopidæ.
- - First posterior cell (R<sub>5</sub>) closed (Fig. 574), usually an extra vein between the third (R<sub>5</sub>) and fourth (M<sub>1</sub>) veins and crossing the anterior crossvein (r-m); costa continuing around margin or stopping at wing-tip; anal cell closed just before wing-margin, therefore short-petiolate, the vein closing discal cell parallel with margin; head and body usually without bristles; arista dorsal.

	very rarely terminal; male usually holoptic; ocelli always
	present. Characteristically showy flower-flies with yellow
	markings; quick fliers and good hoverers; cosmopolitan, about
	3000 species
a.	Antennæ elongate, porrectb
a.	Antennæ moderate in length, drooping, if elongate and porrect
	not placed on a produced fronte
L	First posterior cell without stump of a vein from the third vein
υ.	First posterior cell with a stump of a vein from the third vein.
	which almost divides the cell into twod
	Antennæ inserted on a strong frontal process; a single red ab-
c.	dominal band. (Psarus) PSARINÆ
	Antennæ inserted on the flattened front; abdomen marked with
	several reddish bands. (Chrysotóxum)CHRYSOTOXÌNÆ
J	Arista dorsal; face rounded and pilose; scutellum usually armed
d.	and emarginate; sometimes antennæ of male split into two or
	four parts; larvæ and pupæ developing in ant nests. (Microdon
	(Fig. 573), <b>Mixogaster</b> , <b>Rhopalosýrphus</b> ). (Including <i>MASARYGIDÆ</i> , neotrop., with antennæ of male split into
	MASARIGIDAE, neotrop., with antennae of male spin into
	lobes)
	Style terminal, antennæ on frontal processes; face not with abundant
	pile. (Ceriòides (=Cèria, = Sphyximórpha)) CERIOIDÌNÆ Anterior crossvein located before middle of discal cell, nearly
e.	
	always rectangular
e	oblique
1.	yellow, the facial profile is parallel with eye-margin and face
	and eyes are pubescent; front of mesonotum pubescentg
	Pale color marks present on head, thorax and abdomen
~	Antennæ with terminal bristle. (Callicera). CALLICERATINÆ
g.	Antennæ with dorsal aristah
h	Third antennal joint very large, orbicular, thick; abdomen con-
11.	cave below, genitalia entirely hidden from dorsal view. (Nausi-
	mástar) NATISICASTRÎNA
	gåster)
	below, the genitalia usually largely visible from above. (Chi-
	lòsia, Chrysogáster, Cnèmodon, Heríngia, Orthoneùra
	Pipìza, Psilòta) CHILOSIÌNÆ
;	Lower part of face strongly projecting
1.	Lower part of face not strongly projecting.
j.	Face distinctly projecting conically downward; third antenna
1.	joint stout, with thickened three-jointed arista. (Pelecócera
	Chamestrabus)  PELECOCERATINA

	Face not projecting downward, but strongly produced forward;
	third antennal joint not stout and not with thickened aristak
k.	
	(Sphegina, Neoáscia) SPHEGÌNÆ
	Alula normal; abdomen short, not constricted at base. (Brachy-
	òpa, Rhíngia)
1.	Humeral calli and the region between them destitute of pile;
•	marginal cell open, vein closing first posterior cell usually
	parallel with margin; arista bare or pubescent
	Humeral calli and the interhumeral region more or less pilose;
	marginal cell closed, vein closing first posterior cell distally
	recurrent; arista heavily plumose; hind coxe with hair behind.
	(Trainestile (Fig. 55%) Generalizations (Fig. 575) Memoriana)
	(Volucélia (Fig. 556c), Copestylum (Fig. 575), Temnócera). VOLUCELLÌNÆ
m.	
	gáster) BACCHÌNÆ
	Abdomen oval, not narrow at base nor clavate. (Sýrphus (Fig.
	576), Dídea, Eriozòna, Leucozòna, Melanóstoma, Pára-
	gus, Platychirus, Pyrophæna, Sphærophòria, Xantho-
	grámma) SYRPHÌNÆ
n.	Third vein bending deeply into first posterior cell (Fig. 574);
	femora with conspicuous patch of black spinules near base.
	(Erîstalis (Fig. 574), Arctosýrphus, Helóphilus, Mallòta,
	Megáspis, Mèrodon (= Lampètia) (M. equéstris, Narcissus
	bulb-fly))
	Third vein not bending deeply into first posterior cello
0.	
	Sericomỹia $(=Cinxia)$ ). $(CINXIINÆ, SERICOMYIINÆ)$ .
	ARCTOPHILÎNÆ
	Arista bare or pubescentp
p.	
	angle. (Eumèrus) EUMERÌNÆ
	Apical crossvein oblique, at most very slightly recurrent at apex. q
q.	Marginal cell closed and petiolate. (Milèsia) MILESIÌNÆ
	Marginal cell open. (Xylòta (=Zélima), Brachypálpus, Calli-
	próbola, Cynorrhina, Criorrhina $(=Penthesilea)$ , Myio-
	lépta, Sphecomyia, Spilomyia, Syrítta, Temnóstoma,
	Tropídia) XYLOTÌNÆ
57.	Proboscis firm, styliform, porrect or short; male genitalia terminal,
	more or less asymmetrical; subcosta evanescent, not reaching
	costa, anal angle of wing more or less rectangular, no alula,
	costa interrupted at fourth vein; arista usually thread-like and
	terminal (Fig. 563). (Subfamily Hybotinæ, see couplet 53, g).
	EMPIDIDÆ, part

58. Antennæ with terminal arista; face small and broad; anal angle of wing more or less full, basal cells small, second basal much shorter than discal cell; hind tibiæ and tarsi dilated, especially in male; head and thorax with bristles; female sometimes bright-colored. About 100 species, principally holarctic, some nearctic, indoaustralian and ethiopian. (Platypèza (=Clýthia) (Fig.



Figs. 580-584. Pipunculidæ, Platypezidæ, Sciadoceratidæ

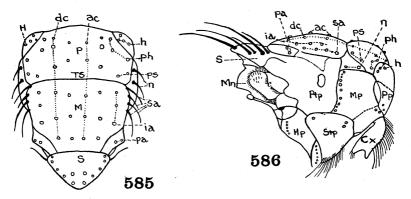
- 580. **Pipunculus,** male (Cole) Pipunculidæ.
- 581. Opetia, wing (Verrall) Platypezidæ.
- 582. Calotarsa, female; a, hind tibia and tarsus of male (Cole) Platypezidæ.
- 583. Platypeza, wing. Platypezidæ.
- 584. Sciadocera (Tonnoir) Sciadoceratidæ.

Antennæ with dorsal arista; face narrow; legs not dilated; head and body without true bristles; head very large, usually spherical, consisting almost wholly of the eyes; anal angle of wings not developed, second basal cell subequal to discal cell in length; anal cell closed near margin; ovipositor large, with bulbous base and long sword-like point, inflexed under abdomen. About 300 species, principally holarctic and australian, some nearctic and ethiopian. (Pipunculus (=Dôrylas) (Fig. 580), Châlarus, Nephrôceros, Verrâllia). (DORYLAIDÆ). PIPUNCÙLIDÆ

## Schizophora

- 60. Second antennal joint with a longitudinal seam along upper outer edge which extends quite to the base; anterior orbits not differentiated above from the lateral vertex plates, bearing a row of convergent lower frontal bristles which are more distant from eye-margin than the upper frontal bristles are (Fig. 594); usually at least lower calypter large; posthumeral and intra-alar bristles usually both present; thorax with a complete transverse suture before wings (Fig. 585); front of male usually narrow or the eyes meeting; subcosta always distinct and ending in costa, first vein (R<sub>1</sub>) never short (Fig. 598); abdominal spiracles at least of segments two to five located in side margins of tergites, very rarely in the membrane. (CALYPTRATÆ, MUSCARIDÆ, MYODARIA SUPERIÒRA, THECOSTÓM-ATA)
  - Second antennal joint without such a seam (except Loxocera, couplet 124); anterior orbits usually separated above from lateral vertex plates, or the latter alone developed and bearing fronto-orbital bristles (if exceptionally the fronto-orbital bristles are located on the orbits the lower ones are closer to the eyemargin than the upper ones are); lower calypter vestigial or undeveloped; posthumeral bristles absent; thorax without a complete transverse suture in front of wings, posterior callosity usually absent; a visible membrane connecting the dorsal and ventral sclerites of the abdomen, in which the spiracles are nearly always located (if spiracles are in tergites, e.g. Chlorop-

idæ and Ephydridæ, the subcosta is imperfect); front of both sexes of nearly equal width, or if wider in female the greater width is due to a widening of the middle stripe; fourth vein (M<sub>1</sub>) nearly straight, never angulate or with an appendage; often very small species. (ACALYPTRATÆ, BORBORÒIDEA, HAPLOSTÓMATA, MYODARIA INFERIÒRA)......71



Figs. 585, 586. Calliphoridæ

585. Calliphora, thorax, dorsal view (Walton) Calliphoridæ.

586. Calliphora, thorax, lateral view (Walton) Calliphoridæ.

Sclerites: Cx, front coxa; H, humerus; Hp, hypopleura; M, posterior portion of mesonotum (metazona); Mp, mesopleura; Mn, metanotum; P, anterior portion of mesonotum (prozona); Pp, propleura; Ptp, pteropleura; S, scutellum; Stp, Sternopleura.

Bristles: ac, acrostichals; dc, dorsocentrals; h, humerals; ia, intra-alars; n, notopleurals; pa, post-alars; ph, posthumerals; ps, presutural; sa, supra-alars.

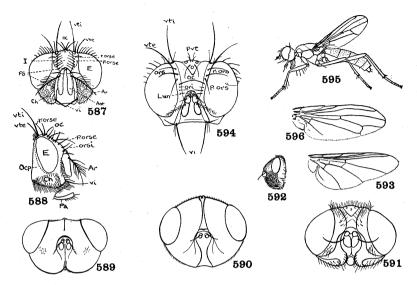
c, calypteres; TS, transverse suture separating prozona from metazona.

**ŒSTRIDÆ** 

a. Head apparently closed below, the small mouth-opening filled by the proboscis with which it is connected by a membrane, mouth-

	parts atrophied or even wanting, the proboscis never angled at
	base; arista always bareb
	Head with a deep groove beneath, mouthparts present, proboscis
	angled at base, withdrawn in the oral groove, palpi not visible;
	arista bare or plumose. (CUTERÉBRIDÆ)d
h	Middle part of face narrow; hypopleuræ with fan of strong hairs.
υ.	( <b>Estrus</b> (Fig. 590), cosmop., in nasal cavities of sheep, ante-
	lope, etc.; Cephalomyia, S. Eur., N. Afr.; Pharyngomyia,
	Eur., in pharynx of elk; <b>Rhinœstrus</b> , Eurasia, Afr., in nasal
	Eur., in pharynx of elk; killiuestrus, Eurasia, Air., in hasai
	cavities of horse and hippopotamus) <b>ESTRÌNÆ</b>
	Middle part of face broad
c.	Middle part of face deeply sunken; hypopleuræ without hairs.
	Elephant stomach bots. (Cobbóldia Fig. 591), Rodhaino-
	mŷia, ethiop.) COBBOLDIÌNÆ
	Middle part of face forming a slightly convex plate; hypopleuræ
	with bundle of hairs. Warbles, widespread, occurring under
	skin of host. (Hypodérma, in ruminants; Œdemágena, in
	reindeer; Œstromŷia, in rodents) HYPODERATMÎNÆ
d.	No facial carina; antennal pit large and deep; antennæ elongate,
	the third joint three times as long as seconde
	Facial carina present; antennal pit small and shallow; antennæ
	short. Rodent parasites, mostly American. (Cutérebra (Fig.
	589). Bogèria, Rogenhòfera) CUTEREBRINÆ
e.	Epistoma rather broad, projecting obliquely forward and down-
	ward between sides of face; arista thickly long-plumose to tip.
	(Pseudogamètes, neotrop.) PSEUDOGAMETINÆ
	Epistoma very narrow, projecting straight downward between the
	sides of face; arista with hairs on upper side only. (Derma-
	tòbia, neotrop., under skin of mammals, including man).
	DERMATOBIÌNÆ
62.	Hypopleuræ and pteropleuræ with one or more vertical rows of
	bristles or hairs (Fig. 586); fourth vein (M <sub>1</sub> ) curving or bending
	forward, narrowing or closing first posterior (apical) cell, often
	with a spur (M <sub>2</sub> ) at the bend (Fig. 598); when three sterno-
	pleural bristles present usually but one behind. (TACHINÒI-
	DEA)
	Hypopleuræ without a vertical series of strong bristles below
	spiracle, if rarely the hypopleural bristles are present there is
	no row of bristles on pteropleura (in Stomoxys, which has a
	porrect rigid proboscis (Fig. 602), both hypopleural and ptero-
	pleural hairs are present); when three sternopleural bristles
	present usually two behind; ventral membrane usually distinct;
	postscutellum not developed convexly

63.	Postscutellum little developed, not convexly prominent (Fig. 586), if more or less prominent the metathoracic spiracular cov-
	ering is not in two parts but covers the entire lower portion,
	leaving a small opening in middle above; middle segments of
	abdomen rarely with rather strong hairs; second ventral seg-
	ment of abdomen more or less overlapping edges of the dorsal
	segments
	Postscutellum strongly developed in the form of a transverse
	rounded ridge often projecting as far as apex of scutellum
	dorsal segments of abdomen with strong bristles in addition
	to finer hairs, their edges overlapping all the ventral seg-
	ments
64.	Hindmost posthumeral bristle located lateral to the presutural
	bristle (Fig. 585) (sometimes absent in <i>Engyzops</i> ); propleura
	and prosternum hairy (bare in Pollenia (Fig. 600) which has
	matted metallic hairs on mesonotum); generally two notopleural
	bristles, rarely three; arista generally long-plumose; body usu-
	ally metallic blue or green; fifth ventral segment of male with a
	split hind margin, sometimes prominently developed; eyes of
	male touching or approximated, of female separated. Cosmo-
	politan CALLIPHÓRIDÆ
a.	Cheeks narrow, about one-fifth eye-height; arista plumose to end;
	curve of fourth vein broadly rounded, basal vein sometimes
	setose; metathoracic spiracle with its front and back ends
	equally rounded; postscutellum usually well developed. (Mes-
	embrinélla, neotrop.) MESEMBRÎNELLÎNÆ
	Cheeks subquadrate, about half the eye-height; curve of fourth
L	vein usually angulate; postscutellum not strongly developed. b
р.	Basal section of radius with distinct setulæ or hairs on posterior upper side
	Basal vein not setulose on posterior upper sided
c.	Lower calypter subtruncate at apex, concave on outer margin,
	haired on part of upper surface; the small rounded callosity
	below base of wing often with erect hairs. (Phórmia, Chry-
	somỳia, Cochliomỳia (C. (Compsomyia) macellària, Screw-
	worm), Protocalliphora (maggots on nestling birds), Pro-
	tophórmia). (CHRYSOMYIINÆ) PHORMIINÆ
	Lower calypter rather narrowly rounded at apex, nearly straight
	on outer margin, bare above; subalar callosity bare or without
	distinct hairs. (Rhínia, Metállea, Rhyncomyia, Stoma-

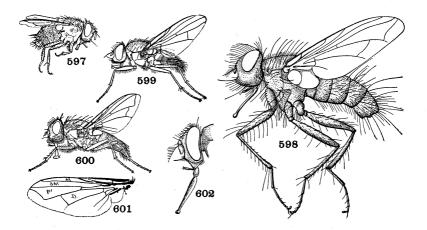


Figs. 587-596. Calliphoridæ, Æstridæ, Gastrophilidæ, Cordyluridæ

- 587. Calliphora, head from front (Walton) Calliphoridæ: Ant, antenna; Ar, arista; Ch, cheek; E, eye; FS, frontal suture; I, interfrontalia; Ocp, occiput; Pa, palpus; oc, ocellar bristles; orsi, inner row of upper orbital bristles; p. orse, proclinate bristle in outer row of upper orbitals; r. orse, reclinate bristles in outer row of upper orbitals; vte, exterior vertical bristle; vti, inner vertical bristle.
- 588. Calliphora, profile of head (Walton) Calliphoridæ. Lettering as for Fig. 587.
- 589. Cuterebra, head from front. Œstridæ.
- 590. Œstrus, head from front. Œstridæ.
- 591. Cobboldia, head from front (Rodhain and Bequaert) Œstridæ.
- 592. Gastrophilus, head from side (Cole) Gastrophilidæ.
- 593. Gastrophilus, wing (Cole) Gastrophilidæ.
- 594. Cordylura, head from front. Cordyluridæ: Lun, frontal lunule; ori, lower set of fronto-orbital bristles; ors, upper set of fronto-orbitals; p.ors, proclinate bristle of upper set; pvt, postverticals; r.ors, reclinate fronto-orbital bristles of upper set; vi, vibrissæ; vte, outer vertical bristle; vti, inner vertical bristle.
- 595. Parallelomma, female (Cole) Cordyluridæ.
- 596. Scatophaga, wing. Cordyluridæ.

- d. Prosternum and propleura hairy; hairs of parafacial stripe not reaching lower margin of eyes. Blow-flies, Blue-bottle flies. (Calliphora (Figs. 585-588), Cynomyia, Lucilia, Onèsia). CALLIPHORÌNÆ
  - Prosternum and center of propleura bare; parafacial hairs extending to lower margin of eyes. (Pollènia (P. rùdis, Clusterfly (Fig. 600); Anthracomỳia, austr.) . . . . . POLLENIÌNÆ
- 65. Calypteres large and round, reaching scutellum; fifth ventral segment of male abdomen with a straight hind margin, or entirely absent; arista generally plumose only on basal half, sometimes bare; eyes bare. Flesh-flies . . . . . . SARCOPHÁGIDÆ
  - - Head not quadrangular in profile; third and fourth sternites less evident, covering margins of tergites; theca of penis usually with spine; arista bare or with very short pubescence; two sternopleural bristles present; front often with orbital bristles b
  - b. Middle tibiæ near the middle with a single extensor bristle; cheeks rather narrow, eyes large, front of both sexes of nearly equal breadth; arista sometimes with weak pubescence; third and fourth sternites completely covered. Larvæ feeding on supplies stored in nests of various bees and wasps. (Miltogramma, Apódacra, Craticùlina, Hilarélla, Metòpia, Opsídia, Pediasiomỳia, Pachyophthálma, Senotaínia). (METOPI-ÌNÆ) ...... MILTOGRAMMATÌNÆ
  - c. Antennæ very short; genitalia large; sternites two to five large, open; eyes very small. (Paramacronýchia, Nemoræa, Eur.).

    PARAMACRONYCHIÌNÆ



Figs. 597-602. Tachinidæ, Dexiidæ, Anthomyiidæ, Muscidæ, Calliphoridæ

- 597. Belvosia. Tachinidæ.
- 598. Ptilodexia. Dexiidæ.
- 599. Limnophora (Cole) Anthomyiidæ.
- 600. Pollenia (Cole) Calliphoridæ.
- 601. Musca, wing. Muscidæ.
- 602. Stomoxys, head. Muscidæ.

66. Ventral membrane more or less evident between the reduced sternites and the margins of the tergites, if not the abdomen is elongate cylindrical (Cylindromyia = Ocyptera), or the

Ventral membrane not visible; abdomen bearing some stout bristles; facial plate flattened, at most slightly produced....67

- - Antennæ inserted above middle of eyes, the arista usually bare, rarely short-pubescent (Fig. 556d); intra-alar bristles usually extending in front of suture, if not the ventral segments broadly visible, or the fifth ventral of male is vestigial; at least two post-humeral and three posterior intra-alar bristles. Tachina flies. Larvæ mostly endoparasitic in caterpillars and other insects. (Táchina (=Larvævora), Áphria, Árchytas, Belvòsia (Fig. 597), Ernéstia, Exorísta, Frontina, Gònia (Fig. 556d), Masícera, Phorócera, Peletèria, Siphòna, Stúrmia, Winthemia, Zeníllia). (Including EXORÍSTIDÆ, MASICÉR—IDÆ, HISTRICÌIDÆ, etc.). (LARVÆVÓRIDÆ).

#### TACHÍNIDÆ!

68. Fourth vein (M<sub>1</sub>) often bending forward to narrow the apical cell at the margin (Fig. 601); if the apical cell is not narrowed then the eyes are not widely separated, or cruciate bristles are present on the front, and the lower calypter is longer than the upper, and the abdomen proper contains only five segments; if the eyes are widely separated (the females and some males) the oval,

<sup>1</sup>This dominant group has been divided by several taxonomists into about sixty so-called families, which do not have the rank of the dipterous subfamilies presented in the previous portions of the key. Because the limits of these groups have not been agreed on by the students of the muscoid flies and because there is no satisfactory published key no attempt is here made to present subdivisions of the Tachinidæ. Even the distinctions between the conventional families Dexiidæ and Tachinidæ have broken down of recent years with the discovery of annectant genera. Perhaps the best solution is to consider but two subfamilies, Dexiinæ and Tachininæ, with a unique army of legions, cohorts and tribes of genera comprising the latter. (See page 9).

BRUES AND MELANDER: CLASSIFICATION OF INSECTS	319
more or less bristly abdomen is distinctive; scutellar suture plete. (MUSCÒIDEA)	0.006 contal ypter n the pefore 0.006 $0.006$ $0.$
Front femora and tibiæ beneath at most with a single row of bris	tles.b
Front legs raptorial, their femora beneath with a double robristles, front tibiæ with one or two rows; first vein bare a (Noréllia, Plethochæta, Pycnoglóssa)NORELLI First vein usually setose above; propleural bristle present. (dylùra (Fig. 594), Acicéphala, Hexamitócera, Meg thálma, Orthochæta, Parallelómma (Fig. 595)).	ow of bove. I <b>INÆ</b> ( <b>Cor-</b>

a.

h.

# CORDYLURÎNÆ

First vein bare above; propleural bristle absent. (Scatóphaga (=Scopeùma) (Fig. 596), Acanthocnèma, Hydromyza, Microprosòpa, Pogonòta, Spathióphora, Tricopálpus). (SCO-PEUMÌNÆ) ..... SCATOPHAGÌNÆ

- 69. Either the hypopleural or pteropleural bristles or hairs present (Fig. 586); basal bristles of abdomen reduced; fourth vein bending or curving forward (straight in Eqinia); arista feathered to tip...70 Neither the hypopleural nor pteropleural hairs or bristles present;
  - abdomen usually bristly; fourth vein curving backward (if curving forward the arista is not feathered to the tip); arista
  - a. Anal vein complete, faint apically but reaching margin . . . . . . b Anal vein not distinctly traceable to margin.................c
  - b. Eyes of male close together on the narrowed front; calvpteres large. (Anthomỳia, Chortóphila, Ègle, Hammomỳia, Hylemyia (H. antíqua, Onion maggot, H. brássica, Cabbage maggot, H. cilicrùra, Seedcorn maggot, H. coarctàta, Wheat bulb fly), Hydrophòria, Opsolàsia, Pegomyia (P. hyoscyami, Beet-fly, P. rubívora, Raspberry cane maggot), Phórbia, Pycnoglóssa). (HYLEMYIINÆ, PEGOMYIINÆ).

#### ANTHOMYIÌNÆ

Eves of both sexes widely separated by the broad front; calvoteres small. (Fucéllia, Chiròsia, Mycóphaga, Myopìna). **FUCELLIÌNÆ** 

c. Lower sternopleural bristle wanting, or if present, closer to one of
the upper sternopleurals than to the otherd
Lower sternopleural equidistant from the two upper; front in
both sexes one-third the width of the head, each orbit with one
long backward-directed bristle on upper half; thorax with but
one pair of presutural dorsocentral bristles; lower stigmatal
bristle directed downward; eyes of both sexes usually widely
separated. (Cœnòsia, Chelísia, Atherígona, Limnospila,
Lispocéphala, Schœnomỳza) CŒNOSIĨNÆ
Lispocéphala, Schænomýza)
usually conspicuously dilated; front of both sexes equal to eye-
width, the interfrontalia without cruciate bristles; parafacials
with some hairs on their entire length. (Lispa)LISPINÆ
Pteropleura usually without such group of hairs, but if haired,
the front of the male is narrower than that of the female; palpi
not conspicuously dilated; parafacials bare below base of
antennæe
TT 1.11 0 1 1.1 1 1 1 1 1 1 1 1 1 1 1 1 1
Hind tibiæ of male without such a strong dorsal bristleg
f. Anal vein very short, stopping abruptly, the seventh vein (axil-
lary or A2) more or less distinctly curved forward around the
apex of the first anal; female with wholly convex front, the
broad orbits (and in males with wide front also) bearing two
fronto-orbital bristles on upper half directed outward over
eyes, or the upper one directed slightly backward; middle tibiæ
of male more or less densely pubescent and often swollen on
inner side. (Fánnia, Cœlomŷia, Piezùra) FANNIÌNÆ
Axillary vein not curving around end of anal vein; front of female
more or less projecting forward, orbits not with two upper fronto-
orbitals curving outward over eyes in either sex; middle tibiæ
of male not pubescent or swollen. ( <b>Phaònia</b> (= $Aricia$ ), <b>Allœo-</b>
stỳlus, Diályta, Hèra, Hydrotæa, Ophỳra, Pogonomỳia).
(ARICIÌNÆ, including HYDROTÆÌNÆ) PHAONIÌNÆ
g. Thorax with an uneven number of dark stripes, or unmarked;
scutellum with only the larger basal and subapical bristles; face
and oral margin usually produced; third and fourth veins paral-
lel or slightly convergent, rarely divergent; usually sparsely
short-setose. (Limnophora (Fig. 599), Hebecnema).
LIMNOPHORÎNÆ
LLIVIN OF FLORIN Æ

70. Proboscis needle-like, porrect, at rest completely ensheathed by the long slender palpi; arista strongly plumose with feathered hairs (Fig. 556e); prosternum membranous; middle coxæ separated by the forward-projecting metasternum; abdominal spiracles located in the membrane between tergites and sternites. Tsetse flies. (Glossina, ethiop.) (Fig. 556e).

#### GLOSSÍNIDÆ

Proboscis, if elongate and porrect, not ensheathed by the palpi; hairs of arista not feathered; prosternal plate developed; abdominal spiracles located in second to fifth tergites.

#### MÚSCIDÆ

**EGINIÎNÆ** 

- a. Proboscis of both sexes elongate, rigid, fitted for piercing and sucking blood, the labella not enlarged; arista with long hairs on upper side, bare or pubescent below; lower calypter rounded posteriorly, its inner basal margin well separated from the lateral basal angles of scutellum. (Stomóxys (S. cálcitrans, Stable fly (Fig. 602)), Hæmatòbia, Lyperòsia (L. irritans, Horn fly); Haphóspatha, Eur., Afr.; Hæmatobósca, China).
  STOMOXYDÌNÆ
  - Proboscis not heavily chitinized, the labella fleshy, fitted for lapping......b
- b. Apical cell widely open, the fourth vein gently curving back; hypopleural bristles present above hind coxæ; pteropleural bristles absent beneath root of wing. (**Egínia**, palæarc.).

Apical cell narrowed; hypopleural bristles nearly always absent, pteropleural bristles or hairs often present; lower calypter with posterior curvature more or less transverse, the inner basal angle very close to and often touching or underlying the basal lateral angle of scutellum. (Músca (M. doméstica, Housefly (Figs. 557, 601), Cordylòbia (C. anthropóphaga, Tumbu-fly, ethiop.), Dasýphora, Graphomýia, Mesembrina, Moréllia, Muscina, Myiospila. Pyréllia, Synthesiomýia).

# MUSCINÆ

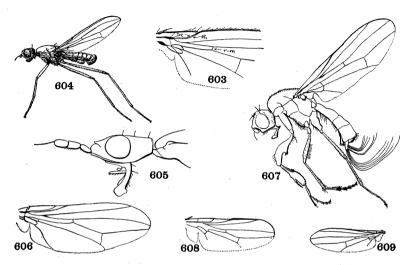
# Acalyptratæ

71. Mouth-opening small, the mouthparts vestigial; antennæ sunken in the facial grooves which form a rounded pocket, arista bare; vibrissæ and bristles absent, no sternopleural or pteropleural bristles; scutellar suture broadly interrupted at middle; ovipositor sturdy and inflexed under abdomen; costa ending at third vein (R<sub>5</sub>) which terminates much before tip of wing, the

	apical cell widely open (Fig. 593). Horse botflies, cosmopolitan. (Gastróphilus (Figs. 592, 593)) GASTROPHÍLIDÆ <sup>1</sup>
	Mouth-opening normal, the mouthparts functioning; third and
	fourth veins usually parallel or converging, very rarely markedly
	diverging; larvæ not internal parasites of horses, etc72
<i>7</i> 2.	Costa entire, no indication of a break near the end of the subcosta
	nor near the humeral crossvein; Sc nearly always distinctly
	separated from R <sub>1</sub> and ending in the costa an appreciable dis-
	tance before R <sub>1</sub> which usually terminates near or beyond middle of wing (see Fig. 608)
	of wing (see Fig. 608)
	mitted light), or if Sc is imperfect the costa is visibly broken or
	constricted before end of first vein (R <sub>1</sub> ) (Fig. 603), or at least
	with an indication of such fracture; sometimes with an addi-
	tional break near the humeral crossvein (Fig. 658)99
73.	
	rarely closely approximated to it, rarely wanting; anal cell
	present
	Subcosta incomplete, developed only at base and continuing as
	an evanescent fold, not attaining the costa; anal cell faint or
74	absent; postvertical bristles divergent
74,	Vibrissæ present at the vibrissal angle of the head, in distinction to peristomial or buccal bristles or hairs
	Definite vibrissæ absent
75.	Palpi well developed
	Palpi very small, vestigial; anterior orbital bristles never devel-
	oped; head spherical, cheeks narrow; arista bare or nearly so;
	posterior spiracle usually with at least one bristle; abdomen
	somewhat elongate and usually narrowed at base; black scav-
	enger flies. Cosmopolitan
a.	Postvertical bristles wanting; head broadened, with projecting
	eyes. (Eurychoromyia)EURYCHOROMYIINÆ
	Postverticals present and diverging; if absent, the head is not
h	broadened b First and second basal cells united. ( <b>Pandôra</b> (Fig. 608), widespr.;
υ.	Saltélliseps, ethiop., asiat.) PANDORÎNÆ
	Basal cells separate

<sup>&</sup>lt;sup>1</sup> The enigmatical family Ctenostylide, erected by Bigot for Ctenostylim from the Amazon River differs in having the arista long-plumose above, mesothorax with a short recurved bristle at each posterior angle and the marginal cell divided into six or seven small cells by thickened crossveins between the costa and the second vein. Bigot located the family near the Conopide from which it differs in the widely open apical cell and atrophied mouthparts. Apparently the only specimens known were collected previous to 1850.

c. d.	Thorax verrucose, subshining, pubescence fine, forming a sheen; dorsal abdominal segments devoid of bristles and setæ. (Toxópoda, Paratoxópoda, mainly ethiop.) TOXOPODÎNÆ Thorax not verrucose, at least mesopleuræ shining, hairs short, not forming a sheen; abdomen often with bristles
	At least mesopleural bristle presente
e.	At least mesopleural bristle presente  Front femora of male more or less excised before end and provided with varying sets of bristles, thorns or prongs; orbital bristle weak or wanting. (Sépsis (Fig. 609), cosmop.; Lasiosépsis, Eur., ethiop.)
f.	bristles but without spinigerous tuberclesf A strong orbital bristle present on each side; abdomen of both
	sexes without bristles; postocular bristle strong. (Meròplius, widespr.)
<b>7</b> 6.	Thorax convex; cheeks, pleuræ and legs not remarkably bristly;
	postvertical bristles divergent
77.	Second antennal joint usually with an angulate projection from the exterior edge (Fig. 635); interfrontal cross-bristles often present; tibiæ usually with preapical bristles; anal vein abbrevi-
	ated, not reaching margin. (See couplets 107 and 129).
	Second antennal joint without angular projection; no interfrontal
	cross-bristles; tibiæ without preapical bristles; anal vein continuing as a fold almost to margin. (See couplet 109). (Acten- óptera, (=Gymnomyza), palæarc.).
78.	NEOTTIOPHÍLIDÆ, part First posterior cell (R <sub>5</sub> ) closed or much narrowed at apex due to
.0.	the convergence of both third and fourth veins (wide in Nothybidæ, couplet 81, which have remarkably long prosternum); abdomen elongate; legs long, or very long and slender79
	First posterior cell widely open, if narrowed, the abdomen is short and the legs are not unusually long and slender



Figs. 603-609. Micropezidæ, Neriidæ, Tanypezidæ, Sepsidæ

- 603. Trichoscelis, basal portion of wing. Trichoscelidæ.
- 604. Calobata, male (Cole) Micropezidæ.
- 605. Nerius, profile of head. Neriidæ.
- 606. Tanypeza, wing. Tanypezidæ.
- 607. Themira, male. Sepsidæ.
- 608. Pandora, wing. Sepsidæ.
- 609. Sepsis, wing (Cole) Sepsidæ.
- 80. Proboscis very long and geniculate; ovipositor elongate; arista terminal. (See couplet 50). (Stylogáster, America, Afr.).

# CONÓPIDÆ, part

- 81. Ocellar and humeral bristles present, though sometimes small; prothorax small; first vein (R<sub>1</sub>) setulose; front of male narrow. (Tanypèza (Fig. 606), Polphopèza, Scipopèza, mainly neotrop.)

  TANYPÉZIDÆ

Ocellar and humeral bristles absent; thorax elongate, the prosternum prominent, the front legs attached behind middle of thorax; hind femora without bristles on posterior edge; first vein bare; front of both sexes wide. (Nothÿbus, malay.).

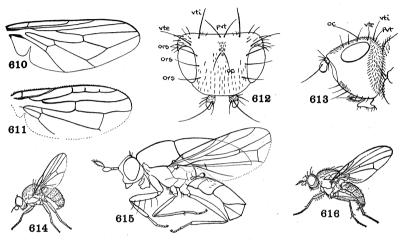
## NOTHÝBIDÆ

- 82. Arista dorsal, located toward base of the third antennal joint; front legs shorter than posterior pairs from which they are widely separated, the front coxæ short; propleuræ scarcely produced in front; second antennal joint without projection; palpi usually small. Mostly tropical . . . . . MICROPÉZIDÆ
  - a. Middle and hind tibiæ uniformly pubescent but without bristles; hind metatarsi with some setæ at base; subcosta evident.
     (Calóbata (= Trepidària) (Fig. 604), Calobatélla, Paracalóbata)
     Middle and hind tibiæ with a row of short extensor bristles; hind
    - Middle and hind tibiæ with a row of short extensor bristles; hind metatarsi without group of setæ at base; subcosta closely approximated to first vein......b
  - - Arista apical or subapical (Fig. 605); front legs longer than posterior pairs, the front coxe lengthened, thus placing the front legs close to the middle pair; propleuræ strongly developed beneath in front of front coxe; second antennal joint with a finger-like process on inside edge; palpi long. Mostly tropical. (Nèrius (Fig. 605), Macrótoma, Telostylus) ....................... NERÌIDÆ
- 83. Eyes prominently bulging, the vertex sunken; scutellum often large and grooved; femora and usually hind tibiæ greatly enlarged; anal cell rather large; prelabrum well developed......84
  Eyes less prominent, vertex not sunken; first posterior cell widely open, if rarely narrowed the femora are not thick.............85
- 84. First vein ending far beyond subcosta, first posterior cell usually narrowed apically due to the angulation of the fourth vein at apex of discal cell; posterior spiracle with a group of bristles; palpi broad. Principally neotropical. (**Rhopalomèra**, **Apophorhýnchus**, **Kræberia**, **Willistoniélla** (Fig. 610)).

### RHOPALOMÉRIDÆ

First vein ending close to subcosta, first posterior cell widely open; posterior spiracle without group of bristles; palpi narrow. (See couplet 119). (**Rhinótora** (Fig. 611), neotrop., ethiop.).

RHINOTÓRIDÆ



Figs. 610-616. Rhopalomeridæ, Rhinotoridæ, Cœlopidæ, Ochthiphilidæ, Celyphidæ, Lauxaniidæ

- 610. Willistoniella, wing. Rhopalomeridæ.
- 611. Rhinotora, wing. Rhinotoridæ.
- 612. Orygma, head from above. For lettering see Fig. 594. Cœlopidæ.
- 613. Orygma, head in profile. Cœlopidæ.
- 614. Leucopis (Cole) Ochthiphilidæ.
- 615. Celyphus. Celyphidæ.
- 616. Minettia (Cole) Lauxaniidæ.

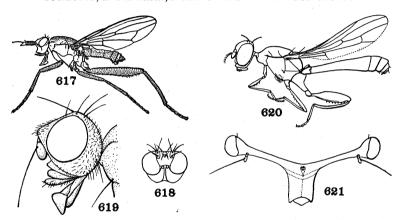
87.	Thorax convex; cheeks, pleuræ and legs not remarkably bristly last tarsal joint not flattened
	Mesonotum and scutellum flattened; head, body and legs coarsely bristly; last tarsal joint flat and enlarged. Seashore species (PHYCODRÓMIDÆ)
a.	Postvertical bristles convergent or parallel; metathoracic spiracl without bristles; propleural bristle absent. (Cœlòpa, Mala comỳza (= Phycodròmia))
	Postverticals divergent; metathoracic spiracle with group of bristles; propleural present. (See couplet 76).(Orýgma (Figs 612, 613)) ORYGMINA
88.	Postvertical bristles convergent or crossing; second antennal join with dorsal bristle; one or two sternopleural bristles and on mesopleural present; lower edge of front femora bearing bristles anal and second basal cells small, anal vein abruptly shortened (Fig. 616); two or one fronto-orbital bristles, the lower often directed inward. Many genera, mostly tropical. (Lauxània)
	Caliòpe, Camptoprosopélla, Homoneura, Minéttia (Fig
	616), Pelomỳza, Sapromỳza, Sapromyzosòma, Steganóp sis, Trigonometòpus.). (Including TRIGONOMETÓPIDÆ
	with front horizontal). $(SAPROM \acute{Y}ZID\cancel{E})$ . LAUXANÌID $\cancel{E}$
	Postverticals parallel or divergent, rarely wanting; second anten nal joint rarely with dorsal bristle; mesopleural and usuall; sternopleural bristles wanting; front femora not bristly be
00	neath; anal vein reaching margin, at least as a fold8

- - Prelabrum vestigial, rarely chitinized, not touching oral margin when the proboscis is extended; first vein ending at middle of

wing; femora setulose, their bristles developed, a characteristic bristle usually present near middle of anterior face of middle femora (Fig. 617). Many genera and species; widespread, largely holarctic. (SCIOMÝZIDÆ) ..... TETANOCERÁTIDÆ

a. Abdomen of female with seven segments, ovipositor not retractile; arista subapical; front femora not bristly. (**Tetanura**).

TETANURÎNÆ
No ovipositor; apical segments of female abdomen, beyond fifth, retractile; arista basal; front femora with bristles......b



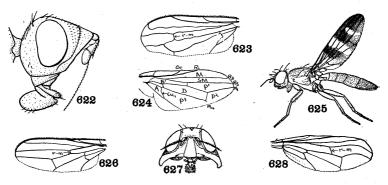
Figs. 617-621. **Tetanoceratidæ, Dryomyzidæ, Megamerinidæ, Diopsidæ** 

- 617. Sepedon (Cole) Tetanoceratidæ.
- 618. Sciomyza, head from front (Hendel) Tetanoceratidæ.
- 619. Neuroctena, head from side. Dryomyzidæ.
- 620. Syringogaster. Megamerinidæ.
- 621. Diopsis, head from front. Diopsidæ.
  - b. Propleural bristle present; front usually without differentiated median polished stripe. (Sciomyza (Fig. 618), Mélina, Pherbéllia (=Ditænia), Pterómicra)......SCIOMYZÌNÆ
     Propleural bristle wanting; front usually with a distinct polished median stripe, rarely subshining. (Tetanócera, Antichæta, Díctya, Élgiva, Hedroneùra, Hoplodíctya, Euthýcera, Límnia, Pœcilógrapha, Renócera, Sépedon (Fig. 617)).
- 90. Head produced on each side into a lateral process bearing the eye at the tip and the antennæ widely distant from each other on

a a	the eye-stalk (Fig. 621), frontal lunule flattened; subcosta weak and parallel with first vein, third vein arising from second near middle of wing, discal cell confluent with second basal; front femora more or less thickened; scutellum bituberculate. A small family, mainly tropical. ( <b>Diópsis</b> (Fig. 621), <b>Sphyracéphala</b> ).
	DIÓPSIDÆ
	Head not produced so that the eyes are stalked and the antennæ
	are distantly separated; third vein arising from second toward base of wing; scutellum not bituberculate91
91.	Anal crossvein (Cu <sub>1</sub> ) not angulate, the anal cell not acutely pro-
<i>J</i> 1.	duced (Fig. 608), first vein bare; ovipositor retractile, not promi-
	nent. (If legs and thorax are unusually elongate, see Nothybidæ,
	couplet 81)
	Anal crossvein usually angulate so that the anal cell is acutely
	produced (Fig. 624), or at least the anal cell apically angled,
	first vein usually setulose above; ovipositor chitinized and more
	or less projecting, usually flattened; post-vertical bristles di-
	verging or parallel; palpi developed. (If indomalayan, elongate
	flies with slender legs and prominent prothorax, see Phytalmidæ, couplet 100). (ORTALÍDIDÆ, s. lat.)
92.	Hind femora long, thickened and spinose beneath; basal cells
02.	lengthened, anal vein usually reaching margin; abdomen
	elongate, slender at base, clavate; postvertical bristles wanting.
	A small family; Europe, N. America, Oceanica. (Megamerina,
	(=Lissa), Syringogaster (Fig. 620), Syrittomyia).
	MEGAMERÍNIDÆ
	Hind femora rarely thickened, not biseriately spinose beneath;
	basal cells short, anal vein abbreviated; postverticals present,
റാ	though sometimes small
93.	Postvertical bristles diverging; palpi vestigial; front legs of male often more or less deformed and bristly; at least abdomen more
	or less shining. (See couplet 75) <b>SÉPSIDÆ</b>
	Postverticals converging, sometimes wanting; palpi developed;
	femora not spinose; usually densely gray-pruinose species, the
	oval abdomen usually pictured with paired brown spots. Europe
	N. America, Asia. (Ochthiphila (=Chamamyia), Acrome-
	tòpia, Leucòpis, (Fig. 614), Leucopomyia, Pseudodínia).
٠,	(CHAMÆMYÌIDÆ) OCHTHIPHÍLIDÆ
94.	The state of the s
	apically narrowed or rarely closed; body often metallic; head large, hemispherical, front broad, eyes not bulging, proboscis
	rarge, nemispherical, front broad, eyes not building, proboscis

stout. (Ulídia, Chætópsis (Fig. 625), Chrysomỳza, Euxésta (Fig. 624), Mosíllus (=Myodìna, =Seóptera), Tímia).

**ULIDÌIDÆ** 



Figs. 622-628. Pyrgotidæ, Pterocallidæ, Ulidiidæ, Richardiidæ, Phytalmiidæ

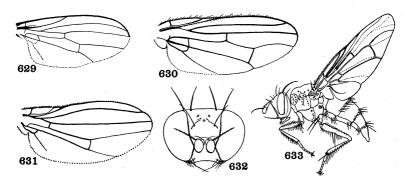
- 622. Pyrgota, profile of head. Pyrogotidæ.
- 623. Pterocalla, wing (Williston) Pterocallidæ.
- 624. Euxesta, wing. Ulidiidæ.
- 625. Chætopsis (Cole) Ulidiidæ.
- 625. Cœlometopia, wing (Williston) Richardiidæ.
- 627. Phytalmodes, head from front (Bezzi) Phytalmiidæ.
- 628. Phytalmodes, wing (Bezzi) Phytalmiidæ.

PYRGÓTIDÆ

96. Third antennal joint round or short-ovate; first vein ending much beyond subcosta; eyes rather protuberant; face vertical, excavated in middle, without antennal grooves, prelabrum small; mesonotum bristly in back only; propleural bristle weak or wanting, sternopleural bristle present; non-metallic flies.

	(Fterocana (Fig. 623), Camopistria, Dasymetopa, Myennis,
	Pseudotephritis, Stylophthalmyia) PTEROCÁLLIDÆ
	Third antennal joint with sharpened apex; antennal grooves dis-
	tinct; subcostal cell not large97
97.	Propleural and usually sternopleural bristles absent, three supra-
01.	alar bristles; mouth-opening very large, clypeus large, proboscis
	heavy, palpi broad. A large group, mainly tropical. (Including
	CEPHALIDÆ)PLATYSTOMÁTIDÆ
a.	Upper occiput usually convex; abdomen stalked, elongate; basal
а.	cells of wing diminutive; sternopleurals usually present; ant-like
	species. (Myrmecomyia, Delphínia, Myrmecothèa, Tri-
	tóxa) MYRMECOMYIÌNÆ
	Upper occiput never convex; abdomen not ant-like, if elongate no
	opper occiput never convex; abdomen not ant-like, if elongate no
1.	sternopleurals present; basal cells not smallb
υ.	Epistome convexly projecting above oral margin. (Traphera,
	Lule, Piara, Xíria) TRAPHERÌNÆ
	Epistome not convexly projecting
c.	third antennal joint much longer than wide, arista not long-
	plumose, the distal part bare; at most one weak fronto-orbital
	bristle. (Stenopterina, Antineura, Duomyia, Elassogáster,
	Lamprophthálma, Xenáspis) STENOPTERÌNÆ
	Abdomen usually elliptical or short-oval, if slender either the
	arista is feathered to apex, or antennæ shorter, or two fronto-
	orbitals present
4	Abdomen spindle-shaped, broadest at middle, or beyond middle.
u.	(Rivéllia, Cleitàmia, Ídana, Laglaisia) RIVELLIÌNÆ
	Abdomen broadly oval, widest before middle, or very small and
	narrowly joined to thorax. (Platýstoma, Achias, Euprosòpia,
	Township of the Lovenship Notice de Belte conthère
	Lamprogáster, Loxoneùra, Naùpoda, Peltacanthina, Scholástes)
	Scholastes) FLAT 1510MATINA
	Propleural and sternopleural bristles present, four supra-alar
	bristles, anterior dorsocentral bristle present; mouthparts less
	developed, cheeks broad. (Ortalis, Anacampta, Hérina,
	Dorýcera, Melièria ( $=Cer\acute{o}xys$ ), Tétanops, Tephronòta).
	(Including DORYCÉRIDÆ) ORTALÍDIDÆ
00	
98.	Discal cell complete, anterior crossvein near middle of wing, costa
	extending only to third vein (R <sub>5</sub> ), second vein (R <sub>3</sub> ) long, ending
	near tip of wing; vibrissæ absent. Europe, America, Ceylon.
	(Períscelis, Cyamops, Marbènia, Neoscutops, Podócera,
	Sphyroperíscelis (Fig. 629), Scùtops) PERISCÉLIDÆ
	Discal cell entirely wanting, anterior crossvein located near base

of wing, costa extending to fourth vein, second vein very short,
ending close to first vein; vibrissæ present. (See couplet 125).
(Astìa (= $Astèia$ ) (Fig. 645) Liomỳza, Sigaloéssa (= $Crepido$ -
$h\acute{a}mma)$ , Stenómicra) ASTÌIDÆ
99. Typically heavy-bodied flies (Fig. 633), with broad, five-segmented
abdomen and with rows of bristles on thorax, abdomen and legs
(Anthophasia with tergites fused and apical bristles alone
present on abdomen); second antennal joint as long as third or
longer, arista bare; vibrissæ present; third vein close to second
and ending much before wing-tip, costa stopping before tip of
wing, first and third veins bristly above at least at base, sub-
costa distinct, obliquely ascending at tip, anal cell prolonged
into a sharp point. Bright-colored flies, 7-18 mm. in length,
with banded wings, resembling stout tachinids. (Tachinisca
(Fig. 633), Peru, Bolivia; Anthophàsia (= Tachinastrus),
Tachiniscídia, ethiop.) TACHINÍSCIDÆ
Not large, heavy-bodied, or very bristly flies
100. Legs long and slender, thorax large, prothorax neck-like, abdo-
men long and clavate, the basal segment as long as remainder
of abdomen; first posterior cell not narrowed; one or no fronto-
orbital bristle, no postvertical, propleural, sternopleural or
dorsocentral bristles, two scutellars; arista long-plumose;
cheeks often produced as lateral processes. Indomalayan.
(Phytálmia (= $Elaphomyia$ ), Angítula, Angitulòides, Ato-
pógnathus, Giraffomỳia, Phytalmòdes (Fig. 627, 628),
Terastiom $\hat{\mathbf{y}}$ ia) PHYTALMÌIDÆ
Legs not unduly long and slender, prothorax not neck-like, body
not suggestive of the Neriidæ101
101. Costa broken only at end of subcosta (Fig. 639)102
Costa broken near humeral crossvein in addition to the subcostal
break (Fig. 658), rarely (Acartophthalmus, couplet 129), the
costa broken only at humeral crossvein
102. Subcosta complete, ending in costa, usually independent of first
radial vein (Fig. 630); second basal and anal cells complete
(except Aulacogastridæ, couplet 106 (Fig. 631), with second
basal and anal cells confluent)
Subcosta incomplete or vestigial, the apical portion represented
as a fold, not ending independently in the costa (Fig. 646)115
103. Vibrissæ present at the vibrissal angle (Fig. 635)
Vibrissæ absent, only peristomial hairs or setæ; no preapical tibia
bristle 112



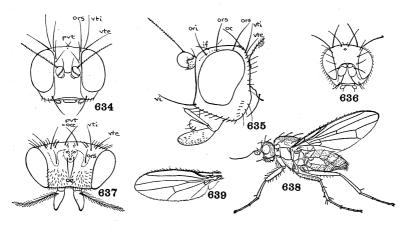
Figs. 629-633. Periscelidæ, Neottiophilidæ, Aulacogastridæ, Tachiniscidæ

- 629. Sphyroperiscelis, wing. Periscelidæ.
- 630. Neottiophilum, wing. Neottiophilidæ.
- 631. Aulacogaster, wing. Aulacogastridæ.
- 632. Aulacogaster, head from front. Aulacogastridæ.
- 633. **Tachinisca** (Kertesz) Tachiniscidæ.
- 106. Second basal and discal cells confluent, anterior crossvein located before middle of cell, costa thin but not broken near humeral crossvein; no postvertical bristles; a graded series of oral bristles in addition to the vibrissæ; tibiæ without preapical bristles. (Aulacogáster (Fig. 631, 632), holarc.).

AULACOGÁSTRIDÆ

107. Frontal orbits reaching the anterior edge of the front and bearing two to four fronto-orbital bristles (Fig. 636); second antennal joint nearly always with a triangular projection on the exterior side; preapical bristles usually present on tibiæ. (See couplet 77). (HETERONEÙRIDÆ)CLUSÌIDÆ
a. Postvertical bristles distant from each other, eyes hairy, arista short; costa broken only at humeral crossvein, Sc diverging from R <sub>1</sub> . (See couplet 129). (Acartophthálmus (Fig. 636), holarc.)
Frontal orbits shortened, fronto-orbital bristles absent, or one or two pairs present; second antennal joint without triangular projection on exterior side
109. Costa spinose, first radial vein hairy above, anal vein reaching margin; four or five sternopleural bristles present; two fronto-orbital bristles; ocellar triangle large. (See couplet 77). (Neottióphilum (Fig. 630), Eur., larvæ parasites of nestling birds).  NEOTTIOPHÍLIDÆ
Costa not spinose, first vein not hairy, anal vein abbreviated; two sternopleurals, one or no fronto-orbital. ( <b>Pióphila</b> , cosmop. ( <i>P. càsei</i> , Cheese skipper); <b>Mycetaùlus</b> , holarc.; <b>Prochylìza</b> ,
nearc.)
a. No propleural bristle; anal vein not reaching margin; the strip bearing the upper fronto-orbital bristle reaching inward from the eye-margin (Fig. 637). (Suíllia (= Helomyza, auct.) (Fig. 637), Allophŷla, Didymochæta, Pórsenus). (HELOMYZ-ÌNÆ auct.)

Propleural bristle present; anal vein nearly or quite reaching margin; the fronto-orbital strip extending only along the eyemargin. (Helomỳza (=Blepharôptera, =Lèria), Amœ. balèria, Anoróstoma, Eccoptomèra, Œcòthea (Fig. 638), Scoliocéntra, Tephrôchlamys). (LERIÌNÆ).. HELOMYZĨNÆ



Figs. 634-639. Clusiidæ, Helomyzidæ, Trichoscelidæ

- 634. Sobarocephala, head from front. Clusiidæ.
- 635. Clusia, head in profile view. Clusiidæ.
- 636. Acartophthalmus, head from front. Clusiidæ.
- 637. Suillia, head from above. Helomyzidæ.
- 638. **Œcothea** (Cole) Helomyzidæ.
- 639. Trichoscelis, wing. Trichoscelidæ.

111. All tibiæ with preapical bristles; two fronto-orbital bristles; propleural bristle present, presutural dorsocentral bristles strong; costa spinulose. (**Trichóscelis** (=Geomyza Loew, = Diástata Malloch) (Fig. 639), **Psiloplàgia**, **Spilóchroa**, **Zagònia**). (TRIXOSCÉLIDÆ, GEOMÝZIDÆ auctt., part).

TRICHOSCÉLIDÆ

Tibiæ without preapical bristles; two or three fronto-orbit	als;
propleurals absent; palpi short; costa not spinulose; grou	ınd
color yellow. Holarctic. (Chyromyia (=Chiromyia, = P	
$t\acute{o}phila$ ). Aphaniosòma)	Æ

112. Second segment of abdomen typically with lateral bristles; first radial vein bare, slightly deflected near apex to form a small stigmal area beyond the subcostal break; femora often thickened and furnished with spines; wings pictured with a few spots or clouds, anal crossvein recurved, anal cell not acutely pointed; eyes sometimes stalked. Many genera and species, mostly tropical. (Richárdia, Cœlometòpia (Fig. 626), Cóniceps, Epiplàtea, Odontomèra, Setéllia, Stenomàcra).

### RICHARDÌIDÆ

Second segment of abdomen without lateral bristles; first vein not forming a characteristic stigma; femora not thickened.....113

114. Head hemispherical in profile, eyes large and vertically semicircular, cheeks narrow, front narrow, in male one-fourth to one-fifth width of head; post-vertical bristles close together; third antennal joint more or less cylindrical; metallic black species. (See couplet 108). (Loncha (Fig. 641)).

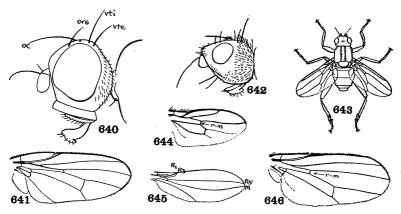
#### LONCHÆIDÆ

Head globular, eyes round, front more than one-third width of head; third antennal joint orbicular; more or less pale-colored species, wings patterned. (Pallóptera (Fig. 640)).

### PALLOPTÉRIDÆ

- 116. Hind metatarsi shortened and incrassate (Fig. 643); no sternopleural bristle, vibrissæ strong, dorsocentral bristles weak.

117. Postvertical bristles converging, presutural dorsocentral bristle present, fronto-orbital bristles directed outward, interfrontal cross-bristles usually present, one sternopleural bristle. (If



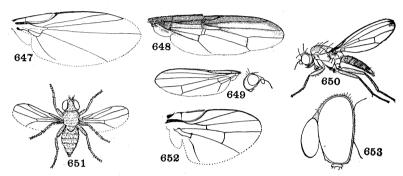
Figs. 640-646. Pallopteridæ, Lonchæidæ, Thyreophoridæ, Borboridæ, Leptoceratidæ, Astiidæ, Canaceidæ

- 640. Palloptera, head from side. Pallopteridæ.
- 641. Lonchæa, wing. Lonchæidæ.
- 642. Thyreophora, head in profile view. Thyreophoridæ.
- 643. Sphærocera, dorsal aspect (Howard) Borboridæ.
- 644. Leptocera, wing (Spuler) Leptoceratidæ.
- 645. Astia, wing. Astiidæ.
- 646. Canace, wing. Canaceidæ.

118. Preapical tibial bristles present; postocellar bristles diverging; two reclinate and one inclinate fronto-orbital bristle present;

	vibrissæ present; one presutural and three postsutural dorso-
	central bristles; terminal segments of female abdomen slender
	and retractile. (Odínia, Neoalticomèrus, Tráginops).
	ODINÎID <i>E</i> E
	Preapical tibial bristles absent, or insect otherwise disagreeing
	from preceding description119
119.	Eyes protuberant; scutellum usually tuberculate and medially
	grooved; front femora thickened; second basal and anal cells
	relatively large; at least base of anal vein firm; two reclinate
	fronto-orbital bristles, vibrissæ present, no postvertical bristles.
	Neotropical and ethiopian. (See couplet 84). (Rhinótora
	(Fig. 611)) <b>RHINOTÓRIDÆ</b>
	Eyes not bulging; scutellum not tuberculate; front femora not
	enlarged; basal cells usually small120
120.	Subcosta uniformly firm though thin, separate from first radial
	vein almost to its tip, second basal and anal cells very small,
	anal crossvein straight, anal vein indicated by a weak fold (Fig.
	646); no sternopleural bristle, postocellars diverging, three to
	five superior orbitals directed outward; ocellar triangle large,
	reaching edge of the front, antennæ well separated, the third
	joint orbicular. (See couplet 113). (If second basal cell is open,
	two fronto-orbitals present, the anterior proclinate and the
	posterior reclinate, no postverticals, but sternopleurals present,
	see Aulacogastridæ, couplet 106). (Cánace (Fig. 646), Xan-
	thocánace) CANACÈIDÆ
	Subcosta apically much weakened
121.	Distinct vibrissæ present at the vibrissal angle of the face122
	Vibrissæ absent, though bristles may occur on the middle part of
	the cheeks; anal vein firm for some distance beyond anal cell,
	except when wings lack anal angle
122,	Second basal cell present; sternopleural bristles present123
	Second basal cell open, fourth vein vestigial beyond discal cell;
	postvertical bristles diverging or parallel; pleuræ without
	bristles except the propleural; hind metatarsi not short. (Cy-
100	pselosòma, Formosa (aberrant genus)) BORBÓRIDÆ, part
123.	Postvertical (postocellar) bristles diverging; anterior fronto-
	orbital present and directed inward; seventh segment of female
	abdomen long and chitinized, not retractile; basal joint of
	arista shorter than broad. A large family, including many
	leaf-mining species; widespread. (Agromyza (A. parvicórnis,
	Corn blotch leaf-miner; A. pusilla, Serpentine leaf-miner; A.

ANTHOMÝZIDÆ



Figs. 647-653. Agromyzidæ, Psilidæ, Opomyzidæ, Chloropidæ, Cryptochætidæ

- 647. Phytomyza, wing. Agromyzidæ.
- 648. Chyliza, wing (drawn by Cole) Psilidæ.
- 649. Psila, wing and profile of head (Cole) Psilidæ.
- 650. Geomyza (Cole) Opomyzidæ.
- 651. Botanobia, dorsal aspect (Lugger) Chloropidæ.
- 652. Cryptochætum, wing. Cryptochætidæ.
- 653. Cryptochætum, head from side. Cryptochætidæ.
- 124. One presutural and two or three postsutural dorsocentral bristles present; postvertical bristles minute or absent; one sternopleural bristle. (**Opomyza**, **Anomalochèta**, **Geomyza** (=Balióptera, not Trichoscelis) (Fig. 650). (GEOMÝZIDÆ, part).

### OPOMÝZIDÆ

- a. Anal cell closed by a straight crossvein; no or one notopleural bristle; third antennal joint elongate oval to very long.....b

- b. Occiput concave; metapleural callus velvety; anal cell distinctly shorter than second basal. (Chyliza (Fig. 648). CHYLIZÎNÆ Occiput convex; metapleural callus bare; anal cell not shorter than second basal. (Psila (Fig. 649), Loxócera) . . . . . PSILÎNÆ
- 125. Ocellar triangle large (Fig. 651); arista bare, pubescent, or heavily feathered; postvertical bristles convergent or absent; second vein (R<sub>3</sub>) long, ending beyond middle of wing. A large, widespread family. (OSCINIDÆ) ..... CHLORÓPIDÆ
  - a. Costa reaching to tip of third vein, or a little beyond. (Chlòrops (= Oscinis), Chloropísca, Eurìna, Ectecéphala, Meromỳza).

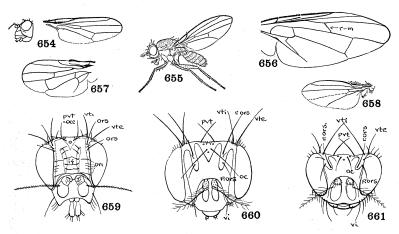
    CHLOROPÎNÆ
    - Costa reaching to tip of fourth vein. (Botanòbia (=Oscinosòma, =Óscinis, auct.) (Fig. 651), Elachíptera, Hippelàtes, Gaùrax, Notonaùlax). (OSCINOSOMÌ NÆ) ..... BOTANOBIÌNÆ
- 126. Subcosta free from first vein, ending steeply in the costal break much before the end of the first vein (Fig. 657), anal cell angular, often drawn out into an acute point, at least first vein setulose, wings usually banded or spotted; inclinate lower frontoorbital bristles present; no vibrissæ, but oral hairs developed; no preapical tibial bristles; seventh segment of female abdomen long and chitinized. A large family, including many species of fruit flies, many tropical. (EURIBÌIDÆ, TEPHRÍTIDÆ, TRYPANÈIDÆ)
  - a. Chætotaxy incomplete, the following bristles lacking, ocellar, inner occipital, postvertical, humeral, presutural, dorsocentral and sternopleural; second basal cell usually widened; antennæ elongate; sixth tergite of female short.....b
    - Chætotaxy complete, preceding bristles usually present; second basal cell not widened; antennæ usually short.................c
  - - Femora not spinose beneath; transverse suture of mesonotum in-

	terrupted in middle; abdomen ovate or clavate. (Dacus (Fig.
	656), palæarc. (D. òleæ, Olive fly); Bactrócera, indoaustr.
	(B. cucurbitæ, Melon fly); Chætodàcus, Pélmatops, indomal.;
	Leptóxyda, ethiop.; Toxotrýpana, neotrop. (T. curvicauda,
	Papaya fly)) DACINÆ
c. S	sixth abdominal tergite shorter than fifth; occipital bristles of
	hind margin of eye slender and pointed; wings banded or
	marked with brown or hyaline. (Trypèta, Acidia, Aciùra,
	Anástrepha (A. lùdens, Orange maggot), Ceratitis (C. capitàta, Mediterranean fruit-fly or Medfly; C. ròsæ, Natal fruit-
	tàta, Mediterranean fruit-fly or Medfly; C. ròsæ, Natal fruit-
	fly), Epòchra (E. canadensis, Currant fruit-fly), Euríbia
	(= Tephritis Latr. 1805, not 1804, = Uróphora), Neaspilòta,
	Platypàrea, Procecidóchares and Œdáspis, Rhagóletis (R.
	cingulàta, Cherry maggot; R. pomonélla, Apple maggot),
	Straussia, Trypànea (= Uréllia, Loew), Zonosèma). (CER-
	ATITINÆ). TRYPETÎNÆ Sixth abdominal tergite of female at least as long as fifth; occipital
,	Sixth abdominal tergite of female at least as long as hith; occipital
	bristles on hind margin of eye usually stout and blunt; wings or at least crossveins with numerous small spots. ( <b>Tephritis</b>
	Latr. 1804, Actinoptera, (= Uréllia), Ensina, Euarésta,
	Eurósta (Fig. 657) (E. solidáginis, Goldenrod gallmaker),
	Eutrèta, Ictérica, Oréllia, Teréllia) TEPHRITÌNÆ
	Dutious, 1000110us, O1011us, 101011us, 12222222222
	Subcosta ending closer to first vein, not steeply bent toward costa,
	or vestigial; lower fronto-orbital usually wanting127
127.	Subcosta complete, ending in costa independently of R <sub>1</sub> , second
141.	basal and anal cells well formed; postvertical bristles diverg-
	ing
	Subcosta incomplete, vestigial, or ending in R <sub>1</sub> , second basal and
	anal cells usually weak or absent
128.	Vibrissæ absent; anal cell more or less acute, often drawn out
	into an acute lobe, second basal cell moderately large; female
	with chitinized ovipositor. Refer to couplet 94, Ortalid series.
	Vibrissæ present; anal cell not produced
129.	Antennæ retractile into deep grooves, face retreating, eyes round
	and small (Fig. 642); scutellum very long, flattened, tipped
	with two setigerous tubercles; two vibrissæ. A small family;
	Eur., Afr., Austr. (Thyreophora (Fig. 642), Centrophle-
	bomỳia, Dasyphlebomỳia, Piophilosòma).
	THYREOPHÓRIDÆ
	Antennæ not retractile, the face flattened, vertical; subcostal
	break not evident. (Acartophthálmus, holarc. (See couplet
	break not evident. ( <b>Acatophitmannus</b> , notate. (See couplet

107)) ...... CLUSÌIDÆ, part

130. Arista lacking, antennæ inserted high on head, the apparent third joint long and leaf-like; no postvertical, orbital, vibrissal or other bristles, but body including mesopleuræ setulose; eyes large, vertical, cheeks linear; proboscis short; scutellum triangular, with sharp margin; calypteres without cilia. A small indoaustralian group, parasitic on scale insects, introduced into America. (Cryptochætum (=Lestóphonus) (Figs. 652, 653)).

CRYPTOCHÆTIDÆ



Figs. 654-661. Ephydridæ, Trypetidæ, Milichiidæ, Carnidæ, Drosophilidæ, Diastatidæ

- 654. Parydra, head in side view and wing (Cole) Ephydridæ.
- 655. Drosophila (Cole) Drosophilidæ.
- 656. Dacus, wing. Trypetidæ.
- 657. Eurosta, wing (Williston) Trypetidæ.
- 658. Meoneura, wing. Carnidæ.
- 659. Desmometopa, head from front. Milichiidæ.
- 660. Cyrtonotum, head from front. Diastatidæ.
- 661. Diastata, head from front. Diastatidæ.

Arista present; fronto-orbital bristles present (*Lipochæta*, couplet 132, a, with no arista or bristles, has small antennæ); scutellum with rounded edge.

	Postverticals convergent or parallel, rarely wanting; vibrissa present
132.	Pubescent, without bristles, scutellum margined with fine bristles cheeks broad and hairy; front hairy, produced as a subconica process over the antennæ. (Sélachops, Eur., an aberran genus)
	EPHÝDRIDA
a.	Antennæ small, widely separated, inserted in cavities; no arista eyes pubescent; bristleless, gray, seacoast flies. (Lipochæta) LIPOCHÆTÌNÆ
b.	Antennæ close together, arista developed
<b>c.</b>	parts usually small
d. 1	No sharp post-buccal ridge, the cheeks rounding into the occipute parafacials paralleling the orbits; lateral frontal bristles proclinate and reclinate
e. :	NOTIPHILÌNÆ Mouth-opening small; eyes usually pubescent. (Hydréllia, Hy- drìna (= Philýgria), Glenánthe, Axýsta) HYDRELLIÌNÆ
	Mouth-opening large; eyes without evident pubescencef Face with median area bare and facial series of bristles parallel with orbits; anterior dorsocentrals absent or undeveloped. (Napæa, Brachydeùtera, Hyadìna, Lytogáster, Párydra
	(Fig. 654), Pelina)

133. Inflexed lower fronto-orbital bristles wanting, the lowest or the

middle of the superior fronto-orbitals may be proclinate, re-
clinate, or directed outward
Inflexed lower fronto-orbitals present; interfrontal cruciate
bristles usually present
134. Second basal and anal cells lacking; no proclinate fronto-orbital
bristles
At least anal cell formed, anal vein present almost to margin;
interfrontal cruciate bristles absent; foremost or middle fronto-
orbital bristle almost always proclinate
135. Hind metatarsi not short and stout; middle tibiæ not bristly;
fronto-orbital bristles reclinate; only one row of acrostichals;
ovipositor large, broadly oval, compressed so that the lateral
margins form narrow ridges. (Pseudopomyza, Eur., aberrant
genus) MILICHÌIDÆ, part
Hind metatarsi short and thickened; middle tibiæ bristly; inter-
frontal cruciate bristles present, fronto-orbitals directed out-
ward; fourth vein continued beyond discal cell only as a fold
(Fig. 644). (Leptócera (=Limosìna) (Fig. 644)).
LEPTOCERÁTIDÆ
136. Subcosta complete, costa usually spinose; mesothorax raised
anteriorly, mesopleuræ with bristles, sternopleural bristles
present DIASTÁTIDÆ
a. Proclinate fronto-orbital bristle arising in front of reclinate ones,
both remote from eyes (Apsinota has only uppermost fronto-
orbital present); arista loosely long-plumose (Fig. 660). (Cyr-
tonòtum (=Diplocentra) (Fig. 660), Apsinòta, Parapsinòta,
Thaumastophila) CYRTONOTÎNÆ
Proclinate fronto-orbital behind the foremost reclinate pair, close
to eyes; arista short-plumose. ( <b>Diástata</b> (=Calopterélla) (Fig. 661), <b>Euthychàta</b> , <b>Tryptochàta</b> )
001), Euthychæta, Tryptochæta) DIASTATINÆ
Subcosta evanescent beyond base, costa not spinose; mesopleuræ
rarely bristly; proclinate fronto-orbital bristle not closer to the
eyes than the reclinate ones are. A large family, mostly tropical.
DROSOPHÍLIDÆ
a. Sternopleural bristle absent, mesopleuræ bristly; hind tibiæ with-
Our prespical pricties, anal cell apically open, metallic colored
out preapical bristles; anal cell apically open; metallic colored.
(Camilla, palæarc.) CAMILLÎNÆ
(Camílla, palæarc.)

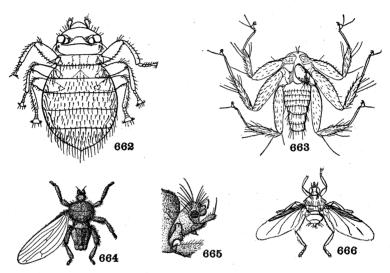
- 137. Postvertical bristles convergent; proboscis usually long and geniculate; oral hairs smaller than vibrissæ. MILICHÎIDÆ
  - - Costa not prolonged as a lappet at the subcostal break, last section of fourth vein at least three times preceding; calypteres rarely with dense cilia; mesopleuræ rarely with bristles. (Madìza, Phyllomỳza, holarc.; Desmometòpa (Fig. 659), widespr.; Aldrichiélla, Eusiphòna, Paramỳia, nearc.).

MADIZINÆ

Postverticals parallel; proboscis short; some oral bristles as strong as the vibrissæ; ocellar triangle wide. (Cárnus (= Cenchridòbia), palæarc.; Meoneùra (Fig. 658) holarc., ethiop.; Hemeromŷia (= Paramadìza) nearc.; Rhodesiélla, ethiop.). . . . CÁRNIDÆ

# Pupipara

139. Head small, capable of folding back into a groove on the flattened and expanded mesonotum; eyes if present represented at most by two facets; wingless, long-legged species, parasitic on bats. Bat-tick flies. (See couplet 169). (Nycteribia (Fig. 663), Bacília, Cyclopòdia, Penicillídia) . . . . NYCTERIBÌIDÆ



Figs. 662-666. Braulidæ, Nycteribiidæ, Streblidæ, Hippoboscidæ

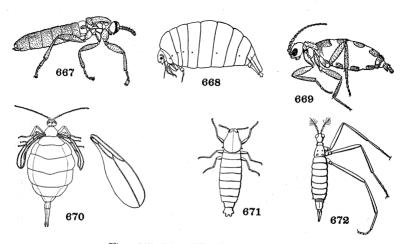
- 662. Braula, dorsal aspect. Braulidæ.
- 663. Nycteribia, dorsal aspect. Nycteribiidæ.
- 664. Trichobius, dorsal aspect (Brues) Streblidæ.
- 665. **Trichobius,** profile of head (Brues) Streblidæ.
- 666. Pseudolfersia, dorsal aspect (Lugger) Hippoboscidæ.

Head not folding back upon the mesonotum which is not strongly flattened and expanded......140

140. Palpi broader than long, projecting leaf-like in front of head; wings when present usually with distinct parallel veins and outer crossveins, at least the principal veins not crowded forward; claws simple; almost always ectoparasites of bats. Bat flies. (See couplet 170). (Strébla, Aspidóptera, Ascodíp-

	teron, Megistópoda, bius (Figs. 664, 665)).		Pterellípsis, Trichò- STRÉBLIDÆ
	Palpi slender and elonga wings if present with th (Fig. 666); tarsal claws small teeth; head and land mammals. (See co	e stronger veins o strong and often body often flatte	crowded along the costa armed with a series of ned: parasites of birds
b. At	the sheep-tick. (Melóp: Echéstypus, ethiop.; $\hat{I}NE$ )	ually not formed wings having gre vings which have hagus (M. ovinus Lipoptèna, holinista, anal cell us Ornithèca, mos Stenópteryx, C	atly reduced venation, almost disappeared in s, Sheep-tick, cosmop.); larc.). (MELOPHAG-LIPOPTENINÆ sually present. (Ornistly indoaustr.; Orniratærrhina, palæarc.) ORNITHOMYIINÆ
c. W W	ings well developed ings vestigial, anal vein	closing anal cell	d. (Allobósca, ethiop.). ALLOBOSCÌNÆ
	ronotum visible from a	espr.) above usually as	ersia, cosmop.; Icósta, OLFERSIÑÆ s a light colored ring. HIPPOBOSCÎNÆ
	Wingless Dipter	a, or with Vestig	rial Wings
142. C	Antennæ and mouthpart Completely submarine; Samoa (see couplet 151 Boring in the skin of bat abdomen undifferentia female (Fig. 676), wid	s present; body as legs vestigial.  (a))	t most pupiform142 s in other adult flies.143 (Pontomÿia, female, HIRONÓMIDÆ, part c-like, head, thorax and tures. (Ascodípteron, et 170)). (ASCODIP— STRÉBLIDÆ, part
I	more or less similar; p developed. (NEMATO Antennæ consisting of t jointed; bristles often p	alpi usually plain OCERA)hree or less joint present. (BRAC	six joints which are all nly jointed; bristles not
144. I			; mesonotum typically145

	No chitinized ovipositor; mesonotum without transverse V-
× .	shaped suture
145.	Flagellum of antennæ short, thin and long-haired. (Chiònea
	(Fig. 672), Snow-flea, apterous, holarc.; Zalùsa, Falkl. Isl.:
	Zaluscòdes, antarc.) LIMONÌIDÆ, part
	Flagellum not much thinner than basal joints of antennæ; wings
	strap-shaped. (See couplet 7). (Típula, female, widespr.).
	TIPULIDÆ, part



Figs. 667-672. Wingless Nematocera

- 667. Thripomorpha (Enderlein) Scatopsidæ.
- 668. **Dahlica** (Dahl) Mycetophilidæ.
- 669. **Pnyxia** (Schmitz) Sciaridæ.
- 670. Termitomastus, dorsal view and wing (Silvestri) Cecidomyiidæ.
- 671. Clunio (Carpenter, Bezzi) Chironomidæ.
- 672. Chionea (Johnson) Tipulidæ.

148. Claws dentate, pulvilli absent; antennal joints clavate and verti-

	cillate. (See couplet 17). (Wasmanniélla, palæarc.).
	CECIDOMYÌIDÆ, part
1.10	Claws simple
149.	Scutellum and halteres present; legs strong. (See couplet 20). (Cobóldia, myrmecophilous, nearc.; Thripomórpha (Fig. 667), palæarc.)
	Small halteres and wing-stumps present. (Austroscìara, female only, in termite nests, Austr.; Aptanógyna, Bértea, Dasyscìara, palæarc.) Halteres and wings wanting. (Epídapus, holarc.; Pnýxia (Fig. 669), Potato-scab mite, nearc.)
150	
150.	Termite guests; antennæ 14-jointed; no ocelli; wings with several veins. (See couplet 14). (Termitadélphus, Termitodípteron,
	neotrop.) PSYCHÓDIDÆ, part
151	Not termite guests
101.	constricted at thorax; halteres present. (See couplet 31).
	(Clùnio (Fig. 671), maritime, palæarc.; Eretmóptera, mari-
	time, nearc.; <b>Pontomyia</b> , male, marine, legs for swimming,
	Samoa; Bélgica, Jacobsiélla, Patagonia). (ERETMOP-
	TÉRIDÆ) CHIRONÓMIDÆ, part
	Mesothorax small, not projecting over head; abdomen broadly
	sessile with thorax; halteres and scutellum undeveloped.
	(Dáhlica (Fig. 668), palæarc.).
	Probably MYCETOPHÍLIDÆ
152.	Empodium pulvilliform, legs long; eyes and ocelli well developed;
	antennæ six-jointed, short, without arista; thorax convex; ap-
	terous. (See couplet 36). (Boreòides, Austral.).
	CHIROMÝZIDÆ, part
	Empodium not pulvilliform
153.	
	anal segments inflexed under venter; arista feathered; mouth-
	parts free; legs not compressed. (Termitoxènia (Fig. 678),
	Ind. Afr.; Termitomỳia, Afr.; Odontoxènia, Java).
	TERMITOXENÎIDÆ
154.	Not physogastrous, or if rarely so, the anus terminal

	segmented; eyes present; antennæ in grooves, the last joint with apical arista. ( <b>Thaumatóxena</b> (Fig. 673), Eur. Afr.).
	THAUMATOXÉNIDÆ
	Head not overlapping thorax
155.	Antennæ consisting apparently of one globular joint, more or less
100.	sunk in cavities on the head, arista long and thin, bare or pu-
	bescent, three-jointed; legs, especially hind pair, robust and
	compressed: mostly myrmecophilous or termitophilous females.
	(See couplet 54). (Puliciphora, Chonocéphalus, widespr.;
	Platýphora (= £nigmàtias = Oniscomyia), holarc.; Acontis-
	tóptera (Fig. 674), Commóptera, Ecitomyia, Adeloptero-
	mỳia, neotrop.; Psyllomỳia, Ænigmatístes, Ænigmato-
	pœus, ethiop.). (STETHOPÁTHIDÆ) PHÓRIDÆ, part
	Antennæ two- or three-jointed; legs never compressed 156
156.	Coxæ not separated by sternum; abdomen segmented; not para-
100.	sitic on warm-blooded animals or on the honey-bee157
	Coxæ separated by sternum; segmentation of abdomen some-
	times obscure; parasitic on birds, mammals, or on the hive-
	bee
157.	No frontal lunule or suture immediately over antennæ; third
	antennal joint more or less tapering
	A Ω-shaped frontal suture over the antennæ; third antennal joint
	more or less ovate, with dorsal arista; palpi at most one-
	jointed
158.	Arista or style usually terminal, if dorsal (Thinodromia) the body
	is heavily gray pollinose. (See couplet 58). (Ariasélla, Dus-
	metina, Pieltània, S. Europe; Thinodròmia, maritime,
	Pacific Coast of N. Am.) EMPÍDIDÆ, part
	Arista dorsal, body metallic (Emperoptera), or brown (Schano-
	philus). (See couplet 58). (Emperóptera (Fig. 675), Hawaii;
	Schenophilus, Antarctic) DOLICHOPÓDIDÆ, part
159.	Hind metatarsi shorter than following joint. (See couplet 116).
	(Aptilòtus, holarc.; Apterina, palæarc.; Ántrops, Anata-
	lánta, Siphlópteryx, Antarct.) BORBÓRIDÆ, part
	Hind metatarsi longer than following joint
160.	
	Chamæbósca, Antarc.) EPHÝDRIDÆ, part
	Mouth-opening normal
161.	Arista loosely plumose. (See couplet 136). (Drosophila, in vesti-
	gial-winged or apterous condition) DROSOPHÍLIDÆ, part
	A minta muhagaant on hana

Falkland Isl.) DRYOMÝZIDÆ, part
Legs not stout or woolly pubescent
163. Mesonotum with well developed bristles on anterior por-
tion
Mesonotum without strong dorsocentral bristles165
674
673
678
675 676 / 677
TI 070 070 TI
Figs. 673–678. Wingless Diptera
673. <b>Thaumatoxena</b> (Trägårdh) Thaumatoxenidæ.
674. Acontistoptera (Brues), dorsal and side views. Phoridæ.
675. Emperoptera, dorsal view (Grimshaw) Dolichopodidæ.
676. Ascodipteron, degenerate female (Adensamer) Streblidæ.
677. Anatalanta (Enderlein, Bezzi) Borboridæ.
678. <b>Termitoxenia</b> , side view (Bugnion) Termitoxeniidæ.

166. Head without bristles, ocellar triangle large. (See couplet 125). (Alómbus, Afr.; Myrmecomórpha, Eur.).

# CHLORÓPIDÆ, part

- Head with vertical and frontal bristles, ocellar triangle not large. 167
  Wings days long a complex but brooking off at base. (See some

wanting or vestigial. (See couplet 140). Bat parasites. (Ascodipteron, male, wings normal but dehiscent at base. (See couplet 142 for female), widespr.; Aspidóptera, Megistópoda, Metelásmus, Paradyschíria, Pterellípsis).

STRÉBLIDÆ, part

# KEY TO THE LARVAE OF THE PRINCIPAL FAMILIES OF DIPTERA<sup>1</sup>

1. Head complete (Figs. 679-704), or the posterior portion with deep longitudinal incisions; mandibles moving horizontally, fitted for chewing; more than three larval stages: body consisting of

<sup>&</sup>lt;sup>1</sup> Based on Malloch: Bull. Ill, State Lab. Nat, Hist., 12, art, 3 (1917).

	acles. (NEMATÓCERA)
	Head incomplete, without a strongly developed upper arcuate
	plate (Figs. 705–737); mandibles moving vertically, mouth-
	parts essentially sucking: body comprising fewer than thirteen
	segments and only exceptionally with as many as nine pairs of
	spiracles. (BRACHÝCERA)
2.	Mandibles normally sickle-shaped, not protruding much beyond
	apices of the well-developed maxillæ, often much shorter, maxil-
	lary palpi distinct (Fig. 686); antennæ well developed, situated
	on the upper surface of a slightly arcuate chitinized dorsal plate;
	no free pharyngeal skeleton within the head capsule, the exo-
	skeleton of the head at least dorsally indicated. (ORTHÓR-
	RHAPHA)
	Mandibles short and hook-like, usually capable of protrusion much
	beyond the poorly developed maxillæ, palpi rarely visible; an-
	tennæ poorly developed or absent, when present situated on a
	membranous surface; a free pharyngeal skeleton present; head
	very poorly developed, completely unchitinized dorsally.
	(CYCLÓRRHAPHA)
3.	Head incomplete, consisting of two parts, the anterior more or less
	chitinized above, the posterior unchitinized and bearing the
	eye spots; thorax and abdomen comprising thirteen segments;
	usually with a "wishbone" or chitin plate on under side of
	second thoracic segment, at least in mature larvæ; larvæ peri-
	pneustic, <i>i.e.</i> with series of lateral spiracles; mandibles apparently absent; antennæ long, two- or three-jointed. Usually
	feeding on plant sap, many larvæ gall makers, some feed on
	aphids, scale insects, mites, etc. Very small insects. (Fig. 679,
	Retinodiplòsis)
	Without such combination of characters4
4.	Abdominal segments with deep constrictions between them (Fig.
	682), venter with series of sucking disks. Clinging to rocks in
	swift-running streams
	Abdomen without a series of large sucking disks for attachment to
	rocks in swift-running water
5.	Ventral sucking disks median, one on the anterior complex seg-
	ment and usually one on each of the five following segments;
	head, thorax and first two abdominal segments fused to form a
	rather small first segment; mandibles three-toothed. (Fig. 682,
	Bibiocéphala) BLEPHAROCERÁTIDÆ

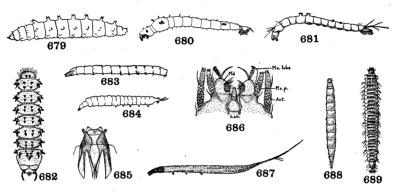
Ventral sucking disks lateral, placed on the prolongations of the abdominal segments; thoracic segments distinct; mandibles deeply incised, the outer arm fringed with a comb of long teeth.

### DEUTEROPHLEBÌIDÆ

- 6. Head incomplete behind, either with three deep wedge-shaped slits, two on dorsum and one on ventral side (Fig. 685), or the ventral surface very poorly chitinized and the dorsal posteriorly in the form of four slender heavily chitinized rods, with a weakly chitinized divided plate on anterior half of dorsum. 7 Head capsule complete, *i.e.* at least bounded above behind and
- - End of abdomen with at most five radiating processes as well as single or double breathing tubes; if six apical processes are present the labium is subdivided centrally; body usually without regularly placed bristles, frequently with dense surface pilosity; head more or less retracted, sometimes weakly chitinized and without distinct labium; antennæ sometimes short and slender and not as long as maxillary lobe; mandibles never with only two teeth. Mostly aquatic, or living in wet ground, sometimes carnivorous, some species are terrestrial and green from feeding on leaves. (Figs. 683, 685, Limònia; Fig. 684, Dicranòta)

The three thoracic segments separate......9

9. A many-toothed chitinized mentum or labial plate (Fig. 694) present on the underside of the head, which is divided into sub-



Figs. 679-689. Nematocerous Larvæ

- 679. Retinodiplosis, side view (Malloch) Cecidomyiidæ.
- 680. Sayomyia, side view (Malloch) Culicidæ.
- 681. Dixa, side view (Malloch) Dixidæ.
- 682. Bibiocephala, dorsal view (Malloch) Blepharoceratidæ.
- 683. Limonia, side view (Malloch) Limoniidæ.
- 684. Dicranota, side view (Malloch) Limoniidæ.
- 685. Limonia, head capsule (Malloch) Limoniidæ.
- 686. Dixa, anterior extremity (Malloch) Dixidæ.
- 687. Bittacomorpha, side view (Hart) Ptychopteridæ.
- 688. **Psychoda,** dorsal view (Malloch) Psychodidæ.
- 689. Psychoda, dorsal view (Malloch) Psychodidæ.
- 10. Posterior portion of abdomen swollen (Fig. 985), the last segment ventrally armed with a sucking disk which bears concentric series of bristles by means of which the larvæ hold on to rocks in the swiftly running streams in which they live; mouth on each side with a large jointed maxilla which bears a fan-like arrangement of long hairs; thorax with a pair of closely fused pseudopods; minute spiracles opening on all abdominal segments, i.e. larvæ peripneustic. (Fig. 985, Simulium).

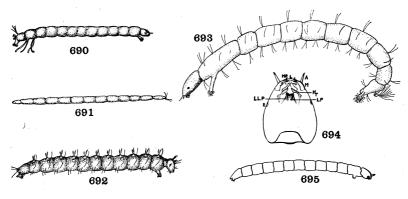
SIMULÌIDÆ

- Body not club-shaped and fitted with terminal sucking disk, nor with large fan-like oral appendages (in Dixidæ the maxillæ bear a small fan); larvæ metapneustic, with only the last pair of spiracles, or amphipneustic, with only the first and last pairs. .11
- 12. Last abdominal segments retractile, terminating in a long slender respiratory tube, ventral pseudopods present and equipped with bristles: aquatic larvæ. (Fig. 687, **Bittacomórpha**).

# **PTYCHOPTÉRIDÆ**

- Mentum not projecting in the form of a cutter; labrum fixed. .15
  15. Either very slender, tapering toward each end, without thoracic or anal pseudopods or surface hairs (Fig. 691), or stouter, with well defined segments which are armed with strong bristles, some of which are lanceolate (Fig. 692); one pair of prothoracic pseudopods present, a single anal pseudopod present or not. Terrestrial, living in manure or under bark, etc., or in water. (Fig. 692, Forcipomyia; Fig. 691, Palpomyia).

**CERATOPOGÓNIDÆ** 



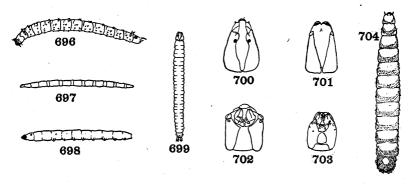
Figs. 690-695. Nematocerous Diptera, Larvæ

- 690. Chironomus, side view (Malloch) Chironomidæ.
- 691. **Palpomyia**, side view (Malloch) Ceratopogonidæ.
- 692. Forcipomyia, side view (Malloch) Ceratopogonidæ.
- 693. **Tanypus**, side view (Brues) Chironomidæ.
- 694. **Protenthes,** ventral view of head (Malloch) Chironomidæ.
- 695. Camptocladius, side view (Malloch) Chironomidæ.

18.	Anal spiracles at the apices of long stalk-like process (Fig. 704); no false segment immediately behind the head. Living in decaying vegetation, rotting fruits, excrement, or under old bark. (Fig. 704, Scatópse) SCATÓPSIDÆ
	Anal spiracles sessile, metathoracic spiracles present; a fully de-
	veloped false segment behind the head armed dorsally with
	spinose processes (Fig. 696). Larvæ principally scavengers living
	in soil, sometimes prevalent about dung, sometimes destructive
	to root crops. (Fig. 696, Bíbio) BIBIÓNIDÆ
19.	Dorsal or clypeal sclerite of head (præfrons) not distinctly taper-
10.	ing behind; antennæ two-jointed <b>BOLITOPHÍLIDÆ</b>
	Dorsal or clypeal sclerite of head conspicuously tapering pos-
	teriorly; antennæ almost indistinguishable
20.	Lateral plates of head meeting on ventral line only for a short
20.	space immediately behind the mouth-opening, then widely
	diverging and not connected at posterior margin; maxillæ and
	mandibles apparently grown together to form a pair of many-
	toothed rasping organs. (Fig. 698, <b>Exèchia</b> ; Fig. 701, <b>Lèia</b> ).
	MYCETOPHILIDÆ
	Lateral plates of head connected for a short space behind the
	mouth-opening and again near posterior margin; maxillæ and
	mandibles distinctly separated from each other, the mandibles
	apically tridentate
21.	
	capsule; body segmentation distinct, especially on venter where
	the outline is moniliform; armed with spines which have two
	or three points
	Clypeal sclerite shorter, not attaining the occipital margin of the
	head-capsule; segmentation distinct but not moniliform; spines
	simple. Living in soil, essentially as scavengers. (Fig. 703,
	Sciara) SCIÁRIDÆ
22.	
	body flattened, each abdominal segment with secondary in-
	cisures, last segment with four fleshy lobes surrounding the
	spiracles. Living in humus soil and under leaves. (Fig. 699,
	Trichócera) TRICHOCERÁTIDÆ
	Clypeal sclerite not tapering behind to a point
23.	Antennæ undeveloped, appearing as pale round spots on sides of
	head; ventral surface of head with sclerites contiguous an-
	teriorly, widely separated posteriorly. Living in slimy webs on
	fungi. (Fig. 702, Ceróplatus) CEROPLÁTIDÆ

Antennæ pedunculate, usually well developed; ventral surface of head with sclerites contiguous on entire length, not separated widely posteriorly; body slender, tapering, abdominal segments each with a single constriction near anterior margin......24

24. Apical segment with five short but distinct processes around the spiracular disk; mandibles apically rounded and furnished with



Figs. 696-704. Nematocerous Diptera, Larvæ

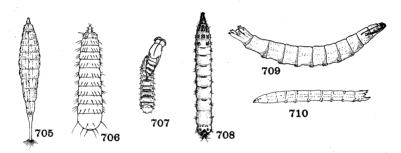
- 696. Bibio, side view (Malloch) Bibionidæ.
- 697. Mycetobia, side view (Malloch) Mycetobiidæ.
- 698. Exechia, side view (Malloch) Mycetophilidæ.
- 699. Trichocera, dorsal view (Malloch) Trichoceratidæ.
- 700. Anisopus, head (Malloch) Anisopodidæ.
- 701. Leia, head (Malloch) Mycetophilidæ.
- 702. Ceroplatus, head, ventral view (Malloch) Ceroplatidæ.
- 703. Sciara, head, ventral view (Malloch) Sciaridæ.
- 704. Scatopse, dorsal view (Malloch) Scatopsidæ.

dense rows of long bristles; the thin ventral chitinous bridge posteriorly connecting the lateral plates not broken. Scavengers, occurring in decaying vegetable matter, manure, and sometimes in sewage. (Fig. 700, Anisopus)

#### ANISOPÓDIDÆ

# Brachycera Orthorrhapha

25.	Posterior spiracles approximated, situated within a terminal or subterminal cleft or chamber, usually concealed, or with a
	terminal breathing tube; body usually shagreened or wholly
	or in part longitudinally striated
	Posterior spiracles rather widely separated, visible, situated on
	apical segment, which may be truncated, chitinized, or armed
	with apical processes, or upon penultimate or antepenulti-
	mate segment; body not shagreened or visibly striated28
26.	Head not retractile, bristly, body 11-segmented, flattened, surface
	finely shagreened, without pseudopods; spiracular fissure trans-
	verse, sometimes rather small; pupæ enclosed in larval skin27
	Head retractile; body 12-segmented, cylindrical, not shagreened,
	usually longitudinally striated, abdomen with a girdle of pseu-
	dopods on each segment; spiracular fissure vertical; pupæ free.
	Usually aquatic, or semiaquatic; predaceous TABÁNIDÆ
27.	Peripneustic, i.e. with lateral abdominal spiracles on most seg-
	ments. Aquatic, or living in semiliquid matter, under bark, in
	manure, or on rotting vegetation. (Fig. 705, Stratiomÿia;
	Fig. 706, Hermètia) STRATIOMYÌIDÆ
	Amphipneustic, i.e. only prothoracic and last abdominal spiracles
	present. Living under bark. (Fig. 707, <b>Xylomỳia</b> ).
	XYLOMYÌIDÆ
28.	Posterior spiracles situated on apical segment
	Posterior spiracles situated on penultimate or antepenultimate
	segment
29.	Projecting portion of head and the flattened apical plate of last
	abdominal segment heavily chitinized, the former cone-shaped,
	entirely closed except at extreme apex, not retractile; apical
	abdominal segment obliquely truncate and with projecting
	processes
	Projecting portion of head more or less retractile, not pointed
	cone-shaped, the movable parts not enclosed; last abdominal
90	segment without a heavily chitinized flattened terminal plate. 31
<b>3</b> U.	Head about twice as long as its greatest width; thoracic segments
	not chitinized above, each with two internal separated chitinized plates wholes without large hoises exist plates your large.
	ized plates; body without long hairs; apical plate very large spiracles vertically elongated, apical paired protuberances
	spiracies vertically elongated, apical paired protuberances small, widely separated, each with a short hair on inner side.
	T 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1
	Living in soil or decaying wood. (Fig. 711, Cœnomyia).



Figs. 705-710. Orthorrhaphous Diptera, Larvæ

- 705. Stratiomyia, dorsal view (Johannsen) Stratiomyiidæ.
- 706. Hermetia, dorsal view (Malloch) Stratiomyiidæ.
- 707. **Xylomyia**, emergence showing withdrawal of pupa from puparium (Greene) Xylomyiidæ.
- 708. **Xylophagus,** dorsal view (Greene) Xylophagidæ.
- 709. Aphrosylus, side view (Wheeler) Dolichopodidæ.
- 710. Dolichopus, side view (Malloch) Dolichopodidæ.
- - Labial plates and rods meeting angularly, grown together in front V-like, in profile appearing bent and usually united behind with the tentorial rods; head mostly membranous, even above to the triangular dorsal plate; usually amphipneustic. Predatory,

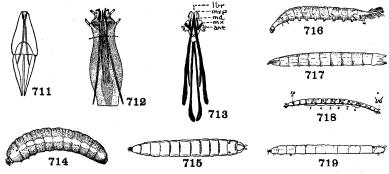
some in decaying vegetable matter.

mostly living in moist earth, some aquatic, some under bark,

	Fig. 713, Drápetis EMPÍDIDÆ
	Fig. 709, Aphrosylus; Fig. 710, Dolichopus.
	<b>DOLICHOPÓDIDÆ</b>
33.	Head capsule long, in large part internal, reaching back to the mesothorax, dorsal plate very long, pear-shaped, broader be-
	hind; hind spiracles ending separately in two plates or tracheal
	gills. (Fig. 716, Athèrix; Figs. 712 and 717, Chrysopìla).
	RHAGIÓNIDÆ
	Head capsule short, the dorsal plate not covering the part inset in
	the thorax, the internal part flat or divided into rods; apical
	abdominal segment without projecting processes, the spiracles
	very small; endoparasites of spiders ACROCERÁTIDÆ
34.	Posterior spiracles situated on the antepenultimate segment, ab-
	dominal segments one to six subdivided, the body apparently
	consisting of twenty segments exclusive of the head; head free,
	slightly longer than broad and not set into the thorax35
	Posterior spiracles situated on penultimate segment; abdominal
	segments simple, the body apparently consisting of eleven or
25	twelve segments exclusive of the head
55.	metacephalic rods lying in prothorax and mesothorax) spatu-
	late at apex; ventral posterior projections in the form of two
	short chitinized rods. Living in soil and decaying wood, preda-
	tory, locomotion serpentine, assisted by the mouthparts. (Fig.
	718, Psilocéphala) THERÉVIDÆ
	Posterior dorsal internal extension of head not spatulate at apex;
	ventral posterior projections absent. Found in fungi, rotten
	wood and in houses, in carpets and furniture; predatory.
	SCENOPÍNIDÆ
36.	Penultimate abdominal segment longer than the ultimate, with a
	deep transverse depression near its apex giving it the appearance
	of two distinct segments; ultimate segment terminating in a
	sharp ridge with a median sharp point, on each side of which
	dorsally and ventrally are situated four very closely approxi-
	mated hairs similar to those of Asilidæ. Living in decaying
	wood, predatory on beetle grubs. (Fig. 715, <b>Mỳdas</b> ).
	MYDÀIDÆ
	Penultimate abdominal segment shorter than ultimate, or if longer
	without a deep transverse depression; apical segment not as above, the hairs not closely approximated
	approximation of the state of t

37. Thoracic segments each with two long hairs, one on each side on ventro-lateral margin; apical segment with six or eight long hairs; head well developed, porrect and more or less coneshaped when viewed from above, appearing flattened when viewed from the side; penultimate segment usually shorter than ultimate or not much longer; body held straight. Living in soil and decaying wood, predaceous. (Fig. 719, **Prómachus**).

ASÍLIDÆ



Figs. 711-719. Orthorrhaphous Diptera, Larvæ

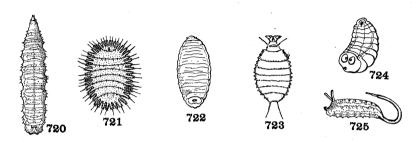
- 711. **Cœnomyia,** cephalopharyngeal skeleton (Malloch) Cœnomyiidæ.
- 712. Chrysopila, cephalopharyngeal skeleton (Malloch) Rhagionidæ.
- 713. **Drapetis**, internal skeleton (Malloch): ant, antenna; lbr, labrum; md, mandible; mx, maxilla; mxp, maxillary palpus. Empididæ.
- 714. **Sparnopolius,** side view (Malloch) Bombyliidæ.
- 715. Mydas, dorsal view (Malloch) Mydaidæ.
- 716. Atherix, side view (Greene) Rhagionidæ.
- 717. Chrysopila, side view (Malloch) Rhagionidæ.
- 718. Psilocephala, side view (Malloch) Therevidæ.
- 719. Promachus, side view (Malloch) Asilidæ.

# Cyclorrhapha

38.	Maxillæ and palpi present, near the very small bent mandibles; body comprising only nine distinct segments, the head and prothoracic and the metathoracic and first abdominal segments appearing grown together; much flattened, seed-like, scarcely twice as long as broad; head bearing underneath on each side a triangular flap, retractile into the thorax; frontal sac opening directly into the upper surface of the head; incisures chitinized above, side margins with many incisions, venter membranous, apical segment with two filiform appendages. Living in mud and about decaying organic matter. (Fig. 723, Lonchóptera).
	Maxillæ not developed, only the mouth-hooks present; frontal sac
	opening in an atrium in the mouth cavity
39.	Upper lip projecting forward between and below the two mouth-
	hooks, distally hooked or toothed, the mouth-hooks porrect,
	single-pointed or more often toothed; body with eleven in-
	cisures, lengthened, anteriorly tapering, posteriorly blunt or
	the eighth abdominal segment tube-like; metapneustic, i.e.
	terminal spiracles alone present. Living in many situations, in
	dung, carrion, ant nests, etc PHÓRIDÆ
	Mentum never extending between the two mouth-hooks as a point
	or hook except in Aphis-eating Syrphidæ, in which case the
	lateral rods of the mentum are produced forward over the
	mentum and fuse to form a projecting pointed upper lip; mostly
	amphipneustic, i.e. anterior pair as well as terminal spiracles
	present
40.	Larva broad and depressed, in dorsal aspect appearing to have
	only nine or ten segments because the head and prothorax are
	inflexed to the under surface, the sides of the body with long
	projecting bristles and sometimes deeply incised and serrate;
	posterior spiracles not or but little prominent, rather widely
	separated, placed at the base of the last segment, prothoracic
	spiracles prominent. Developing in fungi. (Fig. 721, Calli-
	mŷia) PLATYPÉZIDÆ
	Of other conformation
41.	r
	on a common chitin plate some distance before the tip of the
	body; acephalous, mouthparts very indistinct, the mouth-hooks
	sessile; body comprising ten or eleven segments. Endoparasitic
	on Homoptera. (Fig. 722, Pipúnculus) PIPUNCÙLIDÆ

If the posterior spiracles are located on a common chitin plate they are contiguous to each other......42

42. Posterior spiracles close together at the end of a tracheal tube of greater or less length, which is sometimes chitinized and sometimes greatly extensile; mouthparts various, absent in "rattail" larvæ (Fig. 725), developed in the species living in wood, and in the "Aphis-lions" (Syrphus, etc.) the mouthparts are



Figs. 720-725. Aschizous Diptera, Larvæ

- 720. Syrphus, dorsal view (Metcalf) Syrphidæ.
- 721. Callimyia, dorsal view (de Meijere) Platypezidæ.
- 722. Pipunculus, dorsal view (Perkins) Pipunculidæ.
- 723. Lonchoptera, dorsal view (de Meijere) Lonchopteridæ.
- 724. Conops (Brauer) Conopidæ.
- 725. Eristalis, side view (Metcalf) Syrphidæ.

specialized into piercing organs; larvæ apparently acephalous, body with about eleven obscure segments, skin usually rough, form diverse, conical, more or less cylindrical, even hemispherical. Food habits various, under bark, in tuber-roots, about sewage, in ant or bee nests, or free-living and preying on aphids. (Fig. 720, Sýrphus; Fig. 725, Erístalis).

SÝRPHIDÆ

44.	Parasites of insects
	Parasites of mammals, turtles, frogs, etc
45.	Body oval or pyriform, with distinct segments; antennæ wart-like,
	tipped with a chitinous ocellus-like ring; posterior spiracular
	plates large, round or reniform. Living within the abdomen
	of wasps or bees. (Fig. 724, Conops) CONÓPIDÆ
	Body more or less elongate, segmentation less evident
46.	Posterior end of body rather truncate or broadly rounded, without
	long processes
	Posterior end of body terminating in two slender tail-like proc-
	esses, two to five times length of body; young larva hairy on
	posterior half of body. Living within Cottony cushion scale.
	(Fig. 736, Cryptochætum) CRYPTOCHÆTIDÆ
47.	Posterior spiracular plates with a button. Endoparasites of
	various insects in their early stages TACHÍNIDÆ
	Posterior spiracular plates without a button. (Figs. 731 and
	732 D, Sarcóphaga) SARCOPHÁGIDÆ
48.	Body not strongly tapering, often obese, usually provided with
	numerous strong chitinous spines49
	Body strongly tapering in front (Fig. 731), at most with girdles of
	minute spines. (Figs. 731 and 732 D, Sarcophaga).
	SARCOPHÁGIDÆ
49.	Warbles living under skin of various ungulates, rodents, dogs and
	man, or in nasal sinuses or throat of various animals; body with
	strong spines or not. (Fig. 727, Cutérebra) ŒSTRIDÆ
	Bot parasites in the stomach and intestines, principally of horses.
F0	(Fig. 726, Gastrophilus) GASTROPHÍLIDÆ
ου.	Body with lateral and dorsal spinose processes. (Fig. 728, Homal-
	omŷia) ANTHOMYÌIDÆ, part Body without such spinose processes
51	Body truncate or broadly rounded posteriorly
υı.	Body with one or two posterior processes; rather small species57
52	Only one mandibular hook; posterior spiracular plates with sin-
02.	uous slits (Fig. 732 A); no distinct fusiform pads on the side
	margins of the segments; posterior end of body with few if
	any conical processes, anterior end very slender. (Fig. 730,
	Músca)
	With two mandibular hooks; slits of posterior spiracular plates
	r
	not sinuous
53.	not sinuous
53.	not sinuous

	No tubercles above anal area; no distinct processes around spi-
	racular field
54.	Slits in spiracular plates rather short and arranged radially55
	Slits slender, subparallel to each other56
55.	Two tubercles above anal area; posterior spiracular field with
	distinct processes around it. (Fig. 729, Pegomỳia).
	ANTHOMVIDE part

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Figs. 726-732. Cyclorrhaphous Diptera, Larvæ

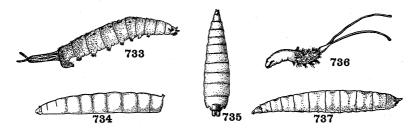
- 726. Gastrophilus, dorsal view (Hadwen and Cameron) Gastrophilidæ.
- 727. Cuterebra, ventral view (Brauer) Œstridæ.
- 728. Homalomyia, dorsal view (Banks) Anthomyiidæ.
- 729. Pegomyia, side view (Frost) Anthomyiidæ.
- 730. Musca, side view (Banks) Muscidæ.
- 731. Sarcophaga, side view (Greene) Sarcophagidæ.
- 732. Left posterior spiracle (Banks): A, Musca, Muscidæ; B, Muscina, Muscidæ; C, Calliphora, Calliphoridæ; D, Sarcophaga, Sarcophagidæ; E, Tritoxa, Ortalidæ.

Four or more tubercles above anal area; slits of spiracular plate usually pointed at inner end. (Fig. 732 B, Muscina).

MÚSCIDÆ, part

### SARCOPHÁGIDÆ

57. Posterior end of body with respiratory and usually forked anal



Figs. 733-737. Acalyptrate Diptera, Larvæ

- 733. Ephydra, side view (Jones) Ephydridæ.
- 734. Agromyza, side view (Frost) Agromyzidæ.
- 735. Drosophila, dorsal view (Banks) Drosophilidæ.
- 736. Cryptochætum, side view (Smith and Compere) Cryptochætidæ.
- 737. Scaptomyza, side view (Frost) Drosophilidæ.

# KEY TO THE PUPAE OF THE PRINCIPAL FAMILIES OF DIPTERA <sup>1</sup>

<sup>&</sup>lt;sup>1</sup> Based largely on Malloch: Bull, Ill. State Lab. Nat. Hist., 12, art. 3 (1917).

Pupa coarctate, *i.e.* enclosed within the indurated last larval moult, and usually barrel-shaped and brown in color; head always retracted, the chitinous portion occupying a position on the inner side of the ventral surface of the puparium; anterior respiratory organs distinct, either protruded from the antero-lateral angles of the head extremity or from dorsum of base of abdomen; adult emerging usually by forcing off the rounded anterior extremity of the puparium in cap-like form (see Fig. 763), or the dorsal half of the thoracic portion — the lines of cleavage being along the lateral margins to a point at base of abdomen; rarely emergence is through rectangular splitting of the dorsum of the puparium (Figs. 760–764).

## CYCLORRHAPHA 1

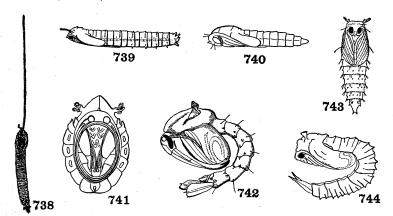
- - Antennæ shorter, projecting downward and outward, not curving over the eyes nor reaching nearly to base of wing; head usually with strong thorns or horns; thoracic respiratory organs sessile, rarely stalk-like; abdomen usually armed with strong spines or bristles, or if unarmed there are only four or five distinct pairs of abdominal segments. (ORTHÓRRHAPHA).............26

- strong thorns or leaf-like elevations, or the legs are recurved

<sup>&</sup>lt;sup>1</sup> The pupe of this large and important group are so imperfectly known that it is impossible to present a key to the families.

Legs short, apices of hind tarsi projecting slightly beyond apices of wings; antennæ short, curved across middle of eye. (Fig. 747, Bíbio)

BIBIÓNIDÆ



Figs. 738-744. Nematocerous Diptera, Pupæ

- 738. Bittacomorpha, dorsal view (Hart) Ptychopteridæ.
- 739. Pachyrrhina, side view (Malloch) Tipulidæ.
- 740. Monardia, side view (Malloch) Cecidomyiidæ.
- 741. Deuterophlebia, ventral view (Pulikovsky) Deuterophlebiidæ.
- 742. Anopheles, side view (Howard) Culicidæ.
- 743. Psychoda, ventral view (Malloch) Psychodidæ.
- 744. Dixa, side view (Malloch) Dixidæ.

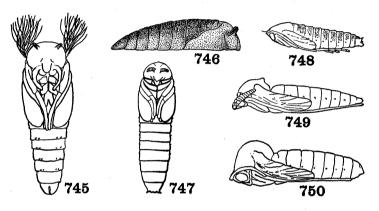
- 7. Thorax conspicuously swollen, almost globose, its anterior profile declivous; sternopleura concealed. (Fig. 750, Lèia).

MYCETOPHÍLIDÆ

	Thorax not conspicuously swollen, the anterior profile not sloping downward8
8.	Scape of antennæ much swollen, globose; abdominal spiracles small or absent; sternopleura enlarged, not concealed by fore coxæ and femora
	Scape of antennæ not much swollen; abdominal spiracles distinct; sternopleura not visible, concealed by the large coxæ and femora of the fore legs.
	(Fig. 740, Monárdia) CECIDOMYÌIDÆ, part (Fig. 748, Sciara)
	Thoracic respiratory organs slender, long and tube-like; legs
9.	straight, extending well beyond apices of wings; body without
	armature except a pair of hairs on anterior margin of head;
	sternopleura concealed CECIDOMYÌIDÆ, part
	SCIÁRIDÆ, part
	Species without such combination of characters, abdomen usually
	with hairs or spines, or the sternopleura is exposed10
10.	Pupa in a pocket-shaped or slipper-shaped cocoon consisting of
	coarse threads, from the wide, open end of which project the
	thoracic respiratory filaments: aquatic, living in swiftly running
	streams. (Fig. 745, Simulium) SIMULĪIDÆ
	Pupa free, or if enclosed or partly so the cocoon is not pocket-like
11.	and respiratory organs do not consist of tube-like branches. 11 Pupa when seen from above oval or rounded in outline, the abdo-
11.	men at base not conspicuously narrower than thorax, so that
	the lateral outline is continuous; dorsal surface with strong
	integument; venter with sucking disks for adhering to rocks in
	running streams where the pupæ occur
	Pupa with abdomen well differentiated from thorax, the dorsum
	membranous, or if strong and almost chitinized surface spines
10	are developed
12.	
	ter; three pairs of lateral sucking disks; respiratory filaments short tubular. (Fig. 741, <b>Deuterophlèbia</b> ).
	Short tubular. (Fig. 741, Deuteropmenia).  DEUTEROPHLEBÌIDÆ
	Antennæ not twice coiled; ventral sucking disks median13
13.	Thoracic respiratory organs lamelliform, consisting of four flat
	plates, the broad sides of which are contiguous. (Fig. 746,
	Bibiocéphala) BLEPHAROCERÁTIDÆ
	Thoracic respiratory organs simple, tube-like. (Maurina, N. and S. Am.)

14. Apical abdominal segment terminating in two or four paddle-like or fin-shaped organs which are fringed on all or part of outer surface by strap-like hairs; or if the apical segment terminates in two long subconic processes the tarsi are recurved against the ventral surface of the base of the abdomen and apex of thorax so that they do not extend beyond apices of wings..........15

Apical segment of abdomen obtuse, armed with short or elongate spines or thorns, or if ending in a pair of long, slender processes



Figs. 745-750. Nematocerous Diptera, Pupæ

- 745. Simulium, ventral view (Malloch) Simuliidæ.
- 746. Bibiocephala, side view (Malloch) Blepharoceratidæ.
- 747. Bibio, ventral view (Malloch) Bibionidæ.
- 748. Sciara, side view (Malloch) Sciaridæ.
- 749. Ceroplatus, side view. Ceroplatidæ.
- 750. Leia, side view (Malloch) Mycetophilidæ.

Thoracic respiratory organs consisting of a single stem, in some cases with a few long, or many short, scale-like surface hairs, but never terminating in numerous thread-like filaments; occasionally the thoracic respiratory organs not elevated.....16

16.	Thoracic respiratory organs not elevated; sternopleura exposed.  CHIRONÓMIDÆ
	Thoracic respiratory organs conspicuously elevated
17.	Thoracic respiratory organs situated close to anterior margin of
	thorax; no stellate hairs on thorax and abdomen.
	CHIRONÓMIDÆ, part
	Thoracic respiratory organs situated close to middle of thoracic
	dorsum
18.	Apical abdominal segment ending in two or four broad, flat,
	paddle-like plates. (Fig. 742, Anopheles; Fig. 997, Culex).
	CULÍCIDÆ, part
	Apical abdominal segment ending in two long subconical proc-
	esses
19.	Apical processes armed with short hairs at apices and on middle
	of outer margin CULÍCIDÆ, part
	Apical processes unarmed. (Fig. 744, <b>Díxa</b> ) <b>DÍXIDÆ</b>
<b>20</b> .	Apices of legs not extending beyond apices of wings
	Apices of posterior legs at least extending beyond apices of wings.22
21.	Apical segment of abdomen ending in two conical processes.
	CERATOPOGÓNIDÆ
	Apical segment of abdomen ending in two upper and two lower
	short thorns. (Fig. 743, Psychòda) PSYCHÓDIDÆ
22.	Thoracic respiratory organs long, bifid; apical abdominal segment
	rounded, without processes; abdominal spiracles pedunculate.
	SCATÓPSIDÆ
	Thoracic respiratory organs simple; apical abdominal segment
กก	not rounded, generally armed with protuberances23
23.	Thoracic respiratory organs elevated but little above disk of
	thorax; tarsi of fore legs overlapping those of middle pair,
	which overlap the hind ones, all rather closely fused together and to wings
	Thoracic respiratory organs very conspicuously elevated; legs
	otherwise formed
24.	Thoracic respiratory organs equal in length, rarely one twice
- 1.	as long as the other; all tarsi distinct
	Thoracic respiratory organs of very different length, one short,
	the other very long; front tarsi overlapping middle pair. (Fig.
	738, Bittacomórpha) PTYCHOPTÉRIDÆ
25.	Abdominal segments each with one transverse row, sometimes
	with two rows, of thorn-like protuberances; palpi recurved at
	apices. (Fig. 739, Pachyrrhina) TIPÙLIDÆ

Abdominal segments rarely with distinct thorn-like protuberances, usually with weak hairs; palpi straight, not recurved at apices.

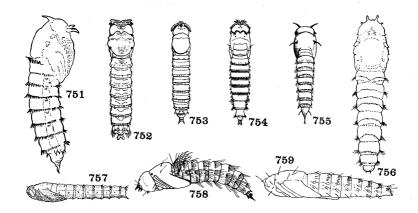
# LIMONÎIDÆ

# Orthorrhapha

26.	Pupa enclosed within the last larval moult
	Pupa free
27.	Thoracic segments one and two each with a smooth plate on
	dorsum; apical segment with a transverse series of short teeth
	near base on ventral surface; pupal skin largely or entirely with-
	drawn from puparium during emergence. (Fig. 707, Xylomỳia).
	XYLOMYÌIDÆ
	Thoracic segments without smooth dorsal plates; pupal skin not
	withdrawn from puparium during emergence. STRATIOMYIDÆ
90	Prothorax with a large aperture mesad of and connected with the
28.	spiracle
	Prothorax without such an aperture
20	Head without strong forwardly directed thorns, at most with one
20.	thorn on base of antenna which is directed to the side; abdomi-
	nal armature weak, becoming gradually stronger toward apex;
	wings short, extending to or slightly beyond apex of basal ab-
	dominal segment; apices of hind tarsi at most extending slightly
	beyond apices of wings; abdomen with seven pairs of spir-
	acles
	Head usually with strong thorns, or if these are absent the ab-
	dominal armature is stronger on basal or second segment than
	it is on apical, or there are less than seven pairs of abdominal
	spiracles; apices of hind tarsi usually extending distinctly be-
	yond apices of wings
30.	Antennal sheaths much thickened at base, apical portion slender,
	styliform, the whole directed almost straight downward. (Fig.
	757, Chrysopila) LÉPTIDÆ
	Antennal sheaths thickened throughout their length, the apical
	portion generally more or less distinctly annulated, the whole
	directed either straight sideways or in a slightly downward direction
31.	rection
υ1.	753, Rhachicerus) LÉPTIDÆ, RHACHICERÌNÆ
	Antennal sheaths showing not more than ten annulations32
	Antomai Showing not more than ten annulations02

32. Antennal sheaths very stout, not over twice as long as their basal breadth; face with a small sharp protuberance on each side a little mesad of the vertical line of apices of antennæ and slightly above middle of face, and at the base of each are two short hairs on their inner side; two very strong postspiracular abdominal bristles on each segment. (Fig. 752, Cœnomŷia).

CŒNOMYÌIDÆ



Figs. 751-759. Orthorrhaphous Diptera, Pupæ

- 751. Mydas, side view (Greene) Mydaidæ.
- 752. Cœnomyia, dorsal view (Greene) Cœnomyiidæ.
- 753. Rhachicerus, dorsal view (Greene) Xylomyiidæ.
- 754. Xylophagus, dorsal view (Greene) Xylophagidæ.
- 755. Psilocephala, dorsal view (Malloch) Therevidæ.
- 756. Dasyllis, dorsal view (Greene) Asilidæ.
- 757. Chrysopila, side view (Malloch) Rhagionidæ.
- 758. Sparnopolius, side view (Malloch) Bombyliidæ.
- 759. Rhamphomyia, side view (Malloch) Empididæ.

Antennal sheaths distinctly annulated, slender, about four times as long as their basal breadth; face without protuberances; post-spiracular abdominal bristles slender, eight to ten on each segment. (Fig. 754, Xylóphagus) . . . . . XYLOPHÁGIDÆ

33. Head without strong thorns; abdomen with three or four distinct pairs of spiracles and without spinose armature.

**ACROCERÁTIDÆ** 

	Head usually with strong thorns, at least with elevated ridge-like
	antennal sheath and several small carinated elevations; abdo-
	men with seven pairs of spiracles and spinose armature34
34.	Head with two thorns
	Head with more than two thorns or with several short tubercles .36
35.	Abdomen with a single transverse series of spines on each dorsal
	segment; wing with a long thorn at base. (Fig. 755, Psilo-
	céphala)
	Abdomen with two transverse series of spines on each dorsal
	segment; wing without thorn at base SCENOPÍNIDÆ
1	
ſ.	

Figs. 760-764. Cyclorrhaphous Diptera. Pupæ

- 760. Didea, dorsal view (Metcalf) Syrphidæ.
- 761. Criorhina, dorsal view (Greene) Syrphidæ.
- 762. Pipunculus, dorsal view (Perkins) Pipunculidæ.
- 763. Sarcophaga, side view (Greene) Sarcophagidæ.
- 764. Exorista. side view (Greene) Ortalidæ.
- 37. Head with strong thorns, or if they are absent the abdomen has the dorsal transverse armature consisting of very strong thorns and intervening long slender hairs; apices of antennæ obtuse 38
  - Head very rarely with strong thorns, two carinate elevations present on upper anterior margin; antennæ with attenuated apices; body without thorns, sometimes with bristles...............39
- 38. Lower median portion of face with a closely approximated pair of stout thorns which are occasionally fused almost to apices; abdomen with the transverse armature of dorsal segments con-

Lower median portion of face without thorns; abdomen with the transverse armature consisting of alternating long and short thorns (except in *Leptogaster*). (Fig. 756, **Dasýllis**). **ASĨLIDÆ** 

DOLICHOPÓDIDÆ

# LITERATURE ON DIPTERA, GENERAL

- ARIAS, J. Classification Catalonian Diptera. Encobet: Entom. de Catalonya, pp. 56 (1917).
- Austen, E. E. British blood-sucking Flies. London, Brit. Mus. Publ., pp. 1-74, 34 plates (1906).
- COQUILLETT, D. W. The type-species of North American genera of Diptera. Proc. U. S. Nat. Mus., 37, pp. 499-647 (1910); Can. Ent., 42, pp. 375-378 (1910).
- Grimshaw, P. H. Guide to literature of British Diptera. Proc. Roy. Phys. Soc. Edinb., 20, pp. 78–117 (1917).
- Hendel, F. Zweiflügler. 2: Allgemeiner Teil. Tierwelt Deutschl., pt. 11, pp. 1-135 (1928)
- Lundbeck, W. Diptera Danica. Kopenhag. 7 vols. (1907–1927) (listed under separate families).
- OSTEN SACKEN, C. R. Western Diptera (U. S.). U. S. Geol. Geog. Surv. Bull. 3, pp. 189–354 (1877).
- Pearce, E. K. Typical Flies (British). London, 3 photographic atlases (1928). Stackelberg, A. Keys to families and genera of blood-sucking Diptera of U.S.S.R. Rev. Microbiol. Epidémiol., 5, pp. 43-56 (1926).
  - Our Flies, Keys to species of Russian Diptera. Govt. Publ. Moscow-Leningrad. 152 pp. (1926).
- WILLISTON, S. W. Manual of families and genera of North American Diptera. 3 ed., New Haven, Conn. 405 pp. (1908).

#### CATALOGUES AND LISTS

ALDRICH, J. M. Catalogue of North American Diptera. Smithson. Misc. Coll., 46, no. 1444, pp. 680 (1905)

- Becker, Bezzi, Kertész and Stein. Katalog der palæarkt. Dipteren. Budapest, 4 vols. (1903–1917).
- Brunetti, E. Catalogue of oriental and South Asiatic Nemocera. Rec. Ind. Mus., 17, 300 pp. (1920).
- Cole, F. R. and A. L. Lovett. Annotated list of Oregon Diptera. Proc. Cal. Acad. Sci. (4), 11, pp. 197–344 (1921)
- JOHNSON, C. W. List of Diptera of New Jersey. Ann. Rept. N. J. State Mus. 1909, pp. 703–814 (1910).
  - List of Diptera of New England. Occ. Papers Bost. Soc. Nat. Hist. no. 7, 326 pp. (1925).
- Kertész, K. Catalogus Dipterorum. Budapest, 7 vols. (1902–1910).

#### SUBORDER NEMATOCERA

- Alexander, C. P. Radial field of wing of Nematocerous Diptera. Proc. Linn. Soc. N. S. Wales, **52**, pp. 42–72 (1927).
  - Nomenclature of radial field in Diptera. 4th Internat. Congr. Entom. Ithaca, Trans., 2, pp. 700-707 (1929).
- Brunetti, E. Fauna Brit. India, Diptera Nemocera, excl. Chironomidæ and Culicidæ. London, 581 pp. (1912).
- Crampton, G. C. Phylogeny of Nematocera. Psyche, **31**, pp. 238–242 (1924). Phylogenetic study of thoracic sclerites of Psychodoid Diptera. Ent. News, **37**, pp. 33–38, 65–70 (1926).
- Edwards, F. W. Phylogeny of Nematocerous Diptera. 3d Internat. Congr. Entom. Verh. 2, pp. 111–130 (1925).
- JOHANNSEN, O. A. Aquatic Nematocerous Diptera. Bull. N. Y. State Mus. no. 68, pp. 327-448 (1903); no. 86, pp. 76-327 (1905).

#### BIBIONIDÆ AND SCATOPSIDÆ

- Edwards, F. W. Synopsis British Bibionidæ and Scatopsidæ. Ann. Appl. Biol., 12, pp. 263–275 (1925).
- McAtee, W. L. Nearctic Bibionidæ. Proc. U. S. Nat. Mus., 60, art. 11, 27 pp. (1922).
- Duda, O. Scatopsidæ exotic to Europe. Konowia, 7, pp. 259–297 (1928).
  Palæarctic Scatopsidæ. Fliegen pal. Reg., Lief. 26, pp. 1–62 (1928).
- Melander, A. L. Nearctic Scatopsidæ. Wash. Agr. Exp. Sta. Bull. 130, 21 pp. (1916).

#### BLEPHAROCERATIDÆ AND DEUTEROPHLEBIIDÆ

- Bezzi, M. Blefaroceridi italiani, etc. Bull. Soc. ent. Ital., 44, pp. 1–114 (1913).
- Curran, C. H. Key to genera of Blepharoceridæ. Can. Ent., 55, p. 267 (1923).

- Edwards, F. W. Neotropical Blepharoceridæ. Dipt. Patagon. and S. Chile, **2**, fasc. 2, pp. 33-75 (1929).
- Kellogg, V. L. Blepharoceridæ of North America. Proc. Cal. Acad. Sci. (3) Zool., 3, pp. 187–226 (1903).

Family Blepharoceridæ. Gen. Ins., fasc. 56, 15 pp. (1907).

- TILLYARD, R. J. Australian Blepharoceridæ. Austr. Zool., 2, pp. 159–172 (1922); N. Zeal. Jour. Sci. Tech., 5, pp. 101–107 (1922).
- TONNOIR, A. L. Australian Blepharoceridæ. Austr. Zool., 3, pp. 47–59 (1923).
- Walley, G. S. Review of Canadian Blephariceridæ. Can. Ent., 59, pp. 112–115 (1927).
- EDWARDS, F. W. Deuterophlebiidæ. Ann. Mag. Nat. Hist. (9), 9, p. 380 (1922).

#### CECIDOMYIIDÆ

Felt, E. P. Key to genera and tribes. Philipp. Jour. Sci. D, 13, pp. 296–324 (1918).

Key to American galls. N. Y. State Mus. Bull. 200, 241 pp. (1918). Résumé, keys and bibliography. N. Y. State Mus. Bull. 257, 239 pp. (1925).

- KIEFFER, J. J. Family Cecidomyiidæ. Gen. Ins., fasc. 152, 346 pp. (1913). Monograph European and Algerian. Ann. Soc. Ent. Fr. 1900, pp. 181–472 (1900).
- RÜBSAAMEN, E. H. and H. HEDICKE. Revision German Cecidomyiidæ. Zoologia, Stuttgart, **29**, Heft 77, Lief. 1, 2, pp. 1–264 (1926).
- Senior-White, R. Cecidomyidæ. Cat. Indian Ins. Govt. India, Centr. Pub. Br., pt. 15 (1927).

#### CHIRONOMIDÆ AND CERATOPOGONIDÆ

EDWARDS, F. W. Revision British Ceratopogonidæ. Trans. Ent. Soc. Lond., 74, pp. 389-426 (1926).

British Chironomidæ. Trans. Ent. Soc. Lond., 77, pp. 279-430 (1929).

Goetghebuer, M. Ceratopogoninæ de Belgique. Mém. Mus. Nat. Hist. Belg., 8, fasc. 3, pp. 1–116 (1920).

Tanypodinæ of France. Faune de Fr., 15, 83 pp. (1927).

- Hoffman, W. A. Nearctic and neotropical Culicoides. Am. Jour. Hyg., **5**, pp. 274–301 (1925).
- JOHANNSEN, O. A. North American Chironomidæ. N. Y. State Mus. Bull. 86, pp. 76–316 (1905).
- Kieffer, J. J. Family Chironomidæ. Gen. Ins., fasc. 42, 78 pp. (1906).
  Formosan Chironomidæ. Ann. Mus. Hung., 14, pp. 81–121 (1916).
  Australian Chironomidæ. Ann. Mus. Hung., 15, pp. 175–208 (1917).
  American Chironomidæ in Hung. Nat. Mus. Ann. Mus. Hung., 15, pp. 292–364 (1917).

African and Asiatic Chironomidæ in Hung. Nat. Mus. Ann. Mus. Hung., 16, pp. 31–139 (1918).

European Chironomidæ in Hung. Nat. Mus. Ann. Mus. Hung., 17, pp. 1–160 (1919).

Ceratopogoninæ. Faun. de France, Paris, 11, pp. 139 (1925).

Key to Ceratopogoninæ. Arch. Inst. Past. Algérie, 4, pp. 96–107 (1926).

- Lenz, F. Metamorphosis in relation to classification of Chironomidæ. Ent. Mitt. Berl., 15, pp. 440–442 (1926).
- Malloch, J. R. Chironomidæ of Illinois. Illinois State Lab. Nat. Hist. Bull. 10, pp. 275–543 (1915).
- THIENEMANN, A. Keys to Swedish larvæ and adults. Arch. Hydrobiol. Suppl., 2, pp. 465–468 (1915); 550–551 (1916).
- Walley, G. S. Canadian Tanypus. Can. Ent., 57, pp. 271-278 (1926).

## CULICIDÆ

- Barraud, P. J. Revision of Culicine mosquitoes of India. Ind. Jour. Med. Res., 14, et seq., numerous parts (1927–).
- Blanchard, R. Les moustiques. Paris, 673 pp. (1905).
- Bonne, C. and J. Bonne-Wepster. Mosquitoes of Surinam. Roy. Col. Inst. Amsterdam, Trop. Med. No. 13, 558 pp. (1925).
- Brug, S. L. Dutch East Indian Anophelines and Culicines. Geneesk. Tijds. Ned. Ind., 65, pp. 661–671 (1925).
- Christophers, Sinton and Covel. Synoptic table Anophelinæ of India. Health Bull. Calcutta, **10**, 22 pp. (1927).
- Cooling, L. E. Synonymic list Australian mosquitoes. Comm. Austr. Dept. Health, Service Publ. 2, pp. 1–61 (1924).
- Coquillett, D. W. North and Central American mosquitoes. U. S. D. A. Ent. Tech. Bull. 11, 31 pp. (1906).
- DYAR, H. G. Canadian mosquitoes. Trans. R. Can. Inst., 13, pp. 71–120 (1921).
  - Mosquitoes of United States. Proc. U. S. Nat. Mus., **62**, art. 1, 119 pp. (1922).
- Dyar, H. G. and R. C. Shannon. American Chaoborinæ. Ins. Insc. Menstr., 12, pp. 201–216 (1924).
  - Subfamilies, tribes and genera of American Culicidæ. Jour. Wash. Acad. Sc., 14, pp. 472–486 (1924).
- Edwards, F. W. Revision palæarctic Culicidæ. Bull. Ent. Res., **12**, pp. 263–351 (1921).
  - Synopsis Australian mosquitoes. Bull. Ent. Res., **14**, pp. 351–401 (1923).
- Felt, E. P. Mosquitoes of New York. N. Y. State Mus. Bull. 79, pp. 241–400 (1904).
- Ferguson, E. W. Mosquitoes of New South Wales. Rept. Dir. Gen. Pub. Health N. S. W. 1925, pp. 187–191 (1924).

- Freeboen, S. B. Mosquitoes of California. Univ. Cal. Publ. Ent. 3, pp. 333-460 (1926).
- GILES, G. M. Handbook of Culcidæ. London, 530 pp. (1902).
- Howard, L. O. Culicidæ of United States. U. S. D. A. Ent. Bull. 25 (1900). Mosquitoes. New York, XVI. and 241 pp. (1901).
- Howard, Dyar and Knab. Mosquitoes of North and Central America. Carnegie Inst. Wash., 4 vols., 1064 pp. (1912–1917).
- JOHANNSEN, O. A. Tables larvæ, pupæ and adults of nearctic Culicidæ. N. Y. State Mus. Bull. 68, pp. 388–429 (1903).
- Kirkpatrick, T. W. Mosquitoes of Egypt. Govt. Press, Cairo, 224 pp. (1925).
- Lang, W. D. Handbook of British mosquitoes. Brit. Mus., 125 pp. (1920).
- Martini, E. Palæarctic Culicidæ (incl. Dixinæ, pp. 20–43). Flieg. Pal. Reg., Lief. 33, 35, 38, 40, 192 pp. (1929, 1930).
- Matheson, R. Handbook of North American mosquitoes. Springfield and London. 268 pp. (1929).
- Matheson, R. and R. C. Shannon. Anophelines of N. E. America. Insec. Insect. Menst., 11, pp. 57–64 (1923).
- Schwetz, J. Synopsis Belgian Congo mosquitoes. Rev. Zool. Afr., Brussels, 15, pp. 271–319 (1927).
- Séguy, E. Key to French mosquitoes. Bull. Soc. Path. Exot., 14, pp. 179–187 (1921).
- Senior-White, R. Indian Culicidæ. Cat. Ind. Mus. Govt. Ind. Centr. Pub. Br., pt. 2, 124 pp. (1923).
- STRICKLAND, C. Anophelines of India, etc. Culcutta and Simla. 19 pp. (1925).
- THEOBALD, F. Monograph Culicidæ. Brit. Mus., 5 vols., 2459 pp. (1901–1910).
- Yamada, S. Anophelines of Japan. Sci. Repts. Govt. Inst. Infect. Dis. Tokyo, **3**, pp. 215–241 (1924); **4**, pp. 447–493 (1925).

#### DIXIDÆ

Edwards, F. W. British Dixidæ, etc. Ent. Mo. Mag., **56**, pp. 264–270 (1920). Johannsen, O. A. North American Dixidæ. Psyche, **30**, pp. 52–58 (1923). Martini, E. Palæarctic Dixinæ. Flieg. Pal. Reg., Lief. 33, pp. 20-43 (1929).

## MYCETOPHILIDÆ, s. lat.

Edwards, F. W. Systematic position of Mycetobia. Ann. Mag. Nat. Hist., 17, pp. 108–115 (1916).

Ditomyinæ. Ann. Mag. Nat. Hist., 7, pp. 431-436 (1921).

Revision of Mycetophilidæ. Trans. Ent. Soc. Lond., 1925, pp. 505–670 (1925).

Australian Ceroplatinæ. Proc. Linn. Soc. N. S. W., **54**, pp. 162–175 (1929).

Edwards, F. W. and D. Keilin. Family Pachyneuridæ. Gen. Ins., fasc. 190, pp. 27–29 (1928).

- Enderlein, G. Lycoriidæ. Arch. Naturg., **77**, Band 1, 3 Suppl., pp. 116–201 (1911).
- JOHANNSEN, O. A. Fungus gnats of North America. Maine Agr. Exp. Sta. Bulls. 172, 180, 196 and 200 (1909–1912).

Family Mycetophilidæ. Gen. Ins., fasc. 93, 141 pp. (1909).

- LANDROCK, K. Tables European Mycetophilidæ. Wien. ent. Ztg., 37, pp. 55-72; 107-120 (1918).
  - Palæarctic Mycetophilidæ. Flieg. pal. Reg., Lief. 12-15, 196 pp. (1926-1927).
- Lengerdorf, F. Lycoriidæ. Flieg. pal. Reg., Lief. 24, 43, pp. 71 (1928–1930).
- Pettey, F. W. Revision species of Sciara. Ann. Ent. Soc. Am., **11**, p. 319–346 (1918).
- Tonnoir, A. L. Synopsis Australian Mycetophilidæ. Proc. Linn. Soc. N. S. W., 54, pp. 584–614 (1920).

#### PSYCHODIDÆ

- ALEXANDER, C. P. Revision of American Bruchomyinæ. Proc. U. S. Nat. Mus., 75, art. 7, pp. 1–9 (1929).
- Banks, N. Eastern (U. S.) Psychodidæ. Can. Ent., 23, pp. 273-275 (1901).
- Haseman, L. Monograph N. American Psychodidæ. Trans. Am. Ent. Soc., 33, pp. 299–333 (1907).
- Kincaid, T. Pacific Coast (N. Am.) Psychodidæ. Ent. News, 10, pp. 30–37 (1899).
- Tonnoir, A. L. African Psychodidæ. Bull. Soc. R. Ent. Egypt., 1921, pp. 80–111 (1921).

European Psychodidæ. Ann. Soc. Ent. Belg., **62**, pp. 49–88, 125–136, 153–181 (1922).

South American Psychodidæ. Dipt. Patagon. Chile, **2**, fasc. 1, 32 pp. (1929).

#### ANISOPODIDÆ

- Edwards, F. W. Key to European Anisopus. Ent. Mo. Mag., **62**, p. 113 (1926).
- Edwards, F. W. and D. Keilin. Protorhyphidæ, Anisopodidæ, Pachyneuridæ and Trichoceridæ. Gen. Ins., fasc. 190, 41 pp. (1928).
- Kerrész, K. Ueber die Familie der Rhyphiden. Termész. Füzet., **25**, pp. 4–6 (1902).

#### SIMULIIDÆ

- Baranoff, N. Classification Simuliidæ. Neue Beitr. syst. Insektenk., 3, pp. 161–164 (1926).
- Coquillett, D. W. Simuliidæ of United States. U. S. D. A. Ent. Bull. 10, pp. 66–69 (1898).

- Dyar, H. G. and R. C. Shannon. North American Simuliidæ. Proc. U. S. Nat. Mus., 69, art. 10, 54 pp. (1927).
- EDWARDS, F. W. British Simuliidæ. Ent. Mo. Mag., **51**, pp. 305–309 (1915); Bull. Ent. Res., **6**, pp. 211–246 (1920).

Malayan female Simuliidæ. Fed. Mal. St. Mus. Jour., 14, p. 59 (1928).

ENDERLEIN, G. Systematic arrangement, Simuliidæ. Zool. Anz., **53**, pp. 43–46 (1921).

Classification Simuliidæ. Arch. f. Klassif. Phylogen. Ent., 1, pp. 77–97 (1930).

- JOHANNSEN, O. A. Simuliidæ of United States. N. Y. State Mus. Bull. 68, pp. 336–388 (1903).
- Malloch, J. R. American Blackflies. U. S. D. A. Bur. Ent. Tech. Bull. 26, 71 pp. (1914).
- O'Kane, W. C. Bionomics of New Hampshire Simuliidæ. Bull. N. H. Exp. Sta. 32, 24 pp. (1926).
- SÉGUY, E. Espèces méditerranéennes, Simulium. Eos, 1, pp. 231-238 (1925).

#### TIPULOIDEA

Alexander, C. P. Neotropical Limnobiinæ. Proc. U. S. Nat. Mus., 44, pp. 481-549 (1913).

Neotropical Tanyderidæ. Proc. U. S. Nat. Mus., 44, pp. 331–335 (1913). Crane-flies of New York. Cornell Univ. Mem. 25, pp. 767–993 (1919);

38, pp. 695–1133 (1920).

Revision Australian Trichoceridæ. Proc. Linn. Soc. N. S. W., 51, pp.

299–304 (1926).

Cylindrotominæ. Gen. Ins., fasc. 187, 16 pp. (1927).

Ptychopteridæ. Gen. Ins., fasc. 188, 12 pp. (1927).

Tanyderidæ. Gen. Ins., fasc. 189, 13 pp. (1927).

Australian Tanyderidæ. Proc. Linn. Soc., N. S. W., **53**, pp. 367–374 (1928).

South American Tipulidæ. Dipt. Patagon. Chile, pt. 1, 240 pp. (1929). Japanese Trichoceridæ. Konowia, **9**, pp. 103–108 (1930).

ALEXANDER, C. P. and W. L. McAtee. Tipuloidea of District of Columbia. Proc. U. S. Nat. Mus., 58, pp. 385–435 (1920).

DIETZ, W. G. North American Pachyrhina. Trans. Am. Ent. Soc., 44, pp. 105–140 (1918).

Edwards, F. W. Craneflies of New Zealand. Trans. N. Zeal. Inst., **54**, pp. 265–352 (1923).

Philippine Tipulidæ. Notulæ Ent., 6, pp. 33-44 (1926).

- Edwards, F. W. and D. Keilin. Trichoceridæ. Gen. Ins., fasc. 190, pp. 30–37 (1928).
- Enderlein, G. Studies in Tipulidæ. Zool. Jahrb. Syst., 32, pp. 1-88 (1912).
- Kuntze, A. Palæarctic Eriopterinæ. Ann. Naturhist. Hofmus. Wien, 28, pp. 361–388 (1914).

Palæarctic Limoniinæ. Zool. Jahrb. Syst., 43, pp. 371–432 (1920).

OSTEN SACKEN, C. R. Studies in Tipulidæ longipalpi. Berl. Ent. Zts., **30**, pp. 153–188 (1886).

Studies in Tipulidæ brevipalpi. Berl. Ent. Zts., **31**, pp. 163–242 (1887). PIERRE, C. Tipulinæ. Gen. Ins., fasc. 186, 68 pp. (1926).

#### THAUMALEIDÆ

- Bezzi, M. Taumaleidi italiani. Boll. Lab. Zool. Gen. Agr., 7, pp. 227–266 (1913).
- Dyar, H. G. and R. C. Shannon. North American Thaumeleidæ. Jour. Wash. Ac. Sci., 14, pp. 432–434 (1924).
- Edwards, F. W. Revision of Thaumaleidæ. Zool. Anz., **82**, Wasmann no., pp. 121–142 (1929)

### SUBORDER BRACHYCERA

#### A. ORTHORRHAPHA

- Bezzi, M. Brachycera and Athericera of Fiji Islands. Brit. Mus. Publ., 220 pp. (1928)
- Brunetti, E. Fauna of British India, Dipt. Brachycera. Lond., 1, 401 pp. (1920).
- Cole, F. R., J. R. Malloch and W. L. McAtee. District of Columbia Tromoptera. Proc. Ent. Soc. Wash., 26, pp. 181–195 (1924).
- HARDY, G. H. Phylogeny of some Diptera Brachycera. Proc. Linn. Soc. N. S. W., 52, pp. 380–386 (1927).
- Shannon, R. and S. W. Bromley. Radial venation in Brachycera. Insec. Inscit. Menstr., 12, pp. 137–140 (1924).

#### ACROCERATIDÆ

- Cole, F. R. North American Cyrtidæ. Trans. Am. Ent. Soc., 45, pp. 1–79 (1919).
- LUNDBECK, W. Danish Acroceridæ. Dipt. Dan., 1, pp. 157-163 (1907).
- Pleske, T. Palæarctic Acroceridæ. Konowia, 9, pp. 156–173 (1930).
- SÉGUY, E. French Oncodidæ. Faune de France, 13, pp. 157-168 (1926).
- Verrall, G. H. British Acroceridæ. Brit. Flies, 5, pp. 447-469 (1909).

#### APIOCERATIDÆ

Hermann, F. Beitrag z. Kenntnis der Apioceriden. Deut. Ent. Zts. 1909, Beiheft, pp. 104-122 (1909).

#### ASILIDÆ

- Back, E. A. Robberflies of North America. Trans. Am. Ent. Soc., 35, pp. 137–400 (1909).
- Banks, N. American species of Dasyllis. Bull. Brkl. Ent. Soc., 12, pp. 52–55 (1917).
- Becker, T. Neue Dioctria. Konowia, **2**, pp. 1–18 (1923). Formosan Asilinæ. Ent. Mitt., **14**, pp. 62, 123, 240 (1925).
- Bezzi, M. Lasiopogon. Boll. Lab. Zool. Gen. Agrar. Portici, 11, pp. 250–281 (1917).
- Bromley, S. W. Asilidæ of Cuba. Ann. Ent. Soc. Am., 22, pp. 272-294 (1929).
- Curran, C. H. Nearctic Cyrtopogon. Can. Ent., **55**, pp. 93 et seq., 5 pts. (1923).
- ENDERLEIN, G. Dasypogoninæ and Archilaphrinæ. Wien. ent. Ztg., 33, pp. 151–174 (1914).
- ENGEL, E. O. Palæarctic Asilidæ. Flieg. pal. Reg., 9 pts., 491 pp. (1925–1930).
- HARDY, G. M. Australian Asilidæ. Proc. Linn. Soc. N. S. W., 51, pp. 305–312, 643–657 (1926); 52, 387–398 (1927); 54, 80–88, 353–360 (1929); 55, 249–260 (1930).
- HERMANN, F. So. Am. Asilidæ. Abh. Deut. Akad. Naturf., 96, pp. 1–275 (1912).
  - Systematik der Asiliden. Zool. Jahrb., **43**, Syst., pp. 161–194 (1920). Leptogastrinæ. Verh. z. b. Ges. Wien, **74**, pp. 140–152 (1926).
- Hine, J. S. Various American genera. Ann. Ent. Soc. Am., 2, pp. 136–170 (Asilus) (1909); 4, 153–172 (Promachus) (1911); 12, 103–157 (Erax) (1919).
- Lundbeck, W. Danish Asilidæ. Dipt. Dan., 2, pp. 1–87 (1908).
- MacAtee, W. L. and N. Banks. District of Columbia Asilidæ. Proc. Ent. Soc. Wash., 22, pp. 13–33 (1920).
- Melander, A. L. Various nearctic genera. Psyche, 30, pp. 136–144 (Lasiopogon); 102–119 (Cyrtopogon); 207–219 (misc.) (1923).
- Melin, D. Biology, etc., of Swedish Asilidæ., Zool. Bidrag., 8, pp. 1–317 (1923).
- Pandellé, L. European Asilus. Rev. Ent. France, 1905, pp. 44-98 (1905).
- RICARDO, G. Australasian Asilidæ. Ann. Mag. Nat. Hist. (8), 9, pp. 473, 585; 10, pp. 142, 350 (1912).
- SÉGUY, E. Asilidæ of France. Faun. d. France, No. 17, 190 pp. (1927).
- Verrall, G. H. British Asilidæ. Brit. Flies, 5, pp. 614-754 (1909).

#### BOMBYLIIDÆ

- Becker, T. Genera of Bombyliidæ. Ann. Mus. Zool. St. Petersb., 17, pp. 421-502 (1913).
- Bezzi, M. South African Bombyliidæ. Ann. S. Afr. Mus., 18, pp. 1–180 (1921). Ethiopian Bombyliidæ. Brit. Mus. Publ., 390 pp. (1924).
- Brunetti, E. Revised catalog oriental Bombylidæ. Rec. Ind. Mus., 2, pp. 437–492 (1909).

- Coquillett, D. W. Various nearctic genera. Can. Ent., **24**, pp. 9-, 123-(1892); Trans. Am. Ent. Soc., **14**, pp. 159-; (1887); **19**, 168-(1892); **21**, 89- (1894).
- Enderlein, G. Systropodinæ. Wien. ent. Ztg., 43, pp. 69–92 (1926).
- JOHNSON, C. W. Bombylius of eastern United States. Psyche, 14, pp. 95–100 (1907).
- LUNDBECK, W. Danish Bombyliidæ. Dipt. Dan., 2, pp. 88-132 (1908).
- Painter, R. H. Review of Heterostylum. Jour. Kans. Ent., 3, pp. 1-7 (1930).
- Paramonow, S. J. Hemipenthes. Encycl. Ent., B. Dipt. 2, pp. 150–190 (1926).

  Monograph palæarctic Bombylius. Mém. cl. sci. phys. math. Acad. Sci. Ukraine, 3, pp. 77–184 (1926).
- Roberts, F. H. S. Revision Australian Bombyliidæ. Proc. Linn. Soc. N. S. W., **53**, pp. 90–144, 413–455 (1928); **54**, 553–583 (1929).
- SÉGUY, E. Bombyliidæ of France. Faune de France, 13, pp. 178–254 (1926). Senior-White, R. Indian Bombyliidæ. Cat. Indian Ins. Govt. Ind. Centr. Pub. Br., pt. 3 (1923).
- VERRALL, G. H. British Bombyliidæ. Brit. Flies, 5, pp. 474–536 (1909).

#### DOLICHOPODIDÆ

- Aldrich, J. M. Keys to various nearctic genera. Trans. Am. Ent. Soc., 30, pp. 269–286 (1904).
  - Nearctic Hydrophorus. Psyche, 18, pp. 45-73 (1911).
- Becker, T. Review palæarctic Dolichopodidæ. N. Acta Leop.-Carol. Akad. Naturf. Halle, **102**, pp. 115–249 (1917); **103**, 205–315 (1918); **104**, 38–212 (1918).
  - American Dolichopodidæ. Abh. z. b. Ges. Wien, **13**, pp. 1–394 (1922). Indo-australian Dolichopodidæ. Capita Zool., **1**, pp. 1–247 (1922). Æthiopische Dolichopodiden. Entom. Mitt., **12**, pp. 1–50 (1923).
- Curran, C. H. South African Dolichopodidæ. Ann. Transv. Mus., 10, pp. 212–232 (1924); Ann. S. Afr. Mus., 23, pp. 377–416 (1926).
  - Nearctic Rhaphium species. Tr. R. Can. Inst. Toronto, **15**, pp. 249–260 (1926); **16**, 99–179 (1927).
    - Ethiopian Chrysosoma, etc. Rev. Zool. Afr., 15, pp. 242–262 (1927).
- Greene, C. T. Nearctic species of Tachytrechus. Proc. U. S. Nat. Mus., 60, art. 17, 21 pp. (1922).
  - Nearctic species of Scellus. Proc. U. S. Nat. Mus., 65, art. 16, 18 pp. (1924).
- HARDY, G. M. Australian Dolichopodidæ. Austr. Zool., 6, pp. 124–134 (1930).
  LOEW, H. Monograph North American Dolichopodidæ. Smithson. Misc.
  Coll. No. 171 (1864).
- LUNDBECK, W. Danish Dolichopodidæ. Dipt. Dan., 4, 407 pp. (1912).
- Parent, O. Reviews of palearctic genera, in Ann. Soc. Sci. Brux., 42, pp. 281–(Chrysotus) (1923); 44, pp. 46– (Thrypticus), pp. 93– (Aphrosylus),

pp. 221- (Diaphorus) (1925); **47**, pp. 1- (Medetera), pp. 83- (Tachytrechus) (1927).

Ethiopian Dolichopodidæ. Bull. Soc. Ent. Egypt, 1929, pp. 151–190 (1929).

Van Duzee, M. C. Reviews of nearctic genera: Neurigona, Ann. Ent. Soc. Am., 6, pp. 22–60 (1913). Asyndetus, Psyche, 23, pp. 88–94 (1916). Medeterus, Proc. Cal. Acad. Sci., 9, pp. 257–270 (1919); Psyche, 35, pp. 36–43 (1928). Diaphorus, Bull. Buff. Soc. Nat. Sci., 11, No. 2, pp. 161–194 (1922). Xiphandrium, Tr. Am. Ent. Soc., 48, pp. 79–87 (1922). Pelastoneurus, Ann. Ent. Soc. Am., 16, pp. 30–48 (1923). Chrysotus, Bull. Buff. Soc. Nat. Sci., 13, No. 3, pp. 52 (1924). Argyra, Proc. U. S. Nat. Mus., 66, art. 23, 43 pp. (1925). Syntormon, Tr. Am. Ent. Soc., 50, pp. 257–287 (1925). Thinophilus, Ann. Ent. Soc. Am., 19, pp. 35–49 (1926). Hydrophorus, Pan.-Pacif. Ent., 3, pp. 4–11 (1927). Sympycnus, Pan-Pacif. Ent., 7, pp. 35– , 51– (1930).

South American Dolichopodidæ. Dipt. Patagon. Chile, 5, fasc. 1, 92 pp. (1930).

VAN DUZEE, M. C., F. R. COLE and J. M. Aldrich. North American Dolichopus. U. S. Nat. Mus. Bull. 116, pp. 1–304 (1921).

Wheeler, W. M. Many nearctic genera. Proc. Cal. Ac. Sci. (3), **2**, Zool. pp. 1–78 (1899).

#### **EMPIDIDÆ**

Bezzi, M. Indo-australian Empididæ. Ann. Mus. Hung., 2, pp. 320–361 (1904).

Neotropical Empididæ. Ann. Mus. Hung., **3**, pp. 424–460 (1905); Nova Act. Akad. Naturf. Halle, **91**, pp. 293–407 (1910).

Collin, J. E. Notes on British Empididæ. Several issues of Ent. Mo. Mag., 62 (1926) and 63 (1927).

New Zealand Empididæ. Brit. Mus. Nat. Hist. Publ., 110 pp. (1928).

COQUILLETT, D. W. North American Empididæ. Proc. U. S. Nat. Mus., 18, pp. 387–440 (1897); Proc. Ent. Soc. Wash., 5, pp. 245–272 (1903).

Engel, E. Das Dipterengenus Atalanta (Clinocera). Deut. ent. Zts. 1918, pp. 1–80, 197–268 (1918).

Frey, R. Rhamphomyia. Notulæ Ent.,  $\mathbf{2}$ , pp. 1–10, 34–45, 65–77 (1922).

Kuntze, A. Table of European Empis. Zts. Hym. Dipt., 6, (1906), 7 (1907) several issues.

Lundbeck, W. Danish Empididæ. Dipt. Dan., 3, pp. 1-324 (1910).

MELANDER, A. L. North American Empididæ. Trans. Am. Ent. Soc., 28, pp. 195–367 (1902).

Family Empididæ. Gen. Ins., fasc. 185, 434 pp. (1927).

MILLER, D. New Zealand Empididæ. Tr. N. Zeal. Inst., 45, pp. 198–206 (1911); 54, 437–464 (1923).

## MYDAIDÆ

- ARIAS, J. Mydaidæ of Spain. Inst. Nac. cienc. fisico-nat. Ser. zool., 15, 40 pp. (1914).
- Bezzi, M. So. African Mydaidæ. Ann. S. Afr. Mus., 19, pp. 191-234 (1924).
- HARDY, G. H. Australian Mydaidæ. Proc. Linn. Soc. N. S. W., 50, pp. 139– 144 (1925).
- JOHNSON, C. W. North American. Proc. Bost. Soc. Nat. Hist., 38, pp. 131– 145 (1926).
- Mackerras, I. M. Australian. Proc. Linn. Soc. N. S. W., **53**, pp. 539–543 (1928).

#### NEMESTRINIDÆ

- Bequaert, J. North American Hirmoneuras. Jour. N. Y. Ent. Soc., 27, pp. 301-306 (1919).
  - Notes on American. Psyche, 37, pp. 286-297 (1930).
- Bezzi, M. South African Nemestrinidæ. Ann. S. Afr. Mus., 19, pp. 164–190 (1924).
- Cockerell, T. D. A. Review of Nemestrinidæ. Tr. Am. Ent. Soc., **34**, pp. 247–253 (1908).
- HARDY, G. H. Australian. Proc. Linn. Soc. N. S. W., 49, pp. 448–460 (1924).
- Lichwardt, B. Beiträge z. Kenntnis der Nemestriniden. Deut. ent. Zts. 1909, 1910.
- MACKERRAS, I. M. Revision Australasian. Proc. Linn. Soc. N. S. W., 50, pp. 489-561 (1925).
- Verrall, G. H. Palæarctic. Brit. Flies, 5, pp. 440-446 (1909).
- Williston, S. W. Synopsis nearctic. Can. Ent., 15, p. 69 (1883).

# PANTOPHTHALMIDÆ

- Austen, E. E. Revision. Proc. Zool. Soc. Lond. 1923, pt. 3, pp. 551-598 (1923).
- Enderlein, G. Keys to species. Zool. Anz., **41**, pp. 97–118 (1912); **52**, p. 228 (1921).

# RHAGIONIDÆ, including XYLOPHAGIDÆ

- ALDRICH, J. M. North American Symphoromyia. Proc. U. S. Nat. Mus., 49, pp. 113-142 (1915).
- Becker, T. Neue Rhagionidæ. Zts. f. wiss. Insektenbiol., 2, pp. 41, 54, 69 (1922).
- Bezzi, M. South African Rhagionidæ. Ann. S. Afr. Mus., **23**, pp. 297–324 (1926).
- Enderlein, G. Xylophagidæ. Zool. Anz., 43, pp. 533-552 (1913).
- Ferguson, E. W. Australian Leptidæ. Jour. R. Soc. N. S. W., 49, pp. 233–243 (1915).

- Leonard, M. D. Nearctic Rhagionidæ. Mem. Am. Ent. Soc. 7, 181 pp. (1930). Lindner, E. Palæarctic. Flieg. pal. Reg., Lief. 1–3, 49 pp. (1924–1925).
- Lundbeck, W. Danish. Dipt. Dan., **1**, pp. 75–82 (Xylophagidæ), 133–157 (Leptidæ) (1907).
- Séguy, E. French. Faun. d. France, 13, pp. 81- (Errinidæ), 88- (Cœnomyiidæ), 90-118 (Rhagionidæ) (1926).
- Verrall, G. H. British. Brit. Flies, 5, pp. 232-319 (1909).

# SCENOPINIDÆ

- CRESSON, E. T. North American. Trans. Am. Ent. Soc., 33, pp. 109–114 (1907).
- Kröber, O. Die Omphraliden. Ann. Hist. Nat. Mus. Hung., 2, pp. 174–210 (1913).
  - Family Omphralidæ. Gen. Ins., fasc. 161, 15 pp. (1914).
  - Egyptian. Bull. Soc. R. Ent. Egypt. 1923, pp. 70–84 (1924).
  - Palæarctic. Flieg. pal. Reg., Lief 4, 8 pp. (1925).
- Lundbeck, W. Danish. Dipt. Dan., 2, pp. 155-160 (1908).
- Séguy, E. Omphralidæ of France. Faune de France, 13, pp. 275-279 (1926).

#### STRATIOMYIIDÆ

- Brunetti, E. Oriental Stratiomyiidæ. Rec. Ind. Mus., 1, pp. 85–132 (1907); 25, pp. 45–180 (1923).
- Curran, C. H. Canadian. Trans. R. Soc. Can. Ottawa, Sec. V (3), 21, pp. 191–228 (1927).
- Enderlein, G. Subfamilies, etc. Zool. Anz., **43**, pp. 289–315, 577–615 (1914); **44**, 1–25 (1915).
  - Stratiomyiidæ and related families. Mitt. Zool. Mus. Berl., 10, pp. 153–214 (1921).
- JOHNSON, C. W. Nearctic Stratiomyia and Odontomyia. Tr. Am. Ent. Soc., 22, pp. 227–278 (1895).
- Kertész, K. Notacanthen. Ann. Mus. Hung., **14**, pp. 123–218 (1916). Revision of Potamidinæ. Ann. Mus. Hung., **18**, pp. 153–165 (1921).
- LUNDBECK, W. Danish. Dipt. Dan., 1, pp. 13-75 (1907).
- Malloch, J. R. Nearctic Pachygasterinæ. Ann. Ent. Soc. Am., 8, pp. 305—320 (1915).
- Melander, A. L. Nearctic Nemotelus. Psyche, **10**, pp. 171–183 (1903). Several nearctic genera. Can. Ent., **36**, pp. 14–24, 53–54 (1904).
- MILLER, D. New Zealand Beridinæ. Tr. N. Zeal. Inst., 49, pp. 172–194 (1917).
- Pleske, T. Palæarctic. Encycl. Ent. B. Dipt. II., 1, pp. 95– (Pachygasterinæ); 105–, 165–, (Clitellarinæ) (1924); 2, pp. 23– (Eulalia) (1925); Palæarctic Errinidæ and Cœnomyiidæ. Encycl. Ent. B. II. Dipt. 2, pp. 161–184 (1926).
  - Studies on palæarctic Stratiomyidæ. Eos, 2, pp. 385-420 (1926).

SÉGUY, E. French. Faune de France, 13, pp. 16–80 (1926).
 VERRALL, G. H. British. Brit. Flies, 5, pp. 51–229 (1909).

#### TABANIDÆ

Bequaert, J. Tabanid Fauna of Belgian Congo. Rept. Harvard Afr. Exped., Harv. Univ. Press, 2, pp. 859–971 (1930).

Efflatoun, H. C. Egyptian. Mem. Soc. R. Ent. Egypt, 4, fasc. 1,114 pp. (1930).
 Enderlein, G. Ein neues Tabanidensystem. Mitt. Zool. Mus. Berl., 10, pp. 333-351 (1922); 11, 253-409 (1925).

FERGUSON, E. W. and G. F. HILL. Australian, Proc. Linn. Soc. N. S. W., 45, pp. 460-467 (1920); 48, 381-435 (1923).

HINE, J. S. Tabanidæ of Ohio. Ohio Acad. Sci. Spec. Pap. 5, 63 pp. (1903).
Tabanidæ of Western U. S. and Canada. Ohio Nat., 5, pp. 217–248 (1904).

Louisiana horseflies. La. Crop Pest Comm. Circ. 6, 43 pp. (1906); La. Exp. Sta. Bull. 93, 59 pp. (1907).

Neotropical. Occ. Papers Mus. Zool. Univ. Mich. 162, 35 pp. (1925).

Knab, F. Stibasoma. Proc. U. S. Nat. Mus., 46, pp. 407-412 (1913).

Kröber, O. Palæaretic Chrysops. Zool. Jahrb., 43, Syst., pp. 41–160 (1920).
Palæaretic. Arch. Naturg., 89, Abt. A, 12 Heft, pp. 55–118; 90, A, 9 Heft, pp. 1–195 (1924).

Egyptian. Bull. Soc. R. Ent. Egyp., 1925, pp. 77-137 (1925).

Neotropical Chrysopini. Konowia, 4, pp. 210-375 (1925).

Palæarctic. Flieg. pal. Reg., Lief. 5-8, 146 pp. (1925).

Nearctic Chrysops. Stett. ent. Ztg., 87, pp. 209-353 (1926).

African Chrysops. Zool. Jahrb., 53, Syst., pp. 175–268 (1927).

Diachlorinæ. Beih. Arch. Schiffs- u. Tropenhyg., 32, pp. 73–123 (1928).

Neotropical. Zool. Anz., **88**, pp. 225–238 (Silvini); 305–312 (Pitocerini); **89**, 211–228 (Pangoniini) (1930).

Patagonian. Dipt. Pat. Chile, 5, fasc. 2, pp. 106–161 (1930).

LUNDBECK, W. Danish. Dipt. Dan., 1, pp. 85–132 (1907).

Lutz, A. Brazilian. Zool. Jahrb. Suppl., 10, pp. 619–692 (1909).

McAtee, W. L. and W. R. Walton. District of Columbia species. Proc. Ent. Soc. Wash., 20, pp. 188–206 (1919).

OSTEN SACKEN, C. R. United States species. Mem. Bost. Soc. Nat. Hist., 2, pp. 365-397, 421-479, 555-560 (1878).

SÉGUY, E. French. Faune de France, 13, pp. 119-157 (1926).

SENIOR-WHITE, R. Indian. Govt. India Centr. Publ. Br., pt. 12, 70 pp. (1927). SURCOUF, J. M. R. Monogr. des Taban. d'Afrique. Paris. 258 pp. (1909).

Family Tabanidæ. Gen. Ins., fasc. 175, 182 pp. Suppl., pp. 183–205 (1921).

Szilady, Z. Palæarctic Chrysops. Arch. f. Nat., 83, pp. 85- (1917).

Verrall, G. H. British. Brit. Flies, 5, pp. 320-439 (1909).

Wehr, E. E. Nebraska Tabanidæ. Univ. Studies, Nebr., 12, pp. 107-118 (1922).

#### THEREVIDÆ

Becker, T. Synopsis of Thereva. Konowia, 1, pp. 16-34 (1922).

Cole, F. R. Revision N. Am. Therevidæ. Proc. U. S. Nat. Mus., 62, art. 4, 140 pp. (1923).

Kröber, O. Therevidæ of S. and Centr. America. Ann. Hus. Hung., 9, pp. 475- (1911).

Palæarctic and ethiopian. Deut. ent. Zts. 1912, 6 parts (1912).

Indoaustralian. Ent. Mitt., 1, p. 115 et seq., 6 parts (1912).

North American. Stett. ent. Ztg., 73, pp. 209-272 (1912).

Family Therevidæ. Gen. Ins., fasc. 148, 69 pp. (1913).

Egyptian. Bull. Soc. R. Ent. Egypt. 1923, pp. 85-107; 1929, 73-80.

Palæarctic. Flieg. pal. Reg., Lief. 1-3, 60 pp. (1925).

Lundbeck, W. Danish. Dipt. Dan., 2, pp. 132–155 (1908).

Mann, J. S. Australian. Austr. Zool., 5, pp. 151-194 (1928); 6, 17-49 (1929).

OSTEN SACKEN, C. R. Tables, etc. Verh. z. b. Ges. Wien, **62**, pp. 289–319 (1912).

Séguy, E. French. Faune de France, 13, pp. 255-275 (1926).

VERRALL, G. H. British. Brit. Flies, 5, pp. 537-589 (1909).

# B. CYCLORRHAPHA, GROUP ASCHIZA CONOPIDÆ

Banks, N. Synopsis of nearctic Zodion and Myopa. Ann. Ent. Soc. Am., 9, pp. 191–200 (1916).

BECKER, T. Neue Dipteren: Conopidæ. Konowia, 1, pp. 195–208; 282–295 (1922).

Kröber, O. Stylogaster. Entom. Mitt., 3, pp. 338–353 (1914).

N. Am. Conops. Arch. Naturg., **81**, A, Heft 5, pp. 121–160 (1915). Katalog Conopidæ (with tables). Arch. Naturg., **83**, A., Heft 8. pp. 1–91; Heft 9, pp. 1–52 (1917).

Egyptian Conopidæ. Bull. Soc. R. Ent. Egypt, 1923, pp. 57–70 (1924). Palæarctic Conopidæ. Flieg. pal. Reg. Nos. 1–3, 48 pp. (1924–1925).

Conopidæ of Philippines. Philipp. Jour. Sci., **34**, pp. 331–347 (1927). Conopidæ of C. Eur. Tierwelt Deutschl., pt. 20, pp. 119–142 (1930).

Séguy, E. Conopidæ of W. Eur. Encycl. Ent. ser. A, No. 9, pp. 9–52 (1928).
VAN DUZEE, M. C. N. Am. Conopidæ. Proc. Cal. Acad. Sci. (4), 16, pp. 573–604 (1927).

WILLISTON, S. W. N. Am. Conopidæ. Trans. Conn. Acad. Sci., 4, pp. 325–342 (1882); 6, pp. 5-, 377- (1883).

#### LONCHOPTERIDÆ

Duda, O. Lonchoptera. Konowia, **6**, pp. 89–99 (1927). Lundbeck, W. Danish Lonchopteridæ. Dipt. Dan., **5**, pp. 1–18 (1916). DE MEIJERE, J. C. H. Palæarct. Lonchopteren. Tijd. Ent., **49**, pp. 44–98 (1906).

#### PHORIDÆ AND TERMITOXENIIDÆ

- Becker, T. Monograph European Phoridæ. Abhandl. z. b. Ges. Wien, 1, pp. 1–100 (1901).
- Brues, C. T. Monograph N. Am. Phoridæ. Trans. Am. Ent. Soc., 29, pp. 331-404 (1903).

Family Phoridæ. Gen. Ins., fasc. 44, 21 pp. (1906).

Synonymic catalogue of Phoridæ. Bull. Wisc. Nat. Hist. Soc., **12**, pp. 85–152 (1915).

Lundbeck, W. Danish Phoridæ. Dipt. Dan., 6, pp. 69-447 (1922).

Malloch, J. R. U. S. Phoridæ. Proc. U. S. Nat. Mus., 43, pp. 411-529 (1913).

Schmitz, H. Myrmecophilous Phoridæ. Zool. Jahrb., **37**, Syst., pp. 509–566 (1914).

Myrmecophilous and termitophilous Phoridæ. Jaarb. Natuurh. Gen. Limburg, 1913, pp. 123–133 (1914).

Neue termitophile Dipteren. Zool. Jahrb., **39**, Syst., pp. 211–266 (1916).

Keys to European Phoridæ. Jaarb. Natuurb. Gen. Limburg, 1917, pp. 79–150; 1919, 91–152 (1919).

Geograph. distribution Europ. Phoridæ. Natuurhist. Maanblad, Limburg, 17, pt. 10, pp. 139–147 (1928).

Revision of Phoridæ. Dümler's, Berlin u. Bonn. 211 pp. (1929).

WASMANN, E. Revision of oriental Termitoxeniidæ. Ann. Soc. Ent. Belg., 57, pp. 16–22 (1913).

## PIPUNCULIDÆ

Becker, T. Revision of European Pipunculidæ. Berl. Ent. Zts., 42, pp. 25–100 (1897).

Tables of new Pipunculidæ. Wien. ent. Ztg., **38**, pp. 123–132; 149–167 (1921).

CRESSON, E. T. N. Am. Pipunculidæ. Trans. Am. Ent. Soc., 36, pp. 267–320 (1910).

HOUGH, G. DE N. Pipunculidæ of U. S. Proc. Bost. Soc. Nat. Hist., 29, pp. 77-86 (1899).

Lundbeck, W. Danish Pipunculidæ. Dipt. Dan., 6, pp. 1-69 (1922).

Perkins, R. C. L. Australian and Hawaiian Pipunculidæ. Bull. Haw. Sugar Plant. Assn., Entom., 1, pp. 123–157 (1905).

Verrall, G. H. British Pipunculidæ. Brit. Flies, 8, pp. 60–126 (1901).

#### PLATPEZIDÆ AND SCIADOCERATIDÆ

Johnson, C. W. Platypezidæ of Eastern N. Am. Occ. Pap. Bost. Soc. Nat. Hist. 5, pp. 51–58 (1923).

LUNDBECK, W. Danish Platypezidæ. Dipt. Dan., 7, pp. 1-39 (1927).

SCHMITZ, H. Sciadoceridæ. Dipt. Patagon. S. Chile, 6, pt. 1, pp. 1-11 (1929).

- Snow, W. A. American Platypezidæ. Kans. Univ. Quart., 3, pp. 143–152 (1894); 205–207 (1895).
- Verrall, G. H. British Platypezidæ. Brit. Flies, 8, pp. 11-59 (1901).

#### SYRPHIDÆ

- Banks, Greene, McAtee and Shannon. Syrphidæ of District of Columbia, Proc. Biol. Soc. Wash., 29, pp. 173–204 (1916).
- Becker, T. Revision of Chilosia. Acta Ac. Germ., **62**, pp. 195–522 (1894). Genus Helophilus. Berl. ent. Zts., **55**, pp. 213–232 (1910).

New Syrphidæ. Mitt. Zool. Mus. Berl., 10, pp. 1-93 (1921).

- Bezzi, M. Syrphidæ of ethiop. region. Brit. Mus. Pub. 146 pp. (1915).
- Brèthes, J. Masarygidæ, n. fam. An. Mus. Buen. Air. (3), 10, p. 439 (1909).
- Brunetti, E. Fauna Brit. India, Lond., **3**, 424 pp. (Pipunculidæ, Syrphidæ, Conopidæ, Œstridæ) (1923).
- CURRAN, C. H. Nearct. Pipiza. Proc. Cal. Acad. Sci. (4), 11, pp. 345–393 (1921).

Review of nearct. Syrphus, pt. Can. Ent., **53**, pp. 152–159 (1921). Revision N. Am. Syrphidæ. Kans. Univ. Sci. Bull., **15**, pp. 1–216 (1925).

S. Afr. Syrphidæ. Bull. Am. Mus. Nat. Hist., **57**, pp. 44–83 (1927). Syrphidæ of Malay Peninsula. Jour. Fed. Mal. States Mus., **14**, pt. 2,

pp. 141–324 (1928).
 Curran, C. H. and C. L. Fluke. Revision of nearct. Helophilus. Trans.
 Wisc. Acad. Sci., 22, pp. 207–281 (1926).

Efflatoun, H. C. Monograph Egyptian Syrphidæ. Mem. Soc. R. Ent. Egypt, **2**, fasc. 1, 123 pp. (1922).

FERGUSON, E. W. Revision Australian Syrphidæ. Proc. Linn. Soc. N. S. W., 51, pp. 517-544 (1926).

Fluke, C. L. Revision Wisconsin Syrphidæ. Trans. Wisc. Acad. Sci., 20, pp. 215–253 (1921).

Hull, F. M. N. Am. Eristalis. Ohio Jour. Sci., 25, pp. 11–42; 285–310 (1925).

JONES, C. R. Syrphidæ of Colorado. Ann. Ent. Soc. Am., 10, pp. 219–231 (1917); Col. Agr. Exp. Sta. Bull. 269, 72 pp. (1922).

Kertész, K. S. Am. Ceria. Termes. Füzet., 25, pp. 85–90 (1902).

Lundbeck, W. Danish Syrphidæ. Dipt. Dan., 5, pp. 18–591 (1916).

METCALF, C. L. Syrphidæ of Ohio. Ohio State Univ. Bull. 17, No. 31, pp. 1–123 (1913).

Syrphidæ of North Carolina. Jour. Elisha Mitchell Soc., **32**, pp. 95–112 (1916).

Syrphidæ of Maine. Maine Exp. Sta. Bull. 253, pp. 193–264 (1916); 263, pp. 153–176 (1917).

MILLER, D. New Zealand Syrphids. Trans. N. Z. Inst., 53, pp. 289–333 (1921). SACK, P. Salpingogaster and Meromacrus. Zool. Jahrb. Syst., 43, pp. 235–272 (1920).

Palæarct. Syrphidæ. Flieg. Pal. Reg., Lief. 30, 32, 34, 144 pp. (1928–1929).

C. Eur. Syrphidæ. Tierwelt Deutschl., pt. 20, pp. 1–118 (1930).

Shannon, R. C. Keys to various groups nearct. Syrphidae. Proc. Ent. Soc. Wash., 18, pp. 101–113 (1916). Proc. Biol. Soc. Wash., 29, p. 203 (1916).
Bull. Brkl. Ent. Soc., 16, pp. 65–72; 120–128 (1921); 17, pp. 30–42 (1922); 18, pp. 17–21 (1923).

N. Am. Chilosia. Insecutor Insc. Menstr., 10, pp. 117-145 (1922).

N. Am. Xylotinæ. Proc. U. S. Nat. Mus., 69, art. 9, 52 pp. (1926).

N. Am. Chrysotoxinæ. Proc. U. S. Nat. Mus., 69, art. 11, 20 pp. (1926).

N. Am. Cerioidinæ. Jour. Wash. Acad. Sci., **17**, No. 2, pp. 38–55 (1927).

Review of S. Am. Syrphidæ. Proc. U. S. Nat. Mus., **70**, art. 9, 34 pp. (1927).

Shiraki, T. Syrphidæ of Japan. Mem. Taihoku Imp. Univ. Ent., 1, 446 pp. (1930).

STACKELBERG, A. A. Palæaret. Cinxiinæ. Acad. Sci. U. R. S. S., **20**, 27 pp. (1927).

Palæaret. Zelima (= Xylota). Deut. ent. Zts., 1925, pp. 279–288 (1925). Palæaret. Cynorrhina. Konowia, 7, pp. 252–258 (1928).

Verrall, G. H. British Syrphidæ. Brit. Flies, 8, pp. 127–676 (1901).

Wehr, E. E. Syrphidæ of Nebraska. Univ. Studies, Nebr., 22, pp. 119–162 (1922).

WILLISTON, S. W. N. Am. Syrphidæ. Bull. U. S. Nat. Mus. 31, xxx. and 335 pp. (1886).

# CALYPTRATÆ, GENERAL

Brauer, F. and Bergenstamm, J. E. Monograph of Schizometopa (tables of about 50 fams.)
Denks. kais. Wiss. Akad. Wien, 56, pp. 69–158 (1889).
Zweiflügler des kaiserlichen Museums Wien. Denks. kais. Wiss. Akad. Wien, 7 pts., 1880–1894
Facsimile reprint, Junk, Berlin, 760 pp. (1923).

GIRSCHNER, E. Beiträge z. Systematik der Musciden. Berl. Ent. Zts., 38, pp. 297–312 (1893).

Ein neues Musciden-System. Ill. Wochenschr. Ent., **1**, pp. 12–16; 30–32, etc., 16 pp. (1896).

SHANNON, R. C. and E. Del Ponte. Argentine muscoids. Rev. Inst. bact. Buen. Aires, 4, pp. 549–590 (1926); 5, pp. 141-147 (1927).

Townsend, C. H. T. Catalog S. Am. Calyptrate Muscidæ. Ann. N. Y. Acad. Sci., 7, pp. 1–44 (1893).

Taxonomy of Muscoidean flies. Smiths. Misc. Coll., **51**, pp. 1–138 (1908).

Muscoideos da regiao humida tropical da America. Rev. Mus. paul., São Paulo, **15**, pt. 1, 205–386 (1927).

#### ANTHOMYIIDÆ

- ALDRICH, J. M. Nearctic Lispa. Jour. N. Y. Ent. Soc., 21, pp. 126–146 (1913).
  Kelp flies (Fucellia) of N. Am. Proc. Cal. Acad. Sci., 8, pp. 157–179 (1918).
- Becker, T. Palæarctic Lispa. Zts. f. Ent., 29, pp. 1-70 (1904).
- HUCKETT, H. C. Anthomyinæ of New York. Cornell Univ. Mem. 77, pp. 1-91 (1923).
- KARL, O. German Anthomyiidæ. Tierw. Deutsch. No. 13, pp. 15–232 (1928).
  MALLOCH, J. R. Subfamilies of Anthomyiidæ. Can. Ent., 49, pp. 406–408 (1917).

Synopsis of various N. Am. genera. Can. Ent., **52**, pp. 253-, 271-(1920); **53**, various parts (1921). Ann. Mag. Nat. Hist. (9), **11**, pp. 74-81, 664- (1923).

Cœnosiinæ. Ent. News, **32**, pp. 106-, 201- (1921). Ann. Mag. Nat. Hist. (9), **10**, pp. 576- (1922).

N. Am. Phaoniine. Trans. Am. Ent. Soc., 48, pp. 227-282 (1922).

Various Australian genera. Proc. Linn. Soc. N. S. W., 50, pp. 36–45 (1925).

- RINGDAHL, O. Nordische Anthomyiden. Ent. Tidskr., **39**, pp. 148–194 (1918). SCHNABL, J. and H. DZIEDZICKI. Die Anthomyiden. Nova Acta Leop., Halle, **95**, pp. 53–358 (1911).
- SÉGUY, E. Anthomyiidæ of S. W. Europe. Faune de France, 393 pp. (1923). STEIN, P. Homalomyia group. Berl. ent. Zts., 40, pp. 1–141 (1895).

Die Anthom. Europas. Arch. f. Natg., **81**, A (10), pp. 1–224 (1916). Revision N. Am. Anthomyiidæ. Berl. ent. Zts., **42**, pp. 161–288 (1897). Arch. Naturg., **84**, pt. 2, A, 8, pp. 1–106 (1920).

Anthomyiidæ of S. Am. Arch. Naturg, 77, pp. 61–189 (1911). Genera of world. Arch. Naturg., 83, A, 1 (1917), pp. 85–178 (1919).

# CALLIPHORIDÆ AND SARCOPHAGIDÆ

- ALDRICH, J. M. N. Am. Sarcophaga and allies. Thos. Say Foundat., 1, 302 pp. (1916).
- Allen, H. W. N. Am. Miltogrammini. Proc. U. S. Nat. Mus. 68, art. 9, 106 pp. (1926).
- Enderlein, G. Sarcophaginæ, s. str. Arch. Klass. Phylog. Ent. Wien, 1, pp. 1-56 (1928).
- Engel, E. O. Rutillidæ, s. lat. Zool. Jahrb. Syst., 50, pp. 339–376 (1925).
   Johnson, T. H. and G. H. Hardy, Australian Sarcophaga. Proc. Linn. Soc.
   N. S. W., 48, pp. 94–129 (1923).
- MALLOCH, J. R. Australian Calliphoridæ. Proc. Linn. Soc. N. S. W., 52, pp. 299-335 (1927).
  - African Calliphoridæ. Ann. Mag. Nat. Hist. (10), **3**, pp. 272–278; **4**, p. 113 (1929).

- PARKER, R. R. Ravinia and Boettcheria of N. Engl. Proc. Bost. Soc. Nat. Hist., 35, pp. 1–77 (1914).
- ROHDENDORF, B. Subfams. Sarcophagidæ. Konowia, 7, pp. 319–321 (1928). Palæarct. Sarcophagidæ. Flieg. pal. Reg. Lief. 39, pp. 1–48 (1930).
- SÉGUY, E. Calliphoridæ of France. Encycl. Ent., A, 9, pp. 89–192 (1928). SENIOR-WHITE, R. Oriental Sarcophagidæ. Rec. Ind. Mus., 26, pp. 193–283 (1924).

Oriental Calliphoridæ. Rec. Ind. Mus., 28, pp. 127–140 (1926).

- SHANNON, R. C. N. Am. Calliphoridæ. Insec. Insec. Menstr., 11, pp. 101–118 (1923); 12, pp. 67–81 (1924). Soc. Ent. Stuttgart, 28, pp. 115–139 (1926). Proc. Ent. Soc. Wash., 28, pp. 115–139 (1926).
- Shannon, R. C. and E. Del Ponte. Argentine Calliphoridæ. Rev. Inst. Bact. Buen. Aires, 4, pp. 583–587 (1926).
- Townsend, C. H. T. Genera of Sarcophagini. Proc. Biol. Soc. Wash., 30, pp. 189–198 (1917).

### MUSCIDÆ AND GLOSSINIDÆ

- ALDRICH, J. M. Neotrop. Mesembrinella. Proc. U. S. Nat. Mus., 62, art. 11, 24 pp. (1922).
- Austen, E. E. Monogr. Tsetse-flies. Brit. Mus. Publ., 319 pp. (1903). Handbook of the Tsetse. Brit. Mus. Publ., 110 pp. (1911).
- Austen, E. E. and E. Hegh. Tsetse flies. Imp. Bur. Ent. Publ., 188 pp. (1922).

Enderlein, G. Stomoxinæ. Zts. angew. Ent., 14, pp. 356-368 (1928).

Hegh, E. Les Tsé-tsés. Bruxelles, 742 pp. (1929).

- Karl, O. German Muscidæ. Tierw. Deutsch. No. 13, pp. 1-15 (1928).
- Malloch, J. R. Australian Muscinæ. Proc. Linn. Soc. N. S. W., **50**, pp. 45, 46 (1925).
- Surcouf, J. M. R. Revision de Muscides testaceæ. Nouv. Arch. Mus. Hist. nat. (5), **6**, pp. 27–124 (1919).
- Townsend, C. H. T. Connectant forms between muscoid and anthomyoid flies. Ann. Ent. Soc. Am., 7, pp. 160–167 (1914).

### CORDYLURIDÆ

- Becker, T. Scatomyzidæ. Berl. ent. Zts., 39, pp. 77-196 (1894).
- Coquillett, D. W. Scatophagidæ. Jour. N. Y. Ent. Soc., **8**, pp. 160-165 (1898).
- Curran, C. H. Several nearct. genera. Can. Ent., 59, pp. 253-261 (1927).
   Johnson, C. W. Several nearct. genera. Psyche, 34, pp. 100-104 (1927).
- MALLOCH, J. R. Generic key, etc. Rept. Can. Arct. Exped. 1913–18, 3, pt. C, pp. 75–81 (1919).

Various nearct. genera. Ohio Jour. Sci., **20**, pp. 286– (1920). Bull. Brkl. Ent. Soc., **17**, pp. 77– (1922). Ent. News, **34**, pp. 139–, 179– (1923). N. Am. Fauna, **46**, pp. 201– (1923).

### ŒSTRIDÆ

- BAU, A. Family Œstridæ. Gen. Ins., fasc. 43, 31 pp. (1906).
- Brauer, F. Œstridæ. Verh. z. b. Ges. Wien, 43, pp. 447-525 (1893).
- Brauer, F. and J. E. Bergenstamm. Tables genera, etc. Denks. kais. Akad. Wiss. Math. Naturw. Wien, **56**, pp. 158–161 (1889); **60**, pp. 89–240 (1893).
- PLESKE, T. Palæarct. Œstridæ. Ann. Mus. Zool. Leningrad, 26, pp. 215–230 (1926).
- RODHAIN, J. and J. BEQUAERT. African Estrinæ. Bull. scient. France, Belg., (7), 50, p. 53–165 (1916–17).
- SÉGUY, E. Genera Œstridæ. Encycl. Ent. B. Dipt., 2, pt. 3, pp. 1-10 (1926).
  Œstridæ of W. Eur. Encycl. Ent. A, 9, pp. 53-88 (1928).
- Townsend, C. H. T. Revision Cuterebridæ. Insecut. Insc. Mens., 5, pp. 23–28 (1917).

#### TACHINIDÆ

- Aldrich, J. M. Metallic green tachinids. Insecut. Insc. Mens., 14, pp. 51–58 (1926).
  - Species with retracted hind crossvein. Trans. Am. Ent. Soc., **52**, pp. 7–28 (1926).
- ALDRICH, J. M. and R. T. WEBBER. N. Am. Phorocera, etc. Proc. U. S. Nat. Mus., 63, art. 17, 90 pp. (1924).
- BAER, W. Classification, etc. Zts. angew. Ent., 6, pp. 185-246 (1920); 7, pp. 97-163; 349-423 (1921).
- Bezzi, M. Fissicorn Tachininæ. Proc. Linn. Soc. N. S. W., 48, pp. 647–659 (1923).
  - Genera with 4 vein obliterated at apex. Ann. Mag. Nat. Hist. (9), 17, pp. 236–238 (1926).
- Coquillett, D. W. N. Am. Tachinidæ. Bull. U. S. Dept. Agr. Tech. Ser. No. 7, 156 pp. (1897).
- CURRAN, C. H. African Tachinidæ. Bull. Ent. Res., 17, pp. 319-340; 18, pp. 103-128 (1927), 237-245 (1928).
  - Am. Archytas. Can. Ent., 60, pp. 201-, 218-, 249-, 275-, (1928).
- ENGEL, E. O. Neotrop. Hystriciidæ. Zool. Jahrb. Syst., 43, 273–328 (1920). LUNDBECK, W. Danish Tachinoidea. Dipt. Dan., 7, pp. 40–560 (1927).
- Malloch, J. R. Exotic Rhiniinæ. Ann. Mag. Nat. Hist. (9), **18**, pp. 496–522 (1926).
  - Catalog Australian Tachinidæ. Proc. Linn. Soc. N. S. W., **53**, pp. 651–662 (1928). Keys, **54**, p. 337 (1929).
    - Ormiini. Ann. Mag. Nat. Hist. (10), 3, pp. 278-280 (1929).
- Müller, A. Determination of German, by 3 genitalia. Nova Acta, Leop., **108**, pp. 1–30 (1926). Konowia, **5**, pp. 72–, 233, 246 (1926).
- Senior-White, R. Oriental Rhiniinæ. Rec. Ind. Mus., 27, pp. 81–96 (1925).

- Tothill, J. D. Nearct. Ernestia. Can. Ent., **53**, pp. 199-, 226-, 247-, 270- (1921).
- Townsend, C. H. T. Tables, etc., N. Am. Trans. Am. Ent. Soc., 19, pp. 88–289 (1892).

Dejeaniini. Psyche, 20, pp. 102-106 (1913).

Relationships of muscoid flies. Can. Ent., 45, pp. 37-60 (1913).

Indian Rhiniinæ. Rec. Ind. Mus., 13, Pt. 4, pp. 185-202 (1917).

Wainwright, C. J. British Tachinidæ. Trans. Ent. Soc. Lond., 76, pt. 1, pp. 139–254 (1928).

### ACALYPTRATÆ, GENERAL

Frey, R. Dipt. schizophora. Act. Soc. Fenn., 48, 245 pp. (1921).

Hendel, F. Papers on various Acalyptrates. Wien. ent. Ztg., 29, pp. 101–127 (1910); 30, pp. 19–46 (1911). Deut. ent. Zts. 1914, pp. 151–176 (American); 1917, pp. 33–47. Entom. Mitt., 5, pp. 294–299 (1916).

Palæarct. Acalyptratæ. Konowia, **1**, pp. 145–160; 253–365 (1922). Tierw. Deutschl., Dipt., pt. 2, 135 pp. (1928).

STURTEVANT, A. H. Notes on Acalyptratæ. Am. Mus. Novitat., No. 76, 12 pp. (1923).

## MICROPEZIDÆ, NERIIDÆ, TANYPEZIDÆ

CRESSON, E. T. N. Am. Micropezidæ. Trans. Am. Ent. Soc., 52, pp. 260–274 (1926).

Enderlein, G. Classification of Tanypezidæ. Zool. Anz., 42, pp. 224-229 (1913).

Classification of Micropezidæ. Arch. Naturg. Berl., A, 88, pt. 5, pp. 140–229 (1922).

Frey, R. Micropezidæ. Not. Ent., 7, pp. 65-76 (1927).

African Tanypodinæ. Ann. Mag. Nat Hist. (10), 3, pp. 313–318 (1929).

VAN DUZEE, M. C. Nearctic Micropeza. Pan-Pacif. Ent., 3, pp. 1-11 (1926).

# CLUSIIDÆ, CŒLOPIDÆ, MEGAMERINIDÆ, SEPSIDÆ

CZERNY, L. Heteroneuriden. Wien. ent. Ztg., 22, pp. 61–107 (1903). Palæarct. Clusiidæ. Flieg. pal. Reg., pt. 28, 12 pp. (1928).

Duda, O. Monograph of Sepsidæ. Ann. Nat. Hist. Mus. Wien, 39, pp. 1–153 (1925); 40, pp. 1–110 (1926).

FREY, R. Finland spp. of Sepsis. Deut. ent. Zts. 1908, 577–588 (1908).
 Palæarct. Sepsidæ. Notulæ Ent., 5, pp. 69–76 (1925).

Hendel, F. Megamerinidæ. Suppl. Entom., 2, pp. 90-93 (1913).

Malloch, J. R. Nearctic Clusiidæ. Proc. Ent. Soc. Wash., 20, pp. 2–8 (1918).Nearctic Coelopa. N. Am. Fauna, 46, p. 214 (1923).

MELANDER, A. L. and NAOMI ARGO. Family Clusiidæ. Proc. U. S. Nat. Mus., 64, art. 11, 54 pp. (1924).

Melander, A. L. and A. Spuler. Sepsidæ and Piophilidæ. Wash. Agr. Exp. Sta. Bull. 143, 103 pp. (1917).

### RHOPALOMERIDÆ, RHINOTORIDÆ, DIOPSIDÆ

- Frey, R. Philippine Diopsidæ. Notulæ Ent., 8, pp. 69-77 (1928).
- LINDNER, E. Revision of Rhopalomeridæ. Deut. ent. Zts. 1930, 122–137 (1930).
- Williston, S. W. Rhopalomeridæ. Psyche, 7, pp. 183–186 (1895).

## LAUXANIIDÆ, CELYPHIDÆ, PERISCELIDÆ, PALLOPTERIDÆ

- FREY, R. Philippine Lauxaniidæ. Act. Soc. F. Fl. Fenn., 56, No. 8, pp. 1–44 (1927).
- Hendel, F. Table of genera of Lauxaniidæ. Encycl. Ent. B. Dipt., 2, No. 2, pp. 103–142 (1925).

Family Lauxaniidæ. Gen. Ins., fasc. 68, 66 pp. (1908).

- JACOBSEN, G. Catalog of Celyphidæ. Ann. M. Z. Acad. Petersb. 1896, pp. 246–252 (1896).
- Kertész, K. Indo-austr. Sapromyza. Termesz. Füz., 23, pp. 254–276 (1900). Lauxaniidæ of Hungary. Math. Termt. Ert., 38, pp. 352–359 (1921).
- MALLOCH, J. R. and W. L. McAtee. Lonchæidæ, Pallopteridæ and Sapromyzidæ of E. U. S. Proc. U. S. Nat. Mus., 65, art. 12, 26 pp. (1925).
- MALLOCH, J. R. Periscelidæ. Proc. U. S. Nat. Mus., 68, art. 21, p. 24 (1926).
  Australian Sapromyzidæ. Proc. Linn. Soc. N. S. W., 52, pp. 399-421 (1927).

Oriental Lauxaniidæ. Proc. U. S. Nat. Mus., 74, art. 6, 97 pp. (1929).

Melander, A. L. Nearctic Sapromyzidæ. Psyche, **20**, pp. 57–82 (1913). Tonnoir, A. L. and J. R. Malloch. New Zealand Sapromyzidæ. Rec. Cant. Mus., **3**, pt. 1, pp. 19–26 (1926).

# TETANOCERATIDÆ, DRYOMYZIDÆ

- Cresson, E. T. Nearctic Sciomyzidæ. Trans. Am. Ent. Soc., 46, pp. 27–89 (1920).
- Frey, R. North-palearct. Tetanocera. Notulæ Ent., 4, pp. 47-53 (1924).
- HENDEL, F. Palæarct. Sciomyzidæ. Abh. z. b. Ges. Wien., 2, pt. 1, 92 pp. (1902).
  - Ethiopian and Indo-mal. Sepedon. Ann. Mus. Nat. Hung., 9, pp. 266–277 (1911).
- Kertész, K. Hungarian Tetanoceridæ. Adlatt. Közlem., 14, pp. 81–126 (1915). Math. Termt. Ert. Budapest, 39, pp. 124–130 (1922).
- MELANDER, A. L. Nearct. Tetanoceridæ. Ann. Ent. Soc. Am., **13**, pp. 305-322 (1920).
- Tonnoir, A. L. and J. R. Malloch. New Zealand Sciomyzidæ. Rec. Cant. Mus., 3, pt. 3, pp. 151–179 (1928).

ORTALID SERIES: PHYTALMIIDÆ, PLATYSTOMATIDÆ, PTERO-CALLIDÆ, PYRGOTIDÆ, RICHARDIIDÆ, TACHINISCIDÆ, ULIDIIDÆ

Bezzi, M. Phytalmiidæ. Rev. Zool. Afr., 12, pp. 225-239 (1924).

Bezzi, M. and J. R. Malloch. Australian Prygotidæ. Proc. Linn. Soc. N. S. W., **54**, pp. 1–31 (1929).

ENDERLEIN, G. Platystominen. Mitt. Zool. Mus. Berl., 11, pp. 97–153 (1924). HENDEL, F. Pyrogotinæ. Gen. Ins., fasc. 79, 33 pp. (1908).

Pterocallinæ. Gen. Ins., fasc. 96, 50 pp. (1909). Deut. ent. Zts. 1909, Beih. S, pp. 1–84 (1909).

Chrysomyza. Zool. Anz., **34**, pp. 612–622 (1909).

Euxesta. Ann. Mus. Nat. Hung., 7, pp. 151-172 (1909).

Ulidiinæ. Gen. Ins., fasc. 106, 76 pp. (1910).

Richardiinæ. Deut. ent. Zts. 1911, pp. 181–270; 367–396 (1911). Gen. Ins., fasc. 113, 56 pp. (1911).

Pyrgotinen. Arch. f. Naturg., 79, Abt. A, pp. 77-117 (1913).

Platystoma. Zool. Jahrb. Syst., 35, pt. 1, pp. 55-126 (1913).

Platystominæ. Abh. z. b. Ges. Wien, **8**, 410 pp. (1914). Gen. Ins., fasc. 157, 179 pp. (1914).

Loew, H. N. Am. Ortalidæ. Monogr. Dipt. N. Am., 3, 209 pp. (1873).

Malloch, J. R. African Pyrgotidæ. Ann. Mag. Nat. Hist. (10), **3**, pp. 249–261 (1929).

Tachiniscidæ. Ann. Mag. Nat. Hist. (10), 7, pp. 334-338 (1931).

### TRYPETIDÆ

Bezzi, M. Ceratitis, Anastrepha and Dacus. Boll. Lab. Portici, 3, pp. 273–313 (1909).

Indian Trypaneids. Mem. Ind. Mus., 3, No. 3, pp. 53-175 (1913).

Philippine Dacus. Phil. Jour. Sci., 15, pp. 411-443 (1919).

Ethiop. Fruit-flies. Bull. Ent. Res., **8**, pp. 215–251 (1918); **9**, pp. 13–46 (1919); **10**, pp. 211–271 (1920), **15**, pp. 73–155 (1924).

S. Afr. Trypetidæ. Ann. S. Afr. Mus., 19, pp. 449-577 (1924).

COQUILLETT, D. W. Genera of Trypetidæ, etc. Jour. N. Y. Ent. Soc., 7, pp. 259-268 (1899).

CRESSON, E. T. N. Am. Rhagoletis. Trans. Am. Ent. Soc., 55, pp. 401–414 (1929).

Efflatoun, H. C. Egyptian Trypetidæ. Mem. Soc. R. Ent. Egypt, 2, No. 2, 132 pp. (1924).

ENDERLEIN, G. Studies on Trypetidæ. Zool. Jahrb. Syst., **31**, pp. 407–460 (1911).

Hendel, F. Bohrfliegen Südamerikas. Abh. Ber. K. Zool. Anthrop. Mus. Dresden, 14, pp. 1–84 (1914).

Table of genera of Tephritinæ. Wien. ent. Ztg., 33, pp. 60-98 (1914).

- LOEW, H. N. Am. Trypetidæ. Monogr. Dipt. N. Am., pt. 1, pp. 49-102 (1862); pt. 3, pp. 211-347 (1873).
  - Europäischen Bohrfliegen. Wien, 128 pp. (1862). Reprint, Junk, Berlin (1923).
- Phillips, V. T. N. Am. Trypetidæ. Jour. N. Y. Ent. Soc., **31**, pp. 119-154 (1923).
- Senior-White, R. Indian Trypetidæ. Cat. Ind. Ins. Govt. Centr. Pub. Br., pt. 4, 33 pp. (1924).

### LONCHÆIDÆ, PIOPHILIDÆ, PSILIDÆ

- Bezzi, M. Lonchæidæ. Bull. Ent. Res. 9, pp. 241-254 (1919); 11, pp. 199-210 (1920).
- Duda, O. Piophila. Konowia, 3, pp. 97–203 (1924).
- Kertész, K. Indo-austral. Lonchæidæ. Termés. Füzet., 24, pp. 82-87 (1901).
- MALLOCH, J. R. and W. L. McAtee. Lonchæidæ, Pallopteridæ and Sapromyzidæ of E. U. S. Proc. U. S. Nat. Mus., 65, art. 12, 26 pp. (1924).
- Melander, A. L. Nearct. Psilidæ. Psyche, 27, pp. 91–101 (1920).
- Nearct. Piophilidæ. Psyche, **31**, pp. 78–86 (1924).

  MELANDER, A. L. and A. Spiller, N. Am. Piophilidæ. Wash. A
- Melander, A. L. and A. Spuler. N. Am. Piophilidæ. Wash. Agr. Exp. Sta. Bull. 143, pp. 53–76 (1917).

## HELOMYZIDÆ, CHYROMYIIDÆ, TRICHOSCELIDÆ

- Aldrich, J. M. Key to genera of Helomyzidæ. Insecut. Insc. Mens., 14, pp. 96-102 (1926).
- ALDRICH, J. M. and P. S. DARLINGTON. N. Am. Helomyzidæ. Trans. Am. Ent. Soc., 34, pp. 67–100 (1908).
- Becker, T. Peletophila. Zts. Hym. Dipt., 4, pp. 129-133 (1904).
- Bezzi, M. Chyromyiidæ. Atti Soc. It. Sci. Nat. Milano, 43, 177–181 (1904).
- CZERNY, L. Palæarct. Helomyzidæ, Trichoscelidæ and Chyromyiidæ. Flieg. pal. Reg., pt. 22, 54 pp. (1927).
  - Palæarct. Helomyzidæ. Wien. ent. Ztg., **23**, pp. 199–244, 263–286 (1904). Konowia, **6**, pp. 35–49 (1927).
- MALLOCH, J. R. Nearc. Chyromyia. Proc. Ent. Soc. Wash., **16**, pp. 179–181 (1914).
  - Nearct. Spilochroa. Proc. U. S. Nat. Mus., 78, art. 15, p. 29 (1931).
- Tonnoir, A. L. and J. R. Malloch. New Zealand Helomyzidæ. Rec. Cant. Mus., 3, pp. 83–100 (1927).

### EPHYDRIDÆ, CANACEIDÆ

- Becker, T. Palæarct. Ephydridæ. Berl. ent. Zts., **41**, pp. 91–276 (1896). Flieg. pal. Reg., pts. 10, 11, 115 pp. (1926).
- CRESSON, E. T. Nearct. Paralimna, etc. Trans. Am. Ent. Soc., 42, pp. 101–124 (1916).

Nearct. Notiphila and Dichæta. Trans. Am. Ent. Soc., 43, pp. 27-69 (1917).

Nearct. Gymnopinæ. Trans. Am. Ent. Soc., 47, pp. 325-343 (1922).

JONES, B. J. Catalog Ephydridæ (with bibliogr.). Univ. Cal. Publ. Ent., 1, pp. 153–198 (1906).

Tonnoir, A. L. and J. R. Malloch. New Zealand Ephydridæ. Rec. Cant. Mus., 3, pp. 1–18 (1926).

### CHLOROPIDÆ, ASTIIDÆ, CRYPTOCHÆTIDÆ

Becker, T. Chloropidæ. I. Palæaret. Arch. Zool., 1, pt. 10, pp. 33–174 (1910).
II. Ethiop. Ann. Mus. Nat. Hung., 8, pp. 377–443 (1911).
III. Indoaustr. Ibid., 9, pp. 35–170 (1911).
IV. Nearet. Ibid., 10, pp. 21–120 (1912).
V. Neotrop. Ibid., 10, pp. 121–156 (1912).

Duda, O. Palæarct. Astiidæ. Deut. ent. Zts. 1927, pp. 113-147 (1927).

Enderlein, G. Oscinosominæ. Zool. Anz., **38**, pp. 10–23 (1911). Sb. Ges. natf. Fr. 1911, pt. 4, pp. 185–244 (1911).

KNAB, F. Cryptochætum. Insecut. Insc. Mens., 2, pp. 33-36 (1914).

MALLOCH, J. R. Botanobiinæ. Proc. U. S. Nat. Mus., 46, pp. 239–266 (1913).
Chloropidæ. Can. Ent., 46, pp. 113–120 (1914).

Australian Botanobiinæ. Proc. Linn. Soc. N. S. W., **52**, pp. 434–445 (1927).

Cryptochætum. Proc. Linn. Soc. N. S. W., 52, pp. 421-423 (1927).

# BORBORIDÆ, LEPTOCERATIDÆ

Duda, O. European Limosina. Abh. z. b. Ges. Wien, 10, pt. 1, 240 pp. (1918).
Zool. Jahrb. Syst., 43, pp. 433–446 (1920). Konowia, 7, pp. 162–174 (1928).

Palearct. Spherocera. Tijds. Ent., 63, pp. 1-39 (1920).

Palæarct. Borborus. Arch. Naturges., **89**, A, pt. 4, pp. 35–112 (1923). Leptocera (non-European). Arch. Naturges., **90**, A, pt. 11, pp. 5–215 (1924).

Malloch, J. R. Costa Rican Borboridæ. Trans. Am. Ent. Soc., 40, pp. 1–24 (1914).

Spuler, A. N. Am. Borboridæ, genera. Proc. Acad. Nat. Sci. Phila., 75, pp. 369-378 (1923).

Nearct. Sphærocera and Aptilotus. Pan-Pacif. Ent., 1, pp. 66–71 (1924).

Nearctic Leptocera. Ann. Ent. Soc. Am., **17**, pp. 106–116 (1924). Psyche, **31**, pp. 121–135 (1924). Jour. N. Y. Ent. Soc., **33**, pp. 70–84, 147–162 (1925). Can. Ent., **57**, pp. 99–104, 116–124 (1925).

Nearct. Borborus and Scatophora. Bull. Brkl. Ent. Soc., 20, pp. 1–16 (1925).

# AGROMYZIDÆ, ODINIIDÆ, ANTHOMYZIDÆ, OCHTHIPHILIDÆ, OPOMYZIDÆ, TETHINIDÆ

Czerny, L. Europ. Geomyza. Wien. ent. Ztg., 22, pp. 123–127 (1903).

Palæarct. Anthomyza. Wien. ent. Ztg., 21, pp. 249–254 (1902).

Ochthiphilidæ. Wien. ent. Ztg., 23, pp. 167-170 (1904).

Palæarct. Anthomyzidæ, Opomyzidæ, Tethinidæ. Flieg. pal. Reg., pt. 28, 31 pp. (1928).

- HENDEL, F. Palæaret. Agromyzidæ. Arch. Naturg., 84, A, pt. 7, pp. 109–174 (1918). Zool. Anz., 69, p. 248 (1927).
- Hering, M. German Agromyzidæ. Tierw. Deutschl., No. 6, 172 pp. (1927). Malloch, J. R. Nearct. Agromyza. Ann. Ent. Soc. Am., 6, pp. 269–336 (1913).

Nearct. Agromyzidæ. Proc. U. S. Nat. Mus., 46, pp. 127–154 (1913).
Nearct. Ochthiphilinæ. Nat. Hist. Surv. Ill. Bull. 13, art. 14, pp. 345–361 (1921).
Ann. Mag. Nat. Hist. (9), 19, pp. 575–577 (1927).

Melander, A. L. Agromyzinæ, Milichiinæ, Ochthiphilinæ and Geomyzinæ. Jour. N. Y. Ent. Soc., 21, pp. 219–273, 283–300 (1913).

### DROSOPHILIDÆ, DIASTATIDÆ, MILICHIDÆ

- Becker, T. Milichiinæ. Ann. Mus. Nat. Hung., 5, pp. 507-550 (1907).
- Duda, O. Europ. Drosophila. Ent. Meddel., 14, pp. 246-313 (1924).

Drosophilidæ. Arch. f. Naturges., 90, A, pp. 172–234 (1924).

Neotrop. Drosophilidæ. Ann. Mus. Nat. Hung., **22**, pp. 149–229 (1925); Arch. f. Naturges., **91**, A, pts. 11, 12, pp. 1–229 (1925).

- Hendel, F. Cyrtonotum. Deut. ent. Zts. 1913, pp. 618-630 (1913). Milichiiden. Entom. Mitt., **8**, pp. 196-200 (1919).
- Kahl, H. Leucophenga. Ann. Carnegie Mus., 11, pp. 364-393 (1917).
- Malloch, J. R. Various Drosophilidæ. Ent. News., 32, pp. 311, 312 (1921).Proc. U. S. Nat. Mus., 68, art. 21, pp. 28-34 (1921).
- Malloch, J. R. and W. L. McAtee. Drosophilidæ of Distr. Col. Proc. Biol. Soc. Wash., 37, pp. 25–41 (1924).
- STURTEVANT, A. H. Nearct. Drosophilidæ. Ann. Ent. Soc. Am., 9, pp. 325–342 (1916).
   Bull. Am. Mus. Nat. Hist., 38, pp. 441–446 (1918).
   Carnegie Inst. Wash. Publ. 301, 150 pp. (1921).

Zygothrica. Proc. U. S. Nat. Mus., 58, pp. 155-158 (1920).

### PUPIPARA

- Aldrich, J. M. Table genera of Hippoboscidæ. Insecut. Insc. Mens., 11, pp. 75–79 (1923).
- DA COSTA LIMA. S. Am. Streblidæ. Arch. Est. Agr. Med. vet. Nictheroy, 5, pp. 25-32 (1921).
- Austen, E. E. Hippoboscidæ. Ann. Mag. Nat. Hist. (7), **12**, pp. 255–266 (1903).

Bequaert, J. Hippoboscinæ. Psyche, 37, pp. 303-326 (1930).

Falcoz, L. Pupipara of France. Faun. de Fr., 14, pp. 21-52 (1926).

Ferris, G. F. Ectoparasites of bats. Ent. News, 27, pp. 433-438 (1916).

New world Nycteribiidæ. Ent. News, **35**, pp. 191–199 (1924).

Samoan Hippoboscidæ. Ins. Samoa, 6, pp. 10-21 (1927).

Ferris, G. F. and F. R. Cole. Hippoboscidæ. Parasitol., **14**, pp. 178–204 (1922). Massonat, E. Pupipara. Ann. Univ. Lyon, N. S. (1), **28**, pp. 1–356 (1909).

Muir, F. Ascodipteron. Bull. Mus. Comp. Zool. Harvard, **54**, pp. 349–366

(1912). Scott, H. Nycteribiidæ. Parasitol., **9**, pp. 593-610 (1917).

Speiser, P. Strebliden. Arch. f. Naturg., 66, pp. 31-70 (1901).

Nycteribiden. Arch. f. Naturg., 67, pp. 11-77 (1901).

Hippobosciden. Ann. Mus. Genova, 40, pp. 553-562 (1901).

Checklist N. Am. Pupipara. Ent. News, 18, pp. 103–105 (1907).

Phylogeny and distribut. of Pupipara. Zts. Wiss. Insektenbiol., 4, pp. 241-, 301-, 420-, 437- (1908).

SWENK, M. H. N. Am. Hippoboscidæ, Jour. N. Y. Ent. Soc., 24, pp. 126– (1916).

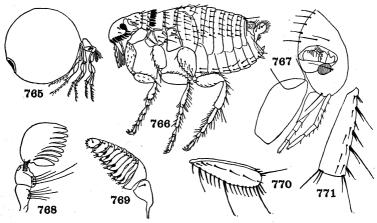
## ORDER SIPHONÁPTERA

# (SUCTÒRIA, APHANÍPTERA, ROPHOTEÌRA)

Small, wingless, strongly compressed, jumping insects, with dark colored, heavily chitinized bristly body and legs; parasitic in the adult condition on mammals, or rarely on birds. Head small, closely articulated with the thorax. Antennæ short and thick, with two large basal joints and an oval or elongate indistinctly jointed apical portion, lying in depressions behind the small, simple eyes which are sometimes wanting. Mouthparts fitted for piercing and sucking, the mandibles setiform, maxillæ blade-like; both pairs of palpi well developed. Thorax small, composed of three similar, freely movable segments. Abdomen large, composed of nine segments; cerci one-jointed. Legs large, stout; coxæ large, tarsi five-jointed with stout claws. Metamorphosis complete; larvæ elongate, cylindrical, legless, with well developed head and biting mouthparts; free living. Pupæ enclosed in cocoons. Fleas.

Head without transverse suture above, the dorsal surface continuous and not thus articulated. (Suborder INTEGRICÍPITA) 4

2.	Head simple at sides, without an articulated, flattened lobe on each side anteriorly; not parasitic on bats
_	ISCHNOPSÝLLIDÆ
3.	Occipital region with a transverse dorsal thickening near the
	middle; vertical suture of head extending below the antennæ
	to the lower margin of the head; posterior margin of front
	usually with a comb-like row of stout spines.
	MACROPSÝLLIDÆ
	Occipital region entire, without such a dorsal thickening; head
	spinose, but without such a comb-like row along the posterior
	margin of the front. (Including CTENOPSYLLIDÆ and
	TYPHLOCERÁTIDÆ) HYSTRICHOPSÝLLIDÆ
4.	
	tergites together longer than the first abdominal segment,
	sides of metathorax sometimes extending over two or three
	abdominal segments; labial palpi with three or more false
	joints; maxillary palpi almost always shorter than the front
	coxæ; third joint of antennæ with nine more or less distinctly
	separated false joints
	Thoracic segments strongly shortened and constricted, their ter-
	gites together shorter than the first abdominal segment; labial
	palpi without false joints; maxillary palpi almost always ex-
	tending beyond the front coxæ; third antennal joint without
	distinctly separated false joints; abdomen of fully matured
-	female greatly distended8
5.	Abdominal tergites provided with numerous setæ arranged in two or more rows; eyes greatly reduced in size, or usually absent . 6
	Abdominal segments with few setæ, forming only a single row on
	all or on most of the abdominal tergites; eyes almost always
	present. (Figs. 766, 767, 768, 770). (Pùlex, Ctenocephálides
	(=Ctenocéphalus) (C. fèlis, Cat-flea; C. cànis, Dog-flea), Ar-
	chæopsýlla, Xenopsýlla (=Læmopsýlla) (X. cheòpis, Plague
	flea), Hoplopsýllus, Ornithopsýlla). (Including ARCHÆO-
	PS ÝLLIDÆ) PULÍCIDÆ
6.	Head without a comb of spines on each cheek
0.	Head with a comb of from two to six stout bristles or spines on
	each cheek. (Ctenophthálmus, Rhadinopsýlla, Micro-
	psýlla) (NEOPSÝLLIDÆ, part)CTENOPHTHÁLMIDÆ



Figs. 765-771. Siphonaptera

- 765. Tunga, mature female (Butler) Tungidæ.
- 766. Ctenocephalides (Patton and Cragg) Pulicidæ.
- 767. Xenopsylla, side view of head (Fox) Pulicidæ.
- 768. Ctenocephalides, antenna (Patton and Cragg) Pulicidæ.
- 769. Ceratophyllus, antenna (Fox) Dolichopsyllidæ.
- 770. Ctenocephalides, hind tibia (Fox) Pulicidæ.
- 771. Ceratophyllus, hind tibia (Fox) Pulicidæ.

8. Maxillæ with a long, narrow, curved lamina which projects downward and backward; maxillary palpi as long as the front coxæ; head evenly rounded above; side pieces of metathorax extending over only the first abdominal segment; abdomen of fully matured female vermiform. (Hectopsýlla, ind.).

HECTOPS ÝLLIDÆ

### LITERATURE ON SIPHONAPTERA

Baker, C. F. Revision of American Siphonaptera. Proc. U. S. Nat. Mus., 27, pp. 265–469 (1904).

Classification of American Siphonaptera. Proc. U. S. Nat. Mus., 29, pp. 121-170 (1905).

- Dalla Torre, C. G. Aphaniptera orbis terrarum. Ber. naturw. med. Ver., Innsbruck, **39**, pp. 1–29 (1924).
- Dampf, A. Die Aphanipteren Westdeutschlands. Ber. bot. zool. Ver. Rheinlande, 1911, pp. 73–113 (1912).

Kritisches Verzeichnis der Aphaniptera Deutschlands. Entom. Mitt., **15**, pp. 377–386 (1926).

- Fox, C. Insects and Diseases of Man. London and Philadelphia, 349 pp. (1925) (Kev to genera of Siphonaptera).
- JORDAN, K. and W. ROTHSCHILD. Revision of Sarcopsyllidæ. Thompson Yates and Johnston Lab. Repts., 7 (1906).

Revision of the non-combed eyed Siphonaptera. Parasitology, 1, pp. 1–100 (1908).

Oudemans, A. C. Anteekningen over Suctoria. Entom. Bericht. (numerous parts) (1909–1917).

Neue Ansichten über die Morphologie des Flohkopfes, sowie über die Ontogenie, Phylogenie und Systematik der Flöhe. Novit. Zool., **16**, pp. 133–158 (1910).

Kritisch Overzicht du Nederlandische Suctoria. Tijdschr. v. Entom., 58, pp. 60-97 (1915).

ROTHSCHILD, N. C. Synopsis of British Siphonaptera. Entom. Monthly Mag., **51**, pp. 49–112 (1915).

Taschenberg, O. Die Flöhe. 118 pp. Halle (1880).

WAGNER, J. N. Revue systématique des espèces des Aphaniptères. Hor. Soc. Entom. Ross., 39, pp. 508–569 (1910).

## ORDER COLEÓPTERA

# (ELEUTERÀTA; ELYTHRÓPTERA)

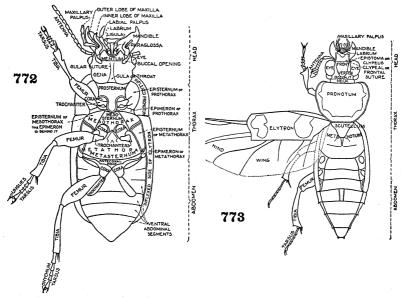
Moderate-sized, small or minute, more rarely very large, hard-bodied insects. Head free, usually prominent, sometimes produced forward to form a snout or beak; mandibles well developed; maxillæ well developed, usually bilobed with the palpi three- to five-jointed; labial palpi shorter, two- or three-jointed; antennæ ten- or eleven-jointed, sometimes less, very rarely more, filiform or variously modified, often with the apical joints enlarged; ocelli nearly always absent. Prothorax free; two pairs of wings, the front pair (elytra) thickly chitinized, sheathing the meso- and metathorax and also nearly always the abdomen, almost always meeting in a straight line down the middle of the back; hind wings occasionally absent, front ones rarely reduced or absent. Legs homonomous, the tarsi usually with five or four joints; no cerci. Metamorphosis complete, the larvæ mandibulate. A very large and widely distributed group, including Beetles and Weevils.<sup>1</sup>

1.	First ventral segment divided by the hind coxal cavities, so that its sides are separated from the usually very small median part (Fig. 772); the first three sternites (ventral abdominal segments)
	immovably united; antennæ usually filiform or nearly so; hind
	wings with a triangular or oval cell at the apex of an elongate
	discal cell. (Figs. 774, 777). (Suborder ADÉPHAGA)2
	First visible sternite extending for its entire breadth behind the
	coxal cavities; hind wing without closed cell at apex of discal
	cell (except Cupidæ), the latter also often absent. (Figs. 775,
	776, 778). (Suborder POLÝPHAGA)12
2.	Mentum and submentum not separated by a suture3
	Mentum and submentum separated by a distinct suture5
3.	Abdomen with four visible sternites; small myrmecophilous beetles
	with the antennæ remarkably thickened and the apical joints
	usually fused into a club or into a broad, more or less unseg-
	mented lamina (Fig. 780). (Paussus Afr., Indomal.; Ceráp-
	terus, Afr.; Arthrópterus, Austr.). (PAUSSÒIDEA).
	PAUSSIDÆ
	Abdomen with six visible sternites; antennæ filiform (if antennæ

<sup>1</sup> A few highly degenerate forms of more or less larviform appearance that will not otherwise run out in the key, may be identified by reference to couplet 198.

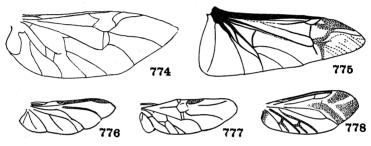
 4. Head with antennal grooves beneath, between the eyes and the maxillary fissures. Terrestrial species. (Pseudomórpha, Am.; Sphallomórpha, Silphomórpha, Austr.; Adelótopus, Austr. and E. Ind.; Hydroporomórpha, Afr.).

# PSEUDOM ÓRPHIDÆ



Figs. 772-773. Coleoptera

- 772. Harpalus, underside (Hayward) Carabidæ.
- 773. **Necrophorus,** wings spread on left side and removed on the right (Hayward) Silphidæ.



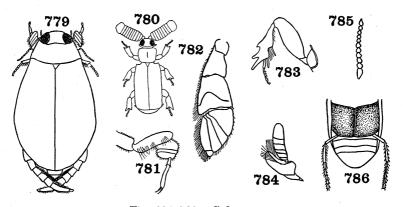
Figs. 774-778. Coleoptera

- 774. Hind wing of Adephaga (Kempers).
- 775. Hydrophilus, hind wing. Hydrophilidæ.
- 776. Necrophorus, hind wing (Kempers) Silphidæ.
- 777. Tachypus, hind wing (Kempers) Carabidæ.
- 778. Lygistopterus, hind wing (Kempers) Lycidæ.
  - 8. Antennæ inserted on the front, above the base of the mandibles; eyes prominent; head wider than the thorax, vertical, with large mandibles. Tiger-beetles. (Cicindèla, cosmop.; Tétracha, widespr.; Oxychìla, neotrop.; Mantíchora, Afr.; Tricóndyla, Indo-austr.; Pogonóstoma, Afr.; Òmus, nearc.).

### CICINDÉLIDÆ

Scutellum present. Ground beetles. A very large diverse, abundant and widespread family. (Figs. 772, 777, 783, 818). (Táchys, Hárpalus, Bembídion, cosmop.; Calosòma, Dyschirius, Scarites, Chlènius, Bráchinus, Lèbia, widespr.; Pteró-

	stichus, Amàra, Cárabus, Anophthálmus, Trèchus,
	Nèbria, holarc.; Mormólyce, indomal.; and many other gen-
	era). (Including PASIMÁCHIDÆ, MORMOLÝCIDÆ).
	CARÁBIDÆ
	Scutellum absent; prosternum covering the mesosternum; rounded convex, riparian species. (Ómophron, cosmop., exc. Austr.).
	convex, riparian species. (Omophron, cosmop., exc. Austr.).  OMOPHRÓNIDÆ
10	Metasternum prolonged behind as a triangular process; aquatic
10.	species
	Metasternum not prolonged between the coxæ; scutellum absent;
	antennæ like a string of beads, with globular joints; bark
	beetles (Fig. 785). (If tarsi are two-jointed, see the dubious
	genus Jacobsonium, couplet 119). (Rhysòdes, cosmop.; Cliní-
	dium, widespr.; Rhysodiástes, widespr.) RHYSÓDIDÆ
11.	Antennæ slender, filiform; abdomen with six visible tergites; eyes
	two. Diving beetles. Water tigers. (Figs. 779, 781). (Can-
	thydrus, Laccophilus, Bidessus, Rhantus, cosmop.; Cy-
	bister, Cœlámbus, widespr.; Hydróporus, Deronéctes, Ágabus, Acílius, Dytíscus, holarc.) DYTÍSCIDÆ
a.	Episternum of metathorax not reaching the middle coxal cavity.b
h	Episternum of metathorax reaching the middle coxal cavityd Greatest anterior extension of the hind coxa near the middle
ь.	(longitudinally) of the body; metasternum more or less pointed
	in the middle behind, and not marked by a transverse suture.
	NOTERÎNÆ
	Greatest anterior extension of the hind coxa nearer to the epipleura
c	than to the medial line of the bodyc Prosternal process not reaching the metasternum.VATELLĪNÆ
C.	Prosternal process reaching the metasternum. LACCOPHILINÆ
d.	Prosternum deflected between the front coxæ so that the pro-
	sternal process is placed on a quite different plane of direction
	from that of the prosternum; the latter not incrassate along middle; front tarsi often 4-jointede
	Prosternal process on the same plane of direction as the pro-
	sternum; front tarsi 5-jointedf
e.	Prosternal process much deflected from the plane of direction of
	the prosternum. Front tarsi usually with only four joints.
	HYDROPORÎNÆ Prosternal process but little deflected from the plane of direction
	of the prosternum; front tarsi 5-jointed; scutellum not visible.
	METHLINÆ
f.	Inferior spur of hind tibia not or but little broader than the
	otherg



Figs. 779–786. Coleoptera

- 779. Cybister (Maxwell-Lefroy) Dytiscidæ.
- 780. Arthropterus (Westwood) Paussidæ.
- 781. **Dytiscus,** front tibia and tarsus of male (Kolbe) Dytiscidæ.
- 782. **Gyrinus,** hind leg (Berlese) Gyrinidæ.
- 783. Scarites, front leg (Kolbe) Carabidæ.
- 784. Dineutes, antenna. Gyrinidæ.
- 785. Rhysodes, antenna. Rhysodidæ.
- 786. Haliplus, coxal plate (Maxwell-Lefroy) Haliplidæ.

Antennæ very short, stout and irregular; eyes four; abdomen with seven visible tergites. Whirligig beetles. (Dineûtes, Gyrìnus; Aulonogỳrus, old world; Gyrètes, neotrop.; Orec-

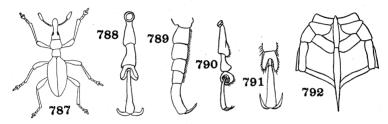
tochilus, palæarc., Indomal.; Orectogyrus, ethiop.; Macro-

	gyrus, neotrop., Indo-Austr.). (GYRINOIDEA).
	GYRÍNIDÆ
12.	Antennæ clubbed or not, if clubbed, not with the club-joints
	lamellate; tarsi frequently with less than five joints; antennæ
	very rarely somewhat lamellate in certain aquatic Hydrophil-
	idæ. (Figs. 793 to 813)
	Antennæ with the last three to seven joints enlarged on one side
	to form a comb-like or lamellate club which can often be opened
	and closed (Figs. 814, 815, 816). Legs often fitted for dig-
	ging; tarsi almost always 5-jointed, the front tarsi very excep-
	tionally reduced or absent; larvæ with thick, curved body and
	well developed legs. Never aquatic. (LAMELLICÓRNIA). 76
13.	Head not prolonged into a beak, gular sutures double, at least
	before and behind; prosternal sutures distinct; proepimera not
	meeting behind the prosternum
	Head generally prolonged in front of the eyes and snout-like
	(Figs. 787, 873, 878); gular sutures confluent along the median
	line or obsolete; proepimera united behind the prosternum,
	prosternal sutures wanting; antennæ usually elbowed (Fig.
	879); palpi usually rigid; all tarsi apparently with four or three
	joints (Fig. 788); larvæ legless or with short legs. (RHYNCÓ-
11	PHORA)
14.	Fourth and fifth tarsal joints if present, not immovably united,
	the articulation between them like that between the other
	joints (Fig. 789). (If rarely united, as in some Erotyldiæ, the antennæ are clavate)
	antennæ are clavate)
	densely pubescent below, with the first three joints dilated
	and with a sole, the third joint usually bilobed (Figs. 790, 791);
	antennæ filiform, rarely serrate or thickened apically; larvæ
	vegetarian. (PHYTÓPHAGA, CERAMBYCÒIDEA)147
15.	Hind tarsi with at least as many joints as the others
	Hind tarsi four-jointed; front and middle tarsi five-jointed (rarely
	with the penultimate joint very short so that the hind tarsi
	are apparently three-jointed and the others apparently four-
	jointed; in very exceptional cases with the front tarsi five-
	jointed and both other pairs four-jointed). (HETERÓ-
	MERA)
16.	Maxillary palpi long and slender, almost always as long as or longer $$
	than the short antennæ; antennæ six- to ten-jointed, the outer

joints forming a distinct, pubescent, sometimes asymmetrical club; elytra with an alula; small to large, almost always aquatic species (Figs. 775, 792). Water scavenger beetles. (Hydræna, Cércyon, Enòchrus, Hýdrous (= Hydróphilus auctt.) Beròsus, cosmop.; Ochthèbius, Sphærídium, Laccòbius, widespr.; Hydróphilus, Helóphorus, holarc.; Tropistérnus, Am.). (PALPICÓRNIA, HYDROPHILÓIDEA).

HYDROPHÍLIDÆ

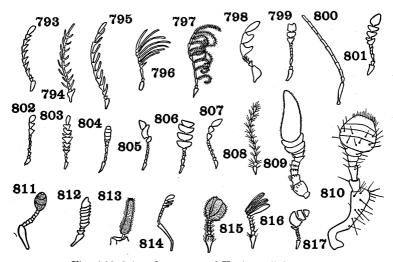
a. Eyes normal, not divided.....b Eyes divided, each into two parts......AMPHIOPÎNÆ



Figs. 787-792. Coleoptera

- 787. Cylas (Pierce) Cyladidæ.
- 788. Camptocerus, tarsus (Hopkins) Scolytidæ.
- 789. Megalodacne, tarsus. Erotylidæ.
- 790. Leptinotarsa, tarsus (Sharp) Chrysomelidæ.
- 791. Saperda, apical part of tarsus. Lamiidæ.
- 792. Hydrophilus, mesosternum (Berlese) Hydrophilidæ.
  - Second joint of posterior tarsi elongate, longer than third; first joint very short; pronotum at base as wide as elytra...........c
     Second joint of posterior tarsi short, about equal to the third..d

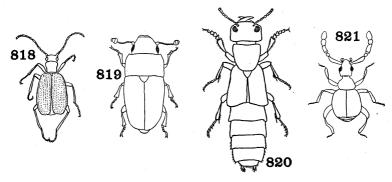
	Maxillary palpi much shorter than the antennæ, if rarely com-
	parable to the antennæ in length the alula is absent and the last
	tarsal joint abnormally long (some Dryopidæ), or only the
	last joint is greatly lengthened (Telegeusidæ)
17.	Elytra short, exposing much of the abdomen; tergites entirely
	corneous in texture; wings usually present and folded beneath
	the elytra in repose; free portion of media atrophied or absent,
	not joining the cubitus to form a long closed axial cell. (If
	external parasites of the beaver, compare Platypsyllidæ, coup-
	let 73). (STAPHYLINIFÓRMIA)
	Elytra covering most of the abdomen, not much shortened and
	covering all or all but one, two or three abdominal segments;
	rarely much shortened in which case the wings either do not
	fold beneath the elytra or are wanting; tergites membranous
	or semimembranous, except sometimes those of the two or three
	apical segments (five in the beaver parasite, Platypsyllus,
	couplet 73)
18.	Abdomen flexible, not enlarged apically, parallel or tapering,
	with six to eight freely movable sternites; antennæ usually
	eleven-jointed; tarsi usually five-jointed; usually small, occa-
	sionally large, slender species. (If wings do not fold beneath
	elytra and maxillary palpi are flabellate, compare Atracto-
	ceridæ (couplet 92) or if palpi are nearly as long as the an-
	tennæ, compare Telegeusidæ (couplet 93); or if body is very
	greatly flattened, compare Cucujidæ (Hemipeplinæ) (couplet
	52)
	Abdomen not flexible, swollen, oval, the segments anchylosed;
	usually only five sternites, rarely with seven or eight; antennæ often with less than eleven joints; tarsi three-jointed; small or
	minute, robust species; abdomen very much wider than the
	prothorax
10	Abdomen with five dorsal segments; antennæ five to eleven-jointed
10.	(usually 11), the last joint never truncate (Figs. 805, 821);
	maxillary palpi usually four-jointed. (Psélaphus, cosmop.;
	Eupléctus, Batrisòdes, widespr.; Bryáxis, Tỳchus, holarc.;
	Eupines, Indo-Austr.) PSELÁPHIDÆ
	Abdomen with three dorsal segments, antennæ with two to six
	joints (Fig. 813); maxillary palpi one-jointed. (Fústiger, Am.;
	Adrànes, nearc.; Cláviger, palæarc.; Artícerus, Austr.).
	CLAVIGÉRIDÆ



Figs. 793-817. Antennæ of Various Coleoptera

- 793. Ludius (Leconte and Horn) Elateridæ.
- 794. **Prionocyphon** (Leconte and Horn) Helodidæ.
- 795. Corymbites (Leconte and Horn) Elateridæ.
- 796. Acneus (Leconte and Horn) Helodidæ.
- 797. Dendroides (Leconte and Horn) Pyrochroidæ.
- 798. Dorcatoma (Leconte and Horn) Anobiidæ.
- 799. Corynetes (Leconte and Horn) Corynetidæ.
- 800. Brontes. Cucujidæ.
- 801. Liodes (Leconte and Horn) Histeridæ.
- 802. Temnochilus (Leconte and Horn) Ostomatidæ.
- 803. Catoptrichus (Leconte and Horn) Silphidæ.
- 804. Colon (Leconte and Horn) Silphidæ.
- 805. Bryaxis (Leconte and Horn) Pselaphidæ.
- 806. Anogdus (Leconte and Horn) Silphidæ.
- 807. Aulicus (Leconte and Horn) Cleridæ.
- 808. Dasycerus (Leconte and Horn) Lathridiidæ.
- 809. Anthrenus (Felt) Dermestidæ.
- 810. Dendroctonus (Felt) Ipidæ.
- 811. Epierus (Leconte and Horn) Histeridæ.
- 812. **Heterocerus** (Leconte and Horn) Heteroceridæ.
- 813. Adranes (Leconte and Horn) Clavigeridæ.
- 814. Lucanus (Leconte and Horn) Lucanidæ.
- 815. Bolboceras (Leconte and Horn) Geotrupidæ.
- 816. Phyllophaga (Leconte and Horn) Melolonthidæ.
- 817. Phymaphora (Leconte and Horn) Mycetæidæ.

20.	Tarsi five-jointed on at least one pair of legs, and almost always
	on all pairs
	All tarsi with less than five joints
21.	Abdomen with five sternites, or less
	Abdomen with at least six sternites
22.	Five abdominal sternites
	Only three visible sternites, the first very long; small beetles living
	in ants' nests; antennæ with only three joints, all but the two
	basal ones fused into a large club-shaped mass. (Gnóstus
	Brazil, Florida) GNÓSTIDÆ
23.	Front coxæ globular or transverse, usually projecting but little
	from the coxal cavities; trochanters never interstitial24
	Front coxæ more or less conical and prominent
24.	Front coxæ transverse, more or less cylindrical25
	Front coxæ globular
25.	Hind coxe grooved to receive the femora26
	Hind coxæ flat, not grooved
26.	Strongly convex beetles with more or less retractile legs; tibia
	dilated and usually grooved near the outer end to receive the
	tarsi; tibial spurs distinct
	Slightly convex, oval species with non-retractile slender legs
	tibial spurs more or less reduced
27.	Antennæ inserted at the sides of the head
	Antennæ inserted on the front; head retracted; third joint of
	tarsi lobed; thorax margined; oval species. (Chelonàrium
	Am.) CHELONARÌIDÆ
28.	Head prominent; mentum large, elongate and sub-elliptical; tarsi
	not lobed. (Nosodéndron, widespr.) NOSODÉNDRIDÆ
	Head retracted; mentum small and quadrate. (Limnichus,
	Pedilóphorus, Syncalýpta, widespr.; Cýtilus, Býrrhus,
	holarc
a	Clypeus not distinct from the front; posterior coxæ almost touch-
а.	ing one another BYRRHÎNÆ
	Clypeus separated from the front by a fine suture; posterior coxe
	more or less widely separatedb
b.	more or less widely separatedb  Prosternum without antennal groovesLIMNICHINÆ
	Prosternum within the front angles with a deep furrow for the
	reception of the antennal club BOTHRIOPHORINÆ
29.	Front coxe with distinctly separated side-piece (trochantin) 30
<i>29.</i>	Front coxe with distinctly separated side-piece (trochantin) 30



Figs. 818-821. Coleoptera

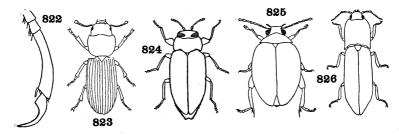
- 818. Brachinus. Carabidæ.
- 819. Glischrochilus (Felt) Nitidulidæ.
- 820. Staphylinus. Staphylinidæ.
- 821. Goniastes (Westwood) Pselaphidæ.

33.	Tarsi more or less dilated, the first joint not shortened; fourth
	joint very small; elytra not usually extending to the tip of
	the abdomen
	Tarsi slender; metatarsus very short; elytra entire, never trun
	cate, covering the abdomen. (Tenebridides, Ancyrona
	widespr.; Péltis, Temnochila, holarc., neotrop.; Nemosòma
	Eur., Am.; <b>Thýmalus</b> , holarc.). (Figs. 802, 823). ( <i>TROGOSÍT</i>
	IDÆ, TEMNOCHÍLIDÆ) OSTOMÁTIDÆ
34.	Maxillæ with well developed inner and outer lobe
	Maxillæ with only a single lobe (Fig. 819). (Carpóphilus, Epu
	ræa, cosmop.; Meligèthes, Cryptárcha, widespr.; Nitídula
	palæarc., Am.; Omosìta, holarc.)NITIDÙLIDA
35.	Antennæ clavate; elytra usually not covering the tip of the ab-
	domen
	Antennæ filiform; elytra entirely covering the abdomen; moder-
	ately large, elongate beetles. (Parándra, widespr.; Archándra
	palæarc., neotrop.; Neándra, nearc.) PARÁNDRIDA
36.	Antennæ 11-jointed; with a three-jointed club; labrum free
	(Brachýpterus, widespr.) BRACHYPTÉRIDA
	Antennæ 10-jointed, with a two-jointed club; labrum fused with
	the clypeus. (Rhizóphagus, Anomóphagus, holarc.).
	RHIZOPHÁGIDÆ
<b>37</b> .	First two or three sternites fused or immovably united38
	All sternites free, or at least separated by equally distinct sutures
	except in very rare cases39
38.	First two sternites connate, the suture between them very weak
	antennæ serrate, very rarely pectinate in the male; tarsi with
	membranous lobes beneath; last tarsal joint not lengthened
	claws moderate or small; active, hard-bodied beetles, of more
	or less metallic color; not aquatic. (Fig. 824). (Acmæódera
	Antháxia, Chrysobóthris, Ágrilus, cosmop.; Psilóptera
	Stigmódera, Bupréstis, widespr.; Bráchys, Am.; Chrysó-
	chroa, Indomal.) BUPRÉSTIDA
a.	Middle coxal cavity formed entirely by the mesosternum (excep-
	in certain South African species of Julodis and Amblysterna)
	Middle coxal cavity formed laterally by the mesosternum and a
	its posterior part by the metasternum
b.	
	Antennal pores concentrated in a depression or fovea on the
	serrate joints
c.	Posterior coxæ slightly dilated on their inner side, their posterior

	margin transverse and slightly sinuate; scutelium invisible;
	antennal pores hidden by silky pubescence. JULODINÆ
	Posterior coxe distinctly dilated on their inner side, their pos-
	terior margin oblique; antennal pores bare.
	THRINCOPYGINÆ
d.	Lateral pieces of the metathorax narrowPOLYCESTINÆ
	Lateral pieces of the metathorax very broad; tergites membran-
	ous SCHIZOPÌNÆ
e.	Lateral branches of mesosternum elongate (except in $Belionota$ )f
	Lateral branches of the mesosternum very short and set back on
	the sides, or invisiblej
f	Antennal pores scattered over the two faces of the serrate joints g
1.	Antennal pores concentrated in a depression or fovea on the ser-
	rate joints
g.	Scutellum absent, or hidden
_	Scutellum visible CHALCOPHORÎNÆ
h.	Front not narrowed at the insertion of the antennæ; eyes not
	very close together, sometimes distant on the vertexi
	Front narrowed at the insertion of the antennæ; eyes strongly
	oblique and closely approaching one another on the upper
	surface CHRYSOBOTHRÎNÆ
i i	Scutellum broad and acuminate behind; mentum large, triangular;
	poriferous foveæ terminal SPHENOPTERÎNÆ
	Scutellum at most moderate, never enlarged in front or acuminate
	behind; mentum strongly transverse; poriferous foveæ terminal
	beinid, mentum strongly transverse, pornerous loveæ terminar
	or inferior BUPRESTÎNÆ
٠j.	Front narrowed at the insertion of the antennæ; antennal cavities
	very large and situated at a considerable distance from the
	eyes; posterior coxæ not dilated on their inner side, with their
	posterior margins horizontal and slightly sinuate; poriferous
	foveæ terminal
	Front not narrowed at the insertion of the antennæ; antennal
	cavities moderate and situated near the eyes; posterior coxæ
	dilated on their inner side, their posterior margin oblique;
	poriferous foveæ variable STIGMODERÎNÆ
k.	
к.	Base of pronotum straight
	Dase of pronotum straight MADIOGENIMA
	First three sternites connate; antennæ slender, slightly thickened
	externally; last joint of tarsi greatly elongated, with very large
	claws; small aquatic beetles. (Fig. 822). (Hélmis, cosmop.;
	Stenélmis, widespr.; Macrónychus, holarc.; Riolus, Lathél-
	mis, palæarc.) HÉLMIDÆ
39.	Prosternum prolonged behind into a median process which is
	received in the mesosternum40

Prosternum without such backwardly directed process (if rarely with such a process, it is not received in the mesosternum). .43

40. Prothorax loosely joined to the mesothorax, freely movable, its hind angles usually prolonged backward into teeth; prosternal spine loosely received in a notch in the mesosternum; front coxal cavities contained entirely in the prosternum............41



Figs. 822-826. Coleoptera

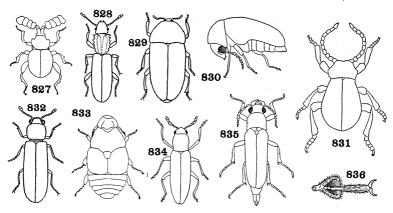
- 822. Macronychus, apical portion of tarsus. Helmidæ.
- 823. Tenebrioides (Back and Cotton) Ostomatidæ.
- 824. Chrysobothris (Chittenden) Buprestidæ.
- 825. Scirtes (Grandi) Helodidæ.
- 826. **Melanotus** (Hyslop) Elateridæ.

### CEROPHÝTIDÆ

42. Labrum visible, free; antennæ arising near the eyes under the frontal margin; last two sternites connected by a membranous suture; prosternum lobed in front; beetles capable of moving the prothorax by its basal joint with a sudden clicking motion. (If the middle coxæ are not distinctly separated, see couplet 76). Click-beetles, Wireworm beetles. (Lacon, Drastèrius, Élater, Cardióphorus, cosmop.; Adelócera, Melanòtus, Álaus, Monocrepídius, widespr.; Pyróphorus, Am., Austr.; Dichró-

nychus, ethiop., Ind.). (Figs. 793, 795, 826). (Including DICHRONÝCHIDÆ) ..... ELATÉRIDÆ

Labrum concealed; no membranous fold between the last two sternites; prosternum not lobed in front; antennæ inserted on the front between the eyes, received in transverse grooves on the front; not able to leap by the prothoracic joint. (**Dro-**



Figs. 827-836. Coleoptera

- 827. Ectrephes (Westwood) Ectrephidæ.
- 828. Silvanus (Chittenden) Silvanidæ.
- 829. Dermestes (Howard and Marlatt) Dermestidæ.
- 830. Lasioderma (Smith) Anobiidæ,
- 831. Mezium (Smith) Ptinidæ.
- 832. Lyctus (Hopkins) Lyctidæ.
- 833. Platypsyllus (Westwood) Platypsyllidæ.
- 834. Languria. Languriidæ.
- 835. Cebrio (Hyslop) Cebrionidæ.
- 836. Monædus, tarsus (Sharp) Monædidæ.

# 

	Antennæ clavate; elytra simple; very small or minute, convex, oval or rounded beetles. (Phálacrus, Ólibrus, Stílbus, widespr.) PHALÁCRIDÆ
45.	Elytra shortened, leaving two segments of the abdomen exposed; antennæ elbowed, very strongly clavate or capitate (Fig. 811); tibiæ compressed, the front pair usually toothed; hard-bodied, smooth and shining; small, convex or much flattened, rarely elongate, beetles
	Elytra entire, covering the pygidium47
46.	Prothorax much wider than the head, its front margin concave; clypeus without projections; tarsi short (Fig. 801). (Saprìnus, Parómalus, cosmop.; Híster, Hololépta, Platysòma, wide-
	spr.; Phelister, Am.) HISTÉRIDÆ
a.	Head not retractile, horizontal, visible from underneath; mouth- parts projecting beyond the front of the prosternumb Head retractile, not visible from underneath when retracted;
	mouthparts covered by the prosternumc
b.	Mandibles projecting; clypeus not prolonged into a rostrum; body more or less flat and depressed
	Mandibles retracted, covered by the long rostriform clypeus; body elongate, cylindrical
c.	Prosternum with a shorter or longer lobe or throat-plate, separated off by a more or less distinct sutured
d.	Prosternum without a lobe or throat-platee Club of antennæ round or oval, pubescent, consisting of four joints, closely compacted, but separated by sutures.  HISTERÎNÆ
	Club of antennæ without sutures, smooth, cylindrical, truncate at apex
e.	Antennæ inserted under the side margin of the front.
•	
	Antennæ inserted on the front ABRÆĨNÆ
	Prothorax as wide as the head, its front margin straight; clypeus with a projection at each side; tarsi very long and slender. (Nipônius, Indomal.)
47.	Antennæ consisting of only two apparent segments, the distal one greatly elongate, enlarged and flattened; minute, more or less globose beetles, living in ants' nests. (Fig. 827). (Éctrephes, Austr.) ECTRÉPHIDÆ Antennæ normal, ten or eleven-jointed, filiform or clavate 48

<b>48.</b>	Elytra each with a pair of large wax-like spots; antennæ short,
	with a four-jointed club; rather large beetles, with the prothorax
	widened behind. (Helòta, As., Indo-malay.) . HELÓTIDÆ
	Elytra not thus ornamented; prothorax not noticeably widened
	behind
49.	Middle coxal cavities open externally, i.e. not closed by the
	meeting of the meso- and metasterna; body elongate, usually
	greatly flattened
	Middle coxal cavities closed externally by the sterna53
50.	Maxillæ covered by corneous plates; front coxal cavities open
	behind. (Passándra, ethiop., neotrop.; Catógenus, widespr.;
	Scalidia, Am.). (Including SCALIDIIDÆ).
	PASSÁNDRIDÆ
	Maxillæ exposed
51.	Tarsi with the third joint simple, not lobed
	Tarsi with the third joint lobed; front coxal cavities closed behind
	(Fig. 839); elytra sometimes shortened. (Hemipéplus, Am.;
	Diagrypnodes, Austr.) Some CUCÙJIDÆ
52.	Front coxal cavities open behind. (Figs. 800, 840). (Cùcujus,
	Læmophlæus, Brontes, widespr.). (Including LÆMO-
	PHLŒIDÆ) CUCÙJIDÆ
	Front coxal cavities closed behind. (Figs. 828, 839). (Silvànus,
	cosmop.; Nausíbius, widespr.) SILVÁNIDÆ
53.	Prosternum not prolonged behind; small, oval, coarsely punctate
	species with the fourth tarsal joint short, the third lobed.
	(Diphýllus, Diplocœlus, widespr.) DIPHÝLLIDÆ
	Prosternum prolonged behind, meeting the mesosternum54
54.	Front coxal cavities open behind (Fig. 840); small or minute
	species. (Lobèrus, Micrámbe, Cryptóphagus, Antheró-
	phagus, widespr.; Cænóscelis, Atomària, holarc.).
	CRYPTOPHÁGIDÆ
	Front coxal cavities closed behind (Fig. 839)
55.	Antennæ with an abrupt club; moderate-sized beetles usually
	black with orange markings. (Fig. 789). (Megalodácne,
	Dácne, widespr.; Epíscapha, palæarc.). Some EROTÝLIDÆ
	Antennæ gradually clavate, short; very small beetles. (Cato-
	pochròtus, Turkestan) CATOPOCHRÓTIDÆ
56.	Hind coxæ dilated into plates which are grooved for the reception
	of the femora
	Hind coxe not thus dilated, not grooved for the reception of the
	femora61

	Front coxal cavities closed behind (Fig. 839)
58.	Second and third joints of tarsi lobed beneath; plate of hind coxe feeble; small pubescent beetles. (Bytùrus, holarc.; Satorýstia, palæarc.) BYTÙRIDÆ
	Tarsi simple, not lobed; ocelli often present; small, coarsely punctured beetles. (Derodóntus, Laricòbius, Peltástica, holarc.) DERODÓNTIDÆ
59.	Antennæ with the last three joints much enlarged, forming a strong club; small or rather small, often scaly beetles. (Derméstes, Attágenus, Trogodérma, cosmop.; Anthrènus,
	widespr.; Cryptorhópalum, neotrop., Austr.). (Figs. 809, 829)
a.	Head without frontal ocellus; mouthparts not covered.
b.	Head with frontal ocellus
	Mouthparts covered by the prosternum or by the coxe and tro-
c.	chanters of the front legs
	Prosternum vertical; hind coxæ reaching the side margins of the body; upper surface bare and glabrous ORPHILÎNA
d.	Form oblong; posterior coxæ contiguous; upper surface with recumbent hairs
e.	Form short, round or short-oval; posterior coxe not contiguous. e Upper surface squamose; head with deep antennal grooves be
	neath
	Antennæ not capitate
60.	Tarsi with a long, hairy pad (onychium) between the claws; tibial spurs present, small; moderate-sized or large, elongate beetles with prominent porrect head; antennæ usually flabellate in the
	male, often with more than eleven joints. (Sándalus, widespr.; Zénoa, nearc.; Callirhìpis, Indo-Austr., neotrop.; Rhipícera,
	Austr.). (SANDÁLIDÆ, RHIPICÉRIDÆ, RHIPIDOCÉR-IDÆ)
	On the last tendence in the la
	Onychium not developed, or very small; no tibial spurs; small, usually oval or cylindrical beetles with the head strongly de-

	flexed. (Figs. 798, 830). (Sitódrepa, Anòbium, cosmop.;
	Ernòbius, widespr.; Hadrobrégmus, Lasiodérma, widespr.;
	Dorcátoma, holarc.) ANOBÌIDÆ
61.	First joint of tarsi very short and indistinctly separated from the
	second
	First joint of tarsi distinct, when rarely very short, the first ventral ${\bf r}$
	segment is not elongate and the head not deflexed64
62.	All the tibiæ dilated and toothed externally; antennæ more or
	less geniculate, with a three-jointed club; head almost as wide
	as the prothorax; clypeus rounded on the sides; elytra not
	covering the tip of the abdomen; large beetles. (Syntèlia,
	Mex., Japan, India) SYNTELÎIDÆ
20	Tibiæ not dilated or toothed
63.	First sternite elongated, always much longer than the second;
	antennæ with a quite distinct, two-jointed club; small elongate
	beetles with prominent head not covered by the prothorax.
	Powder-post beetles. (Fig. 832). ( <b>Lýctus</b> , cosmop.). <b>LÝCTIDÆ</b>
	First sternite not elongated; antennal club three- or four-jointed;
	head usually deflexed and protected by the prothorax; declivity
	of elytra often toothed or spined; elongate, more or less cylindri-
	cal beetles. (APÁTIDÆ) BOSTRÝCHIDÆ
64.	Hind coxe flat or oval, not prominent
OT.	Hind coxæ hat of oval, not profilment
65.	Fourth joint of tarsi extremely short, not visible from above;
00.	small beetles of rather bright colors. (Fig. 799). (Corynètes,
	Necròbia, cosmop.; Pelònium, widespr.; Phyllobænus, Or-
	thopleùra, Am.; Epiphlœus, neotrop.) CORYNÉTIDÆ
	Fourth joint of tarsi not abnormally short
66.	Fifth segment of abdomen conically produced, as long as the three
	preceding ones, elytra not completely covering the abdomen.
	(Scaphidium, Scaphosòma, cosmop.; Bæócera, widespr.).
	SCAPHIDÌIDÆ
	Fifth abdominal segment not elongated, not conically produced .67
67.	Antennæ 11-jointed, with a solid club, consisting of three almost
	entirely fused joints; very small beetles, with tufts of golden
	hairs at the sides of the prothorax or beneath the body, living
	in ants' nests. (Thoríctus, palæarc., ethiop.).
	THORÍCTIDÆ
	Joints of antennal club not thus fused, or antennæ with fewer
	joints

68.	Trochanters attached to the internal margin of the femora. (Fig.
	807). (Tíllus, Opilo, Thanásimus, widespr.; Clèrus, Cyma-
	tódera, Hydnócera, Am.; Trichòdes, holarc.). CLÉRIDÆ
	Trochanters interstitial, i.e. attached to the base of the femora.
	(Fig. 831). (Ptinus, Gibbium, cosmop.; Mèzium, widespr.;
	Sphæricus, holarc.)PTÍNIDÆ
69.	Antennæ capitate, i.e. the last three joints forming a very abrupt
	club; elytra truncate; rather broad, slightly metallic beetles.
	(Sphærites, holarc.)SPHÆRÍTIDÆ
	Antennæ simple, not clubbed
70.	Prothorax very large, oval, longer than the elytra; hind coxæ
	very large, almost dividing the first sternite; antennæ very
	short; hind legs greatly thickened. Large beetles of burrowing
	habits. (Hypocéphalus, neotrop.) HYPOCEPHÁLIDÆ
	Of a different conformation71
71.	Front coxæ with a distinct side-piece (trochantin). (See couplet
	86) Some <b>DAS ÝTIDÆ</b>
	Front coxe without trochantin; long narrow beetles. (Lyméxy-
	lon, Melittómma, widespr.; Hylecœtus, holarc., Austr.)
	(See couplet 88) LYMEXYLÓNIDÆ LYMEXÝLIDÆ
72.	Front coxæ flat, rounded or globular, small and not prominent.73
	Front coxæ conical, prominent, usually large77
73.	, - J
	five abdominal segments; eyes absent; small flattened wingless
	beetles living as external parasites of beavers. (Fig. 833).
	(Platypsýllus, holarc.). (ACREIÓPTERA).
	PLATYPS ÝLLIDÆ
	Front coxæ rounded or globular; not such beetles
74.	Last joint of tarsi not excessively lengthened; tarsal claws not
	enlarged
	Last joint of tarsi greatly lengthened; tarsal claws very large;
	first three sternites connate; small, aquatic beetles. (Pse-
	phènus, Am.; Psephènops, Tychepsephènus, neotrop.;
	Metaeopsephènus, holarc.) PSEPHÉNIDÆ
<i>7</i> 5.	Prosternum prolonged behind into a process which is received in
	a notch in the mesosternum; prothorax loosely attached to the
	mesothorax
	Prosternum without such a backwardly directed process; eyes very
	small or wanting; rare, minute beetles living in the nests of
	rodents. (Leptinus, holarc.; Leptinillus, nearc.).
	T.E.PTÎNID <i>A</i> E

<b>7</b> 6.	Labrum fused with the clypeus; antennæ distant at base; tibial
	spurs well developed. (Fig. 835). (Cèbrio, palæarc.; Scapto-
	lènus, Am.; Cebriorhipis, Indo-malay) CEBRIÓNIDÆ
	Labrum free; tibial spurs very weak. (Plastócerus, Am.; Phylló-
	cerus, palæarc.; Euthysànius, Aplástus, nearc.). (PHYLLO-
	CÉRIDÆ) PLASTOCÉRIDÆ
77.	Abdomen with six sternites
	Abdomen with seven or eight sternites. (MALACODÉRMATA,
	CANTHARÒIDEA)
78.	Fifth segment of abdomen conical, as long as the three preceding
	segments together, the sixth minute. (See couplet 66).
	SCAPHIDÌIDÆ
	Fifth segment not conical nor excessively lengthened79
79.	
	prominent hairy pad between the claws. (See couplet 60).
	RHIPICERÁTIDÆ
	Hind coxæ simple, not grooved80
80.	Hind coxæ flat, not prominent, covered by the femora in repose;
	first joint of posterior tarsi usually very short and indistinct. 81
	Hind coxæ prominent, at least internally82
81.	Tarsi with the fourth joint of normal size; pronotum continuous
	with the propleura. (See couplet 68) CLÉRIDÆ
	Tarsi with the fourth joint very small, indistinct; pronotum sepa-
	rated from the pleura by a marginal line. (See couplet 65).
	CORYNÉTIDÆ
82.	Hind coxæ widely separated83
	Hind coxæ approximated or contiguous
83.	Eyes absent. (See couplet 84) a few <b>SÍLPHIDÆ</b>
	Eyes present, coarsely granulated; small, more or less ovate,
	brown beetles. (Scydmænus, cosmop.; Eucónnus, Steních-
	nus, widespr.; Leptomástax, Cephànium, Neùraphes,
	palæarc.)
84.	Tibial spurs large; antennæ gradually thickened or clavate; hind
	tarsi slender, not widened. Carrion-beetles. (Figs. 773, 776,
	803, 804, 806). (Sílpha, Càtops, Ptomóphagus, widespr.;
	Bathýscia, palæarc.; Necróphorus, nearc.; Liòdes, holarc.).
	(Including CATÓPSIDÆ, LIÓDIDÆ, ANISOTÓMIDÆ, pt.).
	SÍLPHIDÆ
	Tibial spurs small or indistinct
85.	r
	usually soft-bodied species
	Front coxæ without trochantin87

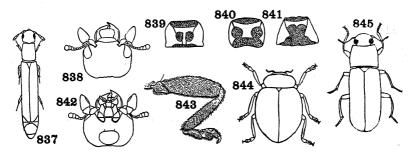
86.	Body with extensible vesicles. (Malthodes, Malachius, holarc.;
	Cóllops, nearc.) MALACHÌIDÆ
	Body without extensible vesicles. (Mélyris, widespr.; Dásytes,
	holarc., neotrop.; Haplocnèmus, palæarc.; Rhádalus, nearc.;
	Astỳlus, neotrop.). (Including RHADÁLIDÆ, (MELÝR-
	IDÆ) DASÝTIDÆ
87.	Epimera of metasternum distinct; maxillary palpi simple; body
	more or less ovate. (Brathinus, nearc.) BRATHÍNIDÆ
	Epimera of metasternum not visible; body elongate88
88.	Elytra shortened, exposing several of the abdominal segments;
	very small species; maxillary palpi simple in both sexes. (Mi-
	cromálthus, holarc.) MICROMÁLTHIDÆ
	Elytra entire; maxillary palpi of the male flabellate. (Hylecœtus,
	holarc., Austr.). (See couplet 71) Some LYMEX YLIDÆ
89.	Antennæ ten- or eleven-jointed, not abruptly capitate and not
	received in cavities; tarsi usually with more than three joints.
	(Fig. 820). (Philónthus, Stènus, Aleóchara, Homalòta,
	Oxýtelus, Lathròbium, Tachýporus, Quèdius, widespr.;
	Staphylinus, holarc., neotrop.) STAPHYLÍNIDÆ
	Antennæ inserted upon the front, near the inner margin of eyes.b
a.	Antennæ inserted upon the front, hear the inner margin of cycs. b  Antennæ inserted on the anterior margin of the head
	Antennæ inserted under the sides of the frontd
b.	Posterior coxæ large, contiguous; antennæ not terminated by a
	distinct club
	Posterior coxæ small, widely separated; antennæ terminated by a
	distinct clubSTENÎNÆ
c.	Antennæ approximate; prosternum developed in front of the
	anterior coxæ
	terior coxe
d.	Prothoracic spiracles conspicuous on removing the front coxæe
٠.,	Prothoracic spiracles difficult to perceive on account of the promi-
	nence of the sides of the prothoraxg
e.	Posterior coxæ transversef
	Posterior coxæ triangular, prominent; antennæ capillary and
	verticillate-pilose
1.	Antennæ filiform, not verticillate-pilose TACHYPORÎNÆ
~	Antennæ capillary, verticillate-pilose TRICHOPHYÎNÆ
8.	Anterior coxæ conical
h.	Anterior coxæ shorti
•	Anterior coxæ large, prominentj
	O. 1 I

i.	Tarsi two-jointed. LEPTOTYPHLÎNÆ Tarsi four-jointed. EVÆSTHETÎNÆ
j.	Vertex without ocelli
k.	Last joint of labial palpi dilated, very large, crescent-shaped.
	OXYPORÎNÆ
1.	Last joint of labial palpi not, or not strongly, dilatedl  Posterior coxæ conical
-	Posterior coxæ transversen
	Palpi with the last joint very small, subulate <b>PÆDERÌNÆ</b> Palpi with the last joint equal to the preceding. <b>PINOPHILÌNÆ</b>
n.	Posterior trochanters small, one-fifth the length of the femora; head with a distinct neck
	Posterior trochanters large, one-third the length of the femora;
	head without a distinct neck PHLŒOCHARÎNÆ
0.	Vertex without ocelli; elytra covering the greater part of the body.  PROTEINÎNÆ
	Vertex with one ocellus; elytra only slightly surpassing the meta- sternumPHLŒOBIĨNÆ
	Antennæ nine-jointed, with a very abrupt club, received in
	cavities on the underside of the prothorax; tarsi three-jointed;
	body rather short, the elytra with several acutely elevated
	1 1 1 1 1 /BWienenderland Wolfensen warne
	longitudinal ridges. (Micropéplus, Kalíssus, nearc.).
90.	MICROPÉPLIDÆ
90.	MICROPÉPLIDÆ  Middle coxæ distant; epipleuræ wanting; elytra usually with a reticulate sculpture; no phosphorescent organs; usually flat
90.	MICROPÉPLIDÆ  Middle coxæ distant; epipleuræ wanting; elytra usually with a reticulate sculpture; no phosphorescent organs; usually flat beetles, widened behind and often with a bold color pattern.
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90.	Middle coxe distant; epipleure wanting; elytra usually with a reticulate sculpture; no phosphorescent organs; usually flat beetles, widened behind and often with a bold color pattern. (Fig. 778) . LÝCIDÆ Middle coxe in contact; epipleure distinct; elytra not reticulate, rarely greatly reduced in size
90. 91.	Middle coxæ distant; epipleuræ wanting; elytra usually with a reticulate sculpture; no phosphorescent organs; usually flat beetles, widened behind and often with a bold color pattern. (Fig. 778) LÝCIDÆ Middle coxæ in contact; epipleuræ distinct; elytra not reticulate, rarely greatly reduced in size
	Middle coxæ distant; epipleuræ wanting; elytra usually with a reticulate sculpture; no phosphorescent organs; usually flat beetles, widened behind and often with a bold color pattern. (Fig. 778)
91.	Middle coxæ distant; epipleuræ wanting; elytra usually with a reticulate sculpture; no phosphorescent organs; usually flat beetles, widened behind and often with a bold color pattern. (Fig. 778) LÝCIDÆ Middle coxæ in contact; epipleuræ distinct; elytra not reticulate, rarely greatly reduced in size91 Antennæ inserted at the sides of the front, before the eyes92 Antennæ inserted on the upper part of the front or at the base of its anterior lobe; phosphorescent organs often present94
	Middle coxæ distant; epipleuræ wanting; elytra usually with a reticulate sculpture; no phosphorescent organs; usually flat beetles, widened behind and often with a bold color pattern.  (Fig. 778)
91.	Middle coxæ distant; epipleuræ wanting; elytra usually with a reticulate sculpture; no phosphorescent organs; usually flat beetles, widened behind and often with a bold color pattern.  (Fig. 778)
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91. 92.	Middle coxæ distant; epipleuræ wanting; elytra usually with a reticulate sculpture; no phosphorescent organs; usually flat beetles, widened behind and often with a bold color pattern. (Fig. 778)

	Maxillary and labial palpi normal; elytra usually complete. Fe-
	male usually larviform. (Drilus, Halacogáster, palæarc.;
	Selàsius, ethiop.; Phrixothrix, neotrop.; Karùmia, Persia;
	<b>Drilocéphalus</b> , neotrop.). (Including KARUMIIDÆ).
	DRÍLIDÆ
94	Head more or less completely covered by the prothorax; episterna
0 1.	of metathorax not sinuate on the inner side. (Luciola, wide-
	spr.; Lucidòta, Photìnus, Photùris, Am.; Lámpyris, pal-
	earc., ethiop.; Lamprocera, neotrop.; Rhagophthálmus,
	As.). (Including RHAGOPHTHÁLMIDÆ). LAMPÝRIDÆ
	Head not at all covered by the prothorax; episterna of metathorax
	sinuate on the inner side; antennæ of male sometimes flabel-
	late. (Cántharis (= Teléphorus), widespr.; Podàbrus, holarc.;
	Chauliognathus, Am.). (Including PHENGODIDÆ) (TEL-
	EPHORIDÆ) CANTHÁRIDÆ
95.	Tarsi four-jointed on all pairs of legs (front ones three-jointed in
	males of some Mycetophagidæ)96
	Tarsi with three joints or less
96.	Wings fringed with long hairs; very small, highly convex beetles97
	Wings not fringed99
97.	Hind coxe in contact, with plates at least partially covering the
	femoraCLÁMBIDÆ
	Hind coxæ distant, transverse, not laminate98
98.	Third joint of tarsi small, concealed in the bilobed second joint.
	(Sàcium, cosmop.; Corylophus, Orthóperus, holarc.; Ár-
	throlips, widespr.). (CORYLÓPHIDÆ, CLYPEASTÉRIDÆ)
	ORTHOPÉRIDÆ
	First three joints of tarsi subequal, each bilobed. (Phænocé-
	phalus, Japan) PHÆNOCEPHÁLIDÆ
99.	Abdominal sternites all free and movable
99.	Sternites one to four firmly united, immovable
100	
100.	J
	second and third joints and a part of the long fourth joint;
	minute elongate beetles with costate elytra. (Fig. 836). (Mon-
	œdus (= Adimerus), neotrop.). (ADIMÉRIDÆ).
	MONŒDIDÆ
	First tarsal joint not thus dilated101
101.	Front coxæ transverse; minute fungus-beetles (Cybocéphalus,
	widespr.). (See couplet 118).
	Tribe Cybocephalini of the <b>NITIDULID</b>
	Front coxe not transverse

102.	Front coxæ globose
103.	Front coxe oval. (If conical, cf. Corynetidæ, couplet 81)105 Tarsi slender, third joint distinct, but shorter than the second;
100.	very small species. (If the cheeks bear projections, see Sil-
	vanidæ, couplet 52; or Cucujidæ, couplet 52, if the body is
	greatly flattened). (Fig. 817). (Mycetæa, holarc., ethiop.;
	Rhànis, nearc.; Liésthes, palæarc.) MYCETÆIDÆ
	Tarsi more or less dilated and spongy beneath; more or less
	elongate beetles with hard body and strongly clubbed antennæ; usually moderate-sized or large species
104	Front coxal cavities closed; metathoracic epimera separated by
101.	a distinct suture; body elongate oval. ( <b>Tríplax</b> , widespr.;
	Erótylus, Ægithus, Brachysphænus, Mycotrètus, neo-
	trop.; <b>Trítoma</b> , holarc.; <b>Cyrtomórphus</b> , Indomal.).
	EROTÝLIDÆ
	Front coxal cavities open; metathoracic epimera not separated;
	body elongate, slender. (Fig. 834). (Languria, Acropteróxys,
	Am.; Adanástus, palæarc., Indomal.; Doubledàya, Indomal.). LANGURÌIDÆ
105	Front coxe almost in contact, prosternum more or less mem-
105.	branous, not visible between them; antennæ nine-jointed; small,
	convex, roughly sculptured beetles. (Georýssus, widespr.).
	GEORÝSSIDÆ
	Front coxæ well separated by the horny prosternum 106
106.	Head more or less concealed by the projecting prothorax; last
	joint of tarsi usually very long; body cylindrical. (Cis, cosmop.;
	Enneárthron, Hendecátomus, holarc.; Rhopalodóntus, palæarc.; Oròphius, palæarc., Austr.). (CIÒIDÆ, CÍSIDÆ).
	palæarc.; Orophius, palæarc., Austr.). (CIOIDÆ, CISIDÆ).
	Head free, not covered by the prothorax; body oval, depressed,
	pubescent. (Litárgus, widespr.; Mycetóphagus, holarc.).
	(TRITÓMIDÆ)
107.	
	simple, not dilated or spinose; not aquatic108
	Antennæ with a large serrate, seven-jointed club (Fig. 812); front
	and middle tibiæ dilated and armed with rows of spines; small,
	subaquatic beetles. (Heterócerus, cosmop.; Litórimus, Mi-
100	cíllus, palæarc.)
108.	tant from the mesosternum. (Colýdium, Aulònium, Cérylon,
	widespr.)

	Antennæ inserted	d on the front; fr	ont coxæ inclose	ed behind by the
	mesosternum.	(Murmídius,	Bothrideres,	Mychócerus,
	holarc.). (Inclu	iding BOTHRII	DÉRIDÆ) N	IURMIDÌIDÆ
109.	Tarsi three-jointe	$\operatorname{ed}$		
	Tarsi with less th	nan three joints.	·	



Figs. 837-845. Coleoptera

837. Cylindrosella (Fouts).

~~.	
838.	Epilachna, head from above (Silvestri) Coccinellidæ.
839.	Prosternum of beetle, showing coxal cavities separated and closed behind
	(Wickham).
840.	Prosternum of beetle, showing coxal cavities separated and open behind
	(Wickham).
841.	Prosternum of beetle, showing coxal cavities confluent and open behind
	(Wickham).
842.	Epilachna, head from below (Silvestri) Coccinellidæ.
843.	Epilachna, hind leg (Silvestri) Coccinellidæ.
844.	Coccinellidæ.
845.	Tenebrio (Girault) Tenebrionidæ.
110.	Wings fringed with long hairs111

	111180 1100 111118000, 01 000 1110000 1110000 1110000
111.	Abdomen with only three sternites. Very small, highly convex
	beetles. (Sphærius, holarc.) SPHÆRÌIDÆ
	Abdomen with six or seven sternites
112.	Antennæ slender, nine- to eleven-jointed, with whorls of long
	hairs; very minute, shining beetles, usually found on foliage.
	(Ptílium, Trichópteryx, Ptinélla, Acrótrichis, palæarc.;
1	$\mathrm{Am.}$ ; Limulòdes, $\mathrm{Am.}$ ). $(TRICHOPTER \acute{Y}GID\pounds)$ .
	THE THE

Wings not fringed, or at most with a short fringe............113

PTILIIDÆ

	Antennæ short, eight-jointed, thickened apically; very small,
	ovate, aquatic beetles. (Hydróscapha, holarc.).
	HYDROSCÁPHIDÆ
113.	Second joint of tarsi dilated; the third joint consisting really of
	two joints, the small, true third joint being fused with the base
	of the last joint, which thus appears as the third114
	Second tarsal joint not dilated115
114.	Tarsal claws usually dilated or toothed at the base; first sternite
	with curved coxal lines; mesothoracic epimeron triangular;
	small, rounded, convex usually brightly spotted beetles. "Lady-
	birds." A large widespread family. (Coccinélla, Chilócoris,
	Hippodamia, Scýmnus, Adàlia, Hyperáspis, Megílla,
	<b>Ánatis, Epiláchna</b> ). (Figs. 838, 842, 843, 844). (Including
	CERASOMMATIDÌIDÆ) COCCINÉLLIDÆ
	Tarsal claws simple; first sternite without coxal lines; meso-
	thoracic epimeron quadrangular; small oblong or oval beetles,
	often with a striking color pattern, usually living in fungi.
	(Endómychus, widespr.; Lycoperdina, holarc.; Aphorísta,
	nearc.; Epípocus, Rhýmbus, Am.; Ámphix, neotrop.;
	Sphærosòma, palæarc.; Amphistérnus, Indomal.).
	ENDOM ÝCHIDÆ
115.	Elytra entire
	Elytra truncate, exposing the last abdominal segment118
116.	Body broadly oval, convex; prothorax much widened behind;
	first three tergites more or less connate; very small beetles.
	(Aphænocéphalus, As.; Discolòma (= Notióphygus), neo-
	trop., Afr.). (APHÆNOCEPHÁLIDÆ, PSEUDOCORY-
	$L\acute{O}PHID$ $ extbf{ ilde{E}}, NOTIOPH\acute{Y}GID$ $ extbf{ ilde{E}}) \dots$ DISCOLÓMID $ extbf{ ilde{E}}$
	Body more elongate, the prothorax narrower, not widened be-
	hind
117.	Abdominal sternites all free; wings with a short fringe of hairs
	(Fig. 808). (Corticària, widespr.; Melanophthálma, cosmop.;
	Latrídius, Cartócere, holarc.; Enícmus, holarc., Austr.).
	LATHRIDÌIDÆ
a	Last three or four joints of the antennæ separately thickened,
	spindle-shaped, and set with long curved hairs. DASYCERÎNÆ
	Antennæ without long hairs at apexb
b	Anterior coxal cavities closed behind
	Anterior coxal cavities open behind HOLOPARAMECINÆ
c	Anterior coxal cavities open behind <b>HOLOPARAMECĪNÆ</b> Anterior coxæ separate; head longer before the eyes; elytra often

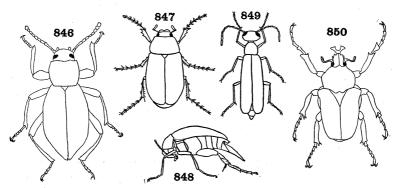
	Anterior coxæ contiguous; head shorter before the eyes; elytra never carinate
]	Basal three sternites connate. (See couplet 108).
	A few COLYDÌIDÆ
118.	Front coxe subtransverse; maxillæ with a single lobe.
	Tribe Smicriptini of the NITIDULIDÆ
	Front coxæ small, rounded; maxillæ bilobed; small flattened bark
	beetles MONOTÓMIDÆ
119.	Tarsi two-jointed; antennæ eleven-jointed; metasternum very
	long; very small, elongate beetles, with the elytra oval. (Jacob-
	sònium, Sumatra) JACOBSONÌIDÆ
	Tarsi consisting of a single joint; antennæ four-jointed; meta-
	sternum not greatly elongated; very small, broad species.
	(Cyathócerus, neotrop.). (If both elytra and wings are want-
	ing and body larviform, see couplet 198) CYATHOCÉRIDÆ
120.	Frontal coxal cavities closed behind (Fig. 839)
	Frontal coxal cavities open behind (Fig. 840)
121.	Tarsal claws simple
	Tarsal claws pectinate; usually elongate, convex, rather soft
	bodied and often thinly silky-pubescent beetles of small or
	moderate size; prothorax widened behind. (Allécula, cosmop.;
	Hymenòrus, palæarc., Am.; Cistèla, widespr.; Ctenìopus,
	palæarc.; Lophópoda, Am.). (ALLECÙLIDÆ) . CISTÉLIDÆ
122.	Ventral segments (abdominal sternites) all freely movable. (If
	the mesosternum is carinate, compare some rare Silphidæ,
	couplet 84)
	First two to four ventral segments more closely joined together,
	more or less fused or immovable
123.	Antennæ 11-jointed
	Antennæ 10-jointed
124.	Prothorax cylindrical; small, soft-bodied beetles with long,
	slender antennæ and protruding eyes; elytra not completely
	covering the abdomen. (Pètria, palæarc.) PETRÌIDÆ
	Prothorax not cylindrical
125.	Prothorax quadrate, not wider than the head; narrow-bodied
	beetles. (If body is greatly flattened, compare males of some
	Cucujidæ, couplet 52, and Silvanidæ, couplet 52). (Elácatis
	(=Óthnius), widespr.; Ábaba, nearc.). (OTHNÌIDÆ).
	ELACÁTIDÆ
	Prothorax greatly expanded at the sides, much wider than the
	head. (Nílio, neotrop.) NILIÓNIDÆ

126.	Elytra entire; small convex beetles. (Sphíndus, holarc.) SPHÍNDIDÆ
	Elytra truncate, exposing the pygidium; small, flattened beetles
	(See couplet 36) Males of RHIZOPHÁGIDÆ
127.	Five ventral segments
	Six ventral segments, the first two immovably united; small black
	beetles. ( <b>Eurystèthus</b> (= Ægialites), nearc., Cali. to Alaska,
	Persia). (ÆGIALÍTIDÆ) EURYSTÉTHIDÆ
128.	Penultimate joint of tarsi spongy pubescent beneath; front coxe
- <b>-</b> 0.	prominent; slender, elongate, usually hairy, soft-bodied species,
	sometimes of metallic color. (Làgria, widespr.; Arthromàcra,
	holarc.; Statira, Am.; Nemostira, Afr., Indomal.).
	LAGRÌIDÆ
	Penultimate joint of tarsi not spongy pubescent beneath; front
	coxe short, not projecting from the cavities
129.	Antennæ filiform or gradually clavate, the joints usually more or
	less bead-like, not concealed under the sides of the head; beetles
	of variable form, oval, elongate, or even pedunculate; usually
	hard-bodied, black or dark colored; moderate, large or small
	species. (Figs. 845, 846). A very large and widely distributed
	family. (Tenèbrio, Strongýlium, Hèlops, Bláps, Ásida,
	Bolitóphagus, Diapèris, Eleòdes, Epítragus, Nyctóbates,
	Platýdema, Zópherus, Tentýria, Tribòlium, Meracántha).
	(Including HELÓPIDÆ, OPÁTRIDÆ, PIMELÌIDÆ, BLÁP-
	IDÆ, DIAPÉRIDÆ) TENEBRIÓNIDÆ
	Antennæ strongly clavate, more or less completely concealed be-
	neath the sides of the head, with large two-jointed club, genicu-
	late at the base, with the first joint very long; middle tarsi some-
	times four-jointed, very small, oval, flattened beetles living in
	ants' nests. (Cossyphòdes, Cossyphodites, ethiop.; Cossy-
	phodinus, India) COSSYPHÓDIDÆ
130.	Antennæ geniculate; elytra truncate behind, exposing two ab-
	dominal tergites; small, oval, hard-bodied species. (Acritus,
	widespr.). (See couplet 46) A few <b>HISTÉRID</b>
	Antennæ not geniculate
131.	Head not strongly or suddenly narrowed or constricted behind
	the eyes; tarsal claws simple or cleft132
	Head strongly and suddenly constricted behind the eyes; if more
	gradually narrowed, the tarsal claws are pectinate138
132.	Middle coxæ not noticeably prominent
	Middle coxe very prominent, contiguous: prothorax without

lateral margin; penultimate joint of tarsi dilated and with a dense brush of hairs beneath; slender, soft-bodied species. (Nacérda, Asclèra, Sessínia, widespr.; Cálopus, Xanthróchroa, holarc.; Óxacis, Am.; Œdémera, palæarc.).

# ŒDEMÉRIDÆ

133. Antennæ received in grooves on the underside of the prothorax; small, oval, flattened beetles with the head partly concealed



Figs. 846-853. Coleoptera

- 846. Eleodes (Hyslop) Tenebrionidæ.
- 847. Phyllophaga (Forbes) Melolonthidæ.
- 848. Mordellistena. Mordellidæ.
- 849. **Epicauta** (Bruner) Meloidæ.
- 850. Goliathus (Westwood) Cetoniidæ.

	in the prothorax; legs retractile. (Monómma, widespr.). (MONÓMMIDÆ) MONOMMÁTIDÆ
	Antennæ free, not received in grooves
134.	Prothorax with a sharp lateral margin
	Prothorax not marginal laterally, narrowed behind, its disk
	without impressions. (Myctèrus, Sphæristes (=Salpingus),
	Lissodérma, widespr.; Pỳtho, holarc.). (SALPÍNGIDÆ).
	PÝTHIDÆ
135.	Epimera of mesothorax not reaching the coxæ, the coxal cavities
	entirely surrounded by the sterna (males of a few genera). (See
	couplet 54)
	Epimera of mesothorax attaining the coxæ
136.	Metasternum long; epimera of metathorax visible137

197	Metasternum quadrate, epimera of metathorax covered. (Males of a number of genera (see couplet 52) CUCŪJIDÆ, part Prothorax widened toward the base, its disk with basal impres-
137.	sions; tarsal claws sometimes cleft or appendiculate; elongate or
	broadly oval species. (Orchèsia, Phlæotrỳia, Serropálpus,
	widespr.; Tetrátoma, Hallómenus, Melándrya, Ósphya,
	holarc.; <b>Pénthe</b> , palæarc., Indomal.). (SERROPÁLPIDÆ).
	melandr vide.
	Prothorax narrowed behind and in front, the sides rounded or
	toothed; without basal impressions; tarsal claws simple; man-
	dibles very large and powerful, porrect. Very large, elongate
	beetles. (Trictenótoma, Ind., E. Ind.; Autócrates, So. As.).
	TRICTENOTÓMIDÆ
138.	
	margined at the sides, as wide as the elytra at base; tarsal
	claws pectinate, with a large appendage at base; medium-sized
	slender beetles. (Cepháloon, holarc.; Typítium, nearc.;
	Sponidium, palæarc.) CEPHALÒIDÆ
	Head suddenly narrowed behind
139.	
	Prothorax rounded on the sides, without a sharp lateral mar-
	gin
140.	
	Antennæ pectinate (male) or subserrate (female); tarsal claws
	serrate or toothed; elytra covering the abdomen. (Evaniócera,
	palæarc., Austr.; <b>Pelecótoma</b> , holarc.).
	Tribe Evaniocerini of RHIPIPHÓRIDÆ
141.	
	trochantin; head placed vertically against the thorax; tarsal
	claws simple, or cleft and pectinate; body usually conically
	narrowed behind, the abdomen often prolonged and pointed at
	tip; small pubescent beetles. (Fig. 848). (Mordélla, Mordel-
	listèna, Anáspis, cosmop.; Tomóxia, widespr.; Tólida,
	palæarc.) MORDÉLLIDÆ
	Hind coxæ transverse; front coxæ with trochantin; body not
	greatly narrowed behind; tarsal claws simple. (Scraptia, cos-
	mop.; Trotommídea, Trotómma, palæarc.; Eválces, neo-
	trop.) SCRAPTÌIDÆ
142	
	Base of prothorax as wide as the elytra; body broad, much
	narrowed behind; elytra usually shortened and narrowed be-

hind; antennæ pectinate in the male, often serrate in the female. Females sometimes much degenerate or even larviform. (Pelecotomòides, Rhipídius, widespr.; Macrosiagon, cosmop.; Rhipíphorus, holarc.; Myodites, neotrop.).

## RHIPIPHÓRIDÆ

- 145. Head constricted far behind the finely granulated eyes. (Macràtria, cosmop.; Pedìlus (=Corphyra), Stereopalpus, holarc.)

  Head constricted just behind the coarsely granulated eyes; tarsi apparently with 4, 4, 3 joints, as the penultimate joint is extremely minute; first two ventral segments immovably united.

(Hylóphilus (=Eùglenes, =Áderus), cosmop.; Sýzeton, Austr.). (XYLOPHÍLIDÆ, EUGLÉNIDÆ, ADÉRIDÆ).

# **HYLOPHÍLIDÆ**

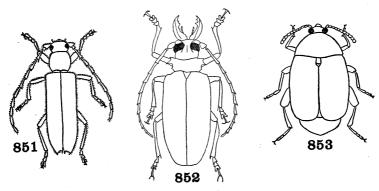
146. Tarsal claws simple; head horizontal; antennæ serrate, often pectinate in the male (Fig. 797); body flattened; moderate-sized beetles. (**Dendròides, Schizòtus,** holarc.; **Pyróchroa,** palæarc.; **Pseudopyróchroa,** palæarc., Indomal.).

# **PYROCHRÒIDÆ**

Tarsal claws toothed or cleft; head deflexed, with the front vertical; elytra sometimes shortened; body plump, usually more or less cylindrical; moderate or large beetles (Fig. 849). Blister beetles. (Epicaûta, Lýtta (=Cántharis), Méloe, Nemógnatha, Cissites, widespr.; Zonites, cosmop.; Macróbasis, Am.; Hórnia, nearc.). (CANTHÁRIDÆ, LÝTTIDÆ).

# MELÒIDÆ

147. Submentum pedunculate; i.e. the mentum supported at its base by a narrow portion or peduncle; antennæ eleven-jointed, serrate, rarely pectinate; head prolonged into a broad muzzle; antennæ and body usually pubescent; elytra shortened, exposing the pygidium. (Fig. 853). Pea and bean weevils. (Mylàbris (=Brùchus, =Lària), cosmop.; Spermóphagus, Pachýmerus,

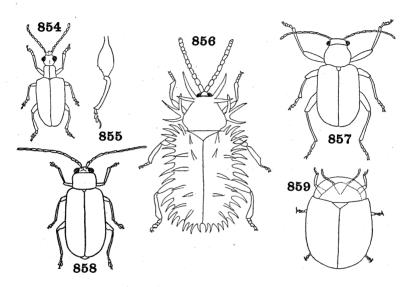


Figs. 851-853. Coleoptera

- 851. Elaphidion (Forbes) Cerambycidæ.
- 852. Acanthophorus (Gahan) Prionidæ.
- 853. **Mylabris** (Felt) Mylabridæ.
- - 149. Prothorax sharply margined at the sides which commonly bear teeth or spines; labrum connate; front coxæ strongly transverse; inner lobe of maxillæ very small or obsolete; large or very large, considerably flattened, usually brown or black beetles. (Fig. 852). (**Priònus**, holarc.; **Macrótoma**, widespr.; **Mállo-**

	don, Orthosòma, Am.; Derancístrus, Pyròdes, neotrop.). PRIÓNIDÆ
	Prothorax very rarely margined at the sides; labrum free; front
	coxæ rounded, rarely strongly transverse; inner lobe of max-
150.	illæ more or less well developed; antennæ never pubescent150 Front tibiæ obliquely grooved on the inner side; front coxæ never
100.	transverse; last joint of palpi usually pointed. (Fig. 791).
	(Monóchamus, Obèrea, widespr.; Sapérda, holarc.; Dorcà-
	dion, palæarc.; Oncideres, Am.; Tetraòpes, nearc.; Bató-
	cera, Afr., Austr.; Tragocéphala, Afr.; Platyomópsis, Austr.).
	(Including BATOCÉRIDÆ)LAMÌIDÆ
	Front tibiæ not grooved; last joint of palpi never acute at tip;
	antennæ pubescent. (Fig. 851). (Leptùra, Callichròma, Xy-
	lótrechus, widespr.; Acmæops, Strangàlia, Phymatòdes,
	Rhàgium, Leptùra, holarc.; Ebùria, Elaphídion, Cyllène,
151	Am.)
151.	vertical
	Mouth inferior, the anterior part of the front prominent, so that
	the mouth is confined to the underside of the head and is small,
	hidden or nearly so
152.	Antennæ widely separated at the base; elytra of hard texture. 153
	Antennæ generally closely approximate at base, not widely separated; elytra more or less soft in texture
153.	Intermediate abdominal sternites not narrowed medially, the
100.	pygidium not exposed behind the elytra
	Intermediate abdominal sternites narrowed medially, the pygidium usually exposed, declivous
154.	Prothorax rounded on the sides, without distinct lateral margin;
	head produced, the eyes prominent; prosternum very narrow. 155
	Prothorax with a distinct lateral margin (if rarely not margined,
	the antennæ are usually short, with the terminal joints trans-
	verse and more or less serrate); head not produced, the eyes
	not prominent; prosternum broad. (If antennæ are strongly clavate, compare a few Erotylidæ, couplet 104)161
155.	Antennæ not separated by the entire front of the head 156
	Antennæ separated by the entire front of the head157
156.	Prosternum very narrow, not distinct; body beneath clothed with
	dense silvery pubescence; first sternite as long as the others
	united; elongate, more or less metallic, semiaquatic beetles.
	(Donàcia, widespr.; Hæmònia, Plateùmaris, holarc.).
	DONACÎID <i>Æ</i>

Prosternum distinct; body beneath not thus densely pubescent; first sternite shorter; less elongate beetles, not aquatic. (Megáscelis, Atelédera, neotrop.) ...... MEGASCÉLIDÆ



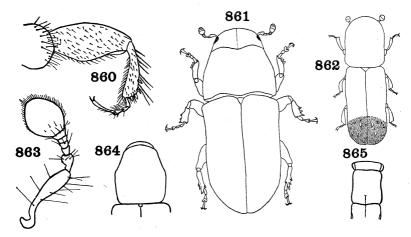
Figs. 854-859. Coleoptera

- 854. Crioceris (Jacoby) Crioceridæ.
- 855. Sagra, hind leg (Jacoby) Sagridæ.
- 856. **Hispella** (Fletcher) Hispidæ.
- 857. Fidia (Johnson and Hammar) Eumolpidæ.
- 858. Diabrotica (Chittenden) Galerucidæ.
- 859. Cassida (Jones) Cassididæ.

lindrical beetles of compact shape, with the head flat, perpen-
dicular and invisible from above. (Cryptocéphalus, cosmop.;
Pachýbrachys, cosmop., mainly Am.; Diorýctus, Indomal.;
Cœnòbius, As., Afr.; Mónachus, neotrop.; Bassàreus,
nearc.)
159. Prothorax without grooves for the reception of the antennæ;
elytra not tuberculate
Prothorax with grooves on the flanks for the reception of the
antennæ; elytra tuberculate; body subquadrate or somewhat
elongate; head flat, deeply inserted in the prothorax, invisible
from above. (Chlàmys, Éxema, widespr.). (FULCIDÁCIDÆ)
CHLAM ÝDIDÆ
160. Apical joint of maxillary palpi pointed; hind femora with one
or two teeth; last joint of tarsi and claws very long; body
elongate, the head not concealed. (Megálopus, neotrop.;
Mastostèthus, neotrop.; Temnáspis, Colobáspis, Afr.,
Indomal.) MEGALOPÓDIDÆ
Apical joint of maxillary palpi more or less truncate; hind femora
without teeth; claw joint normal; elongate, more or less cylin-
drical species with the head deflexed or perpendicular; elytra
generally covering the pygidium. (Clytra, Euras., Afr.; Gyn-
androphthálma, widespr.; Cyaniris, widespr.; Æthomórpha,
Afr., Indo-Austr.; Aspidólopha, Indomal.; Coscinóptera,
nearc.; Bàbia, Am.)
161. Third joint of tarsi deeply bilobed; front coxæ usually
rounded
Third tarsal joint entire, not bilobed; front coxæ transverse;
body oval, convex; antennæ moderately thickened toward
apex. (Fig. 790). (Chrysomèla, Plagiódera, widespr.; Phy-
todécta, Phyllodécta, holarc.; Leptinotársa (L. decemlineàta,
Colorado potato-beetle), Polyspìla, Am.; Timárcha, palæarc.;
Pýrgo, Austr.)
with grooves into which the tibiæ can be placed; metasternum
and abdomen grooved for the reception of the femora; short, very convex beetles, often brilliantly metallic. (Lamprosòma,
neotrop., As.; <b>Lycnóphanes</b> , neotrop., ethiop.).
LAMPROSOMÁTIDÆ

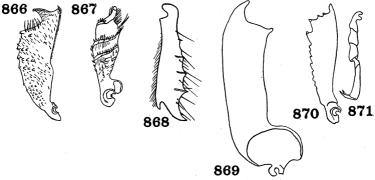
Prothorax generally narrower than the elytra; legs not compressed; abdomen not grooved for the reception of the femora; more or less oblong, convex beetles. (Fig. 857). (Fídia, Co-

	láspis, Am.; Chrýsochus, Adóxus, holarc.; Chrysolámpra,
	As.; Nodóstoma, Indomal.; Colasposòma, Afr., Indomal.;
	Tricliòna, Indomal.; Corynòdes, Afr., Indomal.; Pària,
	Am., As.; Nodonòta, Am., Austr.) EUMÓLPIDÆ
163.	Hind femora slender, adapted for walking; tibiæ usually sub-
	cylindrical; tarsi slender, not retractile. (Fig. 858). (Lupèrus,
	Galerucélla, cosmop.; Galeruca, holarc.; Monóxia, Ceró-
	toma, Diabròtica, Trirhábda, Am.) GALER ÜCIDÆ
	Hind femora greatly thickened, adapted for leaping; tibiæ fre-
	quently sulcate externally, the tarsi retractile. (Altica (= $H\acute{a}l$ -
	tica)). (HALTÍCIDÆ)ALTÍCIDÆ
164.	Head free, not retracted beneath the prothorax; body usually
	spinose, narrowed in front, broad and truncate behind. (Fig.
	856). (Híspa, Afr., As.; Chálepus, Am.; Cephalòlia, neotrop.;
	Cephalodónta, neotrop.; Gonóphora, Callíspa, Dactylíspa,
	Afr., As.)
	Head concealed under the prothorax, which with the elytra, is
	widely margined; body oval or nearly circular in outline. (Fig.
	859). (Cássida, Coptocycla, widespr.; Chelymórpha, Am.;
	Hoplionota, ethiop., Indo-Austr.; Pseudomesophàlia, neo-
	trop.; Aspidomórpha, Afr., Indo-Austr.) CASSÍDIDÆ
165.	Tarsi apparently three-jointed, with the second joint lobed. (Fig.
	877). (AGLYCYDERÒIDEA)
	Tarsi apparently four-jointed, with the third joint lobed167
166.	Head without a distinct beak in either sex, wider than the ante-
	rior part of the pronotum; prothorax subquadrate, the sides
	straight and the pleuræ clearly separated from the notum.
	(Fig. 877). (Aglycýderes, Canary Isl., New Zealand, New
	Caledonia) AGLYCYDÉRIDÆ
	Beak in female well developed; in male rarely attaining a length
	greater than its width; head narrower than the anterior part of
	the pronotum; prothorax oval, the sides bulging outwards and
	the pleuræ indistinctly separated from the notum. (Fig. 872).
	(Proterhinus, Hawaii) PROTERHÍNIDÆ
167.	Rostrum or beak extremely short and broad, scarcely developed;
	tibiæ with a series of teeth externally or the front ones produced
	into a stout curved process at tip; antennæ short with a broad
	club; small oval or cylindrical beetles. (SCOLYTÒIDEA) 168
	Beak of variable length, usually at least broader than long;
	tibiæ simple, without teeth externally or process at tip; an-
	tennæ clubbed or not. (CURCULIONÒIDEA)172



Figs. 860-865. Coleoptera

- 860. Pityogenes, front leg (Felt) Ipidæ.
- 861. Dendroctonus (Hopkins) Ipidæ.
- 862. **Xyleborus** (Hubbard) Ipidæ.
- 863. Ips, antenna (Felt) Ipidæ.
- 864. Hylastes, dorsal outline of head and prothorax (Felt) Scolytidæ.
- 865. Platypus, dorsal outline of head and prothorax (Felt) Platypodidæ.
- 170. Front tibiæ without a prominent process on the outer apical angle. (Figs. 810, 861, 863, 864, 868, 870, 871). (Íps, Pityóphthorus, Hylesinus, Hylástes, cosmop.; Dendróctonus,



Figs. 866-871. Coleoptera

- 866. Scolytoplatypus, front tibia (Hopkins) Scolytoplatypodidæ.
- 867. Platypus, front tibia (Platypodidæ).
- 868. Erineophilus, front tibia (Felt) Ipidæ.
- 869. Camptocerus, front tibia (Hopkins) Scolytidæ.
- 870. Ips, front tibia (Hopkins) Ipidæ.
- 871. **Ips,** tarsus (Hopkins) Ipidæ.

Front tibiæ with prominent rugosities on ventral area. (Fig. 866). (Scolytoplátypus) .... SCOLYTOPLATYPÓDIDÆ

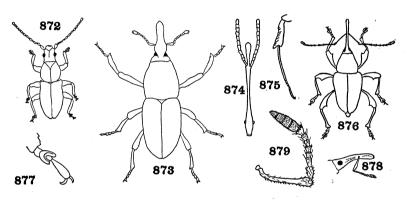
	dorrhýnchus, neotrop.). (BRÉNTHIDÆ) BRÉNTIDÆ
174	Polni florible, lohum present
174.	Palpi flexible; labrum present
	Palpi rigid; labrum wanting CURCULIÓNIDÆ 1
a.	Prothorax with a distinct acute or carinate lateral margin; an-
	tennæ straight, not geniculateb
	Prothorax without a distinct lateral margin, rarely with the weak
	indication of one
b.	Elytra short, exposing three dorsal abdominal segments; short,
	stout species. (Pterócolus, Am.) PTEROCOLINÆ
	Elytra completely covering the abdomen. (Oxycórynus, neo-
	trop.) OXYCORYNÎNÆ
c.	Antennæ straight, not received in grooves; club frequently with
	separated jointsd
	Antennæ geniculate, more or less completely elbowed; the beak
a	with grooves for the reception of the scape; club compactj Antennal club composed of completely separated jointse
a.	Antenna citib composed of completely separated jointse  Antenna with the joints closely united into a compact, oval club.f
	Mandibles flat, toothed on inner and outer sides, tibiæ with short
е.	terminal spurs; tarsal claws free, bifid or acutely toothed.
	RHYNCHITÎNÆ
	Mandibles stout, pincers-shaped; tibial spurs forming two strong
	hooks; tarsal claws connate at base ATTELABÎNÆ
f.	Trochanters elongate, the femora attached to their apices and thus
	separated from the coxæ; elytra completely covering the abdo-
	men, concealing the pygidium; beak porrect, usually long;
	small species. (Apion, cosmop.) APIONÎNÆ
	Trochanters triangular, the femora contiguous with the coxæ. g
g.	Tips of elytra leaving the pygidium exposedh
	Tips of elytra completely covering the abdomen. (Brachýcerus,
	palæarc., ethiop.; Micrócerus, Bròtheus, ethiop.).
	BRACHYCERÎNÆ
h.	First joint of antennæ no longer than the second; beak short,
	broad; middle and hind coxe widely separated; hind legs very
	long, fitted for grasping; broad, short species. (Tachýgonus, Am.)
	First joint of antennæ longer than the secondi
;	Hind femora short, very broad, their outer margin crenulate and
1.	strongly curved; small species with long, curved beak; joints
	of antennal club less closely compacted than usual. (Allo-
	córynus, Am.)
	Ouguand, Lille,

<sup>&</sup>lt;sup>1</sup> This very extensive family has been divided into a long series of subfamilies, the limits of which are not agreed upon by different workers. Those generally recognized may be distinguished by the above key.

Hind femora of the usual elongate, clavate form; beak short, broad; large species. (Ithýcerus, nearc.). (BÉLIDÆ, Leng).

ITHYCERÎNÆ

j. Abdomen of male with an apparent extra segment at tip; the pygidium and anal segment separated by a suture; club of antennæ usually annulated, sensitive and not shining; third joint of tarsi usually deeply bilobed, with a brush beneath (rarely with narrow setose tarsi in some subaquatic species). k



Figs. 872-879. Coleoptera

- 872. Proterhinus (Perkins) Proterhinidæ.
- 873. Sitophilus (Back and Cotton) Curculionidæ.
- 864. Brentus, head from above (Berlese) Brentidæ
- 875. Platypus, tibia and tarsus of front leg (Felt) Platypodidæ.
- 876. Balaninus (Chittenden) Curculionidæ.
- 877. Aglycyderes, hind tarsus (Tillyard) Aglycyderidæ.
- 878. Curculionid Weevil, side of head.
- 879. Sitones, antenna (Silvestri) Curculionidæ.
  - Abdomen similar in both sexes, the pygidium not divided by a suture as the anal segment of the male is at least partly free and retractile; club of antennæ usually with its basal joint enlarged or shining, or both, without or with indistinct sutures......

l. Prosternum not forming a triangular plate in front of the con	
simple or grooved to receive the beak  Prosternum forming a triangular plate in front of the coxæ; be	. m
received in the prosternum when in repose; tarsi usually narr	AK.
and bristly. (Thecestérnus, holarc.; Býrsops, palæar	e.).
(BRYSOPIDE) THECESTERNIN	Æ
m. Trochanters elongated, completely separating the base of	ine
femora from the coxæ NANOPHYIN Trochanters short, triangular; base of femora contiguous w	ith
the coxæthangular, base of femora configuous w	
n. Beak short and stout, received between the front legs in repo	
jumping species with short thickened hind femora.	~~,
ORCHESTÎN	
Beak usually elongate, slender and porrect, rarely conceal	ed;
hind femora not thickened for jumping. CURCULIONIN	Æ
o. Pygidium covered by the elytra; mentum attached to a sh	ort
quadrate gular peduncle	Æ
Pygidium exposed; peduncle of mentum long and narrow,	the
buccal cavity elongate. (Caléndra (=Sphenóphorus)). (CA ÁNDRIDÆ, RHYNCOPHÓRIDÆ) CALENDRÌN	. <i>L-</i> -
175. Beak long, well developed, prothorax without lateral carina	
margin or transverse ridge; elytra completely covering the p	)y-
gidium; anterior coxæ conical. (Rhinómacer, holarc.; Ner	n-
onyx, palæarc.). (DOYDIRHÝNCHIDÆ, NEMONÝC	T-
ID $E$ $)$ RHINOMACERID	Æ
Beak very short; prothorax trapezoidal, with carinate late	ral
margin and usually a transverse raised line or ridge; pygidiu	$\mathbf{m}$
exposed; anterior coxæ globose. (BRUCHÉLIDÆ, CHO	R–
ÁGIDÆ, PLATYRRHÍNIDÆ, PLATYSTÓMIDÆ).	
ANTHRÍBID	
176. Lamellæ of antennal club not capable of closing together, usual	
not flattened, but forming a more or less comb-like mass (F	
814); only five abdominal sternites	
Lamellæ of antennal club movable, flattened and capable of clo	
apposition to form a solid club; six, or more rarely five, a	
dominal sternites	
177. Mentum entire, ligula behind, or at the apex of the menturantennæ not curled in repose	
Mentum deeply emarginate, the ligula large, corneous, filling t	
emargination; labrum free; antennæ straight, curled in repo	
large elongate, somewhat flattened, shining beetles with deep	
lined elytra PASSÁLID	Æ
· · · · · · · · · · · · · · · · · · ·	

178.	Ligula and maxillæ covered by the mentum; antennæ usually
	elbowed. (Fig. 814). Stag-beetles. (Dórcus, Platýcerus,
	Cerùchus, Lucànus, holarc.; Nicàgus, nearc.; Odontólabis,
,	As., Indomal.; Lámprima, Austr.) LUCÁNIDÆ
. ]	Ligula and maxillæ not covered; antennæ straight. (Sinodén-
150 (	dron, holarc.) SINODÉNDRIDÆ
179.	Side pieces of metasternum attaining the coxæ; abdomen usually
٠.	with six tergites
	Side pieces of mesosternum not attaining the coxæ; abdomen with
	five tergites; body heavily sculptured, the elytra usually with
	distinct rows of tubercles; pygidium covered by the elytra;
	small or moderate, rarely large beetles. Skin-beetles. (Tróx,
	cosmop.; Gláresis, holarc.; Cryptogènius, neotrop.). TRÓGIDÆ
100	
180.	Abdominal spiracles placed in a line, each one in the membrane
	between the sternite and tergite and all of them covered by
	the elytra; mentum and ligula separated by a suture181
	At least some of the apical abdominal spiracles located on the sternites, below the connecting membrane; the last one usually
	not covered by the elytra; mentum and ligula usually connate. 191
181.	Hind tibiæ with a single apical spur; scutellum usually invisible;
101.	middle legs widely separated by their coxæ; clypeus expanded so
	as to cover the mandibles and mouthparts; pygidium partly
	exposed; antennæ eight- or nine-jointed. (Onthophagus, cos-
	mop.; Scarabæus, palæarc., ethiop. (S. sàcer, Sacred scara-
	bæus); Gymnopleurus, Afr., As.; Cánthon, Tumble-bugs,
	Deltochilum, Am.; Chærídium, Am.; Pinòtus, neotrop.;
	Còpris, widespr.; Phanæus, Am.). (CÓPRIDÆ).
	SCARABÆIDÆ
*- 7	Hind tibiæ with two apical spurs; scutellum well developed;
	middle legs more approximated; clypeus of variable size, either
	concealing or exposing the mouthparts
182.	Six abdominal sternites
	Five abdominal sternites, or with the sutures between the ster-
	nites effaced
183.	Antennæ with eleven joints
	Antennæ with ten joints or less
	Antennal club with three leaves; mandibles and labrum promi-
	nent; moderate-sized beetles usually with strongly striate elytra.
	(Fig. 815). (Bolbóceras, cosmop.; Athýreus, neotrop., As.;
2- 10	Geotrupes, widespr.; Lethrus, holarc.) GEOTRUPIDÆ

	Antennal club with many (5–7) leaves; moderately large, black,
	hairy beetles. (Pleócoma, Ácoma, nearc.) . PLEOCÓMIDÆ
185.	Antennæ ten-jointed; body more or less hairy186
	Antennæ nine-jointed
186.	Antennal club bare and shining, sparsely hairy; epimera of meta-
	thorax large; conspicuously hairy beetles. (Amphicoma,
	holarc.; Toxócerus, Gláphyrus, palæarc.) . GLAPHÝRIDÆ
	Antennal club, at least on the apical two joints, pubescent and
	dull or opaque
187.	Epimera of metathorax covered; abdominal sternites free; an-
	tennal club simple, lamellate
4	Epimera of metathorax visible; abdominal sternites connate;
	antennal club telescopic, the joints cupuliform. (Hybósorus,
	holarc., ethiop.; Liparochrus, Austr.; Phæochrous, wide-
	spr.) HYBOSÓRIDÆ
188.	Eyes divided in front. (Hýbalus, palæarc.; Órphnus, ethiop.,
	Ind.; Ægídium, neotrop.) ÓRPHNIDÆ
	Eyes entire. (Ochodæus, widespr.) OCHODÆIDÆ
189.	Mandibles concealed beneath the clypeus. (Aphòdius, Atæ-
	nius, Psammòbius, Saprosítes, Oxyòmus, cosmop.; Eu-
	pària, widespr.; Lorditomæus, Afr.) APHODÌIDÆ
	Mandibles not covered by the clypeus. (Ægiàlia, holarc.; Ere-
	màzus, palæarc.) ÆGIALÌIDÆ
190.	Abdomen retractile, whole body together with the legs capable of
	being contracted into a ball; small shining, more or less metallic
	species. (Acanthócerus, Cleonòtus, Am.; Philharmóstes,
	Afr.; Pterorthochætes, Indo-Austr.) ACANTHOCÉRIDÆ
	Body not retractile. Tribe Oncerini of the <b>MELOLONTHID</b>
191.	Abdominal spiracles forming two rows which diverge only
	slightly behind, each row forming a nearly straight line; tarsal
	claws at least of hind legs usually of equal size and with a
	tooth, generally immovable; rarely with only one claw; epi-
	stoma transverse, separated from the front by a suture. (Figs.
	816, 847). A large, widely distributed group. (Melolontha
	(Cockchafers), Oncócerus, Sérica, Diplotáxis, Phylló-
	phaga (=Lachnostérna) (May-beetles, June-bugs), Poly-
	phýlla and many others) MELOLÓNTHIDÆ
٠.	Abdominal spiracles forming two rows which diverge strongly
100	behind, each row forming two lines
192.	Front legs greatly lengthened, especially in the male; sides of
	prothorax dentate: tarsal claws equal: distal leaves of antennal

	club inclosing the proximal ones. (Euchirus, Indomal.;
	Cheirótonus, Propomàcrus, As.) EUCHÍRIDÆ
	Of a different conformation
193.	Tarsal claws, at least of the hind legs, of equal length 194
	Tarsal claws unequal, freely movable, the shorter one not bifid;
	pygidium exposed; usually large, often brightly colored beetles,
	usually no strong dimorphism between the sexes. (Anómala,
	cosmop.; Popíllia, widespr.; Adorètus, Old World; Ani-
	sòplia, palæarc.; Cotálpa, Plusiòtis, Pelidnòta, nearc.).
	RUTÉLIDÆ
194.	Hind legs of male with the femora greatly swollen, the antennæ
	nine-jointed; female without elytra or wings. (Pachypus,
N.	palæarc.) PACHYPÓDIDÆ
	Femora normal; female with elytra195
195.	Mandibles visible from above, more or less widened and blade-
	like; anterior coxæ transverse; large or very large beetles;
	highly dimorphic, the males often with large horns on the head
	and prothorax. Rhinoceros beetles. (Dynástes). DYNÁSTIDÆ
	Mandibles not visible from above, not widened or expanded; an-
100	terior coxæ usually strongly conical
196.	Clypeus emarginate at the sides in front of the eyes, so that
	the base of the antenna is visible from above; antennæ ten-
	jointed
107	(Phænómeris, Oxychìris, Afr.)PHÆNOMÉRIDÆ Epimera of metathorax visible from above between the hind
197.	angle of the prothorax and the humeral callus; lateral edge of
	elytra bent inwards near the base. (Fig. 850). Moderate-sized,
	or often large, frequently brightly colored beetles. (Cetònia,
	palæarc.; Cremastochilus, nearc.; Euphòria, Allorhina,
	Cótinus, Am.; Goliathus (Goliath beetle), ethiop.; and many
	others)
	Epimera of metathorax not visible from above, elytra not sinuate;
	prothorax behind much narrower than the base of the elytra.
	(Tríchius, holarc.; Válgus, widespr.; Osmodérma, holarc.;
	Gnórimus, palæarc.) TRICHÌIDÆ
198.	
	legs larviform, bearing a single claw
	Head prominent, not retracted; legs of normal form with two
	tarsal claws. (See couplet 142).
	A fam famala BUIDIDUÓBID #

- - Body strongly flattened; prothorax triangularly narrowed in front, meso- and metathorax expanded laterally and much wider than the abdomen, the latter with processes extending from the sides of the segments. (**Dulitícola**, Borneo). "Trilobite larvæ." (See couplet 93) . . . . . . . . . . Some females of **DRÍLIDÆ**

# LITERATURE ON COLEOPTERA

#### GENERAL

- Alluaud, C. Catalogue of Coleoptera of Madagascar. In Grandidier, Hist. Madagascar, 21 (1900).
- BLATCHLEY, W. S. Coleoptera or Beetles of Indiana. Nature Publishing Co., Indianapolis (1910).
- Böving, A. G. Classification of beetles according to larval characters. Bull. Brooklyn Entom. Soc., 24, pp. 55–80 (1929).
- Bradley, J. C. Manual of the genera of beetles of America North of Mexico. Ithaca, Daw, Illston & Co., 360 pp. (1930).
- Broun, T. Manual of New Zealand Coleoptera. 1566 pp. Wellington (1880-93).
- Bruch, C. Catálogo de los Coleópteros de Argentina. Rev. Mus. la Plata, 19 (several parts) (1915).
- Calwer, C. G. Käferbuch. 6th edition, 1390 pp., Stuttgart (1916).
- Casey, T. L. Memoirs on the Coleoptera, 1-11 (1910-24). (Some parts are listed separately). (Deals with various groups of North American Coleoptera).
- Forbes, W. T. M. The wing-venation of Coleoptera. Ann. Entom. Soc. America, 15, pp. 328–352 (1922).
  - The wing-folding patterns of the Coleoptera. Journ. New York Entom. Soc., **34**, pp. 42–68; 91–138 (1926).
- FOWLER, W. W. British Coleoptera. 6 vols. London (1887-1913).
  - Coleoptera, general introduction and Cicindelidæ and Paussidæ. Fauna of British India (1912).
- Gahan, C. J. and others. Coleoptera of India. Fauna of British India, 4 vols. (1906–1912). (An extensive work, covering a part of the coleopterous fauna of this region).
- Ganglbauer, L. Die Käfer von Mitteleuropa. Vienna, 4 vols. (1892–1904). (Adephaga, Staphyliniformia and Clavicornia).
- Gemminger, M. and B. Harold. Catalogus Coleopterorum. 12 vols., 3986 pp. Monaco (1868–76).
- Hatch, M. H. Systematic index to keys for the determination of Nearctic Coleoptera. Journ. New York Entom. Soc., **35**, pp. 279–306. (Supplementary to the bibliography in Leng's catalogue).

- HAYWARD, R. Classification of the families of Coleoptera of North America. American Entom. Soc., 37 pp. (1909).
- Henshaw, S. Bibliography, in "Synopsis of North American Coleoptera." Appendix to Leconte and Horn, 1883, pp. 535–552 (1883).
- Horn, G. H. Coleoptera of Baja California. Proc. California Acad. Sci. (2),4, pp. 302-449 (1895); 5, pp. 225-295 (1896); 6, pp. 367-381 (1897).
- HOULEBERT, C. Encyclopédie Scientifique. Les Coléoptères d'Europe, France et Régions voisins. Paris (1922). (A continuing series).
- LACORDAIRE, J. T. Genera des Coléoptères. 13 vols., Paris (1854-76).
- LAMEERE, A. Classification des Coléoptères. Ann. Soc. Entom. Belgique, 44, pp. 355-377 (1900).
- Leconte, J. L. and G. H. Horn. Classification of the Coleoptera of North America. Smithsonian Misc. Coll., No. 507 (1883). (With an extensive list of literature on North American Coleoptera).
- Leng, C. W. Catalogue of the Coleoptera of America North of Mexico. Mount Vernon, N. Y. J. D. Sherman (1920). (With a complete bibliography of the North American Coleoptera).
- Leng, C. W. and A. J. Mutchler. Water Beetles (Dytiscidæ, Hydrophilidæ and Parnidæ) of Florida. Bull. American Mus. Nat. Hist., 38, pp. 73–116 (1918).
- Lucas, R. Catalogus generum et subgenerum Coleopterorum. (Catalogue of such genera and subgenera as have been treated in the Genera Insectorum and the Catalogus Coleopterorum, Junk). Pt. 1, 696 pp., Berlin (1920).
- REITTER, E. Fauna Germanica. Käfer. 5 vols., Stuttgart (1908-16).
- Reitter, E. and others. Bestimmungs-Tabellen der europäischen Coleopteren. (About one hundred parts of this series have appeared in various journals. We are unable to cite them here, but references are contained in the Zoological Record and the Tierwelt Mitteleuropas.)
- Scheerpeltz, O. and A. Winkler. Coleoptera. In Tierwelt Mitteleuropas, 5, Lief. 2, pp. XII 1–272 (1930).
- Schenkling, S. and W. Junk. Coleopterorum Catalogus. W. Junk, Berlin, (1910). (A complete synonymic catalogue of the Coleoptera of the World being published in parts by various authors. It is as yet incomplete and we have listed the parts separately).
- Schultze, W. A Catalogue of Philippine Coleoptera. Philippine Journ. Sci., **11**D, pp. 1–194 (1916).
- Sharp, D., G. C. Champion and others. Coleoptera. In Biologia centrali-Americana. 7 vols. (1887–1909).
- SHARP, D. and PERKINS, R. C. L. Coleoptera. In Fauna Hawaiiensis, 2, pt. 3 (1900).
- Winkler, A. Catalogus Coleopterorum regionis palæarcticæ, pts. 1–8. Vienna, Winkler and Wagner, 752 pp. (1924–28).
- Yakobson, G. G. Key to Beetles of Russia (in Russian). Moscow and Leningrad Govt. Pub., 522 pp. (1927).

## ADEPHAGA

- AHLWARTH, K. Gyrinidæ. Coleop. Cat. Junk, pt. 21 (1910).
- Andrewes, H. E. Carabinæ. Fauna of British India, Carabidæ, 1 (1929).
- ATKINSON, E. T. Catalogue of Oriental Carabidæ. Journ. Roy. Asiatic Soc., 58, pp. 1–126 (1889).
- BARBER, G. W. and W. O. Ellis. Cupedidæ of North America. Journ. New York Entom. Soc., 28, pp. 197–208 (1920).
- Bertrand, H. Les larves et nymphes des Dytiscides, Hygrobiides et Haliplides. Encycl. Entom., part 10, 366 pp. (1928).
- Csiki, E. Carabidæ, Carabinæ. Coleop. Cat. Junk, pts. 91 and 92 (1927). Carabidæ, Mormolycinæ. Coleop. Cat. Junk, pt. 97 (1928).
- Carabidæ, Harpalinæ. Coleop. Cat. Junk, pts. 97, 98, 104 (1928–29).
- Desneux, J. Paussidæ. Gen. Insectorum, fasc. 35, 34 pp. (1905).
- Dupius, P. Carabidæ, Metriinæ and Mystropominæ. Gen. Insectorum, fasc. 116, 4 pp. (1911).
  - Carabidæ, Apotominæ. Gen. Insectorum, fasc. 117, 4 pp. (1911).
  - Carabidæ, Psydrinæ. Gen. Insectorum, fasc. 123, 2 pp. (1911).
  - Carabidæ, Opisthiinæ. Gen. Insectorum, fasc. 126, 2 pp. (1912).
  - Carabidæ, Notiphilinæ. Gen. Insectorum, fasc. 134, 7 pp. (1912).
  - Carabidæ, Pentagonicinæ. Gen. Insectorum, fasc. 145, 4 pp. (1913).
  - Carabidæ, Peleciinæ. Gen. Insectorum, fasc. 146, 4 pp. (1913). Carabidæ, Hexagoniinæ. Gen. Insectorum, fasc. 147, 4 pp. (1913).
- Gestro, R. Rhysodidæ. Coleop. Cat. Junk, pt. 1 (1910); Cupedidæ, Paussidæ. Coleop. Cat. Junk, pt. 5 (1910).
- Grouvelle, A. Synopsis des Rhysodides. Rev. Entom., 22, pp. 85-145 (1903).
- HAYWARD, R. Revision of Bembidium of North America. Trans. American Entom. Soc., 24, pp. 32–143 (1897).
  - Revision of North American Species of Tachys. Trans. American Entom. Soc., 26, pp. 191–238 (1900).
- Heller, K. M. Fauna Sumatrensis. Jacobsoniidæ. Suppl. Entom., **15**, pp. 111–112 (1927); and ibid., **14**, p. 127 (1926).
- HEYNES-WOOD, M. and C. DOVER. Cicindelidæ. Cat. Indian Ins. Govt. India, Centr. Pub. Br., Calcutta, pt. 13 (1927).
- Horn, W. Cicindelidæ. Coleop. Cat. Junk, pt. 86 (1926).
- JEANNEL, R. Monographie der Bathysciinæ. Arch. Zool., 63, pp. 1–436 (1924).
  - Monographie des Trechinæ. Abeille, **33**, pp. 1–592 (1926).
- Kolbe, H. Die Paussiden Südamerikas. Entom. Mitteil., **9**, pp. 131–141; 145–156 (1920).
- LA PONGE, G. V. Carabidæ, Carabinæ. Gen. Insectorum, fasc. 192, 289 pp. (1929–30).
- Leng, C. W. Revision of Cicindelidæ of Boreal America. Trans. American Entom. Soc., 28, pp. 93–186 (1902).

- Leng, C. W. and W. Beutenmüller. Handbook of Cicindelidæ and Carabidæ of Northeastern America. Journ. New York Entom. Soc., **2**, **4** (1894–96). (Several parts).
- Matheson, R. The Haliplidæ of North America, North of Mexico. Journ. New York Entom. Soc., 20, pp. 156–193 (1912).
- Notman, H. Review of Pseudomorphidæ and rearrangement of the Adephaga. Proc. U. S. Nat. Mus., **67**, art. 14, 34 pp. (1925).
- Ochs, G. Gyrinidæ of the Philippine Islands. Philippine Journ. Sci., 24, pp. pp. 81–86 (1924).
- Peringuey, L. Revision of South African Carabidæ. Ann. South African Mus., 23, pp. 579-659 (1926).
- Peschet, R. Dytiscidæ et Gyrinidæ des îles mascareignes. Ann. Soc. Entom. France, **86**, pp. 1–56 (1917).
- REGIMBART, M. Monographie des Gyrinides. Ann. Soc. Entom. France (1882–83).
  - Gyrinidæ. Gen. Insectorum, fasc. 1, 12 pp. (1902).
- Roeschke, H. Monographie der Tribus Cychrini (Carabidæ). Ann. Mus. Hist. Nat. Hungarici, 5, pp. 9–277 (1907).
- ROUSSEAU, E. Carabidæ, Anthiinæ. Gen. Insectorum, fasc. 39, 19 pp. (1906).
  - Carabidæ, Mormolycinæ. Gen. Insectorum, fasc. 40, 5 pp. (1906).
  - Omophronidæ. Gen. Insectorum, fasc. 83, 5 pp. (1908).
  - Carabidæ, Promecognathinæ. Gen. Insectorum, fasc. 84, 4 pp. (1908).
    - Carabidæ, Pamborinæ. Gen. Insectorum, fasc. 85, 3 pp. (1908).
    - Carabidæ, Lorocerinæ. Gen. Insectorum, fasc. 86, 4 pp. (1908).
- Sharp, D. On aquatic Coleoptera or Dytiscidæ. Sci. Trans. Roy. Dublin Soc., (2), **2**, pp. 179–1003 (1882).
- SLOANE, T. G. The Classification of the Family Carabidæ. Trans. London Entom. Soc., 1923, pp. 234–250 (1923).
- Van Dorsselaer, R. Dytiscidæ and Haliplidæ of Belgium. Bull. Soc. Entom. Belgique, 1, pp. 68–119 (1919).
- ZIMMERMANN, A. Die Schwimmkäfer des Deutschen entomologischen Museums, Berlin. Arch. f. Naturg., Jahrg. 83, pp. 68–249 (1917).
  - Amphizoidæ, Dytiscidæ, Haliplidæ and Hygrobiidæ. Coleop. Cat. Junk, pt. 71 (1920).
  - Die Halipliden der Welt. Entom. Blätt., 20, pp. 1–16 et seq. (several parts) (1924).

#### HYDROPHILOIDEA

KNISCH, A. Die exotischen Hydrophiliden des Deutschen Entomologischen Museums. Arch. f. Naturg., Jahrg. 85A, Heft 8, pp. 55–88 (1923). Hydrophilidæ. Coleop. Cat. Junk, pt. 79 (1924).

- D'Orchymont, A. Classification des Palpicornia. Ann. Soc. Entom. France, 88, pp. 105–168 (1919) and Rev. Zool. Africaine, 6, pp. 163–168 (1919). Palpicornia. Cat. Indian Ins. Govt. India, Centr. Pub. Br., Calcutta, pt. 14 (1927).
- REGIMBART, M. Revision des Grands Hydrophiles. Ann. Soc. Entom. France, 1902, pp. 188–232 (1902).

### STAPHYLINOIDEA

- Bernhauer, M. and Schubert, K. Staphylinidæ. Cat. Coleop. Junk, pts. 19, 29, 40, 57, 67, 82 (1910–26).
- CAMERON, M. Indomalayan Staphylinidæ. (New Species of Staphylinidæ from Singapore with Catalogue). Trans. Entom. Soc. London, 1920, pp. 347–413 (1921).

Staphylinidæ. Cat. Indian Ins. Govt. India, Centr. Pub. Br., Calcutta, pt. 6 (1925).

CASEY, T. L. Synopsis of Scaphidiidæ. Ann. New York Acad. Sci., 7, pp. 281–606 (1893).

Synopsis of Scydmænidæ. Ann. New York Acad. Sci., 9, pp. 285–548 (1897).

Synopsis (partial) of North American Pselaphidæ. Ann. New York Acad. Sci., 9, pp. 550–630 (1897).

Revision of Pæderini (Staphylinidæ). Trans. Acad. Sci. St. Louis, 15, pp. 17–248 (1905).

Partial Revision of Aleocharinæ and Xantholinini (Staphylinidæ). Trans. Acad. Sci., St. Louis, **16**, pp. 125–434 (1906).

CSIKI, E. Scaphidiidæ. Coleop. Cat. Junk, pt. 13 (1910).

Platypsyllidæ. Coleop. Cat. Junk, pt. 18 (1910). Scydmænidæ. Coleop. Cat. Junk, pt. 70 (1919).

Desneux, J. Platypsyllidæ. Gen. Insectorum, fasc. 41, 9 pp. (1906).

Fenyes, A. Staphylinidæ, Aleocharinæ. Gen. Insectorum, fasc. 173, 453 pp. (1918–1921).

HATCH, M. H. Studies on the Silphinæ. Journ. New York Entom. Soc., 35, pp. 331–370 (1927).

Liodidæ, Clambidæ. Coleop. Cat. Junk, pt. 105 (1929).

Hetschko, A. Thorictidæ. Coleop. Cat. Junk, pt. 83 (1926).

Horn, G. H. Synopsis of the Silphidæ of the United States. Trans. American Entom. Soc., 8, pp. 219–322 (1880).

Synopsis of the Dascyllidæ of the United States. Trans. American Entom. Soc., 8, pp. 73–114 (1880).

Jeannel, R. and M. H. Hatch. Silphidæ. Coleop. Cat. Junk, pts. 60 and 95 (1914).

JOHANSEN, J. P. Danish Staphylinidæ (in Danish). 600 pp. Copenhagen (1914).

- Olsuffer, G. Synopsis of Leptinidæ. Rev. Russe Entom., 18, pp. 81-90 (1923).
- Portevin, G. Revision des Necrophorini du Globe. Bull. Mus. Hist. Nat. Paris (1923–25). (Various parts).

Les grands Nécrophages du Globe. Encycl. Entom., pt. 6, 270 pp. (1926).

RAFFRAY, A. Australian Pselaphidæ. Proc. Linn. Soc. New South Wales, 25, pp. 131–249 (1900).

Pselaphidæ. Gen. Insectorum, fasc. 64, 487 pp. (1908).

Pselaphidæ. Coleop. Cat. Junk, pt. 27 (1911).

## CUCUJOIDEA

- Bövine, A. G. Remarks on taxonomy of Cucujidæ. Zoologica, New York, 3, pp. 197–221 (1921).
- Casey, T. L. Revision of Cucujidæ of North America. Trans. American Entom. Soc., **11**, pp. 69–112 (1884).

Classification of Cucujidæ. Mem. Coleop., pt. 7, pp. 111 et seq. (1915). RITSEMA, C. Helotidæ. Coleop. Cat. Junk, pt. 34 (1911).

## CANTHAROIDEA

Champion, G. C. Revision of Mexican and Central American Malachiidæ. Trans. London Entom. Soc., 1914, pp. 13–127 (1914).

Revision of Mexican and Central American Chauliognathinæ. Trans. London Entom. Soc., 1914, pp. 126–168 (1914).

Revision of the Mexican and Central American Telephoridæ. Trans. London Entom. Soc., 1915, pp. 16–146 (1915).

Chapin, E. A. Classification of the Philippine components of the Coleopterous Family Cleridæ. Philippine Journ. Sci., 24, pp. 159–286 (1924).

KLEINE, R. Die Lyciden der Philippinen-Inseln. Philippine Journ. Sci., 31, pp. 33–109 (1926).

Coleoptera, Lycidæ. Nova Guinea, Leiden, 15, pp. 91–195 (1926).

Leconte, J. L. Synopsis of Lampyridæ of the United States. Trans. American Entom. Soc., 9, pp. 15–72 (1881).

LENG, C. W. and A. J. MUTCHLER. The Lycidæ, Lampyridæ and Cantharidæ of the West Indies. Bull. American Mus., 46, pp. 413–499 (1922).

OLIVIER, E. Lampyridæ. Gen. Insectorum, fasc. 53, 74 pp. (1907).

Lampyridæ and Drilidæ. Coleop. Cat. Junk, pts. 9 and 10 (1910).

Pic, M. Étude des Malacodermes de l'Indochine. Fauna Indochine Français, fasc. 6, pp. 7–63 (1923).

Lampyridæ, Phengodinæ. Coleop. Cat. Junk, pt. 94 (1927).

Drilidæ (Karumiidæ). Coleop. Cat. Junk, pt. 94 (1927).

Dasytidæ, Melyrinæ. Coleop. Cat. Junk, pt. 103 (1929).

Schenkling, S. Cleridæ. Gen. Insectorum, fasc. 13, 124 pp. (1903).

Cleridæ. Coleop. Cat. Junk, pt. 23 (1910).

Lymexylidæ, Telegeusidæ and Micromalthidæ. Cat. Coleop. Junk, pt. 64 (1915).

## MORDELLOIDEA

Blair, K. G. A Revision of the Pyrochroidæ. Ann. Mag. Nat. Hist., 13, pp. 310–326 (1914).

South African Œdemeridæ. Ann. South African Mus., 23, pp. 353-375 (1926).

Pythidæ and Pyrochroidæ. Coleop. Cat. Junk, pt. 99 (1928).

Borchmann, F. Eurystethidæ. Coleop. Cat. Junk, pt. 2 (1910).

Meloidæ and Cephaloidæ. Coleop. Cat. Junk, pt. 69 (1917).

CASEY, T. Synopsis of Anthicidæ of United States. Ann. New York Acad. Sci., 8, pp. 624–838 (1895).

North American Cephaloidæ. Entom. News, 9, pp. 193-195 (1898).

CSIKI, E. Rhipiphoridæ. Cat. Coleop. Junk, pt. 54 (1913).

Mordellidæ. Coleop. Cat. Junk, pt. 63 (1915).

Horn, G. H. Œdemeridæ of Boreal America. Proc. California Acad. Sci., (2), 6, pp. 382–421 (1896).

MUTCHLER, A. J. and H. B. Weiss. Meloidæ of New Jersey. Circ. New Jersey State Dept. Agric., No. 76, 19 pp. (1924).

Pic, M. Euglenidæ. Gen. Insectorum, fasc. 8, 14 pp. (1902).

Euglenidæ. Coleop. Cat. Junk, pt. 14 (1910).

Pedilidæ. Coleop. Cat. Junk, pt. 26 (1911).

Anthicidæ. Coleop. Cat. Junk, pt. 36 (1911).

SCHENKLING, S. Œdemeridæ, Coleop. Cat. Junk, pt. 65 (1915).

Smith, J. B. Synopsis of Mordellidæ of the United States. Trans. American Entom. Soc., **10**, pp. 73–100 (1882).

Van Dyke, E. C. Genera of North American Meloidæ. Univ. California Publ. Entom., 4, pp. 395–474 (1928).

### ELATEROIDEA

Blanchard, F. Revision of North American Throscidæ. Trans. American Entom. Soc., 43, pp. 1–26 (1917).

Bonvouloir, H. Monographie des Eucnémides. Ann. Soc. Entom. France, (4), 10, supplement, 907 pp. (1871–75).

Buysson, H. Du. Tableaux des Elateridæ de la faune francorhénane. Misc. Ent. Castanet-Tolosan, pp. 1–208 (1910–26).

Candèze, E. de. Monographie des Élatérides. 4 vols. and 3 suppl. Liège (1857–81).

Chamberlin, W. J. Catalogue of the Buprestidæ of North America, North of Mexico. Corvallis, Oreg. 289 pp. (1926).

Dalla Torre, K. W. Cebrionidæ. Coleop. Cat. Junk, pt. 25 (1911). Cebrionidæ. Gen. Insectorum, fasc. 127, 17 pp. (1912).

Fisher, W. S. Revision of West Indian Buprestidæ. Proc. U. S. Nat. Mus., 65, art. 9, pp. 1–207 (1925).

FLEUTHIAUX, E. Études sur les Mélasidæ. Ann. Soc. Entom. Belgique, **60**, pp. 93–104 (1920); **61**, p. 23, etc. (1921).

Catalogue raisonné des Mélasidæ des Îles Philippines. Ann. Soc. Entom. France, **95**, pp. 29–90 (1926).

Horn, G. H. Synopsis of Throscidæ of the United States. Trans. American Entom. Soc., 12, pp. 198–208 (1885).

Monograph of Eucneminæ and Cerophytinæ of the United States. Trans. American Entom. Soc., 13, pp. 5–38 (1886).

Kerremans, C. Buprestidæ. Gen. Insectorum, fasc. 12, 338 pp. (1903). Monographie des Buprestidæ. 7 vols. Brussels (1906–14).

Knull, J. N. The Buprestidæ of Pennsylvania. Ohio Univ. Studies, 2, No. 2, 71 pp. (1927).

OBENBERGER, J. Buprestidæ. Coleop. Cat. Junk, pt. 84 (1926).

Revision of Pachyseloides Trachides (Buprestidæ) of America. Sbornik Entom. Odd. Narod. Praze, **13** (20), pp. 3–149 (1925).

Pic, M. Rhipiceratidæ. Coleop. Cat. Junk, pt. 81 (1925).

SCHENKLING, S. Elateridæ. Coleop. Cat. Junk, pts. 80 and 88 (1925–27).

Plastoceridæ. Coleop. Cat. Junk, pt. 93 (1927).

Melasidæ. Coleop. Cat. Junk, pt. 96 (1928).

Cerophytidæ. Coleop. Cat. Junk, pt. 101 (1928).

Schwarz, O. Elateridæ. Gen. Insectorum, fasc. 46, 370 pp. (1906).

Plastoceridæ. Gen. Insectorum, fasc. 50, 10 pp. (1907).

Elateridæ, pt. Gen. Insectorum, fasc. 51, 4 pp. (1907).

#### DRYOPOIDEA

Barthe, E. Dryopidæ. Misc. Entom., 30, pp. 211-274 (1928).

Carter, H. J. and E. H. Zeck. A Monograph of the Australian Dryopidæ. Australian Zool., **6**, pp. 50–72 (1929).

HORN, G. H. Synopsis of Heteroceridæ of the United States. Trans. American Entom. Soc., 17, pp. 1–17 (1890).

Zaitzev, P. Dryopidæ, Georyssidæ and Heteroceridæ. Coleop. Cat. Junk, pt. 17 (1910).

### DASCYLLOIDEA

BORCHMANN, F. Alleculidæ. Coleop. Cat. Junk, pt. 3 (1910).

CARTER, H. J. Revision of the Australian Cistelidæ. Proc. Roy. Soc. Victoria, 28, pp. 52–104 (1915).

CASEY, T. L. Synopsis of North American Cistelidæ. Ann. New York Acad. Sci., 6, pp. 9–214 (1892). Descriptive catalogue of American Byrrhidæ. Memoirs on the Coleoptera, No. 3, pp. 1–69 (1912).

Dalla Torre, K. W. Byrrhidæ, Dermestidæ and Nosodendridæ. Coleop. Cat. Junk, pt. 33 (1911).

FALL, H. C. Revision of Ptinidæ of Boreal America. Trans. American Entom. Soc., 31, pp. 97–296 (1905).

Grouvelle, A. Byturidæ. Coleop. Cat. Junk, pt. 56 (1913).

JAYNE, H. F. Revision of Dermestidæ of the United States. Proc. American Philos. Soc., 20, pp. 343-377 (1882).

MUTCHLER, A. J. and H. B. Weiss. New Jersey Dermestidæ. Circ. New Jersey Dept. Agric., No. 108, 31 pp. (1927).

Pic, M. Ptinidæ. Coleop. Cat. Junk, pt. 41 (1912).

Dascillidæ and Helodidæ. Coleop. Cat. Junk, pt. 58 (1914).

Westwood, J. O. Description of a new genus of Coleopterous insects. (Gnostidæ). Trans. Entom. Soc. London (n.s.) 3, pp. 90–94 (1854).

#### HISTEROIDEA

BICKHARDT, H. Histeridæ. Coleop. Cat. Junk, pt. 24 (1910).

Histeridæ, part. Gen. Insectorum, fasc. 166a, 112 pp. (1916).

Нетschко, A. Synteliidæ. Coleop. Cat. Junk, pt. 83, (1926).

Horn, G. H. Synopsis of Histeridæ of the United States. Proc. American Philos. Soc., 13, pp. 237–360 (1873).

Portevin, G. Les Liodides de l'Inde. Encycl. Entom., Ser. B, Coleoptera, 1, pp. 75-83 (1926).

### COLYDIOIDEA

Arrow, G. J. Contribution to classification of the Coleopterous family Endomychidæ. Trans. Entom. Soc. London, 1920, pp. 1–83 (1920).

Belon, R. P. Classification des Lathridiidæ. Rev. Entom. 1897, pp. 105–221 (1897).

Récapitulation des Lathridiidæ de l'Amérique méridionale. Ann. Soc. Lyon, **46**, pp. 137–192 (1900).

Lathridiidæ. Gen. Insectorum, fasc. 3, pp. 40 (1902).

Brethes, J. Coccinellides du British Museum, avec une nouvelle famille (Cerasommatidiidæ). An. Mus. Hist. Nat. Buenos Aires, **33**, pp. 195–214 (1925).

CASEY, T. L. Synopsis of Phalacridæ. Ann. New York Acad. Sci., 5, pp. 39–185 (1891).

Revision of American Coccinellidæ. Journ. New York Entom. Soc., 7, pp. 71–163 (1899).

Review of American Corylophidæ, Cryptophagidæ, Tritomidæ and Dermestidæ. Journ. New York Entom. Soc., 8, pp. 51–172 (1900).

Chatterjee, S. N. Nitidulidæ. Cat. Indian Ins. Govt. India, Centr. Pub. Br., Calcutta, pt. 5 (1924).

CSIKI, E. Endomychidæ. Coleop. Cat. Junk, pt. 12 (1910).

Discolomidæ, Orthoperidæ, Phænocephalidæ and Sphæriidæ. Coleop. Cat. Junk, pt. 18 (1910).

Hydroscaphidæ and Ptiliidæ. Coleop. Cat. Junk, pt. 32 (1911).

Dalla Torre, K. W. Ciidæ. Coleop. Cat. Junk, pt. 30 (1911).

Fall, H. C. Revision of the Lathridiidæ of Boreal America. Trans. American Entom. Soc., 25, pp. 101–190 (1899).

FOWLER, W. W. Languriidæ. Gen. Insectorum, fasc. 78, 45 pp. (1908).

Grouvelle, A. Nitidulidæ. Coleop. Cat. Junk, pt. 56 (1913).

Gui, H. L. Coccinellidæ of Kansas. Proc. Kansas Entom. Soc., 1, pp. 26–37 (1928).

Heller, K. M. Erotyliden der indo-australischen Region. Arch. Naturg., Jahrg. 84A, Heft 8, pp. 1–121 (1920).

Hetschko, A. Catapochrotidæ and Monœdidæ. Coleop. Cat. Junk, pt. 83 (1926).

Lathridiidæ. Coleop. Cat. Junk, pt. 85 (1926).

Horn, G. H. Revision of Bostrychidæ and Colydiidæ of the United States. Proc. American Philos. Soc., 17, pp. 555-592 (1878).

Synopsis of the Monotomidæ of the United States. Trans. American Entom. Soc., 7, pp. 257–267 (1879).

Revision of Nitidulidæ of the United States. Trans. American Entom. Soc., 7, pp. 267–336 (1879).

Kraus, E. J. and A. D. Hopkins. Revision of Holarctic Lyctidæ. Bull. U. S. Dept. Agric., Bur. Entom., Tech. Ser. No. 20, pp. 111–138 (1911).

Kuhnt, P. Erotylidæ. Gen. Insectorum, fasc. 88, 139 pp. (1909). Erotylidæ. Coleop. Cat. Junk, pt. 34 (1911).

Lesne, P. Revision des Bostrychides. Ann. Soc. Entom. France, 1896, pp. 95–127 and several later parts (1896–1906).

Les Bostrychides de l'Afrique tropicale française. Encycl. Entom. pt. 3, 301 pp. (1924).

LEVEILLE, A. Temnochilidæ. Coleop. Cat. Junk, pt. 11 (1910).

MATTHEWS, A. Monograph of Trichopterygidæ, 188 pp. London (1872).

Synopsis of North American Trichopterygidæ. Trans. American Entom. Soc., **11**, pp. 113–156 (1884).

Monograph of Corylophidæ and Sphæriidæ. London (1899).

Mequigon, A. Rhizophagidæ. Coleop. Cat. Junk, pt. 61 (1914).

Prc, M. Anobiidæ. Coleop. Cat. Junk, pt. 48 (1912).

Schenkling, S. Derodontidæ. Coleop. Cat. Junk, pt. 64 (1915).

Cryptophagidæ. Coleop. Cat. Junk, pt. 76 (1923).

Languriidæ. Coleop. Cat. Junk, pt. 100 (1928).

Throscidæ. Coleop. Cat. Junk, pt. 10 (1928).

Zaitzev, P. Cyathoceridæ. Coleop. Cat. Junk, pt. 17 (1910).

## TENEBRIONOIDEA

- BLAISDELL, F. E. Revision of the Tribe Eleodini of U. S., Lower California and Adjacent Islands. Bull. 63, U. S. Nat. Mus., 524 pp. (1909).
- Borchmann, F. Elactidæ, Lagriidæ, Nilionidæ, and Petriidæ. Coleop. Cat. Junk, pt. 2 (1910).

Die Lagriinæ. Arch. Naturg., Jahrg. 81A, Heft 6, pp. 46–186 (1916). Die Amerikanischen Gattungen und Arten der Statirinæ (Lagriidæ). Arch. Naturg., Jahrg. 87A, Heft 1, pp. 216–357 (1921).

- CARTER, H. J. Catalogue of Australian Tenebrionidæ. Australian Zoölogist, 4, pp. 117–163, 280; 294 (1926).
- CASEY, T. L. Revision of American Tentyrinæ (Tenebrionidæ). Proc. Washington Acad. Sci., 9, pp. 275–522 (1907).

Revision of American Coniontinæ (Tenebrionidæ). Proc. Washington Acad. Sci., **10**, pp. 51–166 (1908).

- Chatanay, J. Tenebrionidæ, Zophosinæ. Gen. Insectorum, fasc. 176, 50 pp. (1921).
- Csiki, E. Melandryidæ. Coleop. Cat. Junk, pt. 77 (1924).
- Gebien, H. Tenebrionidæ and Trictenotomidæ. Coleop. Cat. Junk, pts. 15, 22, 28, 37 (1910–11).

Monographie der südamerikanischen Camarien (Tenebrionidæ). Arch. f. Naturg. Jahrg. 83A, pt. 3, pp. 25–167 (1919).

Tenebrionidæ of Southwest Africa. Abh. Hamburg Univ., 5, pp. 1–168 (1920).

Ueber einige Gruppen amerikanischen Tenebrioniden. Stettiner Entom. Zeitg., 89, pp. 169–170 (1928).

- Hetschko, A. Cossyphodidæ. Coleop. Cat. Junk, pt. 83 (1926).
- Horn, G. H. Revision of the Tenebrionidæ of North America. Trans. American Philos. Soc., 14, pp. 253–404 (1870).

Synopsis of North American Lagriidæ. Trans. American Entom. Soc., 15, pp. 26–48 (1888).

- Pic, M. Scraptiidæ. Coleop. Cat. Junk, pt. 26 (1911).
- Pouillade, J. Revision of Trictenotomidæ. Insecta, Rennes, 4, pp. 243–251 (1914).
- REITTER, E. Subfamilies and tribes of palæarctic Tenebrionidæ. Wiener Entom. Zeitg., **36**, pp. 53-66 (1917).
- Semenov, A. Synopsis of Platyopinæ (Tenebrionidæ). Horæ Soc. Entom. Rossicæ, **38**, pp. 175–184 (1907).

## CERAMBYCOIDEA

Achard, J. Lamprosomatidæ. Gen. Insectorum, fasc. 159, 13 pp. (1914).

Chrysomelidæ, Chlamydinæ and Sphærocarinæ. Gen. Insectorum, fasc. 160, 25 pp. (1914).

Aurivillius, C. and A. Lameere. Cerambycidæ (s. lat). Coleop. Cat. Junk, pts. 39, 73, 74 (1912–1923).

Boppe, P. Cerambycidæ s. lat., part; Disteniinæ and Lepturiinæ. Gen. Insectorum, fasc. 178, 121 pp. (1921).

CLAVAREAU, H. Chrysomelidæ s. lat. Coleop. Cat. Junk, pt. 59 (1914).

Craighead, F. C. Larvæ of the Prioninæ. Rept. No. 107, Office of Secretary, U. S. Dept. Agric., 24 pp. (1915).

Classification of Larvæ of North American Cerambycidæ. Bull. Canada Dept. Agric. Entom., No. 23, 238 pp. (1923).

HANSEN, V. Chrysomelidæ and Lariidæ. Danmarks Fauna, 31, pp. 1–279, Copenhagen (1927).

Horn, G. H. Revision of the Bruchidæ of the United States. Trans. American Entom. Soc., 4, pp. 311–342 (1873).

Synopsis of the Halticinæ of North America. Trans. American Entom. Soc., **16**, pp. 163–320 (1889).

The Eumolpinæ of Boreal America. Trans. American Entom. Soc., 19, pp. 195–234 (1892).

The Galerucinæ of Boreal America. Trans. American Entom. Soc., 20, pp. 57–136 (1893).

Jacoby, M. Sagridæ. Gen. Insectorum, fasc. 14, 11 pp. (1903).

Chrysomelidæ. Fauna of British India, Coleoptera, 534 pp. (1908).

JACOBY, M. and H. CLAVAREAU. Donaciidæ. Gen. Insectorum, fasc. 21 (1904). Crioceridæ. Gen. Insectorum, fasc. 23, 40 pp. (1904).

Megascelidæ. Gen. Insectorum, fasc. 32, 6 pp. (1905).

Megalopodidæ. Gen. Insectorum, fasc. 33, 20 pp. (1905).

Clytridæ. Gen. Insectorum, fasc. 49, 88 pp. (1907).

LAMEERE, A. Prionidæ. Coleop. Cat. Junk, pt. 52 (1913). Gen. Insectorum, fasc. 172, 189 pp. (1919).

Leng, C. W. Synopsis of Nearctic Cerambycidæ. Bull. Brooklyn Entom.
Soc., 7, and Entomologica Americana, 1-6 (1884–1900) (various parts).
Revision of Donaciidæ of Boreal America. Trans. American Entom.
Soc., 18, pp. 159–176 (1891).

Leng, C. W. and J. Hamilton. The Lamiidæ of North America. Trans. American Entom. Soc., 23, pp. 101–178 (1896).

Maulik, S. Hispidæ and Cassididæ. Fauna of British India, Coleoptera, Chrysomelidæ (1919).

Chrysomelidæ and Halticidæ. Fauna of British India, 442 pp. (1926).

Prc, M. Bruchidæ. Coleop. Cat. Junk, pt. 55 (1913).

Planet, L. M. Histoire naturelle des Longicornes de France. Encycl. Entom. Sér. A, 2. 386 pp. (1924), also suppl. 53 pp. (1927).

Schaeffer, C. Revision of the New World Donaciini. Brooklyn Mus. Sci. Bull., 3, 165 pp. (1925).

Spaeth, F. Cassididæ. Coleop. Cat. Junk, pt. 62 (1914).

Fauna sumatrensis, Cassidinæ. Supplementa Entom., **13**, pp. 1–108 (1926).

- SWAINE, J. S. and R. HOPPING. Lepturini of North America. Pt. I. Bull. Nat. Mus. Canada, No. 52, 79 pp. (1928).
- Webb, J. L. A Preliminary Synopsis of Cerambycoid Larvæ. Tech. Ser., 20, pt. 5, Bur. Entom. U. S. Dept. Agric. (1912).
- Weise, J. Hispidæ. Coleop. Cat. Junk, pt. 15 (1911).

Chrysomelidæ. Coleop. Cat. Junk, pt. 68 (1916).

Galerucidæ. Coleop. Cat. Junk, pt. 78 (1924).

Hispidæ. Gen. Insectorum, fasc. 125, 124 pp. (1911).

#### CURCULIONOIDEA AND SCOLYTOIDEA

- BLACKMAN, M. W. North American Scolytidæ, Subfamily Micracinæ. Bull. Mississippi Agric. Expt. Sta., Tech. Ser. No. 10 (1921).
  - Revision of Pityophthori of North America. Bull. New York Coll. Forestry, Syracuse, Tech. Pub. No. 25 (1928).
- BLATCHLEY, W. S. and C. W. LENG. The Rhyncophora or Weevils of North-eastern America. Nature Pub. Co. (1916).
- Bovie, A. Curculionidæ, Entiminæ. Gen. Insectorum, fasc. 69, 7 pp. (1908). Curculionidæ, Cryptoderminæ. Gen. Insectorum, fasc. 70, 3 pp. (1908). Curculionidæ, Alcidinæ. Gen. Insectorum, fasc. 71, 11 pp. (1908).

Curculionidæ, Læmosaccinæ. Gen. Insectorum, fasc. 89, 6 pp. (1909).

Curculionidæ, Gymnetrinæ. Gen. Insectorum, fasc. 92, 19 pp. (1909).

Curculionidæ, Nanophyinæ. Gen. Insectorum, fasc. 98, 11 pp. (1910).

Curculionidæ, Brachycerinæ. Gen. Insectorum, fasc. 99, 38 pp. (1910).

- Chittenden, F. H. Classification of Nut Curculios of North America. Entom. Americana, 7, pp. 129–207 (1926).
- Dalla Torre, K. W. Aglycyderidæ and Proterhinidæ. Coleop. Cat. Junk, pt. 31 (1911).
- Dietz, W. G. Revision of North American Anthonomini. Trans. American Entom. Soc., 18, pp. 177–276 (1891).

Revision of Ceutorhynchini of North America. Trans. American Entom. Soc.. 23. pp. 387–480 (1896).

- Fall, H. C. Revision of North American Species of Apion. Trans. American Entom. Soc., **25**, pp. 105–184 (1898).
- HAGEDORN, M. Ipidæ. Gen. Insectorum, fasc. 111, 178 pp. (1910). Coleop. Cat. Junk, pt. 4 (1910).
- Hopkins, A. D. Monograph of Pissodes. Bull. U. S. Dept. Agric. Tech. Ser., No. 20, 68 pp. (1911).
  - Monograph of Scolytidæ. Bull. U. S. Dept. Agric., Tech. Ser. No. 17, pt. 1 (1909); pt. 2 (1915).
    - Classification of Cryphalinæ. Rept. 99, U. S. Dept. Agric. (1915).
- Hustache, A. Curculionides de Madagascar. Bull. Acad. Malgache, Tananarive, 7, pp. 1–582 (1924).
- JORDAN, K. Les Anthribides de l'Indochine. Faune Entom. Indochine française, fasc. 6, pp. 71–113 (1923).

KLEINE, R. Die Brenthiden der Nederländischen Ost-Indischen Kolonien. Capita Zool., 2, pt. 4, 86 pp. (1926).

Brenthidæ. Cat. Indian Ins. Govt. India, Centr. Pub. Br., Calcutta, pt. 11 (1926).

Die Brenthiden. Nova Guinea, Leiden, 15, pp. 214–274 (1926).

Brenthidæ. Coleop. Cat. Junk, pt. 89 (1927).

Bestimmungstabelle der Brenthidæ. Ent. Zeit. Frankfort-am-M., **40-42** (1927–28). (In several parts).

Lea, A. M. and A. Bovie. Curculionidæ, Belinæ. Gen. Insectorum, fasc. 91, 13 pp. (1909).

MARSHALL, G. A. K. Curculionidæ. Fauna of British India, Coleoptera (1916).

MURAYAMA, J. On the Platypodidæ of Formosa. Journ. Coll. Agric., Hokkaido Imp. Univ., 15, pp. 197–236 (1925).

Pape, P. Curculionidæ, Brachycerinæ. Coleop. Cat. Junk, pt. 16 (1910).

Petri, K. Monograph of Palæarctic Hyperini. 208 pp., Berlin (1901).

Pierce, W. D. Weevils of the Superfamily Curculionoidea. Proc. Entom. Soc. Washington, 21, pp. 21–36 (1919).

Schenkling, S. and G. A. K. Marshall. Curculionidæ, part. Coleop. Cat. Junk, pt. 106 (1929).

Schönfeldt, H. Brenthidæ. Gen. Insectorum, fasc. 65, 88 pp. (1908); Coleop. Cat. Junk, pt. 7 (1910).

Smith, J. B. Synopsis of Apioninæ of North America. Trans. American Entom. Soc., 11, pp. 41–68 (1884).

STROHMEYER, H. Platypodidæ. Coleop. Cat. Junk, pt. 44 (1912).

Chapuisiidæ. Gen. Insectorum, fasc. 162, 6 pp. (1914).

Platypodidæ. Gen. Insectorum, fasc. 163, 55 pp. (1914).

Swaine, J. W. Catalogue of North American Scolytidæ. Rept. New York State Entom., No. 24, pp. 76–159 (1909).

Classification of Canadian Bark-beetles. Bull. Canada Dept. Agric. Entom., No. 14, pp. 7–143 (1918).

Voss, E. Die Unterfamilien Attelabinæ und Apoderinæ. Stettiner Entom. Zeitg., 85, pp. 1-78; 191-304 (1925).

Wagner, H. Curculionidæ, Apioninæ. Gen. Insectorum, fasc. 130, 109 pp. (1912).

Curculionidæ, Apioninæ. Coleop. Cat. Junk, pt. 6 (1910).

Wolfrum, P. Anthribidæ. Coleop. Cat. Junk, pt. 102 (1929).

#### SCARABÆOIDEA

Arrow, G. J. Fauna of British India. Lamellicornia (Cetoniidæ, Dynastidæ, Rutelinæ, in Fauna British India, 1 (1910–17).

Acanthoceridæ, Glaphyridæ, Hybosoridæ, Ochodæidæ, Orphnidæ, Pleocomidæ and Trogidæ. Coleop. Cat. Junk, pt. 43 (1912).

Notes on the Coleopterous Families Hybosoridæ and Trogidæ. Ann. Mag. Nat. Hist. (9) **15**, pp. 328–331 (1925).

Boucomout, A. Geotrupidæ. Gen. Insectorum, fasc. 7, 20 pp. (1902). Geotrupidæ. Coleop. Cat. Junk, pt. 46 (1912).

Brown, W. J. Revision of North American Ægialiidæ. Canadian Entom., **63**, pp. 9–19 (1931).

Dalla Torre, K. W. Melolonthidæ. Coleop. Cat. Junk, pts. 45, 47, 49, 50 (1912–13).

Dawson, R. W. Scarabæidæ of Nebraska. Univ. of Nebraska Studies, 22, pp. 1–82 (1922).

Fall, H. C. Synopsis of Ochodæidæ of the United States. Journ. New York Entom. Soc., 17, pp. 30–38 (1909).

Fuchs, C. Synopsis of Lucanidæ of United States. Bull. Brooklyn Entom. Soc., 5, pp. 49-52; 57-60 (1882).

GILLET, J. J. E. Scarabæidæ, s. str. Coleop. Cat. Junk, pt. 38 (1911).

Gravely, F. H. Passalidæ. Mem. Indian Mus., 7, pp. 1-144 (1918).

Horn, G. H. Monograph of Aphodiinæ of the United States. Trans. American Entom. Soc., 14, pp. 1–110 (1887).

HOULEBERT, C. and E. MONNOT. Faune entomologique amoricaine, Coléoptères. Lamellicornes. Trav. Sci. Univ. Rennes, 13, pp. 1–171 (1915).

Lea, A. M. Australian Scarabæidæ, s. str. Rec. South Australian Mus., 2, pp. 353–396 (1923).

MOREIRA, C. Insectos Coleópteros Passalideos do Brazil. Fauna brasiliense, N. S. No. 1, 52 pp. (1925).

Ohaus, J. Euchiridæ, Rutelidæ and Phænomeridæ. Coleop. Cat. Junk, pt. 66 (1918).

Olsonfieff, G. Les Phanæides. Insecta, 13, pp. 5-202 (1924).

Pic, M. Eucinetidæ. Coleop. Cat. Junk, pt. 58 (1914).

Roon, G. van. Lucanidæ. Coleop. Cat. Junk, pt. 8 (1910).

Schenkling, A. Cetoniidæ. Coleop. Cat. Junk, pt. 72 (1921). Trichiidæ. Coleop. Cat. Junk, pt. 75 (1922).

SCHMIDT, A. Aphodiidæ. Gen. Insectorum, fasc. 110, 155 pp. (1910).

Aphodiidæ. Coleop. Cat. Junk, pt. 20 (1910).

Ægialiidæ. Coleop. Cat. Junk, pt. 42 (1912).

Scarabæidæ (s. lat. part) Ægialiinæ, Chironinæ, Dynamopinæ, Hybosorinæ, Idiostominæ, Ochodæinæ, Orphninæ. Gen. Insectorum, fasc. 150, 87 pp. (1913).

Aphodiidæ. Das Tierreich. Lief. 45, 333 pp. (1922).

# ORDER STREPSÍPTERA

(RHIPÍPTERA)

Small species parasitic on other insects, the adult males winged and free-living, but the larviform females never leaving the body of their host. Male with the head free. Eyes well developed, sphæroidal, provided with large, highly convex; separated facets. Antennæ with three

to seven joints, one or several of the joints prolonged into a long, lateral process (flabellum). Mouthparts reduced; mandibles often soft or minute; maxillæ fleshy; labium not developed. Prothorax greatly reduced in size, usually ring-like; mesothorax small, strongly transverse; metathorax very large. Fore wings reduced to small clubshaped appendages; hind wings very large and delicate, with a few, fine radiating veins, but without crossveins. Legs rather weak; the coxæ, especially of the front and middle legs very minute; tarsi with from five to two joints; often with the claws absent. Female with the mouthparts and antennæ vestigial; legs and wings absent; head and thorax fused into one strongly flattened piece; sexual openings in the form of segmental, usually unpaired canals opening on several of the abdominal segments. Metamorphosis complete; larvæ undergoing hypermetamorphosis with an active long-legged first stage larva or triungulin; female remaining larviform in the reproductive stage.

1.	Tarsi of male five-jointed, with two claws; pro- and mesothorax
	short, transverse; female unknown. (Superfamily MENGE-
	ÒIDEA) 2
v .	Tarsi of male with four joints or less and without claws3
2.	Antennæ seven-jointed, the third and fourth joints prolonged
	laterally; metathoracic præscutum transverse, reaching the
	humeri and lying entirely in front of the other dorsal plates of
	the metathorax; scutellum broadly rounded in front, longer
	than the præscutum. (Fig. 886). (Triozócera, nearc., neotrop.;
	Méngea, oligocene (fossil)) MENGÈIDÆ
	Antennæ six-jointed, third, fourth and fifth joints prolonged lat-
	erally; sixth elongate; metathoracic præscutum transverse-
	quadrate, not reaching the humeri, depressed and forming a sort
	of neck; lateral lobes of scutum reaching the metathorax;
	scutellum very long, narrowed and rounded in front. (Men-
	genílla, palæarc.; Austrostylops, austr.; Tetrozócera, pa-
	læarc.)
3.	Thoracic spiracles of female more or less distinct, usually promi-
	nent; tarsi of male with four joints (so far as known); species
	parasitic on Orthoptera, Hymenoptera, Hemiptera and Homop-
	ætera
	Thoracic spiracles of female not usually discernible, never promi-
	nent; tarsi of male with two or three joints; parasites of Homop-

4. Female with three longitudinal rows of genital tubes entering the brood canal; males unknown; parasites of Orthoptera.

(Superfamily STICHOTREMATÒIDEA). (Stichôtrema austr.)  Female with a single median series of four or five genital tuber entering the brood canal; tarsi of male four-jointed; pro- and mesothorax short, transverse; parasites of Hymenoptera and Hemiptera. (Superfamily XENÒIDEA).  5. Cephalothorax of female narrow, elongate, with two pairs of spiracles; five genital tubes. Males unknown; parasites of Hemiptera. (Callipharíxenos, Chrysochoríxenos, indomal.  CALLIPHARIXÉNIDA  Cephalothorax of female broader, with only one pair of spiracles parasites of Hymenoptera.
880 881 885 886 886
Figs. 880–886. Strepsiptera  880. Xenos, head of female (Brues) Xenidæ.  881. Stylops, male (Pierce) Stylopidæ.  882. Xenos, wing of male (Kirby) Xenidæ.  883. Anthericomma, antenna of male (Pierce) Halictophagidæ.  884. Cænocholax, antenna of male (Pierce) Myrmecolacidæ.  885. Parastylops, antenna of male (Pierce) Stylopidæ.  886. Triozocera, antenna of male (Pierce) Mengeidæ.  6. Scutellum broadly rounded in front

# MYRMECOLÁCIDÆ

Scutellum longer than the præscutum; antennæ six-jointed, the third joint prolonged laterally. Cephalothorax of female

(Myrmécolax, ind.; Cænócolax, neotrop.)

elongate. Female unknown; parasitic on ants. (Fig. 884).

- - Præscutum not as broad as the mesothorax at base; antennæ four-jointed, the third joint prolonged laterally; fourth joint elongate. Cephalothorax of female variable in shape; four or five genital tubes. Parasites of wasps and bees. (Figs. 880, 882). (Xènos, palæarc., Am.; Pseudóxenos, Eupathócera, holarc.; Vespáxenos, indoaustr.; Halictóphilus, ind.; Belonogastéchthrus, ethiop.; Hómilops, Am.). XÉNIDÆ
- - Tarsi of male two-jointed; head of female with the tubercles ventral, more or less obsolete; three genital tubes entering the brood canal. (Superfamily ELENCHÒIDEA). (Elénchus, neotrop., ethiop., austr.; Dinelénchus, austr.; Liburnelénchus, Elenchòides, Pentagrammáphila, nearc.).

# ELÉNCHIDÆ

10. Antennæ of male four-jointed, the flagellum of the third and the fourth joint elongate, subequal. (Diozócera, palæarc.).

#### DIOZOCERÁTIDÆ

#### LITERATURE ON STREPSIPTERA

- LENG, C. W. Catalogue of N. American Strepsiptera. See Leng, Cat. Coleop., pp. 343–345 (1920).
- Pierce, W. D. A Monographic Revision of the Twisted-Winged Insects Comprising the Order Strepsiptera. Bull. U. S. Nat. Mus., No. 66 (1909). Strepsiptera. Gen. Insectorum, fasc. 121, 54 pp. (1911).

The Comparative Morphology of the Order Strepsiptera, together with Records and Descriptions of Insects. Proc. U. S. Nat. Mus., **54**, pp. 391–501 (1918).

Ulrich, W. Strepsiptera, In Tierwelt Mitteleuropas, 5, Lief. 1, pp. XIII 1–26 (1930).

## ORDER HYMENÓPTERA

Moderate-sized, small or minute, rarely very large; four membranous wings, the fore pair larger and more completely veined; venation rather complete but not complex, sometimes greatly reduced; head free, mandibulate, but the mouthparts usually adapted for lapping liquid food; antennæ variable, sometimes with very many or very few joints, in the higher forms usually with twelve or thirteen joints; eyes usually moderately large; ocelli nearly always present; prothorax not free; legs similar; tarsi nearly always five-jointed; abdomen usually with six or seven visible segments, the first segment fused with the thorax and not forming a part of the apparent abdomen; no cerci; ovipositor usually sting-like, sometimes saw-like, occasionally greatly elongate.

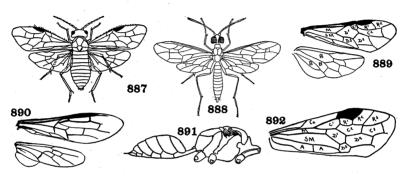
Metamorphosis complete; larvæ caterpillar-like in the more primitive families, legless in the higher forms; pupæ free, sometimes enclosed in a cocoon. Habits varied, phytophagous, predatory, or parasitic. Saw-flies, Wood-wasps, Ichneumon-flies, Ants, Wasps and Bees.

There is no close agreement concerning the family divisions in the Chalastogastra, especially in the higher forms and it is probable that the future will see changes in the grouping of the Diprionidæ and Tenthredinidæ. Among the Terebrantia there is less difference of opinion. The Ichneumonidæ and Braconidæ include many diverse types and are of greater rank than most of the other families, but attempts to divide them have not been satisfactory. The wasps and bees on the other hand have been separated into families on far less important characters. Among the more primitive wasps, some of the family divisions are undoubtedly artificial and must be regarded as tentative only.

#### Adults

	never broadly sessile (Figs. 891, 916, 935); trochanters one- or
	two-jointed; hind wing with less than three basal cells. Sub-
	order CLISTOGÁSTRA (APÓCRITA, PETIOLÀTA)15
2.	Fore wings with three radial cells, $i.e.$ two radial crossveins pres-
۷.	ent; antennæ many-jointed, but with the three basal joints
	strongly developed, the third very long. (Xyèla, holarc.;
	Odontóphyes, Macroxyèla, nearc.) (Fig. 892). XYÉLIDÆ
	Fore wings with only one or two radial cells; third antennal joint
_	only rarely lengthened
3.	0
	antennæ slender, becoming very thin apically, many-jointed;
	radial cell with one crossvein. (Pamphílius (= $L\acute{y}da$ ), Neuró-
	toma, Bactróceros, $holarc.$ ). $(L\acute{Y}DID\pounds)$ PAMPHILÌIDÆ
	Costal cell not divided by a longitudinal vein4
4.	Anterior tibiæ with a single apical spur. (Wood-wasps). Super-
	family SIRICOIDEA
	Anterior tibiæ with two apical spurs. (Saw-flies). (Superfamily
	TENTHREDINOIDEA)8
5.	Fore wings with only one closed cubital cell and one recurrent
	nervure; antennæ ten- or eleven-jointed, inserted much below
	the lower margin of the eyes, beneath a frontal ridge; vertex
	strongly tuberculate. (Orýssus (=0rússus), Eur., N. Am.,
	Austr.; Ophrynòpus, Austr., S. Am.). (IDIOGÁSTRA,
	ORYSSÒIDEA, ORÚSSIDÆ) ORÝSSIDÆ
	Fore wings with two or three closed cubital cells; antennæ inserted
	above the lower margin of the eyes; two recurrent nervures;
	antennæ variable. (Figs. 888, 890) 6
6.	Pronotum nearly truncate or weakly emarginate behind, its an-
٠.	terior portion not forming a vertical surface; abdomen more
	or less compressed. (Fig. 888). (Cèphus, holarc. (C. pygmæus,
	Wheat-stem Saw-fly); Janus, holarc.)
	Pronotum deeply incurved or emarginate behind, its anterior
	part forming a more or less vertical surface; abdomen cylindri-
	cal
7	Parapsidal furrows present; fore wings with a crossvein (Sc) in the
••	costal cell; no triangular plate at the apex of the abdomen;
	prothorax conical, the pronotum a narrow collar. (Xiphýdria,
	widespr.) XIPHYDRÌIDÆ
	Parapsidal furrows absent; fore wings without a crossvein in the
	costal cell; apex of the abdomen with a triangular or spear-
	shaped plate; prothorax subquadrate, the pronotum longer.

- Horntails. (**Trèmex**, cosmop.; **Sìrex**, **Urócerus**, **Xèris**, holarc.). (Fig. 890). (*UROCÉRIDÆ*) ...... **SIRÍCIDÆ**8. Posterior margin of the pronotum straight or nearly so; mesonotum very short, never extending much behind the anterior margins of the tegulæ. (**Megalodóntes**, palæarc.).
  - MEGALODÓNTIDÆ



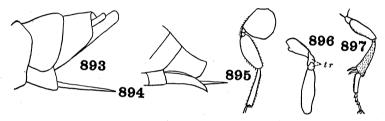
Figs. 887-892. Hymenoptera

- 887. Cladius (Chittenden) Tenthredinidæ.
- 888. Cephus (Marlatt) Cephidæ.
- 889. **Dolerus,** wings: M, median cell; R1, R2, marginal or radial cells; C1, C2, cubital or submarginal cells; D1, D2, D3, first, second and third discoidal cells; SM, submedian cell; B1, B2, B3, basal cells. Tenthredinidæ.
- 890. Tremex, wings. Uroceridæ.
- 891. Chlorion, lateral view of thorax and abdomen (Fernald) Sphecidæ.
- 892. **Xyela,** fore wing. Lettering as in fig. 889; Co, costal cell; A, A, anal cells (MacGillivray) Xyelidæ.

Radial cell not divided; abdomen rounded laterally, not angled

	or carinate; abdomen not greatly swollen. (Pérga, Xylo-pérga, Austr.) PÉRGIDÆ
11.	Antennæ with only three joints, the third very long, sometimes
	split in the male. ( <b>Árge</b> $(=Hyl \acute{o}toma)$ , widespr.; <b>Labidárge</b> ,
	S. Am.; Schizócera, Eur., Am.). (HYLOTÓMIDÆ).
	ÁRGIDÆ
	Antennæ with four or more joints
12.	First discoidal cell petiolate above, i.e. the cubitus arising from
	the basal vein; antennæ four-jointed, the third joint very long.
	(Blasticótoma, one European species).
	BLASTICOTÓMIDÆ
	First discoidal cell almost never petiolate above, the cubitus
	arising beyond the basal vein; antennæ with at least six
1	joints
13.	Radial cell simple, without a crossvein
	Radial cell divided by a crossvein (Fig. 889); antennæ commonly
	filiform, rarely with some of the joints toothed or with pro-
	jections, with at least seven and usually with nine joints. Cos-
	mopolitan but scarce in the tropics. (Monophadnoides (M.
	ribi, Raspberry sawfly); Eriocampòides (E. limacina; E.
	ceràsi, Pear-slugs); Ametastègia (A. glabràta, Dock sawfly);
	Hoplocámpa (H. cooki, Cherry fruit sawfly); Tenthrèdo,
	Tenthredélla, Dólerus, Macròphya, Selándria, Émphytus,
1.4	Fenùsa, holarc.) TENTHREDÍNIDÆ
14.	Antennæ filiform, nine-jointed. (Pachynématus, Pteronídea
	(= Ptéronus), holarc., (P. ribèsi, Currant sawfly), Pontània
	(Willow gall sawflies); <b>Nématus</b> (N. erichsònii, Larch sawfly);
	Diphádnus (D. appendiculàtus, Gooseberry sawfly). (NEM-
	ATÎNÆ)Some <b>TENTHREDÎNIDÆ</b> Antennæ of a different conformation; usually many-jointed, fre-
	quently serrate or pectinate, rarely six-jointed. (Diprion,
	(=Lophyrus) (Pine sawflies)), holarc.; <b>Perreyia</b> , neotrop.;
	Pterygóphorus, Austr.; Acordulécera, Am.). (Including
	PERREYIDÆ, PTERYGOPHÓRIDÆ, LOBOCERÁTIDÆ).
	(LOPHÝRIDÆ) DIPRIÓNIDÆ
15	Last abdominal sternite divided longitudinally, the ovipositor
10.	issuing some distance before the tip of the abdomen (Fig. 893)
	and provided with a pair of narrow sheaths which equal it in
	length. Trochanter divided into two distinct joints, except in
	some forms without stigma; fore wing either with or without a
	costal cell. (TEREBRÁNTIA)

Last abdominal sternite not divided longitudinally, the ovipositor issuing from the tip of the abdomen as a sting (rarely absent) without a pair of exserted sheaths (Fig. 894). Trochanter consisting of a single joint (Fig. 897), or if rarely divided, the second part is very closely attached to and not distinctly separated from the femur (except Trigonalidæ); fore wing always (except Rhopalosomatidæ) with a costal cell. Ants, Wasps and Bees. (ACULEATA)



Figs. 893-897. Hymenoptera

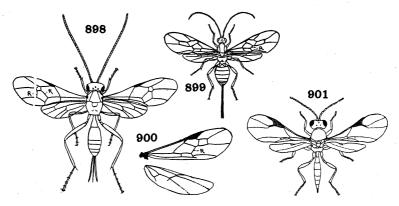
- 893. Ichneumon, apex of abdomen, with ovipositor. Ichneumonidæ.
- 894. Epeolus, apex of abdomen, with ovipositor. Melectidæ.
- 895. Elasmus, basal segments of leg (Silvestri) Elasmidæ.
- 896. Ichneumon, basal segments of leg; tr. two-jointed trochanter (Sharp) Ichneumonidæ.
- 897. Hind leg of a bee (Riley).
- 17. Fore wings with a stigma which is usually triangular or rarely very slender or linear (Fig. 900); costal vein well developed as far as the stigma; abdomen usually with the sternites membranous and with a median fold; antennæ usually with more than sixteen joints; wing venation ordinarily well developed. Ichneumon-flies. (ICHNEUMONÒIDEA) (TRÍSTEGA).....18
- 18. Costal and subcostal (Sc + R) veins separated, enclosing a narrow costal cell; abdominal sternites chitinized (Figs. 904, 905)...19 Costal and subcostal veins confluent, no costal cell (Fig. 900)...26

19.	Mesonotum with a sharp median groove or linear furrow; notauli
	absent; abdomen elongate oval; body more or less cylindrical;
	ovipositor prominent, usually extremely long. (Megályra,
	Austr.; <b>Dinápsis</b> , S. Afr.). (Including <i>DINÁPSIDÆ</i> ).
	MEGALÝRIDÆ
	Mesonotum without median groove; or if with a median impressed
	area, also with notauli
20	Abdomen inserted on the thorax, far above the hind coxæ, com-
20.	monly on a nipple-shaped protuberance; antennæ with 13 or 14
	joints
	Abdomen inserted normally, low down, at the apex of the thorax,
01	and quite close to the hind coxe
21.	
	pletely closed cubital cells, the second one sometimes partly
	open, due to a partial loss of the apical intercubitus. (Fig. 904).
	(Aùlacus, Pristaùlacus, cosmop.; Pammegischia, N. Am.).
	AULÁCIDÆ
	Fore wing with one or no recurrent nervure; only one distinctly
	closed cubital cell, or none
22.	Prothorax long, forming an elongate neck; abdomen long, gradu-
	ally clavate; radial cell of fore wing long, pointed. (Fig. 905).
	(Gasterúption, cosmop.; Pseudofænus, Hemifænus, Austr.)
	GASTERUPTIÓNID <i>Æ</i>
	Prothorax short; body of abdomen short, orbicular, borne on a
	narrow cylindrical pedicel; radial cell short and broad, or
	absent. (Evània, cosmop.; Brachygáster, Eur., Hýptia, Am.).
	EVANÎID <i>Æ</i>
23.	Two or three closed cubital cells
	Only one closed cubital cell, or none
24.	Antennæ with 18 joints or more; head large, quadrate. (See coup-
	let 83)
	Antennæ with 12 or 13 joints. (See couplet 89).
	RHOPALOSOMÁTIDÆ
25.	Antennæ setaceous, multiarticulate, with 30 joints or more; ab-
	domen long and slender; ovipositor long; hind femora swollen
	and toothed before the apex; head tuberculate above. (Sté-
	phanus, cosmop.; Hemistéphanus, S. A., Austr.; Diasté-
	phanus, As., Afr., Austr.) STEPHÁNIDÆ
	Antennæ 14-jointed; abdomen long and slender; ovipositor very
	short; hind femora without teeth; head not tuberculate above.
	(If the body of the abdomen is compressed, rounded $(9)$ or

26.	broadly ovate (♂), borne on a slender cylindrical petiole, compare Roproniidæ, couplet 73.) (Monómachus, S. Am., Austr.; Tetracònus, S. Am.)
	nùsa, Phænocárpa, Aphæreta, Alýsia, cosmop.; Lysiógnatha, N. Am.). (Including LYSIOGNÁTHIDÆ).
	Mandibles attached normally, their tips opposed and meeting
27.	when closed
	armed with a sharp spiniform process. (Agriótypus, holarc.).  AGRIOTÝPIDÆ
28.	Fore wing with two recurrent nervures (except <i>Ophionellus</i> and <i>Hymenopharsalia</i> ); first cubital and first discoidal cells not separated; all the abdominal segments freely movable, except in very rare cases. (Figs. 893, 896, 898, 902). A very large cosmopolitan family. (Including <i>OPHIONÉLLIDÆ</i> , <i>MYER</i> —
	SIIDÆ). This family is commonly divided into a number of subfamilies, but as the current classification involves many exceptions and appears to be highly artificial, a key to these would serve no useful purpose, and be of little practical value.  ICHNEUMÓNIDÆ
	Fore wing with only one, or without any recurrent nervure (Fig. 900); usually with the second and third abdominal segments immovably united above
29.	thorax together, the segments freely movable; tip of propodeum prolonged beyond the hind coxæ. (Hymenopharsàlia, Ophi-
	onéllus, Am.). (See couplet 28). A few ICHNEUMÓNIDÆ Abdomen shorter; propodeum not thus prolonged; venation sometimes considerably reduced. A large cosmopolitan family. (Including APHIDÌIDÆ, CAPITONÌIDÆ, PACHYLOM-MÁTIDÆ, NEORHACÓDIDÆ)
a.	Clypeus semicircularly emarginate below and forming with the mandibles a more or less circular opening or cavityb Clypeus not emarginate below, or at most with a broad, shallow emargination, not forming with the mandibles such an openingi

b. Abdomen sessile, sometimes with the base considerably narrowed, but the first segment not forming a distinct petiole..........c

Abdomen petiolate, the first segment greatly lengthened, at least three times as long as broad at apex, and often very long and slender with the remainder of the abdomen suddenly much wider; antennæ usually very slender. (Spàthius, widespr.; Stephaníscus, Ogmophásmus, ethiop.; Psenóbolus, neotrop.; Cantharóctonus, nearc.). (Including STEPHAN-ISCINÆ)



Figs. 898-901. Hymenoptera

- 898. Cryptus, R, R, recurrent nervures. Ichneumonidæ.
- 899. Microbracon (Hunter and Hinds) Braconidæ.
- 900. Rhogas, wings, R, recurrent nervure. Braconidæ.
- 901. Lysiphlebus. Braconidæ.

	Nervulus originating well beyond the lower end of the basal vein.
	(Exothècus, widespr.; Spinària, ethiop., indomal.; Meso-
	bracon, ethiop.) EXOTHECINÆ
e.	Fore wings with three submarginal cells, all completely formed;
	very rarely without wingsf
	Fore wings with only two submarginal cells, the first sometimes
	incompletely formed; females occasionally winglessh
f.	Head cubical, large, and bulging behind the eyes, rarely slightly
	transverse. (Dorýctes, Dendrosòter, Odontobràcon, wide-
	spr.) <b>DORYCTÌNÆ</b>
	Head clearly transverse, usually much wider than long; not wid-
	ened or bulging behind the eyesg
g.	Subdiscoidal vein arising at the upper corner of the second dis-
O	coidal cell, forming a continuous line with the upper edge of
	this cell. (Hormius, Hormiópterus, widespr.; Chrémylus,
	palæarc.) HORMIÌNÆ
	Subdiscoidal vein originating from the outer side of the second dis-
	coidal cell which is angulate at the point of origin, this vein
	lying below the level of the upper side of the cell. (Fig. 900).
	(Rhògas, cosmop.; Rhýssalus, Clinocéntrus; Gyroneùron,
	ethiop., indomal.) RHOGADÎNÆ
h.	Head, when seen from above, large, cubical; abdominal tergites
	almost always very clearly separated; female always winged;
	hind wing of male often with a stigma-like thickening on the
	costal margin. (Hecábolus, Écphylus, holarc.; Heterospilus,
	widespr.) HECABOLÎNÆ
	Head, when seen from above, strongly transverse, much wider
	than long; sutures of abdomen obsolete above except at the base;
	female commonly wingless; hind wing of male without false
	stigma. (Pámbolus, widespr.) PAMBOLÎNÆ
i.	Abdomen with the tergites separated by distinct sutures, all the
	tergites beyond the third freely movable; if rarely all are fused,
	the abdomen is strongly clavatej
	Abdomen with the tergites fused to form a rigid dorsal shell or
	carapace which is either entirely without sutures or with these
٠.	indicated only as fine groovesw
j.	Abdomen sessile or subsessile; if the first segment is rarely elongate
	its lateral margins are straight; venation of wings not reduced k
	Abdomen petiolate, wing venation often much reduced in the
1	smaller species
k.	Marginal cell either very narrow, or incompletely formed with the
	radial vein weak or wanting apically; second submarginal cell
	usually small or imperfectly formedl
	Marginal cell never much narrowed, second submarginal cell large
	and fully formedp

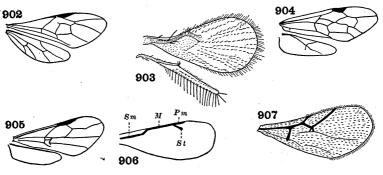
1.	Marginal cell very narrow, the radial vein almost always distinct
	to the tip
	Marginal cell not narrow, more or less incompletely formed
	apically
m.	Paipi very snort; tarsi long, siender, with minute or very indis-
	tinct claws. Minute species parasitic on ants. Neoneurus,
	Elasmosòma, holarc.) NEONEURÌNÆ
	Palpi long, well developed; tarsi not unusually lengthened, with
	large, often toothed or pectinate claws. (Bracon (= $Crémnops$ ),
	Disophrys, Ágathis, Mícrodus, Órgilus, cosmop.; Braunsia.
	ethiop., indomal.; <b>Eárinus</b> , widespr.). (AGATHIDÌNÆ,
	AGATHÎNÆ)BRACONÎNÆ
n.	Second submarginal cell minute or incompletely formed; marginal
	cell wanting except at extreme baseo
	Second submarginal cell large, although more or less weakly de-
	fined apically; apical part of marginal cell weakly defined, but
	the cell is clearly marked by a strongly curved and very deli-
	cately chitinized vein. (Cardiochiles, widespr.; Toxoneuron,
	Am.; Laminitársus, malay.) CARDIOCHILÎNÆ
0.	Radial vein consisting of two nearly equal sections with the stub
	of a third. (Neorhacodes, palæarc.). This group is by some
	placed in the IchneumonidæNEORHACODÎNÆ
	Radial vein composed of only a single basal section. (Microgás-
	ter, Apantèles, Microplitis, cosmop.; Mirax, widespr.; Oli-
ŧ.	goneùrus, neotrop.) MICROGASTRÌNÆ
p.	Fore wings with three submarginal cellsq
	Fore wings with only two submarginal cellsu
q.	Legs long and slender, the spurs of the hind tibiæ often very long;
	body usually slender with the abdomen elongate; head strongly
	transverse. (Macrocéntrus, cosmop.; Zèle, palæarc., Am.;
	Aulacocéntrum, Austrozèle, Australas.; Amicrocéntrum,
	ethiop.; Neozèle, neotrop.) MACROCENTRÎNÆ
	Legs not long and slender, femora stout; abdomen usually stout
	and short, rarely elongate; tibial spurs not elongate; head large
	and thick in all forms with elongate abdomenr
r.	Abdomen inserted low down on the propodeum, between the hind
	coxæ; usually small or minute species
	Abdomen inserted distinctly above the hind coxæ; abdomen often
	elongate; head large, cubical, often with the vertex deeply im-
	pressed; rather large or moderate-sized species. (Hélcon, Gym-
	nóscelus, Aspicólpus, widespr.; Eumacrocéntrus, nearc.;
	Austrohélcon, Schauinslándia, Austr.; Cenocèlius, wide-
	spr.). (Including <i>CENOCŒLIÌNÆ</i> )
a	

	stigma. (Ichneùtes, holarc.; Próterops, widespr.; Ichneut- ídea, Proteropòides, nearc.) ICHNEUTĨNÆ Marginal cell long, much longer than the stigmat
t.	Head above not margined; anal cell of fore wing without trace of
	a crossvein; clypeus often shallowly emarginate, leaving an
	elliptical opening between the mandibles when closed; second
	submarginal cell often much shorter above than below. (Opius,
	cosmop.; Biósteres, Diachásma, Eurýtenes, widespr.; Rhin-
	òplus, ethiop.) OPIĪNÆ
	Head margined above; anal cell of fore wing often with an incomplete crossvein; clypeus not emarginate, fitting close to the
	mandibles when closed; second submarginal cell rarely notice-
	ably shortened above. (Diospilus, widespr.; Dyscolètes,
	holarc.; Eudiospìlus, Neodiospìlus, ethiop.). DIOSPILÎNÆ
u.	Second discoidal cell not completely closed below at apex, the
	posterior part of its outer side without the usual vein. (Bra-
	chístes ( $=Cal\acute{y}ptus$ ), Eubádizon, widespr.). ( $CALYPT\grave{I}N\cancel{E}$ ).
	BRACHISTÌNÆ
	Second discoidal cell completely closed below at apex by a vein . $\cdot \mathbf{v}$
v.	Last section of radial vein curved, marginal cell very short, not
	nearly extending to the wing tip. (Leiophron, Centistes, hol-
	arc.; Centistina, ethiop.) LEIOPHRONINÆ
	Last section of radial vein straight, the marginal cell long, extend-
	ing almost to the tip of the wing. (Blacus, widespr.; Pygóstolus, holarc.) BLACINÆ
w.	Fore wings with three cubital cells. (Chelònus, Chelonélla,
•••	Phanerótoma, Ascogáster, cosmop.; Sphæropyx, palæarc.;
	Minánga, Pachychelònus, ethiop.) CHELONÎNÆ
	Fore wings with two closed cubital cells. ( <b>Triáspis</b> (= $Sigálphus$ ),
	cosmop.). $(SIGALPHIN\cancel{E})$ TRIASPIDÎN $\cancel{E}$
x.	Hind wings with two closed basal cells, the second sometimes
	incompletey
	Hind wings with only one, or without any closed basal cellaa
у.	Fore wings with three submarginal cells, the second one always
	completely closed, their venation not reducedz Fore wings with only two submarginal cells; venation usually
	much reduced, the radial cell usually short. ( <b>Euphorus</b> , hol-
	arc.; Perilitus, Dinocámpus, widespr.; Eustalócerus, Cos-
	móphorus, holarc.) EUPHORÎNÆ
$\mathbf{z}.$	Second submarginal cell about as long as high; antennæ long and
	slender, filiform; abdominal tergites separated by sutures.
	(Meteòrus, cosmop.) METEORÎNÆ
	Second submarginal cell much longer than high; antennæ 18-
	inited the joints toward the apex head-like; second tergite

covering the whole abdomen beyond the petiole. (Helori-mórpha, widespr.) ...... HELORIMORPHÎNÆ

aa. Abdomen inserted low down on the propodeum, between the hind coxæ; fore wings with one, two, or rarely three submarginal cells, the cubitus if present, not arising from the marginal cell. (Fig. 901). (Aphídius (= Încubus), Pràon, Éphedrus, Trióxys, widespr.; Lysíphlebus, holarc.). (INCUBÌNÆ). APHIDIÎNÆ

Abdomen inserted high up on the propodeum, above the level of the hind coxæ; fore wings with two submarginal cells, the cubitus arising from the narrow, triangular marginal cell. (Paxylómma (= Pachylómma) . . . . . . PAXYLOMMATINÆ



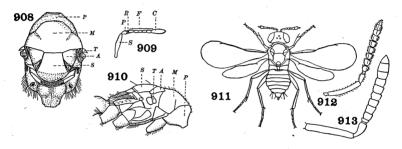
Figs. 902-907. Hymenoptera

- 902. Ophion, wings. Ichneumonidæ.
- 903. Trichogramma, wings (Girault) Trichogrammatidæ.
- 904. Aulacidæ, wings. Aulacidæ.
- 905. Gasteruption, wings. Gasteruptionidæ.
- 906. Chalcid fly, fore wing, diagrammatic. Sm, submarginal vein; M, marginal vein; Pm, postmarginal vein; St, stigmal vein
- 907. Gall wasp, fore wing, diagrammatic (Kieffer).
- 31. Dorsal abdominal plates meeting along the venter and completely enclosing all the ventral plates, except sometimes a part of the

	hypopygium. Parasitic species, cosmopolitan. (Eucòila, Alló-
	tria) FIGÍTIDÆ
	Dorsal abdominal plates usually extending well down on the sides
	of the abdomen, but not meeting along the venter; all, or nearly
	all of the ventral plates visible
32.	Basal joint of the hind tarsi twice as long as the others united,
	second joint with a long process externally which reaches to
	the tip of the fourth joint; abdomen greatly compressed, curved
	like a pruning knife, much longer than the remainder of the
	body. Parasitic species. (Ibàlia, Eur., N. Am.). IBALÌIDÆ
	Basal joint of hind tarsi much shorter; second joint simple, with-
	out process. Mostly gall-making species, almost entirely holarc.
	(Aulacídea, Diástrophus, Rhodites, Disholcáspis, Amphí-
	bolips)
33.	Hind wings exceedingly narrow, linear, the base forming a long
	stalk; ovipositor issuing barely before the tip of the abdomen;
	antennæ with the scape not greatly elongated, usually swollen
	and compressed and without ring joint; very minute species
	with long wing fringe; widespread in distribution. (Polynèma,
	Gonatócerus, Aláptus, Ánaphes) MYMÁRIDÆ
	Hind wings never very narrow, not linear nor with a long stalk
	at the base; ovipositor issuing decidedly before the tip of the
	abdomen; antennæ elbowed (Figs. 909, 912, 913) with long
	scape and usually with from one to three minute ring joints;
	wing fringe almost always much shorter
34.	
	males); axillæ with their anterior margins forming a more or
	less straight line, their sides not extended in front of the tegulæ
	(Fig. 908); spur of front tibia strong, curved
	Tarsi three- or four-jointed (five-jointed or heteromerous only in
	the females of one or two genera); axillæ extended strongly and
	obliquely forward at the sides, well in advance of the tegulæ
	(Fig. 910); spur of front tibia usually weak
35.	
	above; front and hind legs very stout, the middle ones very
	much more slender; males almost always wingless, with stout,
	short, three- to nine-jointed antennæ. (Fig. 917). Tropico-
	politan. Fig-insects. ( <b>Blastóphaga</b> ( $B. ps\`{e}nes = gross\`{o}rum$ , Fig
	caprifier), tropicopolitan; Ágaon, Ceratosòlen, Old World;
	Tétrapus, neotrop.; Pleistodóntes, Austr.) AGAÓNTIDÆ
	Of a different conformation

36. Mesopleura rarely large, with an oblique femoral groove or impression; spur of middle tibia normal, not enlarged...........37 Mesopleura large, entire; flat, without femoral groove in the female and usually in the male; spur of middle tibia usually very large and stout, often fringed with minute spines internally. An extremely varied, abundant, cosmopolitan group. (Eupélmus, Anástatus, Metapélma, Encýrtus, Ageniáspis, Copidosòma, Signíphora). (Including EUPÉLMIDÆ, SIGNIPHÓRIDÆ, and TANAOSTIGMÁTIDÆ).

**ENCÝRTIDÆ** 



Figs. 908-913. Hymenoptera

- 908. **Pteromalus**, thorax from above. P, pronotum; M, mesonotum; T, tegula; A, axilla; S, scutellum. Pteromalidæ.
- 909. Chalcid fly, diagram of antenna. P, pedicel; R, ring-joints; F, funicle; C, club; S. scape.
- 910. Eulophus, thorax seen from side. Lettering as in Fig. 908. Eulophidæ.
- 911. Coccophagus (Howard) Eulophidæ.
- 912. Dibrachys, antenna of female. Pteromalidæ.
- 913. Dibrachys, antenna of male. Pteromalidæ.

#### PTEROMÁLIDÆ

38. Mandibles sickle-shaped, usually with one or two teeth within; thorax greatly elevated; scutellum usually much enlarged and produced behind; abdomen compressed, usually on a long,

	slender pedicel, the second segment very large. (Fig. 920).
	Cosmopolitan, but almost entirely tropical. (Kápala, Ora-
	sèma, Schizaspídia, Thoracántha, Stílbula).
	EUCHARÍDIDÆ
	Mandibles strong and stout, generally with three or four teeth at
	apex; thorax not or very slightly elevated; axillæ separated
	from the mesonotum 39
39.	Hind coxæ very large, long, five or six times larger than the front
	ones
	Hind coxæ never very large, not conspicuously larger than the
	front ones
40.	Hind coxæ more or less triangular in cross-section, sharply ridged
	above; hind femora usually simple, rarely swollen and with a
	tooth beneath; if denticulate beneath, the ovipositor is long . $41$
	Hind coxæ cylindrical, long
41.	Notauli present, ovipositor exserted, usually very long; abdomen
	not coarsely pitted or punctured. Cosmopolitan. (Callimome
	(= Tôrymus), Diámorus, Monodontomèrus, Podágrion,
	Megastígmus (Seed chalcids), Idárnes). (TORÝMIDÆ).
	CALLIMÓMIDÆ
	Notauli absent or obsolete; ovipositor hidden; abdomen of female
	conical, elongate, usually with rows of deep pits or large punc-
	tures. Mainly Eur. and N. Am. (Órmyrus, Tribæus). ORMÝRIDÆ
40	
42.	Hind femora greatly swollen, and toothed or denticulate beneath, their tibiæ curved and oblique at apex; prothorax not elongate
	or narrowed
	Hind femora not swollen, simple; all the legs very long and slender;
	abdomen extremely long and slender; prothorax much elon-
	gated, forming a narrow neck; ovipositor very long. S. Am. and
	Austr. ( <b>Leptofœnus</b> (= $Pelecinella$ ). ( $PELECINELLID$ Æ).
	LEPTOFŒNIDÆ
43.	Fore wings folded longitudinally in repose; ovipositor long, curv-
	ing upwards and forward over the dorsum of the abdomen.
	(Leucóspis, cosmop.; Polistomórpha, S. Am.; Epexo-
	chlænòides, IndAustr.) LEUCOSPÍDIDÆ
	Fore wings not folded; ovipositor only very rarely long, and then
	not thus upcurved; tip of abdomen sometimes drawn out as a
	slender stiff process. (Fig. 914). Cosmopolitan, more abundant
	in the tropics. (Chálcis, Phasgonóphora, Spilochálcis,
	Smicra, Haltichélla, Dirrhinus) CHALCÍDIDÆ

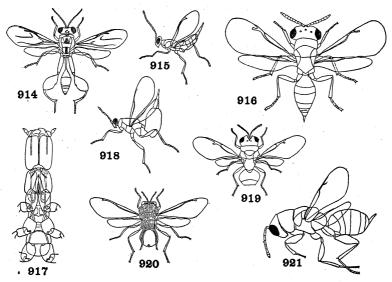
- 45. Abdomen rounded or ovate, more or less compressed, second tergite never very large; the hypopygium usually produced in the female; black or yellowish species. Cosmopolitan. (Harmolita (=Isosòma) (Joint-worms), Eurýtoma, Decátoma, Áxima. Bruchóphagus, Rìleyia). (Figs. 916, 921).

# **EURYTÓMIDÆ**

- - Hind coxæ much enlarged and dilated (Fig. 895), their femora compressed; marginal vein greatly elongated; very small, usually black species. (Elásmus, cosmop.; Euryíschia, Austr.).

#### **ELÁSMIDÆ**

- 48. Tarsi four-jointed (five-jointed or heteromerous in the females of one or two genera); wing hairs not placed in rows or lines; wings usually narrow. A very extensive but poorly known cosmopolitan family. (Figs. 910, 911, 918). (Tetrástichus, Aphelinus, Melittòbia, Ablèrus, Ootetrástichus, Eupléctrus, Eùlophus). (Including TETRASTÍCHIDÆ, APHELÍNIDÆ, ELACHÉRTIDÆ and ENTEDÓNTIDÆ)... EULÓPHIDÆ
  Tarsi three-jointed; wings broad, with the wing hairs usually
  - Tarsi three-jointed; wings broad, with the wing hairs usually arranged in bands or lines; marginal and stigmal veins united to form a strongly recurved stem (Fig. 903). (**Trichográmma** (= Pentárthron, Oóphthora))...... **TRICHOGRAMMÁTIDÆ**



Figs. 914-921. Hymenoptera

- 914. Xanthomelanus (Ashmead) Chalcididæ.
- 915. Trigonoderus (Ashmead) Cleonymidæ.
- 916. Bruchophagus (Urbahns) Eurytomidæ.
- 917. Sycophaga, underside of head and thorax (Grandi) Agaontidæ.
- 918. Paracrias (Ashmead) Eulophidæ.
- 919. **Perilampus** (Ashmead) Perilampidæ.
- 920. Kapala (Ashmead) Eucharididæ.
- 921. Bephratoides (Brues) Eurytomidæ.
- 51. Abdomen short, of normal form, not pointed or enlarged apically. (Wingless males of Idarnini). (See couplet 41).

# Some CALLIMÓMIDÆ

Abdomen broadly sessile, much drawn out into a point apically, or broadened at tip. (Wingless males). (See couplet 35).

AGAÓNTIDÆ

52.	Pronotum as long as the mesonotum and scutellum together;
	abdomen with only four visible segments. (A single species,
	Aliènus, S. Afr.) ALIÉNIDÆ
	Of different conformation
53.	Tarsi five-jointed
	Tarsi four-jointed (see couplet 48) A few <b>EULÓPHIDÆ</b>
54.	Mesopleura large, entire, without a femoral groove. (See couplet
	36) A few <b>ENCÝRTIDÆ</b>
	Mesopleura with an oblique femoral groove or impression $55$
<b>5</b> 5.	Hind coxæ large, more or less triangular in cross-section, ridged
	above. (See couplet 41) A few <b>CALLIMÓMIDÆ</b>
	Hind coxæ small, not ridged above56
<b>56.</b>	Femora of normal size, not greatly swollen on the front or hind
	legs; mesopleura small
	Femora of either the front or hind legs greatly swollen, mesopleura
	large, triangular. (See couplet 46) A few <b>CLEONÝMIDÆ</b>
57.	Hind tibiæ with a single apical spur. (See couplet 37).
	A few PTEROMÁLIDÆ
	Hind tibiæ with two apical spurs
58.	Pronotum wide, quadrate. (See couplet 45).
	A few <b>EURYTÓMIDÆ</b>
	Pronotum narrow, usually narrowed in front or transversely linear.
<b>F</b> 0	(See couplet 46) A few MISCOGÁSTRIDÆ
59.	Abdominal petiole cylindrical, rarely very short; abdomen much
	compressed; antennæ frequently swollen apically or clavate 60
	Abdominal petiole, if well developed, arcuate, or curved toward
60.	the tip, rarely compressed; antennæ never enlarged apically 61 Tergites meeting along the venter and completely enclosing the
00.	sternites. (See couplet 32) Some CYNÍPIDÆ
	Tergites not meeting along the venter; all, or nearly all the ster-
	nites visible. (See couplet 31) A few <b>FIGÍTIDÆ</b>
61.	Second and third tergites immovably united; antennæ with more
01.	than twenty-four joints. (See couplet 29). A few <b>BRACÓNIDÆ</b>
	Second and third tergites freely movable like the other segments;
	very rarely indistinctly separated
62.	Abdominal petiole rather suddenly broadened and bent downward
J	near the tip, its spiracles placed well beyond the middle. (See
	couplet 28)
	Abdominal petiole very short or not of this conformation; very
	small black species. (Some apterous Aphidiinæ). (See couplet
	29) A few <b>BRACÓNIDÆ</b>

63.	Pronotum extending entirely or almost back to the tegulæ, its hind angles usually not lobed, or the tegulæ absent; trochanters occasionally two-jointed
	Pronotum shortened (rarely extended in front as a neck), more or less collar-shaped (Figs. 891, 925), not extending back on the sides to the tegulæ, although each posterior angle is produced to form a lobe; trochanters always one-jointed
64.	First segment of abdomen forming a scale or node (Figs. 922, 923, 924, 926) or the first and second nodiform, and clearly separated both above and below from the gaster, or remainder of
	the abdomen
	First segment of abdomen not scale-like; if nodiform and separated
	from the gaster by a constriction, the second segment forms a part of the gaster and is not separated from it both above and below. 66
65.	
00.	not dimorphic, without sterile worker caste; female with suture
	between mesonotum and pronotum, but without suture or im-
	pression between mesonotum and metanotum or propodeum.
	Insects not living in colonies. (Apterógyna, Afr., As.).
	APTEROGÝNIDÆ
	Hypopygium of male without an upturned spine; female di-
	morphic, the sterile form (worker) wingless, with the thoracic
	sutures lost or much reduced; the fertile form with the thoracic sutures all present. Insects living in colonies. Ants. (FORMI-
	CÒIDEA, HETERÓGYNA) FORMÍCIDƹ
	•
a.	Cloacal orifice slit-shapedb Cloacal orifice round, terminal, surrounded by a fringe of hairs;
	sting not functional; abdominal pedicel consisting of a single
	segment; no constriction between the second and third ab-
	dominal segments; male genitalia not retractile. (Fig. 923). (Prenólepis, cosmop.; Làsius, Formica, holarc.; Polýrha-
	chis, Old World tropics; Camponotus (Wood-ants), cosmop.).
	FORMICÎNÆ
b.	Sting vestigial (except Aneuretus); abdominal pedicel consisting
	of a single segment; no constriction between the second and
	third abdominal segments; species producing a secretion of aromatic rancid odor. ( <b>Dolichodèrus</b> , cosmop.; <b>Dorymỳrmex</b> ,
	aromane raneid odor. (Donottoder us, cosmop., Dorymyrmex,

## DOLICHODERÎNÆ

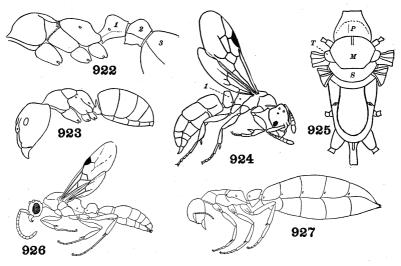
Iridomýrmex (I. hùmilis, Argentine ant), widespread).

<sup>&</sup>lt;sup>1</sup> The ants form a very extensive, widespread, and dominant family divisible into a number of well-marked subfamilies, several of which are undoubtedly of equal rank with many of the families recognized in other groups. A table for their separation is given below, adapted from a recent key by Wheeler.

	Sting developed, sometimes very small, but capable of being exserted. Petiole of abdomen (pedicel) either consisting of two joints, or with the first segment forming a node and the second and third separated by a strong (very rarely weaker) con-
c.	striction
	Pedicel consisting of a single segment, more rarely of two, but in this case the frontal carinæ are very close to each other and do not cover the insertions of the antennæ (Dorylinæ) or the mandibles are linear and denticulate (Myrmecia)e
d.	Antennæ 12-jointed in worker, female and male; clypeus not
	prolonged back between the frontal carinæ, its posterior margin
	rounded; fore wings almost always with two closed cubital cells;
	one of the tibial spurs on the middle and hind legs pectinate;
	ocelli almost always present in the worker. (Pseudomýrma,
	neotrop.; Pachysima, Viticícola, Afr.).
	PSEUDOM YRMÎNÆ
	Clypeus almost always prolonged between the frontal carinæ;
	if not, the spurs of the middle and hind tibiæ are simple or
	absent, or the antennæ are 11-jointed in the worker and female, 12-jointed in the male, and the fore wings have one closed cubi-
	tal cell. ( <b>Pogonomýrmex</b> (Agricultural ants), Am.; <b>Myrmica</b> ,
	holarc., Indo-Mal.; <b>Pheidòle, Monomòrium</b> , tropicopolitan;
	Solenópsis (S. geminàta, Fire ant), cosmop.; Átta (Leaf-
	cutter ants), neotrop.)
е.	Frontal carinæ very close to each other; almost vertical, not at all
•	covering the antennal insertions; abdominal pedicel of one or
	two segments; genitalia of male almost always completely re-
	tractile. Legionary and Driver ants (Fig. 927). (Dórylus,
	Æníctus, Old World tropics; Éciton, neotrop.) DORYLÎNÆ
	Frontal carinæ separated or close together, in the latter case being
	dilated anteriorly to form an oblique or horizontal lamina,
	which covers in part the insertion of the antennæ; abdominal
	pedicel almost always of a single segment. Genitalia of male
	incompletely retractile. (Fig. 924). (Pachycóndyla, neotrop.;
	Myrmècia (Bull-dog ants), Austr.; Stigmatomma, widespr.;
	Ponèra, cosmop.; Cerápachys; Odontómachus, tropico-
	politan). (Including CERAPACHYINÆ) PONERÎNÆ
66.	Winged
	Wingless, or with the wings much reduced in size98
67.	Hind wings without distinct venation and with no closed cells;
	usually small or minute insects. (See couplet 134)68

Hind wings with a distinct venation and with at least one closed cell

68. Antennæ inserted far above the clypeus on a frontal shelf or strong prominence; small or minute, never large species.....69



Figs. 922-927. Hymenoptera

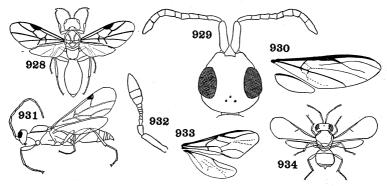
- 922. **Myrmica,** profile view of thorax and base of abdomen; 1, 2, basal nodes of abdomen (first and second segments); 3, third abdominal segment. Formicide.
- 923. Camponotus, lateral outline of body (Wheeler) Formicidæ.
- 924. **Ponera**, female. 1, node or basal segment of abdomen (Wheeler) Formicidæ.
- 925. Chlorion, thorax from above. P, pronotum; M, mesonotum; S, scutellum; T, tegula (Fernald) Sphecidæ.
- 926. Phryacaces, male (Wheeler) Formicidæ.
- 927. Eciton, female (Emery) Formicidæ.

Antennæ not inserted on a frontal shelf or strong prominence, arising just above the clypeus or near the middle of the face. . 70

69. Wing venation very incomplete; radial cell if indicated very small, cubital vein (R<sub>4</sub>+R<sub>5</sub>+M<sub>1</sub>) entirely wanting; antennæ 11- to 14-jointed, usually strongly clavate in the female. (Fig. 932). (Paramèsius, Spilómicrus, Gálesus, Phænòpria, Trichòpria, cosmop.; Hoplòpria, Am.) . . . . . DIAPRÌIDÆ

	Wing venation more complete; radial cell large, but not always
	completely closed; cubital vein well developed except at apex
	antennæ of males 10-jointed, of females 13-jointed. (Embóle-
	mus, Eur.; Myrmecomórphus (= Pedinómma), Eur., N.
	Am.; Ampulicimórpha, N. Am.) EMBOLÉMIDÆ
70.	Antennæ composed of fourteen joints or more71
	Antennæ composed of thirteen joints or less
71.	Basal joint of hind tarsi much shorter than the following joint:
	first abdominal segment as long as the head and thorax together;
	abdomen very long, filiform and composed of equal segments
	in the female; clavate in the male; large insects. (Fig. 930).
	(Pelecinus, Am.) PELECÍNIDÆ
	Basal joint of hind tarsi longer than the second joint; abdominal
	petiole very much shorter72
72.	Head elongate; antennæ 22- to 40-jointed, inserted at the anterior
	margin of the head; females wingless. A small, widespread
	group of rare insects. (Sclerogíbba, Mystrocnèmis, Crypto-
	béthylus) SCLEROGÍBBIDÆ
	Head short, broader than long; antennæ 14- or 15-jointed, inserted
	at the middle of the face, far above the clypeus
73.	Basal vein complete; antennæ 14-jointed; abdomen strongly
	compressed. (Roprònia, N. Am.) ROPRONÌIDÆ
	Basal vein incomplete, its anterior portion wanting; antennæ 15-
	jointed; abdomen not compressed. (Helòrus, Eur., N. Am.)
	HELÓRID <i>Æ</i>
74.	Mandibles very short, with three large teeth, widely separated
	and not meeting when closed; when open, the tips are directed
	laterally; abdomen with only two (female) or four (male) visible
	tergites, the first covering most of the abdomen; sting very
	long, extended forward beneath the body. (Fig. 928). A single
	species. (Vanhórnia, N. Am.) VANHORNÌIDÆ
	Mandibles in normal position, their tips opposing when closed75
<i>7</i> 5.	,
	sexes; fore wings with a broad stigma and a closed, usually very
	small, radial cell; abdomen with a short cylindrical petiole
	the second segment much longer than the others. (Fig. 931)
	(Sérphus (= Proctotrypes), Phænosérphus, widespr.; Exallò-
	nyx, Eur., N. Am.). (PROCTOTRÝPIDÆ) SÉRPHIDÆ
	Antennæ inserted near the mouth, close to the margin of the cly
	peus
76.	Hind wing with a lobe at the anal angle, separated by a deep, slit

Hind wing very narrow without anal lobe or slit-shaped notch 80 77. Abdomen with two to four, rarely five dorsal segments; venter concave; propodeum laterally with sharp keels or teeth; prothorax large, its posterior corners often distinctly separated from the tegulæ; ovipositor tubular, extensile, several-jointed;



Figs. 928-934. Hymenoptera

- 928. Vanhornia (Crawford) Vanhorniidæ.
- 929. Calliceras, head and antennæ. Calliceratidæ.
- 930. Pelecinus, wings. Pelecinidæ.
- 931. Serphus (Brues) Serphidæ.
- 932. Loxotropa, antenna. Diapriidæ.
- 933. Chrysis, wings. Chrysididæ.
- 934. Telenomus. Scelionidæ.

last large tergite frequently dentate; antennæ 13-jointed; body usually with coarse sculpture and of metallic color. Cuckoowasps. (Fig. 933). (Chrysis, cosmop.; Parnòpes, Ellámpus, Hédychrum, widespr.; Allocœlia, S. Afr.). CHRYSÍDIDÆ Abdomen with six to eight visible segments, very rarely with only four or five; venter convex; body rarely with any coarse sculp-

78. Antennæ 10-jointed in both sexes; front tarsi of female usually pincers-shaped (Fig. 937); head broad, transverse, or subquadrate; female frequently apterous with the thorax nodose. (Figs. 937, 938). (Drýinus, Gonátopus, Ánteon, cosmop.; Aphélopus, Eur., N. Am.; Bócchus, N. Am.; Paradrýinus, Austr.)

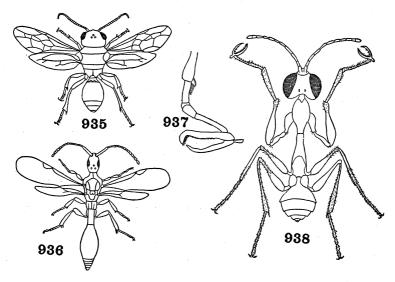
DRYÍNIDÆ

	Antennæ 12 to 13-jointed; front tarsi simple; head elongate or
	rounded
<b>7</b> 9.	Anal lobe of hind wing conspicuous; abdomen with seven or eight
	visible dorsal segments; head usually elongate, oblong; small,
	usually black or bronzed species, the females sometimes wing-
	less. (Béthylus, Mesítius, Perisiérola, widespr.; Sièrola,
	Hawaii; Pristócera, Épyris (s. lat.), Sclerodérma, Goniòzus,
	cosmop.) BETHÝLIDÆ
	Anal lobe of hind wing not conspicuous, separated by a minute
	notch; abdomen with at most six segments visible from above.
	(Cléptes, Eur., Am.) CLÉPTIDÆ
80.	Abdomen acute, or sharply margined along the sides81
	Abdomen rounded on the sides; wings, when present, with the
	radial vein developed, but not complete, leaving the radial cell
	open; no postmarginal vein. (Fig. 929). (Lygócerus, Cono-
	stigmus, Calliceras, cosmop.; Megaspilus, holarc.). (CERA-
	PHRONIDE)CALLICERÁTIDÆ
81.	
	front wings without marginal or stigmal vein and usually with-
	out a subcostal vein also. (Platygáster (including Polygnò-
	tus), cosmop.; Inostémma, Léptacis, Amblyáspis, Eur.,
	Am.). PLATYGÁSTRIDÆ
	Antennæ twelve- or eleven-jointed (if rarely seven- or eight-
	jointed, the club is unjointed, or if ten-jointed, the stigmal vein
	is present); marginal and stigmal veins usually present. (Fig.
	934). (Phanùrus, Telénomus, Calotelèia, Scèlio, cosmop.;
	Tèleas, Prosacántha, Hadronòtus, widespr.; Gryon, Eur.,
	Am.) SCELIÓNIDÆ
82.	
	Antennæ with never more than thirteen joints; twelve-jointed in
	the female and thirteen-jointed in the male, except in rare in-
	stances when the number may be reduced. (VESPOIDEA)85
83.	
	least 16 joints, usually more; hind wing with two large closed
	cells; moderate-sized, often brilliantly colored species. (Fig.
	935). (Trigónalys, palæarc.; Lycogáster, Am., E. Ind.;
	Tapinógalos, N. Am., S. Afr.; Seminòta, neotrop.; Bareo-
	gónalos, N. Am.) TRIGONÁLIDÆ
	Mandibles with not more than three teeth; antennæ 14- or 15-
	jointed
84.	

ing back beyond the insertion of the hind coxæ; antennæ not inserted on a large frontal prominence. (See couplet 25).

#### MONOMÁCHIDÆ

Fore wing always without a closed discoidal cell; propodeum not extended back beyond the insertion of the hind coxæ; antennæ inserted on a conspicuous frontal shelf or prominence; small black or brownish species. (Fig. 936). (Bélyta, Oxýlabis,



Figs. 935-938. Hymenoptera

- 935. Seminota (Schultz) Trigonalidæ.
- 936. Belyta (Fouts) Belytidæ.
- 937. Gonatopus, front tarsus of female. Dryinidæ.
- 938. Gonatopus (Misra) Dryinidæ.

# Aclista, widespr.; Pántoclis, cosmop.; Leptorháptus, holarc.) ..... BELÝTIDÆ

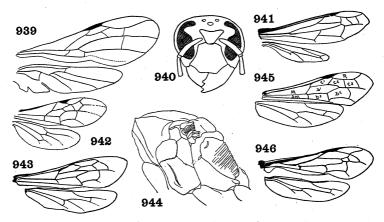
- - First discoidal cell very long, as a rule much longer than the submedian cell; fore wing almost always folded longitudinally when

	in repose; frequently social species, living in colonies. (Figs. 940, 941, 944, 946)
a.	Transverse median vein in hind wing straight or curved, not angulate; fore wing with two or three cubital cells, the marginal cell always truncate at apex; antennæ usually strongly clavate. b
	Transverse median vein angulate; antennæ not noticeably swollen apically
b.	Anal lobe of hind wing elongate, more than half as long as the submedian cell. (Euparagia, N. Am.) EUPARAGINÆ
	Anal lobe of hind wing small, circular or oval, much less than half as long as the submedian cell
c.	Discoidal vein in hind wing obsolete or entirely absent. (Másaris, palæarc.; Pseudomásaris, N. Am.; Trimèria neotrop.; Ma-
	sariélla, Ceràmius, Ceramiòides, Afr.; Paràgia, Austr.). MASARIDÌNÆ
	Discoidal vein in hind wing present, fully developed. (Gayélla,
d.	Paramásaris, neotrop., N. Am.)
	Second cubital cell receiving both recurrent nervurese
e.	Mandibles short and broad, obliquely truncate and toothed at the apex, folding above one another under the clypeus or very slightly crossing
	Mandibles more or less elongate and knife-like, either crossing each other or extended forward to form a beak, their inner
f	margins more or less toothed or notchedj  Tarsal claws bifid or toothed; middle tibiæ with one or two spurs;
1.	clypeus broadly truncate at apex; solitary wasps. (Zèthus, cosmop.; Làbus, Afr., Indo-Mal.) ZETHÎNÆ
	Tarsal claws simple; middle tibiæ with two spurs, very rarely with one; true social wasps, living in colonies, the females often
	dimorphic as fertile females and sterile workersg
g.	Clypeus broadly truncate and more or less emarginate at the apex; first abdominal tergite vertically truncate anteriorly; hind wing without a lobe at the anal angle, the basal third strongly narrowed. Hornets; Yellow-jackets (Véspa, Véspula,
	palæarc., nearc., Indo-Mal.)VESPĪNÆ
	Clypeus pointed at the apex, rarely rounded or straight, in which case the first abdominal tergite is not vertically truncate; hind wing usually with a lobe at the anal angleh

<sup>&</sup>lt;sup>1</sup>This varied and extensive family is divisible into several groups, some of which were formerly regarded as families, but now more generally as of only subfamily rank. A key to these which has been kindly revised by Prof. Bequaert is given below.

h.	Second abdominal segment broadly bell-shaped, its tergite and sternite completely or for the most part fused; first segment much narrower than the second; extensory muscle of the abdomen inserted on the propodeum in a narrow and much compressed slit. (One genus, Ropalídia (= Icària), Afr., Indo-Austr.). (RHOPALIDIÌNÆ)
i.	Second abdominal segment of the usual shape, its tergite and sternite freely articulated
	narrow and much compressed slit; first abdominal segment never narrowed into a stalk; antennæ always of 12 joints in the female and 13 in the male. (Polistes, cosmop.; Gyróstoma oriental)
	antennæ with 11 or 12 joints in the female and 12 or 13 in the male. (Polýbia, Chartérgus, neotrop.; Mischocýttarus, Nectarina, N. Am., neotrop.; Belonogáster, Afr.; Polybiòides, Afr., oriental)
j.	Middle tibiæ with one apical spur (very rarely two or none) clypeus broadly rounded, truncate or emarginate at tip (very rarely pointed). A large cosmopolitan group. (Eùmenes Odynèrus, Symmórphus, Ancistrócerus, Pterochìlus, cosmop.; Synàgris, Afr.; Pachýmenes, widespr.; Monòbia N. Am., neotrop.)
86.	Flagellum of antennæ bare or with very short pubescence, not clothed with conspicuous hairs
	Flagellum clothed with conspicuous hairs, as long as, or much longer than the width of the antennal joints. (Plumàrius (= Konowiélla), Chile, Argentina; Myrmecopterina (= Archihymen), S. Afr.). (KONOWIÉLLIDÆ, ARCHIHYMÉ-IDÆ)
87.	Mesopleura divided by an oblique suture into a lower and upper part; legs, including the coxæ very long; hind femora unusually long; middle tibiæ with two spurs. Spider hunting wasps. (Psammóchares (= Pómpilus), Pépsis (Tarantula hawks), Cerópales, Agènia, widespr.). (CEROPÁLIDÆ, POMPÍL-IDÆ) PSAMMOCHÁRIDÆ Mesopleura not thus divided; legs shorter, the hind femora
	not usually extending to the apex of the abdomen 88

88. Meso- and metasternum forming together a flat plate which is divided by a transverse, more or less sinuous suture, and overlies the bases of the four posterior coxæ; wing membrane, beyond the closed cells, finely longitudinally wrinkled; hypopygium of male with three spines. Large, usually brightly colored wasps. (Scòlia, Campsómeris (=Èlis), cosmop., mainly trop.)



Figs. 939-946. Hymenoptera

- 939. Paniscomima, wings (Enderlein) Rhopalosomatidæ.
- 940. Vespa, head in front view (Schmiedeknecht) Vespidæ.
- 941. Vespa, wings. Vespidæ.
- 942. Sphærophthalma, wings. Mutillidæ.
- 943. Elis, wings of female. Tiphiidæ.
- 944. Polistes, lateral view of thorax (Bequaert) Vespidæ.
- 945. **Elis**; wings of male. R, radial or marginal cell; C1, C2, C3, cubital or submarginal cells; D1, D2, D3, discoidal cells; M, median cell; SM, submedian cell. Tiphiidæ.
- 946. Eumenes, wings. Vespidæ.
- 89. Joints of antennal flagellum long and slender, each bearing at the apex two slender spines; joints of tarsi broadened and deeply lobed (female); hind wing with a prominent anal lobe and a

	deep axillary incision (Fig. 939). (Rhopalosòma, Am.; Panis-
	comìma, Afr.; Hymenochimàra, India).
	RHOPALOSOMÁTIDÆ
90.	Joints of antennal flagellum not spined at tip, tarsi simple90 Mesosternum with two laminæ that overlie or project between the bases of the middle coxæ and usually extend to the mid-line
	where they are separated by a median suture91
	Mesosternum simple, without appendages behind, or with the laminæ reduced to a pair of minute tooth-like projections. 94
91.	Ocelli small
	Ocelli very large; nocturnal insects (males). (Brachycistis, Chyphòtes, Am.). (See couplet 94).
	Brachycistìnæ of the <b>MUTÍLLIDÆ</b>
92.	upcurved spine (if there are 8 ventral abdominal segments and the pygidium is not deeply emarginate, see <i>Dimorphothynnus</i> and <i>Rhagigaster</i> in the Thynnidæ); females with a deep constriction between the first and second ventral abdominal segments. (Figs. 943, 945). ( <b>Típhia</b> , cosmop.; <b>Paratíphia</b> , Am.; <b>Èlis</b> (= <i>Myzìne</i> ), widespr.; <b>Pterómbrus</b> (= <i>Engycýstis</i> ), Am.).
	(Including PTERÓMBRIDÆ and MYZÍNIDÆ). TIPHÌIDÆ
	Males with apex of abdomen not terminating in a single strongly upcurved spine (except <i>Dimorphothynnus</i> and <i>Rhagigaster</i> ),
	otherwise armed or without spines; females (of <i>Anthobosca</i> ) without a deep constriction between the first and second ventral abdominal segments
93.	
	their sockets facing anteriorly or laterally instead of dorsally; wing venation of male complete and extending out to the tips
	of the wings, three closed cubital cells, the first usually divided at least partially by a spur from the first intercubitus; female
	wingless. (Diámma, Rhagigáster, Dimorphothýnnus,
•	
	Austr.; Elaphróptera, Encyrtothýnnus, S. Am.; Glypto-
	metòpa, N. Am.)
	Pygidium of male unarmed; antennæ of male not inserted under a
	frontal prominence, their sockets facing dorsally; female winged.
	(Anthobòsca, S. Am., Austr.) ANTHOBÓSCIDÆ
94.	
	Hind wing without a lobe at the anal angle, at most with an ob-
	tuse emargination at the posterior basal angle

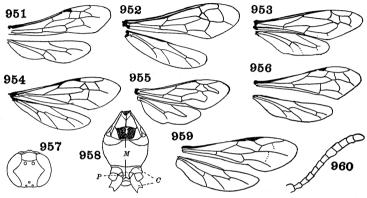
95.	Cubital vein in hind wing originating beyond the transverse median vein; second abdominal segment separated from the first by a strong constriction both above and below; female winged. (Sierolomórpha, nearc., Hawaii).
	SIEROLOMÓRPHIDÆ
	Cubital vein in hind wing not originating beyond the transverse median vein; body almost always conspicuously pilose (Fig. 942); female wingless. Velvet ants. A widespread family occur-
	ring mainly in desert regions. (Mutilla, palæarc., ethiop., oriental; Dasymutilla, Photópsis, Am.; Traumatomutilla,
	neotrop.; Ephutomórpha, austr.; MUTÍLLIDÆ
96.	Abdomen with the several segments separated by strong constrictions (males)
	Abdomen without such constrictions, except between the first
	and second segments; hypopygium of male unarmed; female
	winged; sting long, enclosed in a tubular organ at base; body
	bare, marked with yellow or white. (Sapyga, widespr.; Eusa-
	pỳga, Am.) SAPÝGIDÆ
97.	Hypopygium of male armed with a strong, upcurved spine; female
	wingless, the thorax constricted into three parts. (Methòca,
	widespr.; Dryinópsis, E. Ind.) METHÓCIDÆ
	Hypopygium of male unarmed; female wingless, the thorax di-
	vided into two parts by a transverse suture. (Myrmòsa,
	holarc.)
98.	Thorax clearly divided above into three parts by sutures or
	sharply defined constrictions, or into two by a constriction;
	scutellum nearly always present
	Thorax with only one dorsal sclerite, or with two separated by a
	suture; no scutellum
99.	
	Females of the <b>SCLEROGÍBBIDÆ</b>
	Antennæ with 15 joints or less
100	
	No shelf or strong prominence at the base of the antennæ103
101	petiole; antennæ inserted on a frontal shelf102
	Abdomen without a distinctly separated petiole; antennæ not
	inserted on a shelf. (See couplet 69) Some <b>EMBOLÉMIDÆ</b>
102	Antennæ usually 12-jointed. (See couplet 69).
	Some females of the <b>DIAPRIID</b>
	Antennæ usually 15-jointed. (See couplet 84).
	Some females of the <b>BELÝTID</b> Æ

103.	Abdomen acute or sharply margined along the sides104 Abdomen without an acute or sharply margined edge along the
	sides
104.	Antennæ 12- or 11-jointed (if rarely 7- or 8-jointed, the club is
	unjointed). (See couplet 81).  A few females of the <b>SCELIÓNIDÆ</b>
	Antennæ 10-jointed, rarely with fewer joints, but never more.
	(See couplet 81) A few males of the <b>PLATYGÁSTRIDÆ</b>
105.	Antennæ inserted low down, next to the border of the clypeus. 106
100.	Antennæ inserted at the middle of the face, far above the clypeus.
	(Paracòdrus). (See couplet 75).
	A few females of the <b>SÉRPHIDÆ</b>
106.	Antennæ 10- or 11-jointed
	Antennæ 12- or 13-jointed
107.	Front tarsi of female pincers-shaped; antennæ of male 10-
	jointed. (See couplet 78) Some <b>DRYÍNIDÆ</b>
	Front tarsi of female not pincers-shaped; antennæ of male 11-
	jointed. (See couplet 80) Some CALLICERATIDÆ
108.	Head elongate, usually distinctly longer than wide; thorax
	divided by sutures, or if divided by constrictions, the head is
	much longer than wide. (See couplet 79).  A few females of the <b>BETHÝLIDÆ</b>
	Head almost always oval, slightly broader than high 109
109.	Abdomen with only four visible tergites. (See couplet 52).
100.	ALIÉNIDÆ
	Abdomen normal, with six to eight visible tergites
110.	Wings present, though greatly reduced in size; tegulæ normally
	developed; thorax of normal form, divided by sutures111
	Wings absent; tegulæ indicated only as minute tubercles; thorax
	generally of abnormal form, often divided by constrictions112
111.	Antennæ 13-jointed. (See couplet 95).
	A very few males of the <b>MUTÍLLIDÆ</b>
	Antennæ 12-jointed. (See couplet 87).
110	A very few females of the <b>PSAMMOCHÁRIDÆ</b>
112.	1 /
	between the bases of the middle coxæ. (See couplet 97).  Females of the <b>METHÓCIDÆ</b>
	Mesosternum with two laminæ that overlie or project between
	the bases of the middle coxe
113.	Abdomen with a distinct cylindrical petiole consisting only of the
110.	sternite, the first tergite capping its posterior end. (See couplet
	91) (Females of Chyphôtes) MUTÍLLIDÆ, part
	) (

114.	suture
	Pronotum fused with the remainder of the thorax, the entire thorax without sutures above. (See couplet 95).  Females of the MUTILLIDÆ
115.	Ocelli present. (See couplet 97). Females of the MYRMÓSIDÆ Ocelli absent. (See couplet 91). Females of Brachycístis).  MUTÍLLIDÆ, part
	950
	947 948 949
	Figs. 947-950. Hymenoptera
	948, 949. Hind legs of bees (Smith) Apoidea. Plumose or compound body-hairs of bees (Smith) Apoidea.
116.	Antennæ with 12 (female) or 13 (male) joints; very rarely with 12 in the male
117.	Hind tarsi slender, filiform, the first joint not broadened or thickened; abdomen often petiolate; all hairs on the body simple, unbranched. (SPHECÒIDEA)
	Hind tarsi with the first joint thickened or flattened, often densely hairy; abdomen always sessile; at least a part of the body-hairs branched or plumose. Bees. (Figs. 947, 948, 949, 950, 964, 965).
118.	(APÒIDEA (ANTHÓPHILA))
110.	Middle tibiæ generally with only one well developed apical spur; sometimes with two or none. When a second spur is well de-

119.	Mesosternum produced into a forked process posteriorly; parap-
	sidal furrows distinct; pronotum usually long, conically pro-
	duced in front, usually with a median groove, its posterior lobes
	often approaching close to the tegulæ. (Ampûlex, widespr.;
	Rhinópsis, Dolichùrus, cosmop.) AMPULÍCIDÆ
	Mesosternum not thus produced; parapsidal furrows indistinct
	or wanting
120.	Abdomen with a distinct, usually long, cylindrical petiole, which
	at the base, at least, consists only of the sternite. (Figs. 891, 925,
	951). (Chlorion (=Sphéx, auctt.), Scéliphron (=Pelopæus),
	Sphéx (= Ammóphila), cosmop.) SPHÈCIDÆ
	Abdomen sessile or subsessile, never with a slender, cylindrical
	petiole
121.	Labrum free, well-developed, wider than long, triangular or semi-
	circular, extending beyond the clypeus; sternauli on mesopleura
	not complete. (Stizus, Sphècius, cosmop.; Exeirus, Austr.)
	$(EXE\hat{I}RID\mathcal{E})$ STĪZID $\mathcal{E}$
	Labrum short, small, not or scarcely extending beyond the cly-
	peus. (If the second cubital cell is petiolate and the hind angles
	of the propodeum are neither acute nor spined, see Exeirus,
	couplet 121)
122.	Radial cell broadly truncate at apex and prolonged as a small,
	weakly defined cell; antennæ inserted near the clypeus, very
	close to the clypeal suture; eyes of male usually very large
	and contiguous above. (Dimorpha (= Astata), cosmop.; Dip-
	lopléctron, N. Am.) (ASTÁTIDÆ) DIMÓRPHIDÆ
	Radial cell not appendiculate, pointed at apex123
123.	Antennæ inserted very near to the upper edge of the clypeus,
	close to the clypeal suture; first segment of abdomen usually
	long, slender, nodose at apex and separated from the second by
	a distinct constriction; second cubital cell not receiving a re-
	current nervure. (Méllinus, widespr.) MELLÍNIDÆ
	Antennæ inserted on the face well above the margin of clypeus;
	first segment of abdomen broad and stout; second cubital cell
	receiving at least one recurrent nervure, usually two124
124.	Sternauli on mesopleura complete, usually deep; second cubital
	cell with a distinct upper side, not triangular; propodeum
	rounded; thorax smooth, not coarsely punctate. (Fig. 956).
	(Gorỳtes, cosmop.) GORÝTIDÆ
	Sternauli wanting or indicated only anteriorly; second cubital
	cell triangular or petiolate; propodeum with the upper hind

125. Eyes deeply emarginate; one clearly defined cubital cell, abdomen petiolate and gradually enlarged apically (**Trypóxylon**, cosmop.); or two cubital cells, or three with the second petiolate (**Pison**, cosmop.; **Pisonópsis**, nearc.; **Aulacóphilus**, neotrop.). (Figs. 957, 959). (TRYPOXYLÓNIDÆ). **TRYPOXÝLIDÆ** 



Figs. 951-960. Hymenoptera

- 951. Chlorion, wings. Sphecidæ.
- 952. Eucerceris, wings. Cerceridæ.
- 953. Bembix, wings. Bembicidæ.
- 954. Cerceridæ. Cerceridæ.
- 955. Tachytes, wings. Larridæ.
- 956. Gorytes, wings. Gorytidæ.
- 957. Trypoxylon, front view of head (Kohl) Trypoxylidæ.
- 958. **Tachytes**, underside of thorax. M, mesosternum; P, its posterior process; C, coxæ (Williams) Larridæ.
- 959. Trypoxylon, wings. Trypoxylidæ.
- 960. Niteliopsis, antenna of female (Williams) Larridæ.

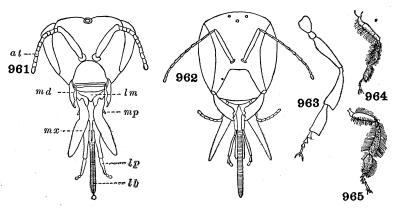
- 126. Fore wings with two or three completely closed cubital cells, the veins enclosing them strong and sharply defined......127

  Fore wings with not more than one closed cubital cell.........133
- 127. Labrum large, free, triangularly elongated beyond the clypeus, much longer than wide; radial cell simple, not appendiculate

	•
•,	at tip; ocelli more or less aborted. (Fig. 953). ( <b>Bémbix</b> (=Bêmbex), cosmop.; <b>Microbémbex</b> , <b>Bicýrtes</b> (=Bembídula), <b>Stíctia</b> (=Monédula), <b>Steniòlia</b> , Am.) <b>BEMBÍCIDÆ</b>
	Labrum small, usually entirely concealed by the clypeus; radial
	cell sometimes with a crossvein clearly defined; ocelli, or at
100	least the anterior one, perfectly formed
128.	Abdomen strongly constricted both above and below between
	the first and second segments
	Abdomen without a strong constriction above between the first
	and second segments
129.	Mesopleura with a vertical furrow which separates the prepectus
	from the rest of the mesopleura; hind femora usually simple at
	apex. (Philánthus, cosmop.; Tráchypus, neotrop.; Aphil-
	ánthops, nearc.) PHILÁNTHIDÆ
	Mesopleura without such a furrow, the prepectus not separated;
	hind femora with a projection below at apex. (Figs. 952, 954).
	(Cercèris, cosmop.; Eucercèris, Am.) CERCÉRIDÆ
130.	Hind femora produced below at apex as a flattened tubercle
	which overlaps the base of the tibia; abdomen sessile. (Alyson,
	holarc., neotrop.; Bothynostèthus, Am.; Scapheùtes, S. Am.).
	ALYSÓNIDÆ
	Hind femora simple at apex, without projection
131.	Marginal cell not appendiculate and mandibles without an
2021	emargination externally; one, two, or three submarginal cells;
	eyes not emarginate on the inner edge. (Stigmus, Psén,
	widespr.; <b>Pemphrèdon</b> , <b>Passalècus</b> , holarc.). (Including
	MIMÉSIDÆ and PSÉNIDÆ) PEMPHREDÓNIDÆ
	Marginal cell appendiculate, or if not, the mandibles are emargi-
	nate externally
132.	
154.	
	usually aborted or deformed. (Figs. 955, 958, 960). (Táchytes,
	Lárra, Táchysphex, cosmop.; Palàrus, Lyròda, widespr.;
	Dinètus, Eur.; Sericophorus, Austr.) LARRIDÆ
	Second cubital cell petiolate, or rarely absent; three perfectly
	formed ocelli; small species. (Míscophus, widespr.; Plenó-
	culus, N. Am.). (NITÉLIDÆ) MISCÓPHIDÆ
133.	project satura
	propodeum above with a long spine or forked process; cubital
	and discoidal cells not distinctly separated. (Oxýbelus, cos-
	mop.; Belómicrus, holarc.) OXYBÈLIDÆ
	Postscutellum and propodeum simple, without scales or spine. 134

- 135. Radial cell appendiculate; black, usually with yellow markings. (Cràbro (including Rhópalum, Solènius, and many subgenera), cosmop., mainly holarc.; Anacràbro, N. Am.).

CRABRÓNIDÆ



Figs. 961-965. Hymenoptera

- 961. **Apis**, head. *at*, antenna; *md*, mandible; *lm*, labrum; *mx*, maxillary palpus; *lp*, labial palpus; *lb*, labium. (Cheshire) Apidæ.
- 962. Head of long-tongued bee (Cockerell).
- 963. Apis, hind leg of worker (Smith) Apidæ.
- 964, 965. Hind legs of bees (Riley).

	Tongue more or less elongate, pointed, not emarginate at the
	tip
138.	Fore wing with three closed cubital cells; moderate-sized, hairy
	bees; tongue split. (Collètes, widespr.; Megacilissa, Am.;
	Gastrópsis, Austr.) COLLÉTIDÆ
	Fore wing with only two closed cubital cells; small bees with very
	little hair, face almost always with white or yellow markings.
	(Prosòpis (= $Hylaus$ ), cosmop.; Hylæòides, Austr.). ( $HY$ -
	$L\!E\!ID\!E\!ID$ PROSOPÍDIDÆ
139.	The cheeks separating the eyes from the mandibles longer than
	the pedicel of the antennæ; large, social, densely hairy species,
	with contrasting black and yellow or sometimes also orange or
	red pile. Bumble-bees. ( <b>Bómbus</b> (=Brèmus), cosmop.; <b>Psí</b> -
	thyrus, holarc., neotrop.). (BRÉMIDÆ) BÓMBIDÆ
	Eyes nearly or quite reaching the base of the mandibles; solitary
	species
140.	Abdomen of female with a dense brush of pollen-collecting hairs
	on the underside; hind legs without a pollen-collecting appara-
	tus; pygidium of female large and broad, without a median flat
	area, very broad and sometimes toothed on the hind margin in
	the male; only two closed submarginal cells; labial palpi with the
	basal joints much elongated, the apical minute. Leaf-cutter
	bees. (Megachile, Lithúrgus, cosmop.; Trachùsa, Eur., Am.;
	Ósmia, Heriades, Anthídium, widespr.). MEGACHÍLIDÆ
	Abdomen of female without such a dense brush; pollen-collecting
	apparatus, if present, consisting of a covering of long, dense
	hairs on the hind legs or of an abnormal widening of the tibiæ;
	absent in the parasitic forms and considerably reduced in cer-
	tain others
141.	
	joint of the hind tarsi strongly enlarged and flattened; body
	conspicuously hairy, especially on the thorax and head; almost
	entirely non-parasitic forms
	No pollen-collecting apparatus present; first joint of hind tarsi
	not conspicuously enlarged or flattened; body without any
	dense vestiture of long hairs and often with a conspicuous
149	color pattern; parasitic species
142.	19 of the state of
	flat median area; tongue short, moderate, or elongate143
	Pygidium without a median triangular area, although sometimes

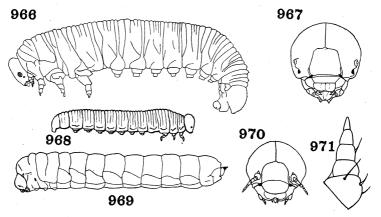
143. Tongue more or less short, dagger-like; radial cell pointed at apex; usually three submarginal cells; maxillary palpi usually six-jointed; pygidium of female with a triangular median area; burrowing bees, sometimes metallic in color. (Andrena, Sphecòdes, Halíctus, cosmop.; Augochlòra, Am.). ANDRÉNIDÆ Tongue elongate, though not so long as in the higher groups; almost always only two submarginal cells; radial cell blunt or truncate at apex, very rarely pointed; rather densely hairy; never brilliantly metallic, though rarely with the head and thorax green; often with yellow markings. (Panúrgus, Panurginus, Halictòides, holarc.: Pérdita, N. Am.), PANÚRGIDÆ Tongue elongate, very long; first two joints of labial palpi greatly lengthened and flattened, sheath-like, last two minute; generally large and densely hairy bees with well-developed pollen collecting apparatus; males often with long antennæ and usually with yellow clypeus. (Podalírius (= Anthóphora), Eùcera (including Melissòdes, Diadàsia, Émphor), cosmop.; Hemísia (= $C\acute{e}ntris$ ), widespr.). ( $ANTHOPH\acute{O}RID\cancel{E}$ ). PODALIRÎIDÆ 144. Hind tibiæ of female greatly widened and posteriorly expanded, smooth on the outer side. Large or moderate sized, often metallic, brilliantly ornamented species. (Euglóssa, Eulèma, neotrop.) . . . . . EUGLÓSSIDÆ Hind tibiæ without smooth corbicula, furnished with hairs for collecting pollen; very large or very small species. (Carpenter 145. Hind tibia and tarsus of female with a dense pollen-collecting scopa; stigma obsolete; large robust bees. (One tropicopolitan genus, Xylócopa) ...... XYLÓCÓPIDÆ Hind tibia and tarsus without distinct scopa, the hairs sparse; stigma large; small species. (Cerátina, widespr.; Allódape, Afr., As., Austr.) ...... CERATÍNIDÆ 146. Apical abdominal tergite of female large and frequently with a median carina; in the male usually emarginate or toothed; only two cubital cells; head large and broad; body generally coarsely sculptured; species of sombre colors. (Stělis, holarc.). STELÍDIDÆ Apical tergite of female not unusually large, sometimes very

- 147. Hind tibiæ greatly broadened, flattened above; large bees of brilliant blue or green metallic color. (Chrysántheda, S. Am.).

#### Larvae

At the present time it is impossible to give any reasonably complete or reliable key for the determination of the larvæ of the numerous families of Hymenoptera. Those of the suborder Chalastogastra have been quite extensively studied by several workers in Europe and in North America, and the grouping of these families below is based to a great extent on the work of Yuasa. Many of the larvæ of the other families have become so greatly specialized, modified, or reduced as a result of parasitism, maternal care, or from life as gall insects that the present, very incomplete knowledge affords no basis for a helpful key. Consequently references to habits have necessarily been introduced as differential characters, for the few of the higher forms that are included. It is thought, however, that the following summary will be helpful and reasonably reliable so far as it extends, although it must be borne in mind that the larve of only a small proportion of the species in any family are known, and that many families cannot be differentiated at all.

1. Body typically caterpillar-like (eruciform) (Figs. 966, 969), commonly with a color pattern; thoracic legs usually well developed, although sometimes much reduced; head much more strongly chitinized than the rest of the body; abdominal prolegs often present, entirely lost in some groups, but the body always retains an eruciform appearance. (Fig. 969). Antennæ and palpi almost always present and consisting of more than one joint. Mandibles strong, almost always with more than one tooth; ocelli frequently present. Alimentary canal continuous



Figs. 966-971. Larvæ of Hymenoptera

- 966. Neodiprion (Middleton) Diprionidæ.
- 967. Pteronidea, head (Yuasa) Tenthridinidæ.
- 968. Phyllotoma. Tenthredinidæ.
- 969. Tremex (Yuasa) Siricidæ.
- 970. Pamphilius, head (Yuasa) Pamphiliidæ.
- 971. Pteronidea, maxillary palpus (Middleton) Tenthredinidæ.

2.	Cerci present as distinctly segmented appendages
	Cerci absent, or present as unjointed vestiges4
3.	Cerci multiarticulate, setiform; larvæ feeding externally on leaves.
	a. Ocelli below and lateral to the antennæ. (Fig. 970).
	PAMPHILÌIDÆ
	b. Ocelli beloworneartheantennæ. MEGALODÓNTIDÆ
	Cerci two-jointed, larva mining in the petioles of ferns.
	BLASTICOTÓMIDÆ
4.	Abdominal prolegs present, well developed and distinctly jointed
	(except in a few leaf-mining forms)5
	Abdominal prolegs wanting; forms either feeding externally or
	boring in the stems or in woody plants or internal parasites
	of other insects; never leaf-miners
5.	Ten pairs of prolegs, one on each abdominal segment; antennæ
	with six or seven joints XYÉLIDÆ
	Six to eight pairs of abdominal prolegs (Fig. 968); reduced or
	absent in leaf-mining and gall-making forms; antennæ with
	five joints or less
6.	
	tarsal claws; prolegs usually well developed7
	Thoracic legs fleshy, indistinctly four-jointed; no tarsal claws;
	prolegs vestigial; larvæ leaf-miners.
	Some PHYLLOTOMINÆ; TENTHREDÍNIDÆ
7.	Prolegs present on abdominal segments 2-8 and 10; antennæ
	elongate, usually five-jointed8
	Prolegs absent on segment 8 or on both 7 and 8; if present on 8,
	the antennæ are one- or two-jointed
8.	Legs five-jointed
	Legs four-jointed
9.	Third abdominal segment divided dorsally by transverse grooves
	into six parts (annulets)
	Third abdominal segment dorsally with more or less than six annu-
	lets
10.	Antennæ conical; five-jointed
	Antennæ not conical; three-jointed, the third joint erect and peg-
	like. (Fig. 966) Most <b>DIPRIÓNIDÆ</b>
11.	
	the femora; tarsal claws short, strongly curved12
	Labrum distinctly asymmetrical, tibiæ longer than the femora;
	tarsal claws slender, only slightly curved.
	ρυντιόπόνη Ε. πενπυρερίκης Ε

12.	Body rather robust, of uniform diameter throughout, with small
	distinct tubercles; tenth abdominal segment usually with sev-
	eral small protuberances above.
	Some BLENNOCAMPINÆ; TENTHREDÍNIDÆ
	Body rather slender, tapering behind, without small distinct tu-
	bercles or protuberances.
	Some EMPHYTINÆ; TENTHREDÍNIDÆ
13.	Third segment with seven annulets on the dorsum; body without
	conspicuous branched spines or tubercles14
	Third segment with five, rarely three or four annulets on dorsum;
	body with conspicuous branched spines or tubercles.
	Some EMPHYTÎNÆ; TENTHREDÍNIDÆ
14.	Antennæ short, one-jointed; labrum divided into three parts by a
	pair of longitudinal sutures
	Antennæ five-jointed; labrum without a pair of longitudinal
	grooves
15.	Prolegs provided with setæ; clypeus with three setæ on each side;
	labrum without a median longitudinal impression. Some
	SELANDRIÌNÆ and EMPHYTÌNÆ; TENTHREDÍNIDÆ
	Prolegs without setæ; clypeus with two setæ at each side; labrum
	with or without median impression
16.	Tibia small, distinctly shorter than the femur.
	Some SELANDRIÌNÆ; TENTHREDÍNIDÆ
	Tibia usually subequal to or longer than the femur.
	TENTHREDINÎNÆ; TENTHREDÍNIDÆ
17.	Prothorax, mesothorax and last abdominal segment above not
	narrowed behind. Some EMPHYTINÆ; TENTHREDÍNIDÆ
	Body not thus ornamented; body often much narrowed behind.
	(Fig. 968) Some PHYLLOTOMINÆ; TENTHREDÍNIDÆ
18.	Thoracic legs five-jointed
	Thoracic legs with six joints, or with the first pair four-jointed
	and the others three-jointed
19.	
	Anal prolegs wanting25
20.	
	five joints
	Anal prolegs fused; antennæ one-jointed.
	SCOLIONEURÎNÆ; TENTHREDÎNIDÆ
21.	Antennæ with five joints; last abdominal segment with several
	protuberances above.
	Some HOLPLOCAMPINÆ; TENTHREDÍNIDÆ

	Antennæ with four, rarely three, joints; last abdominal segment
22.	without protuberances
44.	gland; body often with numerous prominent setæ23
	No eversible glands on the abdomen below; body never conspicu-
ດາ	ously setose
23.	
	setæ of different lengths; setæ microscopically barbed; larvæ external feeders on leaves. <b>CLADIÌNÆ</b> ; <b>TENTHREDÍNIDÆ</b>
	Body usually without setigerous tubercles and never with tubercles
	bearing setæ of more than one size. Larvæ frequently leaf-
04	rollers or gall-makers <b>NEMATÌNÆ</b> ; <b>TENTHREDÍNIDÆ</b>
24.	
	lets Some HOPLOCAMPÎNÆ; TENTHREDÍNIDÆ
	Antennæ one-jointed; third segment with three annulets. (Acor-
25	dulécera) Some <b>DIPRIÓNIDÆ</b> Antennæ three-jointed; third abdominal segment with four
25.	
	annulets; body not depressed; larvæ feeding externally or
	boring in fruits and leaf petioles.  HOPLOCAMPÎNÆ; TENTHREDÍNIDÆ
	Antennæ one- or two-jointed; third segment with two annu-
	lets; body depressed; larvæ leaf-miners.
	FENUSÎNÆ; TENTHREDÍNIDÆ
26.	Thoracic legs large, well developed and distinctly jointed; larvæ
20.	gregarious, feeding on leaves
	Thoracic legs vestigial, indistinctly jointed or entirely wanting;
	abdomen usually with a spine-like process at tip27
27	Ocelli present; antennæ with four or five joints; cerci present as
۷	minute one-jointed appendages; thoracic spiracles present;
	functional legs very small, indistinctly jointed CÉPHIDÆ
	Ocelli absent; antennæ with three joints or less; cerci entirely
	absent
28.	Antennæ with three joints. Larvæ wood-borers. <b>XIPHYDRÌIDÆ</b>
20.	Antennæ with only a single joint
29.	Metathoracic spiracles large, functional; abdomen with a spine at
20.	tip; larvæ wood-borers
	Metathoracic spiracles vestigial, non-functional; abdomen without
	a spine at tip; larvæ parasitic on wood-boring insects.
	ORÝSSIDÆ
30.	Parasitic species feeding internally in the eggs, nymphs, larvæ or
	pupæ of other insects

	Free living or feeding externally on other insects, spiders, etc. or
	on plant materials, sometimes within galls or in specially
	constructed cells or nests
31.	Egg parasites. Various CHALCIDÒIDEA; some SERPHÒIDEA
	especially <b>SCELIÓNID</b> Æ
	Parasites of nymphs, larvæ or pupæ or rarely of adult insects.
	ICHNEUMONOIDEA, many CHALCIDOIDEA, and SER-
	PHÒIDEA; IBALÌIDÆ; some FIGÍTIDÆ
32	Living in galls formed in the tissues of plants. CYNÍPIDÆ and
<b></b> .	some <b>FIGÍTIDÆ</b> that are parasitic on them; a few <b>CHALCI</b> —
	DÒIDEA (some EURYTÓMIDÆ and a few PERILÁMPIDÆ)
	Not in plant galls
99	
33.	
	insects or spiders, sucking the body fluids through openings that
	they cut through the integument, occasionally crawling in-
	side
	Not external feeders attached to the bodies of other insects or
	spiders35
34.	Food-insect and feeder living in the cell, burrow or cavity in which
	the food-insect normally occurs when not parasitized; i.e.
	the food has not been removed from its normal habitat. Most
	of the lower VESPÒIDEA (SCOLÌIDÆ, TIPHÌIDÆ, etc.);
	DRYÍNIDÆ (on Leaf-hoppers); RHOPALOSOMÁTIDÆ;
	PELECÍNIDÆ, BETHÝLIDÆ
	Food-insects or spiders and feeder in a cavity, burrow, or specially
	constructed cell in which the food has been placed after being
	stung by the mother wasp; the cells or individual nests single,
	or if several cells are attached together, each is closed after
	being supplied with food and the larvæ receive no further care.
	Solitary Wasps.
	SPHECÒIDEA, and most non-social VÉSPIDÆ
25	Solitary forms, living in specially constructed cells with a store of
55.	
	honey and pollen; the cells usually single, but if in small groups,
	each is separately sealed after completion and contains a single
	larva(APOÌDEA); a few VÈSPIDÆ (MASARIDÌNÆ)
	Social forms, living in communal nests together with numerous
	others and attended by adult ants, bees or wasps of the same
	species
36.	Living within hexagonal cells of papery material and fed by at-
	tendant wasps on malaxated insect-food Social VESPIDÆ
	Not in hexagonal papery cells37

Nests excavated in soil, in twigs, or other diverse locations, sometimes consisting of papery material or containing silk, but never with hexagonal cells; body usually provided with some simple, hooked or otherwise modified stiff setæ. Ants.

**FORMÍCIDÆ** 

# LITERATURE ON HYMENOPTERA, GENERAL

- André, E. and others. Spécies des Hyménoptères d'Europe et d'Algérie. 116 parts (1882–1920). (An extensive work, covering a number of families; we have not listed the parts separately).
- ASHMEAD, W. H. Hymenoptera. In Fauna Hawaiiensis (1901).
- BINGHAM, C. T. and C. Morley, Hymenoptera. Fauna of British India, 3 vols. (1897–1913).
- Bischoff, H. Biologie der Hymenopteren. Berlin, J. Springer, 598 pp. (1927).
- Börner, C. Stammesgeschichte der Hautflügler. Biol. Zentralbl., 39, pp. 145–186 (1919).
- CAMERON, P. Biologia centrali Americana, Hymenoptera, 1, pp. 1–487 (1883–1900); 2, pp. 1–143 (1888–1900).
  - Hymenoptera orientalia. Mem. Manchester Philos. Soc. (various parts) (1889–1903).
- CRESSON, E. T. Hymenoptera of Cuba. Proc. Entom. Soc. Philadelphia, 4, pp. 1–196 (1865).
  - Synopsis of Families and Genera of Hymenoptera of America North of Mexico. Am. Ent. Soc., 1887, 350 pp. (1887).
- Dalla Torre, K. Catalogus Hymenopterorum. 10 vols. Leipzig (1892–1902).
- Hedicke, H. Hymenoptera. In Tierwelt Mitteleuropas, 5, Lief. 1, pp. XI 1–246 (1930).
- Kirby, W. F. List of Hymenoptera of New Zealand. Trans. Entom. Soc. London, 1881, pp. 35-50 (1881).
- Mocsáry, A. Literatura Hymenopterorum. Termes. Füzetek, 6, pp. 7–122, (1882). (Extensive list, arranged alphabetically by authors). Hymenoptera. Fauna Regni Hungariæ, pp. 7–113 (1918).
- Schmiedeknecht, O. Die Hymenopteren Mitteleuropas. Jena (1930). (An earlier edition appeared in 1904).
- VIERECK, H. L., A. D. MACGILLIVRAY, C. T. BRUES, W. M. WHEELER, and S. A. ROHWER. The Hymenoptera or Wasp-like Insects of Connecticut. Bull. State Geol. Nat. Hist. Surv., Connecticut, No. 22 (1916).

#### SUBORDER CHALASTOGASTRA

- Ashmead, W. M. Classification of Horntails and Sawflies. (Chalastogastra). Canadian Entom., **30-36** (1898, 1903, 1904).
- Bradley, J. C. The Siricidæ of North America. Pomona Journ. Entom. and Zool., 5, pp. 1–30 (1913).
- Cameron, P. A Monograph of the British Phytophagous Hymenoptera. London, Ray Soc., 4 vols. (1882–1893).
- Enslin, E. Tenthredinoidea. In Insekten Mitteleuropas, 3, pp. 95–213, Stuttgart (1914).
- Konow, F. W. Lydidæ. Gen. Insectorum, fasc. 27, 27 pp. (1905).
  Siricidæ. Gen. Insectorum, fasc. 28, 14 pp. (1905).

Tenthredinidæ. Gen. Insectorum, fasc. 29, 176 pp. (1905).

- MORICE, F. D. Australian Sawflies. Trans. Entom. Soc. London, 1918, pp. 247–333 (1919).
- Nielsen, J. C. and K. Henriksen. Danish Siricidæ and Sawflies (in Danish). Danmarks Fauna, pt. 18, 234 pp. (1915).
- Rohwer, S. A. Genera of Pamphiliinæ. Canadian Entom., 42, pp. 215–220 (1910).

Classification of Chalastogastra. Proc. Entom. Soc. Washington, 13, pp. 215–226 (1911).

Studies in Oryssoidea. Proc. U. S. Nat. Mus., 43, pp. 141-158 (1913).

- Rohwer, S. A. and R. A. Cushman. Idiogastra, a new suborder of Hymenoptera. Proc. Ent. Soc. Wash., 19, pp. 89–98 (1917).
- Yuasa, H. A Classification of the Larvæ of the Tenthredinoidea. Illinois Biol. Monogr., 7, No. 4, Urbana (1922).

#### ICHNEUMONOIDEA

- Ashmead, W. H. Classification of the Ichneumon Flies or the Superfamily Ichneumonoidea. Proc. U. S. Nat. Mus., 23, 220 pp. (1900).
- AYYAR, T. V. R. Catalogue of Indian Braconidæ. Rept. Proc. 5th Entom. Meeting, Pusa, pp. 352–362 (1923).
- Baker, C. F. Chelonini of the Philippines (Braconidæ). Philippine Journ. Sci., 31, pp. 451–489 (1926).
- Berthoumieu, V. Ichneumonidæ, Ichneumoninæ. Gen. Insectorum, fasc. 18, 87 pp. (1904).
- Bradley, J. C. Family Roproniidæ. Entom. News, **16**, pp. 14–17 (1905). The Evaniidæ. Trans. American Entom. Soc., **34**, pp. 101–194 (1908).
- Brues, C. T. South African Evaniidæ, Braconidæ, Alysiidæ and Plumariidæ, with catalogue of species. Ann. South African Mus., 19, pp. 1–150 (1924).
  Ethiopian Braconidæ, with a List of the African Species. Proc. Amer. Acad. Arts Sci., 61, pp. 205–436 (1926).
- Cresson, E. T. Species of the Subfamily Ichneumoninæ (Nearctic). Trans. American Entom. Soc., 6, pp. 129–212 (1877).

Cushman, R. A. Revision of American Cremastini. Proc. U. S. Nat. Mus., 53, pp. 503-551 (1917).

North American Lycorini, Polysphinctini and Theroniini. (Ichneumonidæ). Proc. U. S. Nat. Mus., **58**, pp. 7–48 (1920).

Tribe Ephialtini. Proc. U. S. Nat. Mus., 58, pp. 327-362 (1920).

New Oriental and Australian Ichneumonidæ, with key to Genera of Cryptinæ. Philippine Journ. Sci., **20**, pp. 543–595 (1922).

Genera of Paniscini (Ichneumonidæ). Proc. U. S. Nat. Mus., **64**, Art. 20, 48 pp. (1924).

Revision of the North American Ichneumon-flies of Mesostenus and related genera. Proc. U. S. Nat. Mus., 74, Art. 16, 58 pp. (1929).

Cushman, R. A. and S. A. Rohwer. Holarctic tribes of Ichneumonina. Proc. U. S. Nat. Mus., 57, pp. 379–396 (1920).

North American Acænitini (Ichneumonidæ), Proc. U. S. Nat. Mus., **57**, pp. 503–523 (1921).

- DAVIS, G. C. A Review of the Tryphoninæ. Trans. American Entom. Soc., 24, pp. 193–348 (1897).
- DUTT, G. R. Stephanidæ. Cat. Indian Ins. Govt. India, Centr. Pub. Br., Calcutta, pt. 10, (1925).
- ELLIOTT, E. A. Monograph of the Stephanidæ. Proc. Zool. Soc. London, 1922, pp. 705–831 (1922).
- ENDERLEIN, G., Die Braconiden-Subfamilie Mimagathidinæ. Zool. Anz., 28, pp. 449–454 (1905).

Ueber die Klassification der Stephaniden. Zool. Anz., 28, pp. 473–477 (1905).

Zur Kenntnis aussereuropäischen Braconiden (Key to Genera of Agathidinæ). Arch. Naturg., Jahr. 87A, Heft 11, pp. 51–224 (1920).

Fahringer, J. Opuscula Braconologica. Vienna (1925–). A number of parts have appeared and the work is being continued.

Froggatt, W. W. Notes on Megalyra. Proc. Linn. Soc. New South Wales, 31, pp. 399-407 (1906).

Gahan, A. B. Aphididiinæ of North America. Bull. 152, Maryland Agric. Expt. Sta., pp. 147–200 (1911).

Revision of North American Opiinæ (Braconidæ). Proc. U. S. Nat. Mus., 49, pp. 63–95 (1916).

HOOKER, C. W. The Ichneumonidæ of the Tribe Ophionini. Trans. American Entom. Soc., 38, pp. 1–176 (1912).

Kieffer, J. J. Evaniidæ. Gen. Insectorum, fasc. 2, 13 pp. (1902).

Stephanidæ. Gen. Insectorum, fasc. 77, 10 pp. (1908).

Evaniidæ (including Aulacidæ and Gasteruptionidæ). Das Tierreich, Lief. 30, 432 pp. (1912).

Evaniiden der Philippinen. Philippine Journ. Sci., **11**D, pp. 317–346 (1914).

Kriechbaumer, J. Beitrag zu einer Monographie der Joppinen. Berliner Entom. Zeits., 43, pp. 1–166 (1899).

Keieger, R. Revision of Pimpla and Allies (Ichneumonidæ). Ber. naturf. Ges. Leipsig, Jahrg. 1897–98, pp. 47–124 (1898).

Ueber die Ichneumonidengattungen Neotheronia und Theronia. Zeits. Hym. Dipt., Jahrg. **5** and **6** (1905–06) (several parts).

Die Ichneumonidengattung Lissopimpla. Zeits. Hym. Dipt. 1907, pp. 294–302 (1907).

Marshall, T. A. Monograph of the British Braconidæ. Trans. Entom. Soc. London, 1885–1899 (seven parts) (1885–99).

Morley, C. Ichneumonidæ of Great Britain. 3 vols. (1903-08).

Catalogue of British Chalcididæ. London, 74 pp. (1910).

Revision of Ichneumonidæ. London, British Mus. Nat. Hist. 4 pts. (1912–15).

Notes on Braconidæ. XIII. Dacnusides. Entomologist, **57**, pp. 193–198; 250–255 (1924).

MUESEBECK, C. F. W. Revision of North American Apanteles. Proc. U. S. Nat. Mus., 58, No. 2349, pp. 483-576 (1920).

Revision of North American Neoneurinæ and Microgasterinæ. Proc. U. S. Nat. Mus., **61**, No. 2436, art. 15, pp. 1-76 (1922).

Revision of North American Microbracon (Braconidæ). Proc. U. S. Nat. Mus., 67, Art. 8, 85 pp. (1925).

Revision of Braconinæ of North America. Proc. U. S. Nat. Mus., 69, Art. 16, pp. 1–73 (1927).

Rohwer, S. A. Tribe Xoridini (Ichneumonidæ). Proc. U. S. Nat. Mus., **45**, pp. 353–361 (1913).

North American Cenoccelius (Braconidæ). Canadian Entom., pp. 316–322 (1914).

North American Labenini, Rhyssini, Xoridini, Odontomerini, Phytodietini (Ichneumonidæ). Proc. U. S. Nat. Mus., **57**, pp. 405–474 (1921).

Roman, A. Systematische Stellung von Neorhacodes. Entom Tidskr., **44**, pp. 169–174 (1923).

Schletterer, A. Revision of Gasteruptionidæ. Abh. zool.-bot. Ges. Wien, 35, pp. 267-326 (1885).

Die Hymenopteren Gruppe der Evaniiden. Ann. k. k. naturh. Hofmus. Wien, 4, pp. 107–180; 289–338, 373–546 (1889).

Revision of Stephanidæ. Berliner Entom. Zeits., **33**, pp. 71–160 (1889). Die Hymenopteren-Gattungen Stenophasmus, Monomachus, Pelecinus, Megalyra. Berliner Entom. Zeits., **33**, pp. 198–250 (1889).

Schmiedeknecht, O. Opuscula Ichneumonologica. Blankenburg (1902–1931) (numerous fascicles and additions dealing with palæarctic Ichneumonidæ). Ichneumonidæ, Pimplinæ. Gen. Insectorum, fasc. 62, 120 pp. (1908). Ichneumonidæ, Cryptinæ and Ophioninæ, part. Gen. Insectorum, fasc.

34, 71 pp.; fasc. 75, 165 pp.; fasc. 114, 100 pp. (1905, 1908, 1911).

Die Ichneumonidengattung Pimpla. Zeits. angew. Entom., **1**, pp. 396–478 (1914).

Schulz, W. A. Beitrag zur Kenntniss der Familie Pelecinidæ. S. B. Akad. Wiss., München, **33**, pp. 435–450 (1903).

Trigonalidæ. Gen. Insectorum, fasc. 61, 24 pp. (1908).

Systematische Uebersicht der Monomachiden. Proc. 1st Congr. Entom., Brussels, pp. 405–424 (1911).

- STRICKLAND, E. H. North American Pezomachini (Ichneumonidæ). Ann. Entom. Soc. America, 5, pp. 113-140 (1912).
- SZÉPLIGETI, G. V. Braconidæ. Gen. Insectorum, fasc. 22, 253 pp. (1904).
- Tosquinet, J. Ichneumonides d'Afrique. Mém. Soc. Entom. Belgique, 5, 430 pp. (1896).
- UCHIDA, T. Erster Beitrag zur Ichneumoniden Japans. Journ. Coll. Agric. Kokkaido Imp. Univ., 18, pp. 43–173 (1926).
- VIERECK, H. L. Type Species of the Genera of Ichneumon Flies. Bull. 83, U. S. Nat. Mus., pp. 1-186 (1914).

List of families and subfamilies of Ichneumonoidea. Proc. Biol. Soc. Washington, **31**, pp. 69–74 (1918); also ibid., **32**, p. 48 and 198 (1919).

Waterston, J. Dinapsidæ. Ann. Mag. Nat. Hist., (9) 10, pp. 418-420 (1922).

#### CHALCIDOIDEA

- Ashmead, W. H. Classification of Chalcid Flies or the superfamily Chalcidoidea. Mem. Carnegie Mus., Pittsburg, 1, No. 4, pp. 225–551 (1904).
- AYYAR, T. V. R. Check list of Indian Chalcidoidea. Spolia Zeylandica, 13, pp. 235-254 (1925).
- Crawford, J. C. Synopsis of North American Perilampidæ. Proc. Entom. Soc. Washington, 16, pp. 69–76 (1914).
- Gahan, A. B. and M. M. Fagan. Type Species of Genera of Chalcidoidea. Bull. U. S. Nat. Mus., No. 124, 173 pp. (1923).
- Girault, A. A. Australian Hymenoptera Chalcidoidea. Mem. Queensland Mus., 365 pp. (1912–16). (Numerous parts and supplements dealing with all the families).

Systematic monograph of the Signiphorinæ. Proc. U. S. Nat. Mus., 45, pp. 189–233 (1913).

Revision of Trichogrammatidæ. Bull. Wisc. Soc., **11**, pp. 150–179; **12**, pp. 59–71 (1913–14).

- Grandi, G. Catalogo dei Agaonidi. Boll. Lab. Entom. Instit. Sup. Agrar., Bologna, 1, pp. 107–235 (1928).
- Howard, L. O. New Genera and Species of Aphelininæ, with a Revised Table of Genera. Bull. U. S. Dept. Agric. Tech. Ser., No. 12, pt. 4 (1907).
- Kirby, W. F. Synopsis of Genera of Eucharididæ. Journ. Linn. Soc., London, Zool., 20, pp. 28–37 (1886).
- Kryger, J. P. The European Trichogrammatidæ. Entom. Meded., 12, pp. 257–354 (1918–19).
- MAYR, G. Die europäischen Torymiden. Verh. zool.-bot. Ges. Wien, 24, pp. 53–142 (1874).

Die europäischen Encyrtiden. Verh. zool.-bot. Ges. Wien, **25**, pp. 676–778 (1875).

- MERCET, R. G. Fauna Iberica; Encírtidos. Mus. Nac. Cienc. Nat. Madrid, 732 pp. (1921).
- Ruschka, F. Die europäischen Arten der mit Monodontomerus verwandte Gattungen. Zeits. angew. Entom., 9, pp. 395–408 (1923).

Die europäischen Eucharidinæ und Perilampinæ. Deutsch. Entom. Zeits., 1924, pp. 82–96 (1924).

- Schletterer, A. Leucospididæ. Berliner Entom. Zeits., **35**, pp. 141–302 (1890).
- SCHMIEDEKNECHT, O. Chalcidoidea. Gen. Insectorum, fasc. 97, 550 pp. (1909). Weld, C. J. Revision of American Leucospidide. Proc. U. S. Nat. Mus., 61, art. 6, pp. 1–43 (1922).

#### SERPHOIDEA

ASHMEAD, W. H. A Monograph of the North American Proctotrypidæ. Bull. U. S. Nat. Mus., No. 45 (1893).

Classification of the pointed-tailed wasps or the superfamily Proctotrupoidea. Journ. New York Entom. Soc., **10**, p. 241; **11**, pp. 28 and 86 (1902–03).

- Brues, C. T. Scelionidæ. Gen. Insectorum, fasc. 80, 59 pp. (1908), with supplement by J. J. Kieffer.
- Crawford, J. C. A new family of parasitic Hymenoptera (Vanhorniidæ). Proc. Entom. Soc. Wash., **2**, pp. 63–64 (1909).
- FOUTS, R. M. Revision of North American Platygasterinæ. Proc. U. S. Nat. Mus., 63, art. 15a, pp. 1–145 (1924).
- KIEFFER, J. J. Revision des Scélionides. Ann. Soc. Sci. Brussels, 32, pp. 111–250 (1908).

Ceraphronidæ. Gen. Insectorum, fasc. 94, 27 pp. (1909).

Serphidæ. Gen. Insectorum, fasc. 95, 10 pp (1909).

Belytidæ. Gen. Insectorum, fasc. 107, 47 pp. (1910).

Diapriidæ. Gen. Insectorum, fasc. 124, 75 pp. (1911).

Serphidæ. Das Tierreich, Lief. 42, 254 pp. (1914).

Diapriidæ. Das Tierreich, Lief. 44, 627 pp. (1916).

Scelionidæ. Das Tierreich, Lief. 48, 885 pp. (1926).

#### CYNIPOIDEA

- Ashmead, W. H. Classification of Cynipoidea. Psyche, 10, pp. 7, 59, 140 (1903).
- Dalla Torre, W. and J. J. Kieffer. Cynipidæ. Gen. Insectorum, fasc. 9–10, 84 pp. (1902).

Cynipidæ. Das Tierreich, Lief. 24 (1910).

- Fullaway, D. T. Monograph of the Gall making Cynipidæ of California. Ann. Entom. Soc. America, 4, pp. 331–380 (1911).
- Kieffer, J. J. Cynipidæ. In Insekten Mitteleuropas, 3, pp. 1–94, Stuttgart (1914).
- KINSEY, A. C. New Species and Synonymy of American Cynipidæ. Bull. American Mus. Nat. Hist., 42, pp. 293–317 (1920).
  - Phylogeny of Cynipid Genera and Biological Characteristics. Bull. American Mus. Nat. Hist., 42, pp. 357–402 (1920).
- ROHWER, S. A. and M. M. FAGAN. The type species of the genera of the Cynipoidea. Proc. U. S. Nat. Mus., **53**, pp. 357–380 (1917); **55**, pp. 237–240 (1919).
- Weld, L. H. Cynipidæ; Genera of Liopterinæ and Oberthuerellinæ. Philippine Journ. Sci., 21, pp. 323–335 (1922).

## ACULEATA, GENERAL

- ASHMEAD, W. H. Classification of the fossorial, predaceous and parasitic wasps or the superfamily Vespoidea. Canad. Entom., **32**, **34**, **35**, **36** (1900–04). (Various parts published at intervals).
- Berland, L. Hyménoptères Vespiformes. (Vespidæ, Bethylidæ, Dryinidæ, Embolemidæ). Faune de France, 19, pt. 2, 208 pp., Paris (1928).

## BETHYLOIDEA

- Ashmead, W. H. Rhopalosomidæ, a new family of Wasps. Proc. Entom. Soc. Washington, 3, pp. 303–309 (1896).
- Brues, C. T. Note on the Hymenopterous family Rhopalosomatidæ. Psyche, **33**, pp. 18–19 (1926).
- Fullaway, D. T. Revision of Hawaiian Sierola (Bethylidæ). Occ. Pap. Bishop Mus., Honolulu, 7, pp. 57–159 (1920).
- Kieffer, J. J. Dryinidæ. Gen. Insectorum, fasc. 54, 33 pp. (1907). Bethylidæ. Gen. Insectorum, fasc. 76, 50 pp. (1908).

Bethylidæ. Das Tierreich, Lief. 41, 595 pp. (1914).

Perkins, R. C. L. Leaf Hoppers and their Natural Enemies. Dryinidæ. Bull. Hawaiian Sugar Planters Expt. Sta., No. 1, pt. 1 (1905).

#### CHRYSIDOIDEA

- AARON, S. F. The North American Chrysididæ. Trans. American Entom. Soc., 12, pp. 209–248 (1885).
- Bischoff, H. Chrysididæ. Gen. Insectorum, fasc. 151, 86 pp. (1913).
- Bridwell, J. C. Family Alienidæ. Proc. Hawaiian Entom. Soc., 4, p. 117 (1919).
- Ducke, A. As Chrysididas do Brazil. Cat. Faun. Brasiliera Mus. Paulista, São Paulo, 4, pp. 1–31 (1913).

- Mocsáry, A. Monographia Chrysidarum Orbis Terrarum Universi. Budapest, Soc. Franklinianæ (1889).
- Trautmann, W. Die Goldwespen Europas. Weimar, 194 pp. (1927).

#### SCOLIOIDEA

- André, E. Mutillidæ. Gen. Insectorum, fasc. 11, pp. 77 (1903). Bibliographie générale des Mutillides. Rev. Entom., **29**, pp. 134–166 (1910).
- Betrem, J. C. Monographie der Indo-australischen Scoliiden. Treubia, 9, suppl. 388 pp. (1928).
- BISCHOFF, H. Konowiellidæ (Plumariidæ). Entom. Rundschau, Jahrg. 31, pp. 67-69 (1914).
  Monographie der Mutilliden Afrikas. Arch. Naturg., Jahrg. 86A, Heft. 1-5, 830 pp. (1920-21).
- BLAKE, C. A. Monograph of the Mutillidæ of North America. Trans. American Entom. Soc., 13, pp. 179–286 (1886).
- Bradley, J. C. Monograph of North American Mutillidæ. Trans. American Entom. Soc., **42**, pp. 187–214; 309–336 (1916).
  - Review of the Myrmosidæ. Trans. American Entom. Soc., 43, pp. 247–290 (1917).
- Bradley, J. C. and J. Bequaert. Revision of Mutillidæ of Belgian Congo. Bull. American Mus. Nat. Hist., **58**, pp. 63–122 (1928).
- Cresson, E. T. Synopsis of North American species of Sapyga. Trans. American Entom. Soc., 8, pp. xx–xxi (1880).
- Ducke, A. O Genero Pterombrus. Rev. Mus. São Paulo, 9, pp. 107–122 (1914).
- Fox, W. J. The North American Mutillidæ. Trans. American Entom. Soc., 25, pp. 219–292 (1899).
- Guiglia, D. Gli Scoliidei della Liguria. Ann. Mus. Stor. Nat. Genoa, **52**, pp. 424–460 (1928).
- MALLOCH, J. R. Systematic notes on North American Brachycistinæ. Proc. U. S. Nat. Mus., 68, Art. 3, pp. 1–28 (1926).
- MICHA, I. Beitrag zur Kenntnis der Scoliiden. Mitt. Zool. Mus. Berlin, 13, pp. 1–156 (1927).
- MICKEL, C. E. Monograph N. Am. Dasymutilla. Bull. U. S. Nat. Mus., No. 134. pp. 39-340 (1928).
- Turner, R. E. Revision of the Thynnidæ of Australia. Proc. Linn. Soc. New South Wales, **32**, pp. 206–290 (1907).
  - Thynnidæ. Gen. Insectorum, fasc. 105, 62 pp. (1910).

#### FORMICOIDEA

- Arnold, G. Monograph of the Formicidæ of South Africa. Ann. South African Mus., **23**, pp. 191–295 (1926). Also **14**, pp. 159–270 (1916).
- BINGHAM, C. T. Ants of British India. Fauna of British India, Hymenoptera, vol. 2 (1903).

- Borgmeier, T. Catálogo systemático e synonymico das Formigas do Brasil. Arch. Mus. Nac. Rio de Janeiro, **24**, pp. 33–103 (1924) and ibid., **27**, pp. 69–164 (1927).
- Bruch, C. Catálogo de los Formícidos Argentinos. Rev. Mus. La Plata, 19, pp. 211-234; 527-537 (1915).
- Donisthorpe, H. St. J. K. British Ants, their Life History and Classification, London, Routledge & Sons, 1927, 436 pp.
- EMERY, C. Formicidæ, Dorylinæ. Gen. Insectorum, fasc. 102, 34 pp. (1910).
  Formicidæ, Ponerinæ. Gen. Insectorum, fasc. 118, 125 pp. (1911).
  Formicidæ, Dolichoderinæ. Gen. Insectorum, fasc. 137, 50 pp. (1913).

Fauna entomologica italiana, Formicidæ. Bull. Soc. Entom. Ital., 47, pp. 79–275 (1916).

Formicidæ, Myrmicinæ. Gen. Insectorum, fasc. 174, 174A and B, 206 pp. (1921–22).

Formicidæ, Formicinæ. Gen. Insectorum, fasc. 183, 302 pp. (1925).

Forel, A. Les Formicides. In Grandidier, Hist. Madagascar, 20, pt. 2, 280 pp. (1891).

Les Formicides de l'Empire des Indes et de Ceylon. Journ. Bombay Nat. Hist. Soc., 8 and 9 (several parts). (1894–95).

Die Ameisen der Schweiz. Mitt. schweizerischen Entom. Ges. suppl. to 12, 77 pp. (1915).

Cadre synoptique des Fourmis. Bull. Soc. Vaud., **51**, pp. 229–253 (1917). LUEDERWALDT, H. Chave para determinar os Dorylineos Brasilieros. Rev.

Mus. Paulista, **12**, pp. 229–257 (1920). Stitz, H. Formicidæ. Die Insekten Mitteleuropas, **2**, pp. 1–111, Stuttgart

(1914).
WHEELER, W. M. Ants, their Structure and Behavior. 663 pp. Columbia University Press, New York (1910).

Keys to the Genera and Subgenera of Ants. Bull. American Mus. Nat. Hist., New York, **45**, pp. 631–710 (1922).

WHEELER, W. M. and J. W. CHAPMAN. The Ants of the Philippine Islands. Pt. 1. Dorylinæ and Ponerinæ. Philippine Journ. Sci., 28, pp. 47-71 (1925).

#### VESPOIDEA

- Bequaert, J. Revision of the Vespidæ of the Belgian Congo. Bull. American Mus. Nat. Hist., **39**, pp. 1–384 (1918).
- Bradley, J. C. Taxonomy of the Masarid wasps, including a monograph of the North American Species. Univ. California Publ., Tech. Bull., 1, pp. 369–464 (1922).
- Buysson, R. Monographie des Vespides. Ann. Soc. Entom. France, **73** and **74**, several parts, incomplete (1904–06).
- Dalla Torre, K. W. Vespidæ. Gen. Insectorum, fasc. 19, 108 pp. (1904).
  Ducke, A. Révision des guêpes sociales polygames. Ann. Mus. Nat. Hungarici,
  8, pp. 449-544 (1910).

Ueber Phylogenie und Klassifikation der sozialen Vespiden. Zool. Jahrb. Abth. f. Syst., **36**, pp. 303–330 (1914).

Catálogo das Vespas Sociaes do Brazil. Rev. Mus. Paulista, 10, pp. 314–374 (1918).

- Isely, D. Synopsis of North American Eumenidæ. Ann. Entom. Soc. America, **10**, pp. 345–366 (1917).
- Saussure, H. Synopsis of American wasps. Misc. Coll. Smithsonian Institution, Washington, No. 254 (1875).
- ZAVATTARI, E. Materialien zu einer Monographie der Neotropischen Eumeniden. Arch. f. Naturg., 78A, Heft 4, pp. 1-272 (1912).

#### PSAMMOCHAROIDEA

- Banks, N. Psammocharidæ: Classification and Descriptions. Journ. New York Entom. Soc., 19, pp. 219–237 (1911).
- HAUPT, H. Monographie der Psammocharidæ von Mittel Nord- und Osteuropa. Beiheft, Deuts. Entom. Zeits. (1926–27).

Weiterer Ausbau meines Systems der Psammocharidæ. Mitt. Zool. Mus. Berlin, **15**, pp. 109–197 (1929).

Classification of Psammocharidæ with two cubital cells. Mitt. Zool. Mus. Berlin, **16**, pp. 673–797 (1930).

- Lucas, R. Die Pompilidengattung Pepsis. Berliner Entom. Zeits., **39**, pp. 449–480 (1894).
- Sustera, O. Die paläarktischen Gattungen der Psammocharidæ. Verh. zool.-bot. Ges. Wien, **62**, pp. 171–213 (1912).

#### SPHECOIDEA

- Arnold, G. The Sphegidæ of South Africa. Ann. Transv. Mus., 9, 10, 12, 13, 14 (1922–31).
- Ashmead, W. H. Classification of Sphegoidea. Canadian Entom., **31**, p. 145, 161, 212, 238, 291, 345 (six parts) (1899).
- Bouwman, B. E. De Graafwespen van Nederland. Levende Natuur, Amsterdam, 31, 32, 33 (numerous parts) (1927–28).
- Bradley, J. C. North American Nyssonidæ. Trans. American Entom. Soc., 46, pp. 113–132 (1920).
- Cresson, E. T. Monograph of the Philanthidæ of North America. Proc. Entom. Soc. Philadelphia, 5, pp. 85–132 (1865).
- Fernald, H. T. The Digger Wasps of North America, subfamily Chlorioninæ. Proc. U. S. Nat. Mus., **31**, pp. 291–423 (1906).
- Fox, W. J. The North American Pemphredonidæ. Trans. American Entom. Soc., 19, pp. 307–326 (1892).

The North American Larridæ. Proc. Acad. Nat. Sci. Philadelphia, 1893, pp. 467–551 (1894).

The Crabronidæ of Boreal America. Trans. American Entom. Soc., 22, pp. 129–226 (1895).

Kohl, F. Zur Kenntnis der Pemphredonen. Ann. naturh. Hofmus. Wien, **5**, pp. 49-65 (1890).

Die Larriden. Verh. zool.-bot. Ges. Wien,  $\bf 34$ , pp. 171–268, 327–454 (1885).

Monographie der Gattung Sphex. Ann. naturh. Hofmus. Wien, 5, pp. 77–194, 317–462 (1890).

Die Hymenopterengattung Philanthus. Ann. naturh. Hofmus. Wien, **6**, pp. 345-370 (1891).

Über Ampulex Jur. und verwandten Hymenopterengattungen. Ann. naturh. Hofmus. Wien, 8, pp. 455–516 (1893).

Zur Monographie der Gattung Sphex. Ann. naturh. Hofmus. Wien,  $\mathbf{10}$ , pp. 42–74 (1895).

Die Gattungen der Sphegiden. Ann. naturh. Hofmus. Wien, **11**, pp. 233–516 (1896).

Monographie der Gattung Podium. Abh. zool.-bot. Ges. Wien, 1, Heft 4, pp. 1–101 (1902).

Monographie der Gattung Ammophila. Ann. naturh. Hofmus. Wien, 21. pp. 228–382 (1906).

Die Crabronen der paläarktischen Region. Ann. naturh. Hofmus. Wien, 29, pp. 1–453 (1915).

Die Hautflüglergruppe der Sphecinen. IV. Sceliphron. Ann. naturh. Hofmus. Wien, **32**, pp. 1–170 (1918).

MICKEL, C. E. A Synopsis of the Sphecoidea of Nebraska (Hymenoptera). Nebraska Univ. Studies, 17, pp. 342–456 (1918).

Parker, J. B. A Revision of the Bembicine Wasps of America North of Mexico. Proc. U. S. Nat. Mus., **52**, pp. 1–155 (1917).

A Generic Revision of the Fossorial Wasps of the tribes Stizini and Bembecini. Proc. U. S. Nat. Mus., **75**, art. 5, 203 pp. (1929).

- Pate, V. S. L. Studies in Oxybeline Wasps. Entom. News, **40**, pp. 219–222 (1929).
- Rohwer, S. A. North American genera of Nyssonidæ. Proc. U. S. Nat. Mus., **59**, pp. 403–413 (1921).
- Schletterer, A. Die Hymenopteren Gattung Cerceris. Zool. Jahrb. Abth. Syst., 2, pp. 349-510 (1887).
- WILLIAMS, F. X. The Larridæ of Kansas. Kansas Univ. Sci. Bull., 8, pp. 117–213 (1913).

Larridæ of the Philippines. Bull. Hawaiian Sugar Planters Assoc., No. 19, pp. 61–101 (1928).

#### APOIDEA

- ALFKEN, J. D. Die Bienenfauna von Ostpreussen. Schrift. Königsberg, 53, pp. 114–182 (1912).
- ASHMEAD, W. H. Classification of Apoidea. Trans. American Entom. Soc., 26, pp. 49–100 (1899).

Bischoff, H. Zur Kenntnis afrikanischer Schmartozerbienen. Deuts. Entom. Zeits., 1923, pp. 585–603 (1923).

Blüthgen, P. Beiträge zur Kenntnis der afrikanischen Halictinæ. Zool. Jahrb. Abth. f. Syst., **55**, pp. 163–252 (1928).

Cockerell, T. D. A. North American Anthophoridæ. Trans. American Entom. Soc., **32**, pp. 63–116 (1906).

Genera of Colorado Bees. Univ. Colorado Stud., 16, pp. 99-126 (1928).

Ducke, A. Die Bienengattung Osmia. Ber. naturf. Ver. Innsbruck, **25**, pp. 1–323 (1900).

Franklin, H. J. The Bombidæ of the New World. Trans. American Entom. Soc., **38**, pp. 177–486; **39**, pp. 73–200 (1912–13).

FRIESE, H. Monographie der Gattung Ceratina. Termes. Füzetek, 19, pp. 34–65 (1896).

Monographie der Gattung Panurgus. Termes. Füzetek,  $\mathbf{20}$ , pp. 78–102 (1897).

Monographie der Gattung Megacilissa. Ann. Hofmus. Wien, 13, pp. 59–86 (1908).

Monographie der Gattung Euglossa. Termes. Füzetek, **21**, pp. 117–172 (1899).

Monographie der Gattung Exomalopsis und Tetrapedia. Ann. Hofmus. Wien, **14**, pp. 247–304 (1899).

Monographie der Gattung Centris. Ann. Hofmus. Wien, **16**, pp. 237–350 (1901).

Die Bienen Afrikas. Jena. Denkschr., 14, pp. 85-475 (1909).

Die Bienenfauna von Neuguinea. Ann. Mus. Hist. Nat. Hungarici, 7, pp. 179–288 (1909).

Megachilidæ. Das Tierreich, Lief. 28, 440 pp. (1911).

Die europäischen Bienen. Berlin (1922–23).

Frison, T. H. The Bumblebees of the Philippine Islands. Philippine Journ. Sci., 27, pp. 113–121 (1925); 37, pp. 273–281 (1928).

Kirby, W. F. Monographia Apum Angliæ. 2 vols. Ipswich (1882).

LUTZ, F. E. and T. D. A. COCKERELL. Distribution and bibliography of North American Bees (Apidæ, Meliponidæ, Bombidæ, Euglossidæ, Anthophoridæ). Bull. American Mus. Nat. Hist., **42**, pp. 491–641 (1920).

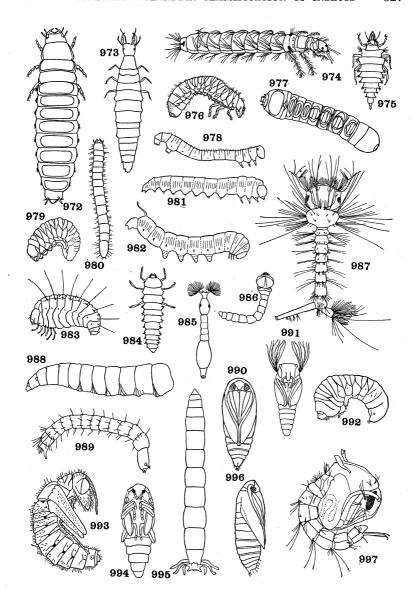
Maidl, F. Die Xylocopen des Wiener Hofmuseums. Ann. naturf. Hofmus. Wien, **26**, pp. 249–330 (1912).

MEYER, R. Monogaphie der Apidæ-Sphecodinæ. Arch. f. Naturg., Jahrg. 85A, Heft 1–2, pp. 79–242 (1920).

ROBERTSON, C. Synopsis of Panurgidæ. Psyche, 29, pp. 159-173 (1922).

SCHMIEDEKNECHT, O. Apidæ Europæ. 2 vols., 1112 pp. Berlin (1882–84).

Smith, H. S. Revision of Ceratinidæ of North and Middle America. Trans. American Entom. Soc., 33, pp. 115–124 (1907).



# Figs. 972-997. Larvæ and Pupæ of Various Insects

- 972. Calosoma, larva (Duncan) Coleoptera, Carabidæ.
- 973. Dytiscus, larva (Maxwell-Lefroy) Coleoptera, Dytiscidæ.
- 974. Caddis worm, larva (Duncan) Trichoptera, Phryganeidæ.
- 975. Chrysopa, larva (Chittenden) Neuroptera, Chrysopidæ.
- 976. Phyllophaga, larva (Forbes) Coleoptera, Melolonthidæ.
- 977. **Mallodon**, larva, ventral view. Coleoptera, Prionidæ.
- 978. Geometrid larva (Packard) Lepidoptera, Geometridæ.
- 979. **Hylastinus**, larva (Chittenden) Coleoptera, Scolytidæ.
- 980. Melanotus, larva (Forbes) Coleoptera, Elateridæ.
- 981. Saw-fly, larva (Maxwell-Lefroy) Hymenoptera, Tenthredinoidea.
- 982. Acherontia, larva (Maxwell-Lefroy) Lepidoptera, Sphingidæ.
- 983. Mylabris, larva (Howard) Coleoptera, Mylabridæ.
- 984. Anatis, larva (Britton) Coleoptera, Coccinellidæ.
- 985. Simulium, larva (Osborn) Diptera, Simuliidæ.
- 986. Chrysobothris, larva (Chittenden) Coleoptera, Buprestidæ.
- 987. Culex, larva (Dyar) Diptera, Culicidæ.
- 988. Musca, larva (Howard) Diptera, Muscidæ.
- 989. Pulex, larva (Chittenden) Siphonaptera, Pulicidæ.
- 990. **Gelechia,** pupa, underside (Hunter) Lepidoptera, Gelechiidæ.
- 991. Simulium, pupa (Miall) Diptera, Simuliidæ.
- 992. Lyctus, larva (Hopkins) Coleoptera, Lyctidæ.
- 993. Sialis, pupa (Davis) Megaloptera, Sialididæ.
- 994. Cyllene, pupa (Hopkins) Coleoptera, Cerambycidæ.
- 995. Tipula, larva (Needham) Diptera, Tipulidæ.
- 996. Gelechia, pupa, ventral side (Hunter) Lepidoptera, Gelechiidæ.
- 997. Culex, pupa (Knab) Diptera, Culicidæ.

# PART II

# OTHER TERRESTRIAL ARTHROPODA

# CONSPECTUS OF THE HIGHER GROUPS OF TERRESTRIAL ARTHROPODS (EXCEPT INSECTS)

# Class ONYCHOPHORA

Order MALACOPODA (Peripatidæ, Peripatopsidæ)

# Class ARACHNIDA

#### Order MICROTHELYPHONIDA

(Kæneniidæ)

## Order PEDIPALPIDA

# Suborder Uropygi

(Thelyphonidæ, Schizomidæ)

# Suborder Amblypygi

(Tarantulidæ, Charontidæ, Phrynidæ)

## Order RICINULEI

(Ricinoididæ)

# Order SCORPIONIDA

(Buthidæ, Scorpionidæ, Ischnuridæ, Diplocentridæ, Chactidæ, Vejovidæ, Bothriuridæ)

# Order SOLPUGIDA

(Galeodidæ, Solpugidæ, Hexisopodidæ)

#### Order CHELONETHIDA

Suborder Heterosphyronida (Chthoniidæ)

# Suborder Diplosphyronida

Neobisioidea (Syarinide, Neobisiide, Hyide, Ideoroncide)

GARYPOIDEA (Menthidæ, Olpiidæ, Garypodidæ)

# Suborder Monosphyronida

Cheliferidæ, Feaellidæ)

#### Order PHALANGIDA

Suborder Cyphophthalmi (Sironidæ)

Suborder Laniatores (Oncopodidæ, Triænorhynchidæ, Assamiidæ, Phalangodidæ, Cosmetidæ, Gonyleptidæ)

# Suborder Palpatores

PHAL/ NGIOIDEA (Phalangiidæ)

Nemastosomatoidea (Acropsopilionidæ, Trogulidæ, Nemastosomatidæ, Ischyropsalidæ)

#### Order ARANEIDA

# Suborder Liphistiomorphæ

LIPHISTIOIDEA (Liphistiidæ)

# Suborder Avicularimorphæ

ATYPODEA (Ctenizidæ, Atypidæ, Migadidæ, Dipluridæ, Paratropididæ, Pycnothelidæ)

AVICULARIOIDEA (Barychelidæ, Aviculariidæ)

# Suborder Arachnomorphæ

Dysderidæ, (Hypochilidæ, Filistatidæ, Dysderidæ, Oonopidæ, Hadrotarsidæ, Telemidæ, Caponiidæ)

Argiopoidea (Œcobiidæ, Urocteidæ, Psechridæ, Tengellidæ, Amaurobiidæ, Agelenidæ, Hersiliidæ, Zodariidæ, Palpomanidæ, Eresidæ, Pisauridæ, Lycosidæ, Oxyopidæ, Senoculidæ, Leptonetidæ, Sicariidæ, Pholcidæ, Dictynidæ, Theridiidæ, Linyphiidæ, Archæidæ, Uloboridæ, Dinopidæ, Argiopidæ, Mimetidæ)

Clubionoidea (Zoropsidæ, Acanthoctenidæ, Ctenidæ, Drasidæ, Ammoxenidæ, Prodidomidæ, Homalonychidæ, Sparassidæ, Selenopidæ, Platoridæ, Thomisidæ, Aphantochilidæ, Clubionidæ, Attidæ)

#### Order ACARINA

EUPODOIDEA (Cryptognathidæ, Bdellidæ, Eupodidæ)

Trombidiodea (Trombidiidæ, Anystidæ, Erythræidæ, Cæculidæ, Tetranychidæ, Cheyletidæ)

Hydrachnoidea (Halacaridæ, Hydrachnidæ, Hydrovoltziidæ, Limnocharidæ, Eylaidæ, Thermacaridæ, Protziidæ, Sperchonidæ, Pseudohydryphantidæ, Hydryphantidæ, Limnesiidæ, Arrhenuridæ, Lebertiidæ, Atractididæ, Hygrobatidæ, Brachypodidæ, Ewingiidæ)

IXODOIDEA (Argantidæ, Ixodidæ)

Parasitoidea (Holothyridæ, Spelæorhynchidæ, Spinturnicidæ, Halarachnidæ, Uropodidæ, Parasitidæ, Dermanyssidæ)

Oribatoidea (Labistommatidæ, Hoplodermatidæ, Oribatidæ)

Acaroidea (Tarsonemidæ, Canestrinidæ, Tyroglyphidæ, Listrophoridæ, Analgesidæ, Cytoleichidæ, Acaridæ)

Demodicoidea (Ériophyidæ, Demodicidæ)

#### Order PENTASTOMIDA

CEPHALOBÆNOIDEA (Cephalobænidæ) LINGUATULOIDEA (Linguatulidæ)

#### Class TARDIGRADA

#### Order **HETEROTARDIGRADA**

(Halechiniscidæ, Echiniscidæ)

#### Order EUTARDIGRADA

(Milnesiidæ, Macrobiotidæ)

#### Class PAUROPODA

#### Order HETEROGNATHA

(Brachypauropodidæ, Pauropodidæ, Eurypauropodidæ)

## Class DIPLOPODA

Subclass PSELAPOGNATHA

(Polyxenidæ)

Subclass CHILOGNATHA

Division OPISTHANDRIA

Order LIMACOMORPHA

(Glomeridesmidæ)

Order ONISCOMORPHA

(Sphærotheriidæ, Glomeridæ, Gervaisiidæ, Glomeridellidæ)

Division PROTERANDRIA

Superorder EUGNATHA

## Order POLYDESMOIDEA

Suborder **Polydesmidea** (Polydesmidæ, Mastigodesmidæ, Cryptodesmidæ, Stylodesmidæ, Oniscodesmidæ, Vanhoeffeniidæ, Peridontodesmidæ)

Suborder **Strongylosomatidea** (Strongylosomatidæ, Sphærotrichopidæ, Leptodesmidæ, Oxydesmidæ, Platyrhacidæ, Gomphodesmidæ, Sphæriodesmidæ, Rhacodesmidæ, Fontariidæ)

#### Order **NEMATOPHORA**

Suborder **Chordumidea** (Trachysomatidæ, Chamæosomatidæ, Metopidothrigidæ, Conotylidæ, Diplomaragnidæ, Caseyidæ, Underwoodiidæ, Chordeumidæ, Orobainosomatidæ, Faginidæ, Heteroporatiidæ, Verhæffiidæ, Heterolatzelidæ, Anthogonidæ, Brachychæteumidæ, Anthroleucosomatidæ, Neatractosomatidæ, Pseudoclididæ, Rothenbuehleriidæ, Attemsiidæ, Opisthocheiridæ, Haplobainosomatidæ, Craspedosomatidæ)

Suborder **Stemmiulidea** (Stemmiulidæ)

Suborder **Striariidea**(Striariidæ)

Suborder **Lysiopetalidea** (Lysiopetalidæ, Dorypetalidæ, Callipodidæ)

#### Order JULIFORMIA

Suborder Julidea (Blaniulidæ, Julidæ)

Suborder **Spirobolidea** (Spirobolidæ, Rhinocricidæ, Trigoniulidæ, Pachybolidæ, Spiromimidæ)

Suborder **Spirostreptomorpha** (Spirostreptidæ, Harpagophoridæ, Odontopygidæ, Cambalidæ, Pseudonannolenidæ, Pericambalidæ, Cambalopsidæ, Physiostreptidæ)

Superorder COLOBOGNATHA

Order **PLATYDESMIFORMIA** (Polyzoniidæ, Siphonocryptidæ, Siphonophoridæ, Platydesmidæ, Siphoniulidæ)

#### Class CHILOPODA

Subclass ANAMORPHA

Order SCUTIGEROMORPHA (Scutigeridæ)

Order LITHOBIOMORPHA

LITHOBIOIDEA (Lithobiidæ, Henicopidæ) CERMATOBIOIDEA (Cermatobiidæ) CRATEROSTIGMOIDEA (Craterostigmidæ)

# Subclass EPIMORPHA

Order **GEOPHILOMORPHA** (Himantariidæ, Schendylidæ, Oryidæ, Mecistrocephalidæ, Geophilidæ, Soniphilidæ, Neogeophilidæ, Gonibregmatidæ, Sogonidæ)

Order SCOLOPENDROMORPHA (Scolopendridæ, Cryptopidæ)

# Class SYMPHYLA

## Order CEPHALOSTIGMATA

(Geophilellidæ, Scutigerellidæ, Scolopendrellidæ)

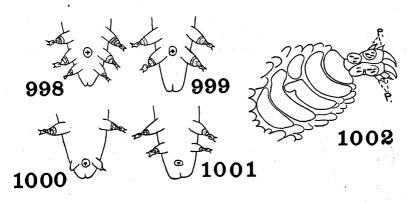
# CLASS ONYCHÓPHORA

# (MALACÓPODA, POLÝPODA, PROTRACHEÀTA)

Moderate-sized or rather large, soft bodied, caterpillar-like species. Body elongate, gently narrowed at the ends, convex above, flattened below, bearing from fourteen to more than forty pairs of short, similar, annulate, but not distinctly jointed legs. Integument transversely wrinkled, each fold bearing many somewhat regularly disposed papillæ. One pair of annulated antennæ. Mandibles blade-like, toothed and denticulate on the edge. Legs with a series of several transverse pads below near tip and two prominent apical claws. Excretory organs opening by a pore on the fourth and fifth pairs of legs. Sexual orifice near the posterior end of the body. Respiration by means of many long, unbranched tracheæ that open by minute spiracles scattered over the body.

patus, malav.) ..... PERIPATOPSÌNÆ

- a. Legs bearing from three to seven small, more or less distinctly jointed, conical papillæ at tip above, near the base of the claws.
   (Fig. 1002). (Macroperípatus, Epiperípatus, Plicatoperí-



Figs. 998-1002. Onychophora

- 998. Peripatus, apical part of body (Bouvier) Peripatidæ.
- 999. Peripatoides, apical part of body (Bouvier) Peripatopsidæ.
- 1000. Peripatopsiy, apical part of body (Bouvier) Peripatopsidæ.
- 1001. Paraperipatus, apical part of body (Bouvier) Peripatopsidæ.
- 1002. **Oroperipatus**, underside of fourth leg. P, P, papillæ (Bouvier) Peripatidæ.

#### LITERATURE ON ONYCHOPHORA

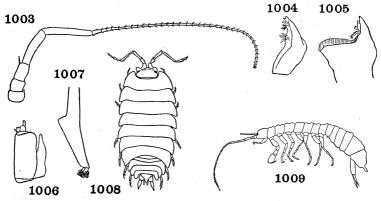
Bouvier, E. L. Monographie des Onychophores. Ann. Sci. Nat., (9) **2**, pp. 1–383, **5**, pp. 61–318 (1905–07).

CLARK, A. H. Distribution of Onychophora. Smithsonian Misc. Coll., 65, No. 1, pp. 1–25 (1915).

# CLASS CRUSTÀCEA

# ORDER ISÓPODA

As this group of Crustacea includes a small number of terrestrial forms, these have been included, especially since a few species occur abundantly in places where insects are commonly found. Two suborders are represented among the actually terrestrial species.



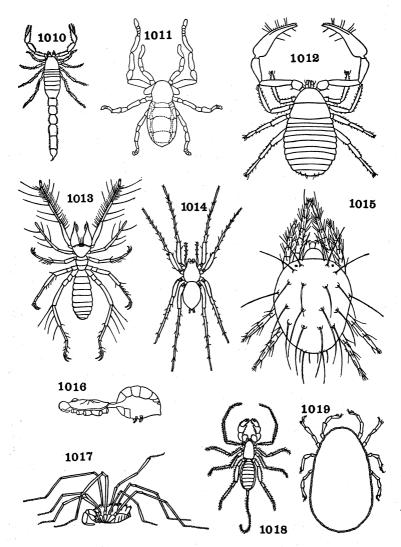
Figs. 1003–1009. **Isopoda** 

- 1003. Ligidium, second antenna (Richardson) Ligiidæ.
- 1004. Philoscia, mandible (Richardson) Oniscidæ.
- 1005. Haplophthalmus, mandible (Richardson) Trichoniscidæ.
- 1006. Porcellio, maxilliped (Richardson) Oniscidæ.
- 1007. Philoscia, first maxilla (Richardson) Oniscidæ.
- 1008. Porcellio (Richardson) Oniscidæ.
- 1009. Phreatoicus (Richardson) Phreatoicidæ.

3.	Flagellum of second antennæ multiarticulate. (Fig. 1003).
	(Lìgia, Ligidium). ( $LIG\acute{Y}DID\rlap{/}E$ ) LIG $\rat{IID}\rlap{/}E$
	Flagellum of second antennæ with not more than six joints4
4.	Mandible with the grinding surface well developed. (Fig. 1005)
	(Trichoníscus, Tithanètes, Brackenridgia, Haplophthál-
	mus, Androníscus) TRICHONÍSCIDÆ
	Mandible with the grinding surface small, poorly developed.
	(Fig. 1004)
5.	Inner lobe of first maxillæ bearing only two or three plumose
	process at its tip. (Fig. 1007)
	Inner lobe of first maxillæ bearing four or more (usually many)
	plumose processes. (Eubèlum, Éthelum) EUBÉLIDÆ
6.	Terminal joints of maxillipeds small. (Fig. 1006)
	Terminal joints of maxillipeds large, lamellar. Scýphax, Scy-
	phacélla)SCYPHÁCIDÆ
7.	Body capable of being rolled into a perfect ball; head deeply im-
	mersed in the first thoracic segment; uropods short, not reach-
	ing beyond the tip of the abdomen. (Cubaris (= Armadillo),
	Armadillídium, Péntheus, Sphæroníscus, Uropòdias).
	ARMADILLÍDID <i>Æ</i>
	Body not capable of being rolled into a ball; head only slightly
	immersed in the first thoracic segment; uropods elongate, reach-
	ing beyond the apical abdominal segments (Figs 1004 1006

# CLASS ARÁCHNIDA

Moderate-sized, small or very small, rarely large Arthropoda. Body formed of two groups of segments, a cephalothorax and an abdomen, which are usually clearly separate although sometimes entirely fused. Cephalothorax rarely provided with any movable sutures; abdomen usually more or less freely articulated and formed of several similar segments, the apical portion sometimes forming a tail-like prolongation, or (spiders and mites) showing no sutures. Cephalothorax bearing six pairs of appendages, the first usually chelate; the second often hooked and fitted for chewing or crushing prey; second pair sometimes sensory or fitted for walking, usually terminated in claws. No antennæ. No compound eyes; several simple eyes (two to twelve) usually present. No wings. Respiration by tubular tracheæ or by tracheal book-lungs or both. Development direct. Spiders, Mites, Ticks, Scorpions, Harvestmen, Tongue-worms.



Figs. 1010-1019. Various Arachnida

- 1010. Buthus (Kraepelin) Scorpionida.
- 1011. Cryptocellus (Ewing) Ricinulei.
- 1012. Chelifer (Ewing) Chelonethida.
- 1013. Galeodes (Dufour) Solpugida.
- 1014. Tegenaria (Emerton) Araneida.

<ul> <li>1015. Tetranychus (Woodworth) Acarina.</li> <li>1016. Liphistius (Warburton) Araneida.</li> <li>1017. Protolophus (Banks) Phalangida.</li> <li>1018. Thelyphonus (Kraepelin) Pedipalpida.</li> <li>1019. Argas (Bishopp) Acarina.</li> </ul>
2020. — 6. m. ( – 2020 pp.)
KEY TO THE ORDERS OF ARACHNIDA
1. Abdomen distinctly segmented, no silk-spinning organs
2. Abdomen with the posterior segments forming a contrasting long tail-like prolongation
3. Post-abdomen consisting of six segments, terminating in a prominent bulbous sting; abdomen broadly joined to the unsegmented cephalothorax; second ventral segment with a pair of comb-like organs; four pairs of book-lungs opening on the third to sixth
sternites; pedipalpi stout, terminating in large pincers; legs fitted for walking, tarsi three-jointed. Distribution widespread in warm dry countries. (Fig. 1010). Scorpions.
Post-abdomen very slender and many-jointed, not ending in a sting; abdomen narrowed at base, no comb-like ventral organs tarsi of first legs many-jointed.
4. Pedipalpi slender, similar to the walking legs; three pairs of booklungs, opening on segments 4, 5 and 6 of abdomen. Minute
species, under 3 mm. in length. Delicate, pale colored, tropical species. (Fig. 1020). MICROTHELYPHÓNIDA (Page 539)
Pedipalpi very stout, contrasting with the very long first pair of legs; two pairs of book-lungs, opening on segments 2 and 3 Moderate to large species. Tropical. Whip-scorpions.  PEDIPÁLPIDA (Page 540)
5. Abdomen constricted at base; front legs very long and with long
tarsi. Tropical
Abdomen broadly joined to the cephalothorax; front tarsi not elongate
6. Pedipalpi with large pincer-like claws. Small flat species, usually living under the bark of trees. (Fig. 1012). Pseudoscorpions. Book-scorpions
Pedipalpi without pincer-like claws

- 7. Head distinct from the three-parted thorax; cheliceræ relatively large, their pincers opening up and down. Pale-colored, mostly nocturnal species, inhabiting hot dry regions. (Fig. 1013). Wind-scorpions, Solpugids . . . . . . . SOLPŪGIDA (Page 546) Cephalothorax not distinctly divided into head and segments;
- 8. Abdomen nine-segmented as indicated by the tergites; cheliceræ usually exposed; two eyes usually present, often on tubercles; legs usually excessively long and slender. (Fig. 1017). Cosmopolitan, abundant. Harvestmen, Daddy-long-legs.

PHALÁNGIDA (Page 551)

Abdomen apparently four-segmented, with lateral as well as dorsal plates, and with a small terminal several-jointed piece; cheliceræ concealed by a large, pendant hood; eyes absent; legs only moderately long. (Fig. 1011). Tropical, rare species.

RICINÙLEI (Page 542)

9. Abdomen joined to the cephalothorax by a narrow short stalk. (Figs. 1014, 1016). Cosmopolitan. Spiders.

ARANÈIDA (Page 554)

- 10. Adult always with legs; usually with four, rarely two pairs; young frequently with three pairs; body surface very rarely bearing numerous transverse annulations; rarely living as internal parasites. (Figs. 1015, 1019). Mites, Ticks..ACARINA (Page 565)
  - Adult legless, body long; finely transversely annulate; internal parasites of vertebrates. (Fig. 1083). Tongue worms.

PENTASTÓMIDA (Page 579)

#### LITERATURE ON ARACHNIDA

Treatises on the Whole Class, or Dealing with More Than a Single Oder

- Banks, N. Synopsis of North American Scorpions, Solpugids and Pedipalpi. American Naturalist, **34**, pp. 421–427, (1900).
- Comstock, J. H. The Spider Book. Doubleday, Page & Co., Garden City, N. Y. (1912). (Contains also brief synopses of the North American representatives of several Orders of Arachnida in addition to the Araneida).
- Косн, L. Arachniden Australiens. 1489 + 274 pp. Nürnberg (1869-89).
- LAWRENCE, R. F. Arachnida of Southwest Africa, Pt. 2, Ann. S. Afr. Mus., 25, pp. 217–312 (1928).

Pocock, R. J. Arachnida. Fauna British India (1900).

Arachnida. In Biologia centrali-Americana. 71 pp. (1902).

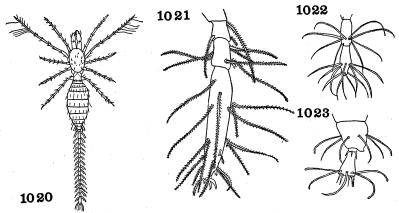
Simon, E. Arachnides de France, 1-7 (1874-1914).

Thorell, T. Studi sui Ragni Malesi e Papuani. 4 parts. Genoa (1877–90). Warburton, C. Arachnida, In Cambridge Natural History, 4, pp. 297–473 (1909).

# ORDER MICROTHELYPHÓNIDA

(PALPIGRÀDI; PALPIGRÀDA)

Minute, delicate, elongate species with a long, jointed median appendage at the tip of the abdomen. Cephalothorax above consisting of a large carapace which bears the pedipalps and the first three



Figs. 1020-1023. Microthelyphonida

- 1020. Kænenia (Wheeler) Kæneniidæ.
- 1021. Kænenia, apical part of third leg (Börner) Kæneniidæ.
- 1022. Kænenia, two joints of flagellum (Börner) Kæneniidæ.
- 1023. Kænenia, apex of abdomen (Börner) Kæneniidæ.

pairs of legs, and a small, separate posterior section which bears the fourth pair of legs. Cheliceræ large, chelate; pedipalps leg-like, nine-jointed, bearing a pair of claws at tip. First pair of legs the longest, twelve-jointed, the tarsi consisting of several segments, terminated by a pair of claws. Second and third legs seven-jointed; fourth eight-jointed, all with a pair of claws at tip. Eyes absent. Mouth anterior.

Respiration by three pairs of lung sacs, opening on the fourth to sixth abdominal segments below. Abdomen elongate-oval, consisting of eleven segments, the third to seventh segments larger than the others. Apex of abdomen bearing a long, slender median appendage consisting of nine to fifteen moniliform joints, bearing sparse bristly hairs. (Figs. 1020, 1021, 1022, 1023).

# LITERATURE ON MICROTHELYPHONIDA

HANSEN, H. J. Biospeologica LIII. Palpigradi. Arch. Zool. Paris, 65, pp. 167–180 (1926) (with bibliography).

Kraepelin, K. Palpigradi. Das Tierreich, Lief. 12, pp. 1-3 (1901).

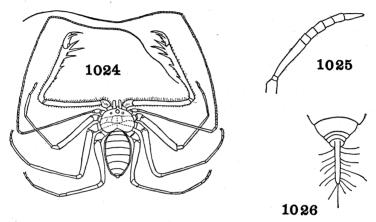
# ORDER PEDIPÁLPIDA

(PEDIPÁLPI)

Large or moderate-sized species with heavily chitinized integuments. Cephalothorax forming a solid carapace, or with two small thoracic segments separated by sutures. Cheliceræ claw-like. Pedipalpi six-jointed, strong, sometimes very stout, clawed at apex or imperfectly chelate, sometimes very long; first pair of legs sensory in function, forming a long, many-jointed filament; other legs stouter and fitted for walking, the tarsi with claws at tip. Abdomen consisting of ten or eleven segments; sometimes with a long slender many-jointed median apical appendage or filament, sometimes rounded at apex and without appendage. Respiration by book-lungs in the second abdominal segment and sometimes also in the third. Restricted to warm countries. Whip-scorpions and Tailless whip-scorpions.

- - Cephalothorax short and broad, more or less expanded laterally, the lateral margins strongly convex; abdomen blunt and rounded at apex, without a median appendage at apex; femur and tibia of pedipalpi strongly spinose. (Suborder AMBLYPYGI).....3

2. Cephalothorax dorsally with an undivided shield; eyes well developed, two close together on a tubercle in front and a group of three large ones toward each lateral margin; abdominal appendage long, filiform, consisting of many movable joints; tarsus of first pair of walking legs (second legs) seven-jointed and at least four times as long as the metatarsus; second and



Figs. 1024–1026. Pedipalpida

- 1024. Damon (Pocock) Tarantulidæ.
- 1025. **Schizomus,** metatarsus and tarsus of first pair of legs of female (Graveley) Schizomidæ.
- 1026. Schizomus, apex of abdomen. Schizomidæ.

third segments of abdomen each with a pair of book-lungs. (Fig. 1018). (Mimoscórpius, Tetrabàlius, Hypóctonus, malay.; Thelýphonus, indomal.; Typopéltis, As.; Mastigopróctus, Thelyphonéllus, Am.). . . . . THELYPHÓNIDÆ

- - Tibia of the fourth pair of legs not divided, or consisting of only two sections; the tarsus four-jointed; apical spurs of tibiæ of pedipalpi directed forward. (Fig. 1024). (Tarántula (= Phrynichus) ethiop., ind.; Dâmon, Titanodâmon, ethiop.). (PHRYNICHÌNÆ, Kræpelin 1899) .... TARANTÜLIDÆ
- 4. Tarsi five-jointed, with a pulvillus at the base of the claws; second abdominal tergite sometimes distinctly margined. Old World species. (Chàron, indomal.; Charìnus, ethiop., malay.; Catagèus, Stygophrynus, ind.; Sàrax, austromal.).

## CHARÓNTIDÆ

Tarsi four-jointed, without pulvillus; second tergite not margined. New World species. (Phrynus (= Tarántula), Hemiphrynus, Acanthophrynus, Heterophrynus, Am.)... PHRÝNIDÆ

#### LITERATURE ON PEDIPALPIDA

BÖRNER, C. Pedipalpida. Zoologica, Heft 42 (1904).

Graveley, F. H. Revision of the Oriental subfamilies of Tarantulidæ. Rec. Indian Mus., 11, pp. 433–455 (1915).

Indo-australian Thelyphonidæ. Rec. Indian Mus., **12**, pp. 59–85 (1916).

Kraepelin, K. Revision der Tarantuliden. Abh. naturh. Ver. Hamburg, 13, pp. 1–53 (1895).

Revision der Uropygi. Abh. naturh. Ver. Hamburg, **15**, 60 pp. (1897). Pedipalpida. Das Tierreich, Lief. 8 (1899).

## ORDER RICINÙLEI

# (MERIDOGÁSTRA, RHIGNOGÁSTRA)

Small or very small species with extremely thick, hard integument. Body elongate oval, without long tail-like prolongation at the apex of the abdomen but with a very short three segmented projection. Cephalothorax bearing anteriorly a large movable hood which when folded down completely hides the cheliceræ; cheliceræ chelate. Eyes absent. Pedipalpi five-jointed, short; geniculate at the apex of the femur; tarsus minute, opposed to a small projection at the tip of the tibia. Legs rather stout, the second pair the longest; tarsi of second to fourth legs five-jointed. Abdomen oval, broadly joined to the

cephalothorax, consisting of nine segments, basal ones very short; three to six much longer; six to nine forming a very short retractile apical tubercle. Respiration by tubular tracheæ opening by a pair of spiracles at the sides of the cephalothorax above the third coxa. (Fig. 1011).

The living species form a single family. (Ricinòides (=Crypto-stémma), ethiop.; Cryptocéllus, neotrop.). (CRYPTOSTEMMÁT-IDÆ, HOLOTÉRGIDÆ, CURCULÒIDIDÆ, POLIOCHÉRIDÆ).

RICINÒIDIDÆ

#### LITERATURE ON RICINULEI

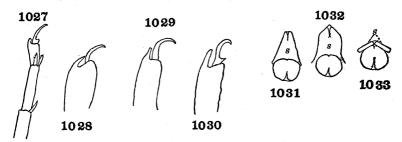
Hansen, H. J. and W. Sörenson. Two Orders of Arachnida. 178 pp. Cambridge, Eng. University Press (1904).

Ewing, H. E. A Synopsis of the Order Ricinulei. Ann. Entom. Soc. America, 22, pp. 583-600 (1929).

## ORDER SCORPIÓNIDA

(SCORPIÒNES)

Large or very large species. Cephalothorax and abdomen broadly attached; cheliceræ chelate; pedipalpi very large, bearing a large swollen chela at the apex. Cephalothorax with a pair of median eyes near the middle and a group of two to five lateral eyes at each side; rarely absent. Basal portion of abdomen broad, the five apical segments much narrower, forming a highly flexible postabdomen, the last segment of which bears a sting at apex and contains a poison gland. Four pairs of legs, each consisting of seven segments; tarsi three-jointed. Underside of abdomen at base with a pair of movable pectinate organs. Respiration by book-lungs opening by paired slits on the third to sixth abdominal segments. Mainly tropical species. Scorpions.



Figs. 1027-1033. Scorpionida

- 1027. Centruroides, tarsus of fourth leg. Buthidæ.
- 1028. Pandinus, tip of last tarsal joint. Scorpionidæ.
- 1029. **Vejovis,** tip of last tarsal joint. Vejovidæ.
- 1030. Iomachus, tip of last tarsal joint. Ischnuridæ.
- 1031. Centruroides, sternum. Buthidæ.
- 1032. Pandinus, sternum (Kraepelin) Scorpionidæ.
- 1033. Bothriurus, sternum (Kraepelin) Bothriuridæ.
- - Postabdomen with only one median ventral keel. (Uródacus, Austr.: Hemiscórpion, palæarc.). (See couplet 5).

# SCORPIÓNIDÆ, part

- 4. Last segment of postabdomen with a distinct thorn or tubercle below the sting; chela flat or rounded; last joint of tarsi with two rows of spines beneath. (Diplocéntrus, Oiclus, neotrop.; Nèbo, palæarc.) ...... DIPLOCÉNTRIDÆ Last segment of postabdomen simple, without thorn or tubercle
- 5. Tip of last tarsal joint rounded or acute and produced on the sides to form a lobe the edge of which forms an acute angle

with the apical projection that lies dorsally between the claws. (Figs. 1028, 1032). (Scórpio, palæarc., ethiop.; Pandinus, Opisthophthálmus, ethiop.; Heteromètrus, indomal.).

#### SCORPIÓNIDÆ

- Tip of last tarsal joint more or less truncate, without rounded lobes, its apical edge forming a right angle with the dorsal projection between the claws; last tarsal joint below with spines or bristles. (Fig. 1030). (Ischnùrus, Hadógenes, ethiop.; Opisthacánthus, ethiop., neotrop.; Hormùrus, indoaustr.; Iómachus, ethiop., ind.) ............................... ISCHNÙRIDÆ

This family is divisible into four well-marked subfamilies as follows:

- a. Group of two lateral eyes without a pale spot just behind the eyes; movable finger of cheliceræ without well developed teeth, or with only a single tooth on its lower surface......b
  - A pale spot behind each group of two lateral eyes; movable finger of cheliceræ with a number of teeth on its lower surface. (Chærilius, indomal.). (CHÆRILIIDÆ) . . . . . . CHÆRILIINÆ
- b. Four basal segments of postabdomen with a ventral keel on the median line; ventral surface of body granular or minutely tuberculate. (Megacórmus, neotrop.) .. MEGACORMÎNÆ Four basal segments of postabdomen without any distinct keels below, or with a pair, one on each side of the median line....c
- c. Hand (basal thickened portion of the apical joint of the pedipalpi) with a strong keel above which divides the surface into two surfaces that meet almost at a right angle. (Euscórpius, palearc.) ..... EUSCORPIÌNÆ
- 7. Sternum more or less triangular, strongly narrowed in front; first tarsal joint of third and fourth pair of legs often with a thorn at tip; three or five lateral eyes; last segment of postabdomen often bearing a spine beneath the sting. (Figs. 1010, 1027, 1031). (Bùthus, palæarc., ethiop., indomal.; Isomètrus, widespr.; Tityus, Centruròides, Am.; Anánteris, neotrop.; Uropléctes, Archisomètrus, ethiop., indoaustr.; Isometròides, austr., Buthèolus, palæarc., Parabùthus, ethiop.).

Sternum with parallel sides, usually wider than long, with a deeply impressed median groove; three lateral eyes; last segment of postabdomen always without a spine beneath the sting. (Fig. 1029). (Vejõvis, Hadrūrus, N. Am.; Scórpiops, ind.; Hadrurðides, Carabóctonus, neotrop.; Iùrus, palæarc.). VEJÓVIDÆ

#### LITERATURE ON SCORPIONIDA

EWING, H. E. Scorpions of the Western United States. Proc. U. S. Nat. Mus., 73, No. 2730, 24 pp. (1928).

Gough, L. H. Key to Egyptian Scorpions. Cairo Ministry Agric. Egypt. Bull. 76, 7 pp. (1927).

HEWITT, J. Survey of the Scorpion Fauna of South Africa. Trans. Roy. Soc. South Africa, 6, pp. 89–192 (1918).

Kästner, A. Scorpione (Scorpionida). In Tierwelt Mitteleuropas, 3, Lief. 1, pp. II 1–3 (1928).

Kraepelin, K. Revision der Skorpione. Jahrb. Hamburg. wiss. Anst., 8, pp. 1–144, (1891); 11, pp. 1–248, (1894); 12, pp. 1–24 (1895).
Scorpionida. Das Tierreich. Lief. 8 (1899).

## ORDER SOLPÜGIDA

# (SOLÍFUGÆ)

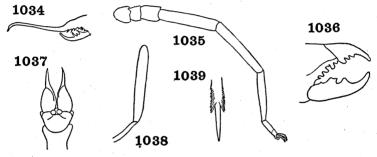
Large or moderately large, pale-bodied, very active species. Cheliceræ very large, chelate, moving in a vertical plane. Cephalothorax bearing only one pair of legs, separated from three distinct thoracic segments to each of which a pair of legs is attached. Abdomen elongate, oval, consisting of ten segments, without terminal filament or appendage. Pedipalpi long, leg-like, six-jointed. Tarsus of pedipalpi and first pair of legs one-jointed; second to fourth legs with the tarsi often several-jointed; tarsi usually provided with claws. Respiration by tracheæ that open through spiracles behind the first pair of legs on both the first and second abdominal segments. Distribution tropical and subtropical, most generally in dry or arid regions.

1. Fourth pair of legs ending in a pair of claws as do the second and third pairs; basal portion of fourth pair of legs (including the coxæ and two trochanters) clearly shorter than the remainder of the leg (femur, tibia and two tarsal joints); fourth legs with the basal section of femur very much shorter than the apical section.

Fourth pair of legs without claws, these replaced by short spines; basal portion of fourth leg almost as long as the remainder of the leg; basal section of femur of fourth leg longer than the apical section. (Hexisopus, S. Afr.) ..... HEXISOPÓDIDÆ

2. Spiracles of second and third abdominal segments covered by a plate which has a finely serrated posterior margin; tarsus of pedipalpi freely jointed and capable of folding against the metatarsus. (Galeòdes, palæarc., ind.; Paragaleòdes, palæarc.).

GALEÓDIDÆ



Figs. 1034-1039. Solpugida

- 1034. Gnossipus, chelicera in lateral view (Kraepelin) Solpugidæ.
- 1035. Solpugid, third leg (Ewing).
- 1036. Eremobates, chelicera in lateral view (Putnam) Solpugidæ.
- 1037. Eremobates, head in dorsal view (Putnam) Solpugidæ.
- 1038. Eremobates, tip of pedipalpus (Putnam) Solpugidæ.
- 1039. Solpuga, tip of flagellum of pedipalpus (Kraepelin) Solpugidæ.

Spiracles of the second and third abdominal segments not covered by serrated plates, freely visible or concealed by lateral folds; tarsus of pedipalpi immovably attached to the metatarsus although separated by an apparent suture. (Figs. 1034, 1036, 1037, 1038, 1039). (Solpuga, Dæsia, Gnosíppus, Ceròma, ethiop.; Rhagòdes, ethiop., As.; Dínorhax, indomal.; Sarónomus, neotrop.; Ammótrecha, Eremóbates, Am.; Gylíppus, Kárschia, palæarc.) SOLPÜGIDÆ

#### LITERATURE ON SOLPUGIDA

Kraepelin, K. Zur Systematik der Solifugen. Mitt. naturh. Mus. Hamburg, 16, pp. 197–258 (1899).

Solpugida. Das Tierriech, Lief. 12, pp. 4-159 (1901).

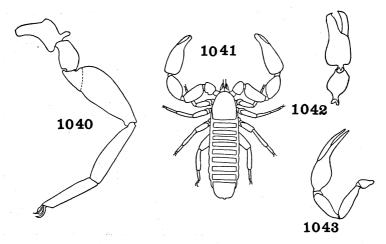
- Hewitt, J. A Short Survey of the Solifugæ of South Africa. Ann. Transvaal Mus., 7, pp. 1–76 (1919).
- Putnam, J. D. Solpugida of America. Proc. Davenport Acad. Nat. Sci., 3, 149 pp. (1882).
- SIMON, E. Essai d'une Classification des Galéodes. Ann. Soc. Entom. France, 1879, pp. 93–154 (1879).

## ORDER CHELONÉTHIDA

# (PSEUDOSCORPIÓNIDA; PSEUDOSCORPIÒNES, CHERNETÍDEA)

Small or very small Arachnida with flattened body. Cephalothorax unjointed, although sometimes with distinct transverse grooves; abdomen with 11 or 12 tergites, each often composed of a pair of plates, narrowly separated along the median line. Cheliceræ twojointed, chelate. Pedipalpi very large, six-jointed, much longer than the walking legs, their tarsi forming a much enlarged pincers-like grasping organ. Legs six-jointed, the tarsi one or two-jointed and the femur sometimes divided into two sections; tarsi with a pair of apical claws. Cephalothorax narrower than the abdomen which is often much widened; abdomen without any apical prolongation, joined to the cephalothorax along its entire base. Eves two or four, placed near the sides of the carapace; sometimes wanting. Respiration by tracheæ opening by four spiracles placed just behind the second and third abdominal plates at the sides of the body. Genital aperture on the first sternite. Slow-moving carnivorous species of retiring habits, living under bark, stones, etc. False-scorpions; Book-scorpions.

- 2. Tarsi two-jointed. (Suborder DIPLOSPHYRÓNIDA)......3
  Tarsi one-jointed (Fig. 1040). (Suborder MONOSPHYRÓN-IDA).....9
- - Movable finger of chelicers with a single subapical lobe or tooth which is rarely secondarily subdivided, but never multidenticu-



Figs. 1040-1043. Chelonethida

- 1040. Chelifer, fourth leg (Ewing) Cheliferidæ.
- 1041. Chelifer (Tullgren) Cheliferidæ.
- 1042. Chelifer, pedipalpus (With) Cheliferidæ.
- 1043. Ideoroncus, pedipalpus (With) Ideoroncidæ.
  - 5. Pleural membrane of abdomen smoothly longitudinally plicate, never granulate; femur of fourth pair of legs with an oblique cross-suture (rarely nearly perpendicular to the longitudinal axis). (Chitra, Syarinus, Hyarinus, nearc.). SYARÍNIDÆ Pleural membrane granulate or granulate-striate; femoral articulation of fourth pair of legs truly vertical to the longitudinal axis of the femur-patella. (Neobísium, Microbísium, holarc.; Blòthrus, Rôncus, palæarc.) . . . . . . NEOBISÌIDÆ
  - 6. Pleural membrane of abdomen granulate; claw with twelve tactile setæ on the fingers. (Hŷa, indomal.) . . . . . . . HŸIDÆ

<b>7.</b> ,	Pleural membrane of abdomen evenly longitudinally plicate; fingers of claw usually with many more than twelve tactile setæ. (Fig. 1043). (Ideoróncus, Bóchica, neotrop.; Albìorix, N. Am.; Shravàna, indomal.) IDEORÓNCIDÆ Venom apparatus present in both the movable and fixed fingers of the claw; no sheathing structure on either claw to receive the venom-tooth
	Venom apparatus present in the fixed finger of the claw only; a
	well developed sheathing structure present on the movable finger to receive the venom-tooth. ( <b>Ménthus</b> , nearc., neotrop.).
	miger to receive the venom-tooth. (Mentinus, nearc., neotrop.).  MÉNTHIDÆ
8	Pleural membrane of abdomen smoothly and evenly plicate;
0.	carapace not or rarely noticeably narrowed in front; sides of
	abdomen more or less parallel, the abdomen not greatly wider
	than the cephalothorax. (Olpium, palæarc.; Garypinus,
	Am.; Seriànus, nearc., neotrop., indomal.; Solinus, nearc.,
	Austr.; Hòrus, ethiop.; Xenólpium, indoaustr.) OLPÌIDÆ
	Pleural membrane prickly, granular, or irregularly wrinkled; cara-
	pace narrowed in front, clearly triangular; abdomen subovate,
	much broader than the cephalothorax. (Gárypus, palæarc.,
	neotrop.; Geogárypus, palæarc., indomal.; Lárca, holarc.;
	Anagárypus, austromal.; Maorigárypus, austr.). GARYPÓDIDÆ
0	Four well developed eyes
9.	Two eyes or none. (Figs. 1040, 1041, 1042) CHELIFÉRIDÆ
10	Carapace triangular, narrowed in front, without horn-like proc-
10.	esses in front of the eyes; abdomen without pleural plates.
	(Synsphyrònus, Austr.) GARYPÓDIDÆ, part
	Carapace not triangular, with two or more prominent horn-like
	processes in front of the eyes FEAÉLLIDÆ

### LITERATURE ON CHELONETHIDA

processes in front of the eyes ....

Balzan, L. Revisione dei Pseudoscorpioni de Paraguay. Ann. Mus. Civ. Genova (2), **29**, pp. 401–451 (1890). Pseudoscorpiones de Venezuela. Ann. Soc. Entom. France, 60, pp. 497-

552 (1891).

Chamberlin, J. C. Synoptic Classification of False Scorpions, Part I. Ann. Mag. Nat. Hist. (10), 4, pp. 50–80 (1929), 5, pp. 1–48, and pp. 585–618 (1930).

Coolidge, K. R. List of North American Pseudoscorpionida. Psyche, 15, pp. 108-114 (1908).

Ellingsen, E. Die Pseudoscorpione des Berliner Museums. Mitt. zool. Mus. Berlin, 4, pp. 355–423 (1910) (extensive bibliography).

Pseudoscorpions of South Africa. Ann. South African Mus., 10, pp. 75–128 (1912).

- Kästner, A. Afterscorpione (Pseudoscorpionida). In Tierwelt Mittleuropas, 3, Lief. 1, pp. IV 1–13 (1928).
- Kew, H. W. Synopsis of False Scorpions of Britain and Ireland. Proc. Roy. Irish Acad., 29, Sec. B, No. 2, (1911), and Supplement, ibid., 33 (1916).
- With, C. J. Chelonethida of the Australian Region. Ann. Mag. Nat. Hist., (7) 15, pp. 94–143 (1905).

Indian False-Scorpions. Kgl. Danske Vid. Selsk. Skrifter., 7, pp. 1–124 (1906).

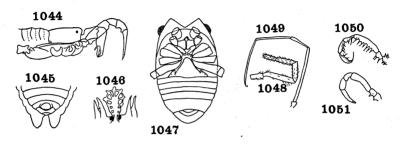
## ORDER PHALÁNGIDA

# (OPILIÒNES)

Usually moderate-sized species with long or very long legs. Cephalothorax not separated from the abdomen by a constriction, forming an unsegmented or very indistinctly segmented dorsal carapace. Abdomen segmented, with nine dorsal plates and fewer ventral ones. Head with two eyes, often on stalk-like tubercles. Cheliceræ three-jointed, chelate; rather long and sometimes thickened. Pedipalpi similar to the legs, but usually much shorter. Legs very long and slender; coxæ very large and firmly attached to the body. Respiration by tracheæ; one pair of spiracles opening ventrally near the base of the abdomen. Harvestmen, Daddy-long-legs.

- - of the cephalothorax. (Fig. 1045). (Suborder CYPHOPH-THÁLMI). (Siro, palæarc.; Ogòvea, ethiop.; Miópsalis, Stylocéllus, Indomal.; Péttalus, ind.). . . . . . . SIRÓNIDÆ
- 2. Pedipalpi stout, their tarsi with a strong grasping claw; apical joint of the tarsus of first and second pairs of legs with only one simple claw; the third and fourth tarsi with either two, or with a tridentate claw; sternite of first abdominal segment much reduced, not reaching in front of the hind coxæ; hind legs usually the longest. (Fig. 1044). (Suborder LANIATÔRES)...3

Pedipalpi slender, palpiform, their tarsi with a weaker claw which is occasionally lacking; apical joint of all four tarsi with only



Figs. 1044-1051. Phalangida

- 1044. Oncopus, anterior part of body in profile (Roewer) Oncopodidæ.
- 1045. Pettalus, apical part of abdomen (Roewer) Sironidæ.
- 1046. Ortholasma, anterior part of cephalothorax (Roewer) Trogulidæ.
- 1047. Acropsopilio, underside of body (Roewer) Acropsopilionidæ.
- 1048. **Trogulidæ,** pedipalpus (Roewer) Trogulidæ.
- 1049. Phalangiidæ, pedipalpus (Roewer) Phalangiidæ.
- 1050. Gonyleptidæ, pedipalpus (Roewer) Gonyleptidæ.
- 1051. Oncopodidæ, pedipalpus (Roewer) Oncopodidæ.

## TRIÆNONÝCHIDÆ

5. Pedipalpi carried with the tips curved backwards behind the patella, weakly armed; anterior margin of the cephalothorax below with five erect, conical teeth. (Assamia, Metassamia, indomal.; Trionyxélla, ind.; Mérmerus, mal.; Macrodampètrus, Dampètrus, Mosòia, Austr.) .... ASSAMÌIDÆ

	Pedipalpi not curved, held straight forward or at most bent to the side of the cheliceræ; anterior margin of the cephalothorax sometimes sharply excavated, but never with five erect, coni- cal teeth
6.	Tarsi of the third and fourth legs each with a false claw (pseudonychium) below the true claws
	Tarsi of third and fourth legs without a pseudonychium. (Bel-
	oníscus, Obaloniànus, indomal.; Scotolèmon, palæarc.
	Phalangòdes, Am.; Zalmóxis, Austr.; Metabiántes, ethiop.).
-	PHALANGÓDIDÆ
1.	Pedipalpi weak, their femur, tibia and tarsus broadly flattened and keeled, curved in front of and to the side of the cheliceræ
	all joints of the pedipalpi almost unarmed, without stout spines.
	(Libitòides, Metacynórta, Am.; Paravanònes, Cynórta,
	Cynórtula, Metarhaucus, Flírtea, Ergínulus, neotrop.).
	(PALPÍNIDÆ) COSMÉTIDÆ
	Pedipalpi very stout, their femur not flattened, their tibia and
	tarsus rounded, at most flattened beneath between the stout
	tibial and tarsal spines, not keeled; the pedipalpi porrect and
	not curved toward the cheliceræ. (Fig. 1050). (Discocýr
	tus, Metagynòdes, Eusárcus, Pachylòides, Wèyhia, Gony- léptes, Goniosòma, Cælopỳgus, neotrop.).
	GONYLÉPTIDA
8.	· · · · · · · · · · · · · · · · · · ·
-	without, or with only a very minute claw; maxillary lobe of the
	second coxa very small or absent, never freely movable; no ac
	cessory spiracles on the legs. (Superfamily NEMASTOSOMA-
	TOIDEA)
	Tarsi of the pedipalpi always longer than the tibiæ and always with a distinct, simple or pectinate claw; maxillary lobe of the
	second coxa distinct, freely movable, long and narrow; two
	accessory spiracles on the first to fourth tibiæ. (Fig. 1049)
	(Lacínius, Phalángium, Opílio, holarc.; Cáddo, nearc. Mítopus, Odiéllus, palæarc.; Rhampsinitus, ethiop.; Gag-
	rélla, indoaustr.). (Superfamily PHALANGIÒIDEA).
	PHALANGIDA
9.	Eyes large, highly convex, widely separated, not on a common
	ocular tubercle, placed one at each side of the cephalothorax

Both eyes placed on a common median ocular tubercle......10

**ACROPSOPILIÓNIDÆ** 

(Fig. 1047). (Acropsopílio, neotrop.).

- - Ocular tubercle placed at the anterior margin of the cephalothorax and forming with it a large lobe which covers the cheliceræ and pedipalpi from above. (Figs. 1046, 1048), (Trógulus, Dicranolásma, palæarc.; Ortholásma, Dendrolásma, nearc.).

#### TROGULIDÆ

11. First and second thoracic tergites fused together in the cephalothorax; first to fourth coxe each with a series of tubercles before and behind. (Nemastosòma, holarc.; Crósbycus, nearc.).

## **NEMASTOSOMÁTIDÆ**

First thoracic tergite fused with the cephalothorax and forming a prosoma; second thoracic tergite free, not fused with the cephalothorax nor the first tergite; first to fourth coxe without an anterior and posterior row of tubercles. (Ischyrópsalis, palæarc.; Táracus, Sábacon, Tomicomèrus, nearc.).

ISCHYROPSÁLIDÆ

#### LITERATURE ON PHALANGIDA

- Banks, N. Synopsis of North American Phalangida. American Naturalist, **35**, pp. 669–679 (1901).
- Bishop, S. C. and C. R. Crosby. Phalangida of the Southeastern United States. Journ. Elisha Mitchell Soc., 1924, pp. 8–26 (1924).
- Roewer, C. F. Die Weberknechte der Erde. 1116 pp. Jena, G. Fischer. (1923).

Weitere Weberknechte. Abh. Naturw. Ver. Bremen, **26**, pp. 261–402 (1927).

Weberknechte (Opiliones). In Tierwelt Mitteleuropas,  $\bf 3$ , Lief. 1, pp. V 1–10 (1928).

Walker, M. E. Revision of Phalangida of Ohio. Ohio State Univ. Studies, Biol. Surv., Bulletin 19, 4, No. 4, pp. 153-175 (1928).

## ORDER ARANÈIDA

Small or moderate-sized, rarely large Arachnida. Cephalothorax usually oval, the head frequently separated by an indistinct suture. Usually eight simple eyes, sometimes less or occasionally none. First pair of appendages (cheliceræ) hooked or chelate, each provided with a poison-gland opening near the tip. Second pair of appendages (pedipalpi) six-jointed, with one-jointed tarsi; similar to, but usually much

shorter than the four pairs of walking legs which are seven-jointed. Tarsi two-jointed, clawed at tip and often with additional claws or bristles used for manipulating the web. Abdomen almost always entirely unsegmented, its integument thinly chitinized, attached to the cephalothorax by a very strongly constricted base. Respiration by book-lungs opening by one or two pairs of slits on the underside of the abdomen; tracheæ usually present in addition to book-lungs, opening by one or two spiracles on the abdomen below. Silk-spinning glands present, opening by short tubular organs on the abdomen below. Development direct, the young essentially like the adults. Spiders.<sup>1</sup>

- 1. Abdomen entirely unsegmented in the adult (OPISTHO-Abdomen segmented above and showing several distinct segments posteriorly between the anal tubercle and spinnerets which are separated by a considerable space; six to eight spinnerets; fangs of cheliceræ moving up and down; two pairs of lungs. (Suborder LIPHISTIOMÓRPHÆ (=MESOTHÈLÆ)). (Liphístius, Heptathèla, Anadiastothèle) ..... LIPHISTÌIDÆ 2. Fangs of cheliceræ moving up and down; two pairs of lungs. (Suborder AVICULARIMÓRPHÆ  $(=MYGALOM \hat{O}RPHÆ)$ ). 3 Fangs of cheliceræ moving in and out. (Suborder ARACHNO-3. Tips of tarsi beneath the claws without claw-tufts. (NELÍ-Tarsi with claw-tufts, i.e. a bundle of hairs at the tip, just beneath the claws; two claws with a single row of teeth, or claws smooth; maxillary lobes wanting: lip free. (HYPODÉMATA).....9 4. All tarsi with a brush of hairs (scopula) beneath; two claws pectinate in two rows: cheliceræ without rastellum: maxillary lobes wanting; lip immobile; four spinnerets, the anterior pair small. close together, the posterior pair heavier and longer; eight eyes in a compact group, heterogeneous. (Pycnothèle, Lycinus). PYCNOTHÉLIDÆ At least the third and fourth tarsi without a scopula......5
- or double row; four (rarely six) comparatively short spinnerets,

  ¹ The accompanying key follows that given by Professor Petrunkewitch in his recent "Systema Aranearum," with a veryfew changes in nomenclature, some modifications of arrangement and the frequent omission of characters of lesser diagnostic value or more difficult recognition.

5. Basal segment of cheliceræ with a rastellum, *i.e.* armed on outer side near tip with several rows of stout teeth; maxillary lobes wanting: three claws, upper claws similar, pectinate in a single

	the anterior pair more or less approximated; eight eyes, hetero-
	geneous. (Actinopus, Cteniza, Ídiops, Acáttyma, Brachy-
	bóthrium, Pachylomèrus). Trap-door spiders. CTENÍZIDÆ
	Cheliceræ without a rastellum
6.	Lip free; maxillary lobes wanting; three claws, the upper ones
	pectinate in a single or double row; four or six spinnerets, the
	posterior pair very long; eight or six eyes, heterogeneous;
	species spinning webs. (Brachythèle, Evàgrus, Haplothèle,
	Hexùra, Diplùra, Áname, Tréchona, Macrothèle).
	DIPLURIDÆ
	Lip immobile
7.	Six spinnerets, the anterior pair close together; maxillary lobes
	well developed; three claws, the upper ones pectinate in a single
	row; eight eyes in three groups, heterogeneous. (Atypus,
	Calómmata, Microhexùra)
	Four or two spinnerets
8.	Four claws, the upper ones with one or several teeth, sometimes
	dissimilar; maxillary lobes wanting; four spinnerets; eight eyes
	in two rows, heterogeneous; body with simple hairs. (Cádmon,
	Migas, Moggridgea, Pœcilomigas). $(MIGID\cancel{E})$ .
	MIGÁDIDÆ
	Two claws with a single tooth each, sometimes with a vestigial
	third claw; four or two spinnerets; coxe of pedipalpi with a
	vestigial maxillary lobe; eight eyes in a compact group, hetero-
	geneous; body with scales and clubbed hairs. (Parátropis,
	Anisáspis, Anisaspòides) PARATROPÍDIDÆ
9.	Last joint of posterior spinnerets very short; four or two spin-
	nerets; cheliceræ often with several rows of stout teeth ex-
	nally at apex (rastellum); claws similar, smooth or with a few
	teeth in a single row; eight eyes, heterogeneous. (Barychèlus,
	Diplothèle, Leptopélma, Sàson) BARYCHÉLIDÆ
	Last joint of posterior spinnerets as long as, or often longer than
	the preceding joint; four spinnerets; claws similar, pectinate
	in a single row; cheliceræ without a rastellum; eight eyes in
	a compact group, heterogeneous. Tarantulas. (Theraphòsa,
	Aviculària, Eurypélma, Grammóstola, Eumenóphorus,
	Selenocósmia, Ornithóctonus) AVICULARÌIDÆ
10.	Abdomen with two pairs of spiracles (TETRASTÍCTA)11
	Abdomen with a pair of spiracles leading into book-lungs and a
	single median spiracle behind them, leading into a tracheal
	tube

11.	Cribellum and calamistrum present, at least in the female12
	Cribellum and calamistrum absent in both sexes
12.	Two pairs of lungs; cribellum entire, not divided; eight eyes in
	two rows, heterogeneous. (Hypochilus, Ectatostícta).
	HYPOCHÍLIDÆ
	One pair of lungs; cribellum divided; lip immobile; eight eyes in
	a compact group, heterogeneous. (Filístata). FILISTÁTIDÆ
13.	The first pair of spiracles leading into book-lungs, the second pair
	into tracheal tubes14
	Book-lungs entirely absent; both pairs of spiracles leading into
	tracheal tubes
14.	Three or two claws pectinate in a single row; lip free, long; six eyes,
	the antero-median ones (direct eyes) wanting or all eyes absent;
	third pair of legs sometimes reversed in position, directed for-
	ward; coxæ of four anterior legs long and cylindrical. (Dýsdera,
	Segéstria, Ariàdna, Harpáctes, Stalita) DYSDÉRIDÆ
	Two claws pectinate in a double row
15.	Six eyes or none; six spinnerets, the anterior ones approximated.
	(Orchéstina, Gamasomórpha, Òonops, Epéctris, Scaphi-
	élla) OONÓPIDÆ
	Eight eyes, heterogeneous; four spinnerets in a chitinous ring.
	(Hadrotársus, Gmógala) HADROTÁRSIDÆ
16.	Three claws, dissimilar, pectinate in a single row; cheliceræ with-
	out boss, their oblique margins with teeth; lip immobile; colulus
	(a slender appendage in front of the spinnerets) present; eyes
	wanting; six spinnerets, the anterior pair longest. (Telèma,
	Apneumonélla) TELÉMIDÆ
	Two or three claws; upper ones similar, pectinate in a single row;
	cheliceræ without boss, with smooth margins; lip free; colulus
	wanting; eight or two eyes, the anterior median eyes always pres-
17	ent. (Capònia, Nóps, Caponina, Diploglèna). CAPONIIDÆ
17.	Three claws, rarely the third vestigial in which case there are only
	two spinnerets and the first pair of legs are exceptionally heavy.
	(TRIÓNYCHA)       18         Two claws (DIÓNYCHA)       42
18.	Anal tubercle large, two-jointed, with a fringe of long hair19
10.	Anal tubercle normal
19.	Cribellum and calamistrum present; eight eyes in a compact group,
IJ.	heterogeneous. ( <b>Ecòbius</b> ) <b>ECOBÌIDÆ</b>
	Cribellum and calamistrum wanting; eight eyes in two rows,
	heterogeneous. (Uróctea)
	TOUR OF CHOOLS. (CICOLON) CICOLINIA

20. Cribellum and calamistrum present in the female and in the im-
mature male
Cribellum and calamistrum wanting
21. Tarsi with claw tufts (a bundle of hairs at tip, just beneath the
claws); three claws, the upper ones pectinate in a single row;
eight eyes in two rows, homogeneous, diurnal. (Pséchrus,
Fecènia, Stiphídion, Matáchia) PSÉCHRIDÆ
Tarsi without claw-tufts
22. First and second tarsi and metatarsi with a brush of hairs (scopula)
beneath; tarsi with two rows of trichobothria; six spinnerets;
cheliceræ with teeth on both margins; three claws, the upper
ones pectinate in a single row; eight eyes in two rows. (Ten-
gélla, Themàcrys) TENGÉLLIDÆ
All tarsi without scopula; no trichobothria, or with a single row . 23
23. Eyes homogeneous, diurnal
Eyes heterogeneous; only the anterior median ones diurnal26
24. Cribellum divided; six spinnerets, the posterior pair longer and
heavier than the anterior pair; three claws, the upper pair dis-
similar, pectinate in a single row; eight eyes in three rows, of
which the first is formed of four eyes. (Erèsus, Dórceus,
Adonèa, Stegódyphus) ERÉSIDÆ
Cribellum entire; spinnerets more nearly equal; claws similar25
25. Femora with trichobothria; eight eyes in two rows; orb-weaving
species. (Ulóborus, Sýbota, Hyptiòtes, Miagrámmopes,
Avellópsis) ULOBÓRIDÆ
Femora and tarsi without trichobothria; eight eyes in three or
two rows, the first row formed of four eyes, those of the second
row often very large. (Dinòpis, Ménneus) DINÓPIDÆ
26. Tarsi with a single row of trichobothria; cribellum divided.
(Amauròbius, Amphigỳrum, Badúmna, Titanœca).
AMAUROBÌID <i>Æ</i>
Tarsi without trichobothria; cribellum usually entire. (Dictyna,
Scotolathys, Lathys, Myrópsis, Altélla, Chærèa).
DICTÝNIDÆ
27. Trochanter of pedipalp inserted on the dorsal surface of the maxil-
lary lobe; head conspicuously elevated over the thorax; cheli-
ceræ inserted far above the mouth; eight or six eyes. (Archæa,
Mecysmauchènius)ARCHÆIDÆ
Trochanter of pedipalp inserted on the outer edge of the maxillary
lobe; cheliceræ inserted nearer to mouth; claw tufts wanting . 28
28. Six spinnerets; three claws

	Only two spinnerets, the middle and hind pair wanting; pedipalp of female without claw; lip free; two or three claws, the upper ones dissimilar, pectinate in a single row; eight eyes, heterogeneous. (Lûtica, Otíothops, Palpimānus, Stenochilus, Hermíppus)
29.	Anterior lateral edge of the first and second tibia and metatarsus with a row of long spines, the interspaces between which are occupied by a row of much shorter, curved spines; tarsi with serrated bristles; eight eyes in two rows, heterogeneous, the
	lateral eyes contiguous. (Mimètus, Ero, Gélanor, Oárces,
	Phobetinus) MIMÉTIDÆ
	First and second metatarsi and tibiæ without such spines 30
30.	Fourth tarsi with a ventral row of serrated bristles forming a distinct comb; legs without spines; upper claws similar, pectinate
	in a single row or smooth; eight, rarely six or four eyes in two
	rows, heterogeneous. (Steatòda, Latrodéctus, Argyròdes,
	Spintharus, Theridion, Monèta, Crustulina, Nicódamus).
	THERIDÌIDÆ
	No ventral comb of serrated bristles on the fourth tarsi31
31.	Cheliceræ immovably fused or united at the base
	Cheliceræ free
32.	Usually six nocturnal eyes in three groups of two each; if eight
	eyes, then the direct eyes are also nocturnal and eyes are ar-
	ranged in two rows of four each; lip long. (Sicàrius, Scytòdes, Loxósceles, Plectreùrys, Diguètia, Drymùsa). (SCYTÓD-
	IDÆ) SICARÌIDÆ
	Usually eight eyes, heterogeneous, direct eyes alone diurnal or
	absent when there are only six eyes; nocturnal eyes in two
	groups of three eyes each, or all eyes in a single compact group;
4.5	lip wide. (Phólcus, Modísimus, Spermóphora, Psilóchorus,
	Physoglènes, Ninètis) PHÓLCIDÆ
33.	Serrated bristles present on tarsi, forming at least one pair of
	spurious claws
	Tarsi without serrated bristles, or at least not in the shape of spurious claws
34	Six eyes in a compact group, nocturnal; tarsi with onychium; one
<b>01.</b>	pair of spurious claws; lip immobile, short; legs with spines.
	(Leptonèta, Merizócera, Ochyrócera, Psilodérces, Usòfila).
	LEPTONÉTIDÆ
	Eight eyes, rarely six or none; lip rebordered
35.	Cheliceræ with stridulating ridges on the external surface; upper

	claws similar, pectinate in a single row; legs with fine spines
	or smooth; eyes heterogeneous. Sheet-web weavers. (Boly-
	phántes, Drapetísca, Labúlla, Linýphia, Eriógone, Tapi-
	nòpa, Lophocarènum, Màso) LINYPHÌIDÆ
	Cheliceræ without stridulating ridges; upper claws similar or dis-
	similar, pectinate in a single row; legs usually with stout spines;
	eyes usually homogeneous, diurnal, rarely heterogeneous.
	Orb-weavers. (Argiope, Arànea ( $=Epeira$ ), Mèta, Néphila,
	Leucauge, Tetrágnatha, Theridiosòma, Gasteracántha).
	(EPEÍRIDÆ) ARGIÓPIDÆ
36.	Cheliceræ without a boss
	Cheliceræ with a boss
37.	Colulus present; last joint of posterior spinnerets always long;
	anterior spinnerets shorter than the posterior pair; eight eyes
	on a tubercle, heterogeneous; the anterior lateral ones noc-
	turnal, all others diurnal. (Hersília, Hersiliola, Murrícia,
	Tàma) HERSILÌIDÆ
	Colulus wanting; last joint of posterior spinnerets short; anterior
	pair of spinnerets usually considerably longer than the pos-
	terior pair; eyes in two or three rows. (Zodarion, Storena,
	Làches, Cryptothèle, Cithæron) ZODARÌIDÆ
38.	Anterior median eyes not far in advance of all the other eyes39
	Anterior median eyes small, approximated, placed far in advance
	of the other eyes near the edge of the clypeus; eight eyes, hetero-
	geneous; body with plumose hairs. (Senóculus).
	SENOCÜLIDÆ
39.	Trichobothria on the tarsi either in two rows or irregularly dis-
	tributed
	Trichobothria on the tarsi in a regular row; legs without scopulæ;
	body with plumose hairs; eight eyes (rarely six or none) in two
	rows of four each, usually heterogeneous, only the anterior
	median eyes diurnal; eyes sometimes almost homogeneous. (Fig.
	1014). (Agalèna, Tegenària, Cœlòtes, Argyronèta, Cy-
40	bæus, Hahnia, Rhoicinus) AGALÉNIDÆ
40.	All trochanters strongly notched; posterior row of eyes always
	recurved
	Only the fourth trochanter with a slight notch; eyes often grouped
	in a hexagon, the posterior four forming a procurved line;
	body with scale-like hairs; eight eyes, all diurnal. (Oxyòpes,
	Hamatáliva, Oxyopeidon, Peucètia, Tappònia).

41.	Upper claws with numerous teeth in a single row; lower claw with two or three teeth; body always bearing plumose hairs; legs
	rather long, sometimes formed for running sideways; female carrying her egg-cocoon in her cheliceræ, or if attached to the
	spinnerets, it is hemispherical. (Chiasmòpes, Pisaùra, Than-
	atídius, Dolómedes, Thaumàsia, Thalássius).
	PISAURIDÆ
	Upper claws with few teeth; lower claw either smooth or with a
	single tooth; body usually with only simple hairs; legs shorter, always formed for running forwards; cocoon globular, carried
	attached to the spinnerets. Wolf spiders. (Lycòsa, Pardòsa,
	Allocòsa, Arctòsa, Piràta, Evippa, Hippasa) . LYCÓSIDÆ
42.	Cribellum and calamistrum present
	Cribellum and calamistrum absent
43.	First and second tarsi and metatarsi with a scopula; claw tufts
	wanting; cribellum divided; eight eyes in two rows, homo-
	geneous, diurnal. (Zorópsis, Rècius, Zorócrates).
	ZOR ÓPSID <i>Æ</i>
	None of the tarsi or metatarsi with scopula; claw-tufts present;
	cribellum divided or entire; eight eyes in three rows, homo-
	geneous, diurnal. (Acanthóctenus) . ACANTHOCTÉNIDÆ
44.	1
	Cheliceræ immovably attached at base; with a boss and a row of
	teeth along the entire inner edge; lip immobile; eyes eight,
45.	in a compact group. (Ammóxenus) AMMOXÉNIDÆ Eight eyes in three rows, homogeneous, diurnal, the first row of
40.	four, second and third of two each; sometimes with first row so
	strongly curved that four rows are apparently formed; legs
	formed for running forwards and jumping; body often scaly.
	Jumping spiders. A very large family. (Lyssómanes, Sálti-
	cus, Myrmaráchne, Marpíssa, Phidíppus, Dendryphántes,
	Pellènes, Áttus)
	Eyes either in two rows, or if in three rows, the second is com-
	posed of four eyes, or else the eyes are distinctly heterogene-
	ous
46.	Sternum wider than long, broadly truncate behind; body very flat;
	middle pair of spinnerets between the anterior pair. (Plator,
	Véctius, Doliomàlus)
	Sternum normal; middle pair of spinnerets not thus placed47
47.	Six eyes in the first row; eight eyes in two rows, heterogeneous,
	only the posterior median ones nocturnal body flat legs

	adapted for running sideways; anterior pair of spinnerets close together. (Selènops) SELENÓPIDÆ
	First row never composed of six eyes
48.	Anterior pair of spinnerets wide apart49
	Anterior pair of spinnerets contiguous or nearly so50
49.	Eight eyes in two rows of four each, if only six eyes are present,
	the direct eyes are wanting; cheliceræ with boss, the margins
	toothed; pedipalp of female with claw. (Drássus, Drassodes,
	Zelòtes, Gnaphòsa, Callílepis, Anágraphis, Hemiclèa).
	DRÁSSIDÆ
	Eight eyes in a compact group, the first row composed of four eyes;
	cheliceræ without boss, with smooth, oblique margins; pedipalp
	of female without claw. (Prodídomus, Zimìris, Elelèis,
	Pródida) PRODIDÓMIDÆ
50.	Sternum oval or long; posterior coxæ approximated; tarsi without
	spurious claws
	Sternum wide, pointed behind; posterior coxæ far apart; tarsi
	with one pair of spurious claws; eight eyes in two rows of four,
	homogeneous, the posterior row strongly recurved and much
	wider than the anterior one. (Homalonychus). HOMALONÝCHIDÆ
E 1	All tarsi without scopula; colulus present; cheliceræ with boss,
31.	but without scopa, their margins usually smooth
	At least the first and second tarsi with scopula; colulus absent;
	cheliceræ with boss and scopula, the margins toothed53
52.	
04.	posteriorly, not reaching beyond the third coxæ; fourth meta-
	tarsi longer than the second; pedipalp of female without claw.
	(Aphantochilus, Bucrànium, Majélla).
	APHANTOCHÍLIDÆ
	Lip and sternum not thus, of the usual form; second metatarsi
	longer than the fourth; pedipalpi of female with claw. (Strò-
	phius, Stephanópsis, Philódromus, Dièta, Coriaráchne,
	Misùmena, Oxýptila, Synèma, Xýsticus). THOMÍSIDÆ
53.	Legs formed for running forwards; apex of metatarsus with a
	chitinous rim
	Legs formed for running sideways; apex of metatarsi with a soft,
	trilobate membrane; pedipalpi of female with a claw; eight
	eyes in two rows of four each. (Delèna, Spariánthis, Mi-
	crómmata, Heterópoda, Palístes, Clástes, Staianus).
	SPARÁSSIDÆ

#### LITERATURE ON ARANEIDA

- Ausserer, A. Die Arachniden-Familie der Territelariæ. Verh. k. k. zool.-bot. Ges. Wien, **21**, pp. 117–224 (1871), and **25**, pp. 156–206 (1875).
- Banks, N. Families and Genera of Araneida. American Naturalist, 39, pp. 293–323 (1905).

Catalogue of Nearctic Spiders. Bull. U. S. Nat. Mus., No. 72 (1910).

- Bishop, S. C. Revision of Pisauridæ of the United States. Bull. N. Y. State Mus., No. 252, 140 pp. (1924).
- BÖSENBERG, W. Die Spinnen Deutschlands (1901-03).
- CAMBRIDGE, F. P. Spiders of the Genus Latrodectus. Proc. Zool. Soc. London, 1892, pp. 247–261 (1892).
- Chamberlin, R. V. Revision of North American Lycosidæ. Proc. Philadelphia Acad. Nat. Sci., 1908, pp. 157–318 (1908).
  - North American Gnaphosidæ. Proc. Biol. Soc. Wash., **35**, pp. 145–172 (1922).
- CHYZER, C. and L. Kulczynski. Araneæ Hungaricæ. 2 vols. Budapest (1892–97).
- Comstock, J. H. The Spider Book. Doubleday Page & Co., Garden City, N. Y. (1912).
- Dahl, F. Die Spinnen Norddeutschlands (1883).
  - Die Lycosiden Deutschlands. Abh. K. Leop. Car. Deutsch. Akad. Naturf., 88, pp. 179–678 (1908).
    - Salticidæ in Tierwelt Deutschlands, 3, Lief. 1, 55 pp. (1926).
- Dalmas, Compte de. Araignées de Nouvelle Zélande. Ann. Soc. Entom. France, 86, pp. 317–430 (1918).
  - Synopsis des araignées de la famille des Prodidomidæ. Ann. Soc. Entom. France, 87, pp. 279–340 (1918–19).
- EMERTON, J. H. Therididæ of New England. Trans. Conn. Acad. Arts Sci., 6, pp. 1–86 (1882).
  - Epeiridæ of New England. Trans. Conn. Acad. Arts Sci., 6, pp. 295–342 (1884).
  - New England Lycosidæ. Trans. Conn. Acad. Arts Sci., 6, pp. 481–505 (1884).

Dictynidæ of New England. Trans. Conn. Acad. Arts Sci., 7, pp. 443–460 (1888).

Drassidæ, Agalenidæ and Dysderidæ of New England. Trans. Conn. Acad. Arts Sci., 8, pp. 1–40 (1890).

New England Spiders of the Family Thomisidæ. Trans. Conn. Acad. Arts Sci., 8, pp. 359–381 (1892).

Common Spiders of the United States. Boston (1902).

Supplement to New England Spiders. Trans. Conn. Acad. Arts Sci., 14, pp. 173–236 (1909).

Graveley, F. H. Indian Lycosidæ. Rec. Indian Mus., 26, pp. 587-613 (1924). Hentz, N. M. Spiders of the United States. Collected writings, edited by

J. H. Emerton. Boston Soc. Nat. Hist. (1875).

KEYSERLING, E. Die Spinnen Amerikas. Nürnberg. 1, Laterigradæ (1880); 2, Therididæ (1884–86); 3, Brazilian Spiders (1891), 4, Epeiridæ (1892–93). Koch, L. Die Arachnidenfamilie der Drassiden. (1866).

Lessert, R. Araignées du Kilimandjaro et du Mérou. Rev. Suisse Zool., 27, pp. 99–234 (1919). See also Rept. Kilimandjaro-Meru Exped. (1915).

Mello-Leitão, M. Aphantochílidas e Thomísidas do Brasil. Arch. Mus. Nac. Rio de Janeiro, **31**, pp. 1–359 (1929).

Peckham, G. W. and E. G. Revision of North American Attidæ. Trans. Wisconsin Acad. Sci., 16, pp. 355–596 (1909).

Petrunkewitch, A. Catalogue of American Spiders. Bull. American Mus. Nat. Hist., 29, pp. 1–809 (1911).

On Families of Spiders. Ann. Acad. Sci., New York, 29, pp. 145–180 (1923).

Arachnida from Panama. Trans. Conn. Acad. Sci., 27, pp. 51–248 (1925).

Systema Aranearum. Trans. Conn. Acad. Arts Sci., 29, pp. 1–270 (1928).

Spiders of Porto Rico. Trans. Conn. Acad. Arts Sci., **30**, pp. 1–355; **31**, pp. 5–191 (1930).

RAINBOW, W. J. Census of Australian Araneida. Rec. Australian Mus., 9, pp. 107–319 (1911).

Terretelariæ of Australia. Rec. Australian Mus., 10, pp. 187–270 (1914).

RAINBOW, W. J. and R. H. PULLEINE. Australian Trap-door Spiders. Rec. Australian Mus., 12, pp. 81–169 (1918).

Reimoser, E. Catalog der paläarktischen Spinnen. Abh. zool.-bot. Ges. Wien, **10**, pp. 1–280 (1919).

ROEWER, C. F. Spinnen (Araneida). In Tierwelt Mitteleuropas, 3, Lief. 2, pp. VI 1–144, (1928).

Simon, E. Aviculariides de l'Amérique du Nord. Act. Soc. Linn. Bordeaux, 44, pp. 307–339 (1892).

Histoire naturelle des Araignées. Paris, 2 vols. (1892-03).

SMITH, C. P. Theraphosidæ of California. Ann. Entom. Soc. America, 1, pp. 207–250 (1908). Thorell, T. Descriptive Catalogue of Spiders of Burma. London, British Museum, 404 pp. (1895).

Tucker, R. W. E. Drassidæ of South Africa. Ann. South African Mus., 19, pp. 251-438 (1923).

## ORDER ACARÌNA

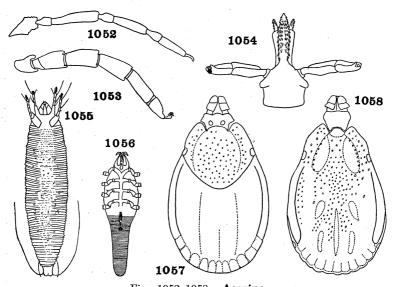
## $(AC\acute{A}RIDA)$

Usually small or minute species of rounded or oval form, with the body not divided by any deep constrictions. Abdomen not segmented although occasionally with a large number of minute transverse ridges; very broadly joined to the cephalothorax from which it is sometimes separated by a suture. Cheliceræ usually two-jointed, frequently chelate. Pedipalpi with five joints or less, usually clawed or chelate, occasionally long and serving as tactile organs. Adults usually with four pairs of legs, occasionally with only two; larva almost always with three pairs. Legs with five to seven joints, usually ending in one or two claws. Eyes usually present, commonly as one or several at each side of the body; rarely with a median one. Respiration by tubular tracheæ opening by one or several pairs of spiracles, or through the integument only. Habits varied, vegetarian or carnivorous, frequently parasitic. Mites and Ticks.

The Acarina are an extremely large and diverse group, but have received comparatively little attention. The classification given below follows in general that of Banks, while the arrangement of the water mites is that adopted by Viets. Many families and subdivisions in use by other acarologists have been merged with the larger and better known families.

- Body with four legs placed near the anterior end, each composed of five joints; plant-inhabiting species, usually forming deformations or galls. (Fig. 1055). (Eriôphyes (= Phytóptus) (E. pyri, Pear blister mite), Phyllocóptes, Anthocóptes, Tegonòtus. Epitrímerus). (PHYTÓPTIDÆ).

**ERIOPH YIDÆ** 



Figs. 1052–1058. Acarina

- 1052. Holosiro, leg (Ewing).
- 1053. Rhipicephalus, leg (Ewing) Ixodidæ.
- 1054. Ixodes, mouthparts (Nuttall) Ixodidæ.
- 1055. Eriophyes (Kendall) Eriophyidæ.
- 1056. **Demodex** (Banks) Demodicidæ.
- 1057. Rhipicephalus, female (Nuttall) Ixodidæ.
- 1058. Rhipicephalus, male (Nuttall) Ixodidæ.
  - - No such spiracle on a stigmal plate on this part of the body. .12
  - 4. Hypostome (median portion of mouthparts) large, provided below with a large number of recurved teeth or barbs; ventral surface of body posteriorly with at least one pair of furrows extending

	backwards from the genital pore; skin leathery; large species usually parasitic on mammals, more rarely on birds or reptiles. (Superfamily IXODÒIDEA)
	Hypostome small, not toothed or barbed; venter without furrows; body often with coriaceous shields; posterior margin never
	crenulated nor fluted; eyes absent. (Superfamily PARASI-TÒIDEA)
5.	Body above with a chitinous plate or scutum which almost covers
	the body of the male, but is reduced to a small plate near the front of the body in the female; body below sometimes with
	ventral shields; mouthparts of adult clearly visible from above; stigmal plate behind the coxæ; tarsus with a pulvillus. (Figs.
	1053, 1054, 1057, 1058). (Ixòdes, Dermacéntor, Hæmaphỳ-
	salis, Margáropus, Amblyómma) IXÓDIDÆ
	Body without scutum in either sex; without any ventral shield;
	mouthparts of adult almost entirely concealed from above; stigmal plate between coxæ of third and fourth legs; tarsi
	without pulvillus. (Fig. 1019). (Árgas, Ornithódoros).
	ARGÁNTIDÆ
6.	Palpi not enlarged at tip; spiracles behind the coxe of the third
	legs
	Palpi with the last joint enlarged; a spiracle above the coxa of the third leg; moderately large species. (Holòthyrus, ethiop.,
	indoaustr.)
7	Genital aperture near the anus; mouthparts capable of being re-
٠.	tracted within a large buccal cavity, which is near the anterior
	end of the body and separated from the rest of the body by a
	suture; living on bats. (Spelæorhýnchus, neotrop.).
	SPELÆORHÝNCHIDÆ
	Genital aperture not or only slightly behind the hind coxæ; an-
	terior part of body not separated by a suture; body rather broad and flat, with relatively short legs
8.	Spiracle and peritreme clearly on the ventral side of the body.
	(Fig. 1060)
	Spiracle and peritreme situated on the dorsum, or very nearly so;
	legs short and very bristly, all with large caruncles, larva
	living in body of female. (Fig. 1066). (Spintúrnix (= Pteróp-
	tus), Periglischrus, on bats; Ptilonýssus, on sparrows).
0	(PTEROPTÌNÆ) SPINTURNÍCIDÆ
9.	Anus surrounded by a chitinous plate or shield; rarely parasitic
	on mammals
	·

Anus without a chitinous plate or shield; internal parasites in the respiratory tract of mammals. (**Halaráchne**, in bronchi of seals; **Pneumonýssus**, in lungs of monkeys). (Fig. 1062).

## **HALARÁCHNIDÆ**

- - First pair of legs inserted in the mouth cavity with the oral tube; dorsum of body projecting beyond the camerostome; male genital opening in the sternal plate. Species often occurring on insects. (Urópoda, Discopòma, Cílliba, Uroplitélla, Polyáspis, Tráchytes, Dínychus, Deraióphorus).

### UROPÓDIDÆ

- 11. Claw of cheliceræ usually without teeth, often stylate or needle-like, the fixed arm always without a seta; tarsi of first pair of legs with claws or caruncles; second legs of male without processes; anus of female usually at anterior end of anal shield; all the chitinous shields rather weak and often not evident. Species parasitic on birds, mammals and reptiles. (Dermanyssus (D. gallina, Chicken mite); Liponyssus, on mammals; Ophionyssus, on snakes) . . . . . . . . . . . . . DERMANÝSSIDÆ

  - Three well-defined subfamilies may be distinguished as follows:
  - a. Body usually elongate; first pair of legs with or without claws; genital opening of male usually in front of the sternal plate; usually free-living species; rarely myrmecophilous......b
    - Body circular in outline, or nearly so; conspicuously hairy; first pair of legs greatly elongated and without claws; genital opening of male in the sternal plate; usually myrmecophilous species living externally on the bodies of ants. (Antennóphorus, Echinomegístus, Antennomegístus).

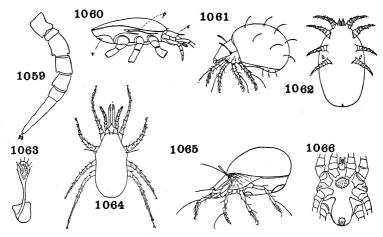
#### ANTENNOPHORÎNÆ

b. First pair of legs with claws or caruncles, used for walking; second pair of legs often enlarged in the male; species free-living or parasitic, often on vertebrates. (Figs. 1060, 1064). (Parásitus (=Gámasus), Làlaps, Echinolàlaps, Hàmogámasus).

#### PARASITÎNÆ

First pair of legs without claws or caruncle, as long as or longer than the body and serving as sensory organs; second pair of

12. Body usually coriaceous, with few hairs; with a specialized seta arising from a pore near each posterior corner of the cephalothorax (pseudostigmatic organ, Fig. 1063); eyes absent, mouthparts and palpi usually very small; ventral openings of abdo-



Figs. 1059-1066. Acarina

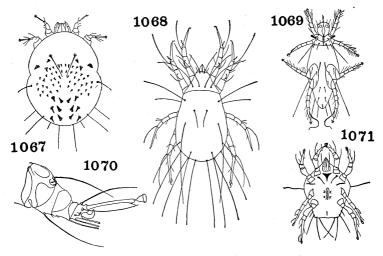
- 1059. Macrocheles, leg (Ewing) Parasitidæ.
- 1060. Parasitus (Banks); e, epistoma; p, peritreme; v, anal plate. Parasitidæ.
- 1061. Hoploderma (Banks) Hoplodermatidæ.
- 1062. Pneumonyssus (Banks) Halarachnidæ.
- 1063. Oribatidæ, pseudostigmatic organ (Banks) Orbatidæ.
- 1064. Parasitus (Banks) Parasitidæ.
- 1065. Oribatella (Banks) Oribatidæ.
- 1066. Spinturnix, underside (Banks) Spinturnicidæ.

LABIDOSTOMMÁTIDÆ

14.	Cephalothorax loosely attached to the abdomen, so that it is freely
	movable; palpi four-jointed. (Fig. 1061). (Hoplodérma,
	Phthirácarus, Protolóphora, Mesolóphora).
	HOPLODERMÁTIDÆ
	Cephalothorax immovably united with the abdomen; palpi five-
	jointed. (Figs. 1063, 1065). (Oríbata, Oribatélla, Galúmna,
	Pèlops, Hypochthònius, Trizètes, Hermánnia).
	ORIBÁTIDÆ
15	Aquatic species, living entirely in and laying their eggs in water
10.	and nearly always in fresh water; palpi four or five-jointed;
	parasitic or free-living. Superfamily HYDRACHNÒIDEA16
	Terrestrial or parasitic species, never living in water, except
	certain parasitic species with two-jointed palpi and chelate
	hind legs
16.	Basal attachment of the third and fourth pairs of legs strongly
10.	lateral in position, visible from above next to the lateral margin
	of the body, cephalothorax with two large posterior and two
	small lateral plates
	Basal attachment of the third and fourth pairs of legs not visible
	from above
17	Palpi composed of five joints; red species. (Hydrovólzia).
17.	HYDROVOLZÎIDÆ
	Palpi with only three or four joints; species living on marine alge,
	not adapted for swimming. (Halácarus, Porohalácarus, Wal-
	terélla, Soldanellònyx) HALACÁRIDÆ
10	
18.	Eyes placed near the middle of the vertex, connected by a trans-
	verse median chitinized structure
	Eyes placed at the sides of the body, not connected across the
	vertex by a chitinized structure, although sometimes with
10	chitinous plates between them
19.	Chitinized plate connecting the eyes forming a median band, i.e.
	much higher than wide; no swimming hairs on any of the legs; very soft-bodied species with thinly chitinized integument.
٠.	(Limnóchares) LIMNOCHÁRIDÆ
	Chitinized plate connecting the eyes forming a transverse band
	that is much wider than high; three anterior pairs of legs with
20	swimming hairs. (Eylàis) EYLÀIDÆ
20.	Legs six-jointed
	Legs five-jointed; body with a dorsal shield; two lateral eyes and
	a median one. (Thermácarus, holarc., in hot springs).
	THERMACÁRIDÆ

21.	Lateral eyes placed together on a chitinized tubercle or plate at each side of the head
	Lateral eyes not placed together on a chitinized plate, although
	sometimes close together as a contiguous pair
22.	Eye-plate free, not attached to any other plate. (Piersígia).
	(See couplet 19) EYLÂIDÆ, part
	Eye-plate fused laterally to a chitinous plate which is produced
	medially behind
23.	Body without a dorsal shield; legs without swimming hairs.
	(Prótzia, Calònyx, Partnùnia, Wandèsia) PROTZÌIDÆ
	Body usually with dorsal shield; legs frequently with swimming
	hairs
24.	Cheliceræ styliform; beak produced, tubular. (Hydráchna,
	Bárgena) HYDRÁCHNIDÆ
	Cheliceræ with basal segment and opposable claw, not styliform;
	beak not produced
25.	
	Second joint of palpi with a sharp projection inwardly. (Spér-
	chon, Pseudospérchon) SPERCHÓNIDÆ
26.	
	phántes) PSEUDOHYDRYPHÁNTIDÆ
	Body with a dorsal shield. (Hydryphántes, Thỳas, Viètsia,
	Pánisus, Mamérsa, Eupàtra, Georgélla).
07	HYDRYPHÁNTIDÆ
27.	Eyes separated, not forming contiguous double-eyes
90	Eyes united to form a double-eye at each side of the body. 29
48.	Fourth pair of legs with apical claws; palpi distinctly chelate at tip. (Diplodóntus, Oxyópsis) . HYDRYPHÁNTIDÆ, part
	Fourth pair of legs without apical claws; palpi not chelate at tip.
	(Limnèsia, Limnesiélla, Duralimnèsia) LIMNESÌIDÆ
29.	
-0.	Last joint of palpi forming a stout hook which is opposable to the
	broad tip of the preceding joint. (Arrhenurus, Thoraco-
	phorácarus, Mundamélla, Wùria) ARRHENÙRIDÆ
30.	Epimera of the fourth pair of legs with a large pore near the inner
	angle; tarsus of fourth leg pointed, without claws. (Teutònia)
	LEBERTÌIDÆ, part
	Epimera of the fourth leg without a pore
31.	Second joint of palpi with a strong bristle inwardly which often
	arises from a sharp projection32
	Second joint of palpi without a sharp projection or bristle34

32.	Third joint of palpi bearing five to seven stout bristles that are longer than the joint. (Lebértia, Pilolebértia, Hexalebértia).  LEBERTIIDÆ
	Third joint of palpi without such bristly hairs
99	Challenge land hash like an extensile within a tube formed by the
აა.	Cheliceræ long, beak-like or extensile within a tube formed by the
	epistoma; carapace composed of several plates. (Atractides,
	Pseudotorrenticola) ATRACTÍDIDÆ
	Cheliceræ of a different conformation. (See couplet 32). (Dártia,
0.4	Bandàkia) LEBERTÌIDÆ, part
34.	Body strongly compressed from the sides, or narrowly elongate-
	oval; legs inserted one above another near the anterior end of
	the body; fourth pair of legs without apical claws. (See couplet
	33). (Óxus, Gnaphíscus, Frontípoda).
	ATRACTÍDIDÆ, part
	Body not compressed nor narrowly elongate; legs inserted one
	behind another in the usual position
35.	Epimera of the first pair of legs free, not fused medially behind
	the maxillæ36
	Epimera of the first pair of legs completely fused. (Hygróbates,
	Mégapus) HYGROBÁTIDÆ
36.	First pair of legs bearing blunt bristles, usually placed in pairs
	on tubercles. (Unionícola, Neumánnia, Huitféldtia).
	HYGROBÁTIDÆ, part
	First pair of legs not thus bristly
37.	
	bearing shields; usually convex species. (Féltria, Nauta-
	ráchna, Foréllia, Piòna, Hydrochoreùtes, Acércus).
	HYGROBÁTIDÆ, part
	Body with pore-bearing plates; usually flattened. (Midea, Mi-
	deópsis, Brachypoda, Atùrus, Álbia). BRACHYPÓDIDÆ
38.	Palpi two-jointed, partly enclosing the cheliceræ; third and
	fourth pairs of legs in both sexes greatly thickened, the last two
	joints opposable to form grasping organs; tarsi without suckers;
	parasitic in the gills of crabs. (Ewingia, nearc.). EWINGIIDÆ
	Palpi with three or more joints; rarely the fourth pair of legs en-
	larged to form a grasping organ in the male only, in which case
	the other tarsi bear suckers39
39.	Palpi small, three-jointed, adhering for some distance to the beak;
	body usually with ventral suckers at the genital opening or near
	the anal opening; eyes absent; tarsi often with suckers at tip;
	frequently parasitic (Superfamily ACARÒIDEA) 40



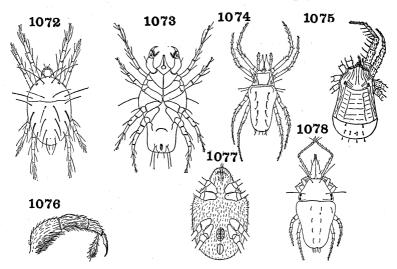
Figs. 1067-1071. **Acarina** 

- 1067. Acarus (Banks) Acaridæ.
- 1068. **Tyroglyphus** (Banks) Tyroglyphidæ.
- 1069. Pediculoides (Banks) Tarsonemidæ.
- 1070. Acarus, leg (Banks) Acaridæ.
- 1071. Canestrinia (Banks) Canestriniidæ.

42.	Legs short; the tarsi of the first and second pairs without clavate
	hairs, terminating in suckers; species parasitic on insects.
	(Fig. 1071). (Canestrínia, Hemisarcóptes, Linòbia, Coleop-
	$ ext{ter\'ophagus})$
	Legs longer; the tarsi of the first and second pairs of legs ex-
	ternally near the base with a clavate hair; usually free-living,
	rarely parasitic on insects. (Fig. 1068). (Tyróglyphus, Rhizó-
	glyphus, Aleuròbius, Histiosòma, Glycíphagus, Monie-
	ziélla, Trichotársus) TYROGLÝPHIDÆ
<b>4</b> 3.	External parasites of mammals; legs or mouthparts modified to
	form clasping organs to cling to the hairs of the host. (Lis-
	tróphorus, Labidocárpus, Schizocárpus, Myocóptes, Tri-
	chècius) LISTROPHÓRIDÆ
	Species not living externally on the hair of mammals and without
	such specialized clinging apparatus
44.	External parasites of birds, living on the plumage. (Análges,
	Freyàna, Pterólichus, Pterodéctes, Allóptes).
	ANALGÉSIDÆ
	Internal parasites living in the skin or tissues of birds, and in or
	on the skin of mammals
45.	Genital aperture of female longitudinal in position; species living
	in the skin or internal tissues of birds. (Cytoleichus, Lamin-
	osióptes). $(CYTODOTIDÆ)$ CYTOLEÌCHIDÆ
	Genital aperture of female transverse; skin parasites of mammals
	or birds. (Figs. 1067, 1070). (Acarus (= Sarcoptes) (Itch
	mites), Psoróptes, Chorióptes, Cnemidóptes). $(SARCÓP-$
	TID $E$ ) ACÁRID $E$
46.	Last joint of palpi opposable like a thumb to the preceding joint
	which is nearly always claw-like at tip (Fig. 1076); body often
	bearing many hairs. (Superfamily TROMBIDÒIDEA)47
	Palpi not thus modified; simple or rarely modified to hold prey;
	body with few hairs. (Superfamily EUPODOIDEA)52
47.	
	with inconspicuous chitinous shields48
	First and second pairs of legs with processes bearing spines;
	cephalothorax with a large, sculptured, median chitinous shield,
	abdomen with two or more similar shields; coxæ contiguous
	(Fig. 1075). (Cæculus)
48.	1 1
	posable to the basal portion
	Cheliceræ stylet-shaped or needle-like, retractile50

49. Last joint of first pair of legs usually enlarged; cephalothorax with a median dorsal line or groove; legs inserted in two groups, the coxe of the first two pairs placed far forward and those of the two posterior pairs in a second group far removed from the first. (Fig. 1076). Harvest mites. (**Trombidium**).

## TROMBIDÌIDÆ



Figs. 1072-1078. Acarina

- 1072. Tetranychus (Banks) Tetranychidæ.
- 1073. Cheyletus (Banks) Cheyletidæ.
- 1074. Rhagidia (Banks) Eupodidæ.
- 1075. Cæculus (Banks) Cæculidæ.
- 1076. Trombidium, palpi (Banks) Trombidiidæ.
- 1077. Smaris (Banks) Erythræidæ.
- 1078. Bdella (Banks) Bdellidæ.

Last joint of first pair of legs not enlarged, often long; cephalothorax without median line or groove; coxæ of all legs usually closely approximated, forming a single group. (Anýstis, Tarsótomus, Gekòbia, Pterygosòma) .................................. ANÝSTIDÆ

Last joint of first and fourth pairs of legs enlarged; cephalothorax usually with a median dorsal groove; adults free-living, usually

active and with long slender legs. (Fig. 1077). (**Erythræus** (= Rhyncholophus), **Smàris, Fessònia, Átomus, Eatoniàna**). (Including  $SM\acute{A}RID\pounds$ ). ( $RH\acute{Y}NCHOL\acute{O}PHID\pounds$ ).

## ERYTHRÆIDÆ

- - Predaceous or parasitic; claws often absent on one or more pairs of legs; body bristles often branched or pectinate. (Fig. 1073). (Cheylètus, Cheylètia, Cheyletiélla, Myòbia, Picòbia). CHEYLÉTIDÆ
- 52. Cephalothorax and abdomen completely fused, the body showing no division into these two parts; no bristles on body above; palpi simple; last joint of first pair of legs not noticeably longer than the preceding joint; mouthparts covered by a large hyaline hood. (Cryptógnathus, palæarc.). CRYPTOGNÁTHIDÆ
  - Cephalothorax and abdomen more or less distinctly separated; upper surface of body furnished with bristles; palpi sometimes geniculate; mouthparts without hood-like covering.......53
- 53. Last joint of first pair of legs longer than the preceding joint, often twice as long; cephalothorax with four bristles, a pair in front and a more widely separated pair behind. (Bdélla (Fig. 1078), Cunáxa, Cŷta, Eùpalus, Scírula). (Including CU-NÁXIDÆ)
  - Last joint of first pair of legs not or only slightly longer than the preceding joint; bristles of cephalothorax not arranged as above. (Eupòdes, Linopòdes, Notophállus, Tỳdeus, Rhagídia (Fig. 1074), Ereynètes, Álichus, Bimichælia, Teneríffia). (Including TE NERIFFÌIDÆ) ..... EUPÓDIDÆ

#### LITERATURE ON ACARINA

- André, M. Contribution a l'étude des Acariens libres. Trombididæ. Bull. Soc. Zool. Paris, **51**, pp. 175–228 (1926).
- Bagnall, R. S. and J. W. H. Harrison. A Catalogue of the British Eriophyidæ. Ann. Mag. Nat. Hist., (10) 2, pp. 427–445 (1928).
- Banks, N. Oribatoidea of the United States. Trans. Amer. Entom. Soc., 5, pp. 1-16 (1895).
  - Catalogue of the Acarina of the United States. Proc. U. S. Nat. Mus., 32, pp. 595-625 (1907).

Revision of Ixodoidea of the United States. Bull. U. S. Dept. Agric., Bur. Entom. Tech. ser., No. 15 (1908).

The Acarina or Mites. Rept. No. 108, U. S. Dept. Agric. 153 pp. (1915).

Berlese, A. Acari, Miriopodi e Scorpione Italiani. Prostigmata, Cryptostigmata, Mesostigmata, Sarcoptidæ. 4 vols. (1882–1910).

Monografia del Genere Gamasus. Redia, 3, pp. 68-304 (1905).

Trombidiidæ. Redia, 8, pp. 1–291 (1912).

- Canestrini, G. Prospetto dell' Acarofauna Italiana. Padova, 7 parts (1885–97).
- Canestrini, G. and P. Kramer. Demodicoidea. Das Tierreich, Lief. 7, pp. 1–3 (1899).

Sarcoptidæ. Das Tierreich, Lief. 7, pp. 4-193 (1899).

Ewing, H. E. Oribatoidea of Illinois. Bull. Ill. State Lab. Nat. Hist., 7, pp. 337–389 (1909).

Synopsis of Beetle-mites (North America). Ann. Entom. Soc. America, **10**, pp. 117–132 (1917).

Classification of Mites. Ann. Entom. Soc. America, **15**, pp. 213–222 (1922).

- FIELDING, J. W. Australasian Ticks. Service Publ. Australia, 9, pp. 1–114 (1926).
- Jacot, A. P. Annotated Bibliography of the Oribatoidea. Tsingtao, pp. 1–60 (1929).

Oribatid Mites of the Subfamily Phthiracarinæ of Northeastern United States. Proc. Boston. Soc. Nat. Hist., 39, pp. 209–261 (1930).

- JAKOB, E. Die Verwandschaft der Zeckengattungen. Zeits. Morph. Ökol., 1, pp. 309–372 (1924).
- Koenicke, F. Nordamerikanische Hydrachniden. Abh. naturw. Ver. Bremen, 13, pp. 167–226 (1900).

Acarinen- insbesondere Hydracarinen- System. Abh. naturw. Ver. Bremen, **20**, pp. 121–161 (1910).

Acarina. Süsswasserfauna Deutschlands, Heft 12, (1916).

- Lundblad, O. Süsswasseracarinen aus Dänemark. Vid. Selsk. Skr. Math.natv. Afd. Copenhagen, (8) 6, No. 2, 150 pp. (1920).
- MARSHALL, R. The Arrhenuri of the United States. Trans. Am. Microscop. Soc., 28, pp. 851-40 (1908).
- McGregor, E. A. Red Spiders of America. Proc. U. S. Nat. Mus., 56, pp. 641-679 (1919).
- MICHAEL, A. D. Oribatidæ. Das Tierreich, Lief. 53, 93 pp. (1898). British Oribatidæ. 2 vols. London, Ray Soc. (1883–87).

British Tyroglyphidæ, 2 vols. London, Ray Soc. (1901–03).

Nalepa, A. Eriophyidæ. Das Tierreich, Lief. 4 (1898).

Die Systematik der Eriophyiden. Verh. zool.-bot. Ges. Wien, 67, pp. 12–38 (1917).

Index nominum Eriophyidarum. Marcellia, Avellino, **20**, pp. 25–66 (1923).

- Neuer Catalog der bisher beschriebenen Gallmilben. Marcellia, **25**, pp. 67–183 (1929).
- Neumann, L. G. Revision de la Famille des Ixodides. Mém. Soc. Zool. France,
  9, pp. 1–44; 10, pp. 324–420; 11, pp. 120–230; 12, pp. 249–372 (1896–01).
  Ixodidæ. Das Tierreich, Lief. 26, 166 pp. (1911).
- Nuttall, G. H. F. and C. Warburton. Ticks of the Belgian Congo. Bull. Entom. Res., 6, pp. 313–352 (1916).
- NUTTALL, G. H. F., C. WARBURTON, W. F. COOPER, and L. E. ROBINSON. A Monograph of the Ixodoidea. Cambridge University Press, London, pt. 1 (1908); pt. 2 (1911); pt. 3 (1915).
- OUDEMANS, A. C. New List of Dutch Acari. Tijdschr. v. Entom., 43 (1901); 45 (1902).
  - Revision des Chélétines. Mém. Soc. Zool. France, 18, pp. 36–144 (1906). Analytical Key for the Classification of Families and Genera of Diacrotricha. Entom. Berich., 6, pp. 226–235 (1924).
  - Kritisch historisch Overzicht der Acarologie. Tijdschr. Entom, Suppl. to 69, 500 pp. (1926).
- Paoli, G. Monografia dei Tarsonemidi. Redia, 7, pp. 215–281 (1911).
- Piersic, R. Deutschlands Hydrachniden. Zoologica, Heft 22 (1897–1900). Hydrachnidæ. Das Tierreich, Lief. 13, 336 pp. (1901).
- RAINBOW, W. J. A Synopsis of Australian Acarina. Rec. Australian Mus., 6, pp. 145–193 (1906).
- Schulze, P. Ixodoidea. Biol. Tiere Deutschlands, Lief. 2 (1923). Zecken (Ixodoidea). Tierwelt Mitteleuropas, **3**, Lief. 4, pp. X 1–10
- (1929).
  Sellnick, M. Die Tardigraden und Oribatiden der ostpreussischen Moosrasen.
  Schr. physik. Ges. Königsberg, 49, pp. 317–350 (1908).
  - Oribatei (Oribatoidea). In Tierwelt Mitteleuropas, 3, Lief. 4, pp. IX 1–42 (1929).
- SOAR, C. D. Acarina or Mites. Watson's Micros Rec., London, Nos. 6-9 (various parts).
- Soar, C. D. and Williamson, W. The British Hydracharina, 3 vols. Ray Soc. London (1925–29).
- Thon, K. Monographie der Hydrachniden Böhmens. Arch. naturw. Landesdurchforsch. Prag, 12, pt. 2, pp. 1–84 (1906).
- Thor, S. Zur Systematik der Acarinenfamilien Bdellidæ, Eupodidæ und Cunaxidæ. Ver. k. k. zool.-botan. Ges. Wien, **52**, pp. 159–165 (1902).
- Trägårdh, I. Monographie der arktischen Acariden. Fauna Arctica, 4, pp. 1–78 (1904).
- Trouessart, E. L. Revision des Genres des Analgesinæ. Bull. Soc. Zool. Paris, 40, pp. 207–223 (1916).
- VIETS, K. Fortschritte in der Kenntnis der Hydracarinen (Bibliography).
  Arch. Hydrobiol. u. Planktonk., 8, pp. 589-629 (1913); 9, pp. 550-578 (1914).
  Versuch eines Systems der Hydracariden. Zool. Anz., 69, pp. 188-199 (1926).

Die Halacaridæ der Nordsee. Zeits. wiss. Zool., **130**, pp. 83–173 (1927). Wassermilben (Aquatic Acarina). In Tierwelt Mitteleuropas, **3**, Lief. **4**, pp. VIII 1–57 (1928).

Halacaridæ. In Die Tierwelt der Nord- und Ostsee. XI C. 72 pp. Grimpe and Wagner (1928).

VITZTHUM, H. Acari aus Spanien. Senckenbergiana, Frankfurt-am-M., 8, pp. 30–39 (1926).

Malayische Acari. Treubia, 8, pp. 1-198 (1926).

Landmilben (Terrestrial Acarina). In Tierwelt Mitteleuropas, 3, Lief. 3, pp. VII 1-112 (1928).

Walter, C. Die Hydracarinen der Schweitz. Rev. Suisse Zool., 15 (1907).
Wolcott, R. H. A Review of the Genera of Water Mites. Trans. Am. Microscop. Soc., 26, pp. 161–243 (1905).

## ORDER PENTASTÓMIDA

# (LINGUATÙLIDA)

Rather large, elongate, vermiform, cylindrical or somewhat flattened animals. Body divided externally by constrictions into a large number of annuli or apparent segments; straight, bent or sometimes coiled. Mature form legless, but the embryo and very young animal with two or three pairs of short, imperfectly jointed legs. No antennæ. Mouth opening rounded or oval, situated close to two pairs of claw-like, movable chitinous hooks that arise from shallow pits. Respiratory organs wanting. Genital opening of male always at the base of the abdomen, of the female either at the base or apex of the abdomen; ovary or uterus either large and sac-like, or long, tubular and strongly sinuous or coiled, lying either above, below or twined about the alimentary tract. Metamorphosis slight, but evident. Parasitic; the adult form usually in the respiratory tract of reptiles, the immature one frequently in the tissues or internal organs of mammals. Tongueworms.

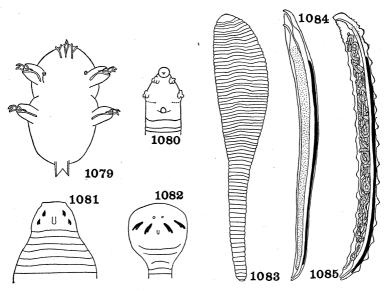
The following classification is similar to that developed by Sambon, except that his subfamilies are regarded as families to render the divisions of this group more nearly on an equality with those of related orders.

1. Mouth opening lying in front of the oral hooks; genital opening of female at the anterior end of the abdomen, the uterus large and sac-like; salivary glands of moderate size; larva with three pairs of short legs. (Figs. 1080, 1084). (RAILLIETIÉLLIDÆ).

CEPHALOBÆNIDÆ

Three subfamilies may be distinguished as follows:

a. Anterior end of body forming a short, blunt beak. (Cephalobæna, neotrop., in lungs of snakes).... CEPHALOBÆNÎNÆ
 Anterior end of body not forming a beak-like projection, rounded.b



Figs. 1079-1985. Pentastomida

- 1079. "Pentastomum," young larva (Stiles) Linguatulidæ.
- 1080. Raillietiella, anterior extremity (Sambon) Cephalobænidæ.
- 1081. Sebekia, anterior extremity (Sambon) Linguatulidæ.
- 1082. Kiricephalus, anterior extremity (Sambon) Linguatulidæ.
- 1083. Linguatula (Sambon) Linguatulidæ.
- 1084. **Cephalobænidæ**, diagrammatic vertical section of body (Sambon) Cephalobænidæ.
- 1085. Linguatulidæ, diagrammatic vertical section of body (Sambon) Linguatulidæ.
  - b. Body with two apical projections at the posterior end; posterior pair of oral hooks larger than the anterior ones. (Fig. 1080). (Raillietiélla, in reptiles and batrachians, widespr.).

#### RAILLIETIELLÎNÆ

Body without apical projections, bent downwards at posterior end; both pairs of oral hooks very small. (Reighárdia, in birds, neotrop.) ...... REIGHARDIĨNÆ

Mouth opening lying between or behind the oral hooks; genital opening of female at posterior end of abdomen, the uterus tubular, greatly elongated and irregularly coiled; salivary glands as long as the body; larva with only two pairs of legs. (Figs. 1079, 1081, 1082, 1083, 1085). (POROCEPHÁLIDÆ).

## LINGUATULIDÆ

This family includes three well-marked subfamilies, as follows:

*Note*: **Pentastoma** and **Pentastomum** have in the past been used as more or less inclusive generic names for the adult and immature stages of various species.

#### LITERATURE ON PENTASTOMIDA

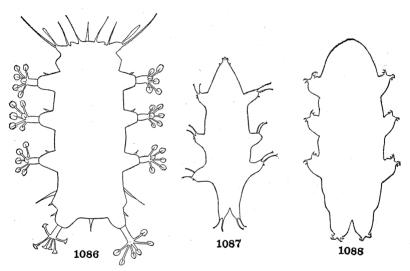
Heymons, R. Pentastomida. Kükenthal und Krumbach, Handbuch der Zoologie, 3, pt. 1, pp. 69–131 (1926).

Zungenwürmer, in Tierwelt Mitteleuropas, 3, Lief. 1, pp. 1–2 (1926).

Sambon, L. W. A Synopsis of the Family Linguatulidæ. Journ. Trop. Med., 25, pp. 188-206; 391-428 (1922).

# CLASS TARDÍGRADA

Very small or minute species, usually less than one mm. in length. Body more or less elongate oval, consisting of a head and four bodysegments; integument tough and more or less transparent, not chitinous. Four pairs of short, fleshy, unjointed legs, each terminated by several claws or clavate vesicles. Cuticula usually forming distinct plates; often granulate or sometimes spinose. Mouthparts tubular, provided with a central piercing organ or stylet. Antennæ absent. Eyes absent, or present as two ocelli. Terrestrial or aquatic species; no special respiratory organs. Species capable of excessive and prolonged dessication without injury. Sexes separate; development from the egg direct, without metamorphosis. Water Bears.



Figs. 1086-1088. Tardigrada

- 1086. Batillipes (Marcus) Halechiniscidæ.
- 1087. Milnesium (Marcus) Milnesiidæ.
- 1088. Macrobiotus (Marcus) Macrobiotidæ.
- Head bearing in front a pair of central and a pair of lateral cirri, in addition to two sensory appendages at each side. (Fig. 1086). (Order HETEROTARDÍGRADA)
   Head usually without such sensory appendages, or at least with only the four anterior ones present; the two at each side always wanting. (Order EUTARDÍGRADA)
- 2. Middle portion of leg capable of being withdrawn or telescoped within the basal part, the two parts separated by a distinct fold;

#### LITERATURE ON TARDIGRADA

Amman, J. Schweizerische Tardigraden. Berne (1908).

Carlzon, C. Schwedische Tardigraden. Zool. Anz., 34 (1909).

Forel, F. A. Tardigrades. Le Léman, Monographie Limnologique, 3 (1904). MARCUS, E. Tardigrada. Tierwelt Deutschlands, 12, 230 pp. (1928).

Tardigrada. In Bronn's Klassen und Ordnungen des Tierreichs, 5, Abth. 4, Buch 3, 608 pp. (1929).

NEDERSTRÖM, P. Die aus Finland bekannten Tardigraden. Act. Soc. Faun. Flor. Fennica, 46, No. 8 (1919).

Rahm, G. Tardigrada, Bärtierchen. Biol. Tiere Deutschlands., pt. 22, pp. 1–56, Geb. Bornträger, Berlin (1927).

RICHTERS, F. Tardigrada. In Kükenthal und Krumbach, Handbuch der Zoologie, 3, pp. 1–68 (1926).

# CLASS PAURÓPODA

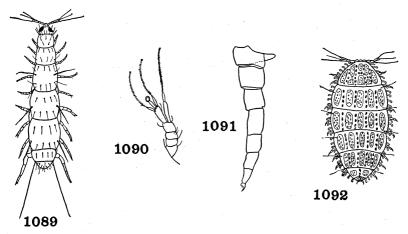
Very small species of elongate, rounded and more or less flattened form. Body composed of from six to ten segments, as indicated by the tergites, and bearing nine pairs of legs. Antennæ branched apically; composed of four stout basal joints, the fourth bearing two elongate joints, one of which bears at tip a whip-like many-jointed flagellum and the other bears two such flagella. Mouthparts composed of a pair of mandibles, and one pair of maxillæ. Cerci bristle-like. Tracheæ absent; no respiratory organs.

 with a few bristles above; anal segment exposed, not covered by the preceding tergite. (Brachypaùropus, palæarc.).

#### BRACHYPAUROPÓDIDÆ

2. Integument thinly chitinized, with a few bristles above; usually white or pale colored; head and anal segment free, not covered above. (Figs. 1089, 1091). (Paùropus, Allopaùropus, Stylipaùropus, Hemipaùropus, Scleropaùropus).

PAUROPÓDIDÆ



Figs. 1089-1092. Pauropoda

1089. Pauropus (Latzel) Pauropodidæ.

1090. Eurypauropus, antenna (Latzel) Eurypauropodidæ.

1091. Pauropus, leg (Ewing) Pauropodidæ.

1092. Eurypauropus (Latzel) Eurypauropodidæ.

#### LITERATURE ON PAUROPODA

Kenyon, F. C. The Morphology and Classification of the Pauropoda. Tufts College Studies, No. 4 (1895).

LATZEL, R. Die Myriapoden der Œsterreichisch-Ungarischen Monarchie. Vienna, Alb. Hölder (1880-84).

## CLASS DIPLÓPODA

Body long, comparatively narrow and of generally uniform width; cylindrical or at least not usually noticeably depressed or flattened. Eleven or more body segments or somites; thirteen or more pairs of legs; usually considerably more. Legs always more numerous than the dorsal segments, most of the apparent segments bearing each two pairs of legs, but the first three or four and the last one or two with only a single pair. Head with a pair of short, usually seven-jointed, antennæ, a pair of lateral eyes and a pair of mandibles; other mouth-parts forming a plate-like structure (gnathochilarium). Eyes two, consisting of a group of ocelli. Legs usually six or seven-jointed, bearing an apical claw. Genital ducts opening between the second and third pairs of legs. Respiration through spiracles opening into paired pockets, one or two pairs corresponding to each of the somites. Second or fourth legs of male usually modified into copulatory appendages. Somites often with paired repugnatorial glands, opening laterally.

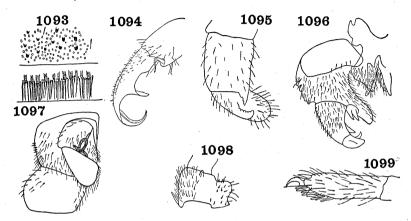
Slow moving, usually vegetarian animals. A large group, more abundant in the tropics. Millipedes.

The System adopted here follows that of Attems in his recent "Myriopoda of South Africa."

- 2. Sternites divided into two parts, tracheæ dichotomously branched; male with one or two pairs of modified copulatory appendages (telopods) at the posterior end of the body, but without modified legs on the seventh segment. (OPISTHÁNDRIA)......3

  Sternites not divided; tracheæ not branched, but arising in tufts from the tracheal trunks; male with one or two pairs of modified legs (gonopods) on the seventh body segment. (PROTERÁN-

DRIA) ...... 7



Figs. 1093-1099. Diplopoda

- 1093. **Sphærotherium,** pygidium and marginal bristles (Attems) Sphærotheriidæ.
- 1094. Sphæriodesmus, gonopod (Chamberlin) Sphæriodesmidæ.
- 1095. Sphærotherium, anterior gonopod (Attems) Sphærotheriidæ.
- 1096. Sphærotherium, posterior gonopod (Attems) Sphærotheriidæ.
- 1097. Sphærotherium, vulva (Attems) Sphærotheriidæ.
- 1098. Sphærotherium, coxa of fourth leg (Attems) Sphærotheriidæ.
- 1099. Sphærotherium, last joint of a leg (Attems) Sphærotheriidæ.

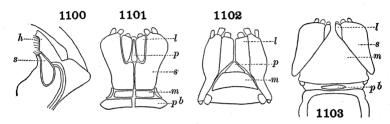
  - 4. Seventeen pairs of legs, not including the telopods of the male; ocelli in a single row or wanting. (GLOMERÍDIA).......5
    - Twenty-one pairs of legs, not including the telopods of the male; ocelli numerous, in several rows. (Figs. 1095, 1096, 1097, 1098, 1099). (SPHÆROTHÈRIA). (Sphærothèrium, Globothèrium, ethiop.; Sphæropèus, Arthrosphæra, indomal.; Cyliosòma, austr.) ..... SPHÆROTHERÌIDÆ

5.	Seventeenth pair of legs of male with one to three small joints
	beyond the large coxa. (Glómeris, palæarc.; Rhopalómeris,
	Apiómerus, indomal.) GLOMÉRIDÆ
	Seventeenth pair of legs of male with four or five small joints be-
	yond the large coxa (Fig. 127)
6.	Nineteenth pair of legs in male with the second and third joints
	(prefemur and femur) simple, without a bristly projection;
	body smooth. (Glomeridélla, Typhloglómeris, Glomerel-
	lìna, palæarc.) GLOMERIDÉLLIDÆ
	Nineteenth pair of legs in male with a bristle-bearing projection on
	the prefemur and femur; body often tuberculate or carinate.
	(Gervàsia, Dodèria, palæarc.) GERVAISÌIDÆ
7.	First pair of legs of the seventh somite of the male modified into
	gonopods, often the second pair of this somite also and the
	adjacent pairs on the sixth and eighth segments similarly modi-
	fied; mouthparts with three pairs of palpal lobes. (EU-
	GNATHA)
	First pair of legs of the seventh somite of the male normal, not
	modified to form gonopods; mouthparts without palpal lobes.
	(COLOBÓGNATHA)
8.	Anal segment without spinning styles or spinning glands; body
	of less than 23 or more than 40 somites9
	Anal segment with two or three pairs of spinning glands, opening
	by one to three pairs of spinning styles; body of 26 or more
	somites. (NEMATÓPHORA)
9.	Nineteen to 22 body somites (usually 20); only the first pair of legs
	on the seventh somite of the male modified into gonopods; the
	gonopods inserted in a completely closed circular opening; body
	cylindrical, or flattened with lateral expansions. (POLYDES-
	MOIDEA)
	More than 40 body somites; both pairs of legs of seventh somite of
	the male modified into gonopods, or the second pair absent;
	opening in which the gonopods are inserted never closed in
	front; body cylindrical. (JULIFÓRMIA)
10.	Coxe of the gonopods rather widely separated, but connected by
	broad, medially coalescent processes; the median part raised
	and keel-shaped. (POLYDESMÍDEA)
	Coxe of the gonopods close together, free or more or less connected
	but not by broad, medially keel-like, raised processes. (STRON-
1 1	GYLOSOMATÍDEA)
11.	Coxæ of the gonopods with a hooked process on the external

	margin; tergites broadly expanded at the sides, the lateral mar-
	gins produced, with a number of sharp teeth, each tooth bearing
	a bristle. (Peridontodésmus, neotrop.).
	PERIDONTODÉSMIDÆ
	Of a different conformation
12.	Head not or incompletely covered by the first tergite, which is
	generally small, not lengthened or enlarged
	Head completely or almost completely hidden by the first tergite
	when seen from above; this tergite large16
13.	Coxa of gonopod bearing a long, annulate flagellum; posterior
	portion of each tergite with three transverse rows of bristles,
	which usually arise from small tubercles. (Mastigonodésmus,
	Schedoleiodésmus, palæarc.) MASTIGONODÉSMIDÆ
	Of a different conformation14
14.	Second tergite enlarged; posterior portion of tergites smooth or
	granulate, often densely clothed with setæ. (Oniscodésmus,
	Trigonóstylus, Oncodésmus, neotrop.; Doratodésmus,
	mal.; Hyperóthrix, ethiop.) ONISCODÉSMIDÆ
	Second tergite not enlarged15
15.	Gonopod with a brush of hairs inwardly near the tip and a seminal
	pouch which opens at the base of this brush. (Fig. 1101).
	(Polydésmus, holarc.; Brachydésmus, Epanerchòdus, pal-
	æarc.; Opisthotrètus, mal.; Opisthoporodésmus, austro-
	mal.) POLYDÉSMIDÆ
	Gonopod without brush of hairs or seminal pouch. (Vanhoef-
	fènia, ethiop.; Gyrophállus, nearc.; Trichopolydésmus,
	palæarc.) VANHŒFFENÎIDÆ
16.	
	transparent projections from the sides of some of the abdominal
	tergites (usually 5th, 7th, 9th, 10th, 12th, 13th and 15th). (Sty-
	lodésmus, ethiop.; Myrmecodésmus, Corypherépsis, Sy-
	noptùra, neotrop.; Myxodésmus, mal.). STYLODÉSMIDÆ
	Openings of the repugnatorial glands very minute or not visible.
	(Cryptodésmus, neotrop.; Phenacóporus, austromal.; Cryp-
	tocórypha, ethiop., mal.; Atopodésmus, austr.).
	CRYPTODÉSMIDÆ
17.	
	ess
	Coxa of the gonopod without any such process; 20 body somites.
	(Rhacodésmus, Am.; Acutángulus, Rhachidomórpha,

18. Certain joints of the anterior (or more) legs furnished with peculiar bristles that have a bulbously swollen base, and long slender or short nipple-shaped tip. (Sphærotrichopus, austr.; Pleonaràius, Semnosòma, neotrop.; Scytonòtus, nearc., austr.; Icosidésmus, ethiop., austr.; Gnoméskelus, ethiop.).

SPHÆROTRICHÓPIDÆ



Figs. 1100-1103. Diplopoda

- 1100. Polydesmus, gonopod: h, hair-brush; s., seminal vesicle.
- 1101. **Blaniulus**, gnathochilarium: l. lingual lamellæ; p., promentum; s., stipes; pb., prebasilare. Blaniulidæ.
- 1102. Xiphochætoporatia, gnathochilarium. Lettering as in Fig. 1101.
- 1103. Rhinocricus, gnathochilarium. Lettering as in Fig. 1101.

- 21. Coxe of the gonopods large, connected by a narrow band; femur with a large process that is often longer than the tibia and tarsus together. (Leptodésmus, Cyclorhábdus, neotrop.; Ísaphe, nearc.; Devillea, palæarc.) ..... LEPTODÉSMIDÆ
  - Coxæ of the gonopods free, not connected with one another. (Oxydésmus, Metaphóricus, Orodésmus, ethiop.).

OXYDÉSMIDÆ

22.	Lateral expansions of the tergites well developed or large, more or less horizontal
	Lateral expansions of the tergites very strongly declivous, almost
	vertical; body very strongly convex above, its form highly
	adapted for curling up tightly. (Sphæriodésmus, Cyclodés-
	mus, Colobodésmus, neotrop.; Desmònus, nearc.).
	SPHÆRIODÉSMIDÆ
23.	Legs of male with fleshy lobes beneath the claws; gonopod articu-
	lated at the end of the coxa; tergites without striking sculpture.
	(Gomphodésmus, Antíphonus, Aulodésmus, Ulodésmus,
	Astrodésmus, Elaphógonus, ethiop.). GOMPHODÉSMIDÆ
	Legs of male without lobes beneath the claws
24.	Tibia and tarsus of gonopods not separated. (Fontària, holarc.;
	Rhysodésmus, neotrop., palæarc.; Mélaphe, palæarc.; Pachy-
	désmus, nearc.; Tuberculàrium, ethiop.; Asphalidésmus,
	austr.) FONTARÌIDÆ
	Tibia and tarsus of gonopods usually separated by a distinct con-
	striction; gonopod articulated at the end of the coxa. (Platý-
	rhacus, neotrop., indoaustr.; Amplinus, Pycnótropis, neo-
	trop.; Polýlepis, mal.) PLATYRHÁCIDÆ
25.	Body of $26-32$ (usually $30$ ) somites; repugnatorial glands absent . $26$
	Body of 39 or more somites; repugnatorial glands present on the
	fifth and following somites
26.	First tergite small; each somite above with three pairs of bristles
	arising from minute tubercles. (Superfamily CHORDEUMÒI-
	DEA)
	First tergite very large, its sides partly enclosing the head; body
	capable of being rolled into a spiral; some tergites with strong
	longitudinal carinæ; a few ocelli at each side of the head.
	(Superfamily STRIARIÒIDEA). (Striària, nearc.).
O.	STRIARÎIDÆ
27.	First four joints of antennæ short, subequal in length; fifth much
	longer and thicker, forming a club together with the apical
	joints; dorsum tuberculate or spinulose, very strongly convex; legs short and stout. (TRACHYZONA)
	First three joints of antennæ increasing in length, the second and
	third each much longer than the preceding joint; fourth at
	least twice as long as the first; legs long and slender. (XESTO-
	ZÒNA)
28.	
	leinosòma, palæarc.) TRACHYSOMÁTIDÆ
	. r

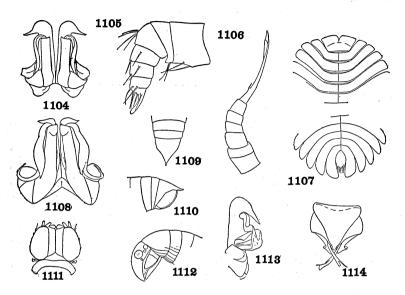
	Body surface spinulose, dark in color; 28 somites. (Chamæo-sòma, Achrochórdum, palæarc.). CHAMÆOSOMÁTIDÆ
29.	Promentum present as a separate, more or less triangular sclerite
	in front of the mentum (Fig. 1102)30
	Promentum absent
30.	Hind gonopod forming a large, stout, unjointed club
	Hind gonopod never forming a stout, unjointed club; usually one-
	or two-jointed, or sometimes more or less like a walking leg, with
	three to six joints
31.	Sides of body longitudinally striate; tergites without lateral
	expansions, (Caseya, nearc.) CASEYIDÆ
	Sides of body not longitudinally striate; anterior tergites with
	carinate lateral expansions. (Underwoodia, nearc.).
	UNDERWOODÌID <i>Æ</i>
32.	Hind gonopod with a long thread-like or flagelliform process33
	Hind gonopod without such long process
33.	Front gonopod with a filiform process like that of the hind gono-
	pod; second pair of legs of female with the joints beyond the
	coxæ vestigial. (Heteroporàtia, Tessinosòma, Haploporà-
	tia, palearc.) HETEROPORATIIDÆ
	Front gonopod without such process; second pair of legs of female
	normally developed, similar to the other legs. (Verhoeffia,
9.4	palæarc.)
34.	Posterior pair of legs of eighth somite with coxal glands
	gina, palearc.) FAGÍNIDÆ
25	Tarsi of male without papille
00.	Tarsi of third to seventh pairs of legs of male furnished with
	papille
36.	Hind gonopods one- or indistinctly two-jointed
ου.	Hind gonopods three- or four-jointed. (Brachychæteùma,
	Scutógona, Macrochæteùma, palæarc.).
	BRACHYCHÆTEÙMIDÆ
37.	Tracheal pockets (the cavities next to the spiracles, leading into
	the tracheal tube) fused with the sternite and not movable.
	(Anthógona, Cranógona, palæarc.) ANTHOGÓNIDÆ
•	Tracheal pockets not anchylosed with the sternite, movable.
	(Anthroleucosòma, Pródicus, palæarc.).
	ANTHROLEUCOSOMÁTIDÆ
38.	Coxæ of the front gonopods not forming a ring
	Coxæ of the front gonopods partly fused, forming together a ring

· ·	which bears the unsegmented, movable part of the gonopod. (Heterolatzélla, palæarc.) HETEROLATZÉLLIDÆ
39.	Coxæ of the front gonopods without an elongate, flagelliform process or long, stalked pencil of hairs
	Coxæ of front gonopods each bearing an elongate, flagelliform
	process or a long, stalked pencil of hairs. (Neatractosòma,
	Trimeróphoron, palæarc.) NEATRACTOSOMÁTIDÆ
40.	1
	horn-like projection internally; hind gonopods highly modified, wanting or one- to three-jointed, rarely four-jointed, and not resembling walking legs
	Coxæ of second pair of legs of the eighth segment with a horn-like
	projection internally; hind gonopods four- to six-jointed and
	with claw, retaining the appearance of walking legs. (Pseùdo-
	clis, palæarc.; Cleidógona, Am.; Pseudotrèmia, nearc.).
	PSEUDOCLÍDIDÆ
41.	Sternite bearing the second pair of legs of the sixth somite in the
	male, produced into a long, rod-shaped process; coxæ of front
	gonopods elongate, free or nearly so. (Rothenbuehlèria,
	palearc.) ROTHENBUEHLERÌIDÆ
	This sternite without rod-shaped prolongation; coxæ of front
	gonopods shorter, not extending beyond the remainder of the
40	gonopod, frequently fused together
42.	Coxæ of the second pair of legs of the sixth somite of the male elongate; rounded inwardly or without distinct process medi-
	ally 43
	Coxæ of second pair of legs of sixth somite of the male transverse,
	with one or two processes which extend to the apex of the ster-
	nite. (Attémsia, Syngonopòdium, Dendromonómeron,
	Haasea, palæarc.) ATTEMSÌIDÆ
43.	Sternite of the front gonopod obsolete or absent, never with a
	median process; the coxe firmly united44
	Sternite of the front gonopod very large, bearing a long median
	process; the coxe not fused with one another. (Opisthocheiron, palearc.)
11	Coxe of the front gonopods flat, not prominent; hind gonopods
44.	two- to four-jointed. (Haplobainosòma, Brælemanneùma,
	Hispaniosòma, palearc.) HAPLOBAINOSOMÁTIDÆ
	Coxe of front gonopods prominent, forming a simple or two-parted
	elevation. (Craspedosòma, Oxydáctylon, Macheiriópho-
	ron Caratosòma nalmaro) CRASPEDOSOMÁTIDÆ

45.	Coxe of the ninth pair of legs with glands; tarsi of the third to seventh pairs of legs almost always bearing papille46
	Coxæ of the ninth pair of legs without glands; tarsi usually with-
	out papille
46.	Body composed of 32 somites; tergites with prominent lateral
	expansions. (Diplomarágna, Syntelopodeùma, palæarc.).
	DIPLOMARÁGNIDÆ
	Body composed of 28 or 30 somites; tergites without or with very
	slight lateral expansions
47.	Hind gonopods two-jointed; second pair of legs on the sixth and
	eighth somites modified into accessory gonopods, not resem-
	bling walking legs. (Chordeùma, Chordeumélla, Micro-
	chordeùma, Orthochordeumélla, palæarc.) CHORDEÙMIDÆ
	Hind gonopods three- to five-jointed; second pair of legs on the
	sixth somite normal, similar to the other walking legs. (Oro-
	bainosòma, Brachybainosòma, palæarc.).
	OROBAINOSOMÁTIDÆ
48.	First pair of legs of the eighth somite highly modified, reduced to
	two- or three-jointed stumps; vertex of male bearing one or
	two tufts of hairs. (Metopidothrix, Schedotrigona, indo-
	austr.) METOPIDOTHRÍGIDÆ
	First pair of legs of the eighth somite normal, not modified or re-
	duced; vertex of male without tufts of hairs. (Conotyla, Tri-
	chopétalum, Zygónopus, nearc.; Apodígona, neotrop.;
40	Japanosòma, palæarc.)
49.	Promentum present as a separate sclerite in front of the mentum
	(as in Fig. 1102); ocelli numerous. (Superfamily LYSIOPETA-
	LÕIDEA)
	(Figs. 1111, 1112). (STEMMIULÒIDEA). (Stemmiùlus,
	Prostemmiùlus, Diopsiùlus, ethiop.) STEMMIÙLIDÆ
50.	Second pair of legs of female completely formed, similar to the
	other walking legs; tracheal pockets of gonopods small, fused
	to the gonopod. (Cállipus, palæarc.) CALLIPÓDIDÆ
	Second pair of legs of female greatly reduced, the segments beyond
	the coxæ not developed; tracheal pockets of gonopods large,
	not fused to the gonopod
51.	Gonopods very long, slender and strongly bent. (Dorypétalum,
	Silvéstria, Cyphocállipus, Dorycállipus, palæarc.).
	DORYPETÁLID <i>Æ</i>

Gonopods not noticeably long, not bent. (Lysiopétalum, Apfelbéckia, Broelemánnia, Callipodélla, palæarc.).

LYSIOPETÁLIDÆ



Figs. 1104-1114. Diplopoda

- 1104. Diaporus, gonopod (Chamberlin) Spirostreptidæ.
- 1105. Siphonophora, anterior gonopod (Chamberlin) Siphonophoridæ.
- 1106. Siphonophora, posterior gonopod (Chamberlin) Siphonophoridæ.
- 1107. Platydesmus, anterior and posterior ends of body (Chamberlin) Platydesmidæ.
- 1108. **Gymnostreptus,** gonopods (Chamberlin) Spirostreptidæ.
- 1109. Paraiulus, posterior end of body of male (Chamberlin) Blaniulidæ.
- 1110. **Paraiulus,** side view of posterior end of body of male (Chamberlin) Blaniulidæ.
- 1111. Prostemmiulus, gnathochilarium (Chamberlin) Stemmiulidæ.
- 1112. Prostemmiulus, head and first tergite (Chamberlin) Stemmiulidæ.
- 1113. Orthoporus, gonopod (Chamberlin) Spirostreptidæ.
- 1114. Rhinocricus, gonopods (Chamberlin) Rhinocricidæ.

	Stipites not meeting along the median line, widely separated by the mentum or promentum; promentum if present in contact with the mentum
53.	
	striation (Julus)
	Gonopods free, not sunk in depressions; first pair of legs of male
	usually four- to six-jointed; tergites without any longitudinal
	striation. (Figs. 1109, 1110). (Blaniùlus, Choneiùlus, palæ-
	arc.; Nopoiùlus, holarc., neotrop.; Paraiùlus, Am.; Uro-
	blaniùlus, nearc.) BLANIÙLIDÆ
54.	Promentum absent; mentum large, triangular, extending between
	the prebasilare and stipites which it separates widely (Fig. 1103); last joint of legs of male generally padded, but not the
	fourth and fifth joints. (Superfamily SPIROBOLÒIDEA)55
	Promentum present or absent; mentum surrounded at the sides
	by the stipites which extend backwards and lie in contact with
	the prebasilare. (SPIROSTREPTOMORPHA)59
55.	Hind gonopods connected to each other at base by the sternite
	and membranes; pores of repugnatorial glands usually open-
	ing on the anterior portion of the tergites. (TRIGONIU-
	LÍDEÆ)56
	Hind gonopods not united at base, sternite completely absent;
	pores of repugnatorial glands almost always opening on the
-	posterior portion of the tergites. (SPIROBOLÍDEÆ) 58
56.	Coxæ of hind gonopods completely chitinized
	meeting at a right angle with a membranous portion between
	them. (Trigoniùlus, Eucárlia, Cherástus, ethiop., indo-
	austr.; Mystálides, ethiop., malay.; Allopocóckia, neotrop.).
	TRIGONIÙLIDÆ
57.	Front gonopods short and broad. (Pachýbolus, ethiop.; Micro-
	spiróbolus, Caríbolus, neotrop.; Trachelomégalus, Eu-
	centróbolus, malay.) PACHYBÓLIDÆ
- 1	Front gonopods long and slender. (Spiromimus, Pygodon,
	ethiop.)
58.	Hind gonopods consisting of only a single joint, the coxa not dis-
	tinctly separated or vestigial; first tergite more or less nar-
	rowed laterally. (Spiróbolus, holarc.; Tylóbolus, nearc.;
	Messicóbolus, neotrop.; Pseudospiroboléllus, indomal.;

	Hind gonopods distinctly two-jointed; first tergite broadly rounded
	at the sides. (Fig. 1114). (Rhinócricus, neotrop., austromal.
	Polyconóceras, Dinematócricus, austromal.; Eurhinócricus,
	Cubóbolus, neotrop.) RHINOCRÍCIDÆ
59.	Mentum broadly triangular; its anterior angle lying far behind the
	front margin of the stipites. (Superfamily SPIROSTREP-
	TÒIDEA) 60
	Mentum elongate-triangular, its anterior angle extending far for-
	ward, dividing the gnathochilarium completely to its anterior
	margin; first pair of legs of male modified. (Superfamily CAM-
	BALÒIDEA)
30.	Both sternites of the gonopod-bearing segment present, in spite
	of the atrophy of the hind gonopods. (ODONTOPYGIDEÆ).
	(Odontopýge, Haplothýsanus, Prionopétalum, Spino-
	társus, ethiop.) ODONTOP ÝGIDÆ
	Sternite of the posterior part of the gonopod-bearing segment
	and the gonopod absent. (SPIROSTREPTÍDEÆ)61
61.	End of the gonopod flattened, almost always bearing on the edge
	a row of long, curved bristles; last somite without any spinose
	projection. (Fig. 1104, 1108, 1113). (Spirostréptus, Gymno-
	stréptus, Diáporus, Orthóporus, Scaphiostréptus, Alló-
	porus, neotrop., ethiop.) SPIROSTRÉPTIDA
	End of the gonopod not thus flattened, without such bristles
	anal segment almost always with a tail-like projection. (Harpa-
	góphora, ethiop.; Poratóphilus, ethiop., neotrop.; Thyro-
	pỳgus, ethiop., indomal.; Rhynchopróctus, malay.).
	HARPA GOPHÓRID <i>A</i>
32.	Promentum present, separated from the mentum63
	Promentum not separated from the mentum; repugnatorial pores
	present on the fifth somite
33.	Promentum divided into two halves by a longitudinal suture64
	Promentum not divided by a longitudinal suture. (Cámbala,
	Sámichus, austr.; Nannolène, Epinannolène, neotrop.
	Julomórpha, widespr.). (Including NANNOLÉNIDÆ and
	EPINANNOLÉNIDÆ) CAMBÁLIDÆ
64.	Repugnatorial pores absent on the fifth somite; hind gonopods
	three-jointed; front ones one-jointed, without bristles. (Peri-
	cámbala, ind.) PERICAMBÁLIDÆ
	Repugnatorial pores present on the fifth somite; hind gonopods want-
	ing, the front ones two-jointed, the second joint strongly bristled
*	(Pseudonannolène, neotrop.). PSEUDONANNOLÉNIDÆ

65. Posterior portion of tergites smooth; hind gonopods absent. (Physiostréptus, Holopodostréptus, neotrop.).

#### PHYSIOSTRÉPTIDÆ

Posterior portion of the tergites with stout longitudinal carinæ or tubercles; hind gonopods present. (Cambalópsis, Cambalomórpha, Trachviùlus, ind.). (TRACHYIÙLIDÆ).

## CAMBALÓPSIDÆ

- - Tergites, pleurites and sternites completely coalesced; body cylindrical. (Siphoniùlus, malay.) . . . . . SIPHONIÙLIDÆ
- - Gnathochilarium possessing most of the parts typical of the Diplopoda (as in Fig. 1101). (Fig. 1107). (Platydésmus, N. Am.; Fiòria, Dolístenus, palæarc.) ...... PLATYDÉSMIDÆ
- - Ocelli absent; body constricted at the base of each somite; repugnatorial pores opening on keels or tubercles. (Figs. 1105, 1106.) (Siphonóphora, indoaust., ethiop., neotrop.; Siphonorhìnus, malay., neotrop.) . . . . . . . . . . . . . SIPHONOPHÓRIDÆ
- 69. Repugnatorial pores opening on the lateral lobes of the tergites; tergites divided by a median suture; head completely concealed by the first tergite. (**Siphonocrýptus**, malay.).

# SIPHONOCR YPTIDÆ

Repugnatorial pores opening on the body of the tergites, far from the lateral margin; tergites without median suture; head partly free and visible. (**Polyzonium**, **Orsíboe**, palæarc.; **Siphonòtus**, austromal., neotrop.; **Burínia**, ethiop.).

# **POLYZONÌIDÆ**

# LITERATURE ON BOTH DIPLOPODA AND CHILOPODA (MYRIAPODA)

Attems, C. Die indo-australischen Myriapoden. Arch. f. Naturg. Jahrg. 80A, Heft 4, pp. 1–398 (1914).

Chilopoda and Diplopoda in Kükenthal's Handbuch der Zoologie, 4, Lief. 1–4, pp. 17–402 (1926).

The Myriapoda of South Africa. Ann. South African Mus., 26, pp. 1–431 (1928).

- Die Myriapodenfauna von Albanien und Jugoslavien. Zool. Jahrb. Abth. f. Syst., **56**, pp. 269–356 (1929).
- Leptodesmidæ und andere Polydesmiden. Zoologica, Heft 79, 149 pp. (1931).
- Bollman, C. H. Myriapoda of North America. Bull. U. S. Nat. Mus., No. 46 (1893).
- Chamberlin, R. V. The Chilopoda and Diplopoda of the West Indies. Bull. Mus. Comp. Zoöl. Harvard, **62**, pp. 151–262 (1918).
  - The Myriapoda of the Australian Region. Bull. Mus. Comp. Zoöl. Harvard, **64**, pp. 1–269 (1920).
- Cook, O. F. The Myriapoda of Northwestern North America. Pap. Harriman Alaska Exped., 8, pp. 49–77 (1910).
- LATZEL, R. Die Myriapoden der Oesterreichisch-Ungarischen Monarchie. Vienna, Alb. Hölder (1880–84).
- Underwood, L. M. The North American Myriapoda. Entomologica Americana, 1, pp. 141–151 (1885).
- Verhoeff, K. W. Results of Dr. E. Mjöberg's Swedish Sci. Exped. to Australia. Ark. Zool., 16, No. 5, 142 pp. (1924).
- Wood, H. C. Myriapoda of North America. Trans. American Philos. Soc., 13, pp. 137–248 (1865).

#### LITERATURE ON DIPLOPODA

- Attems, C. System der Polydesmiden. Denkschr. math.-naturw. Kl. Kais. Akad. Wiss. Wien, **67**, pp. 1–262 (1898); *ibid.*, **68**, pp. 1–186 (1899).
  - Ueber paläarktische Diplopoden. Arch. Naturg. Jahrg. 92A, No. 1, pp. 1–144; No. 2, pp. 145–256 (1927).
  - Diplopoden des Belgischen Congo. Polydesmoidea. Rev. Zool. Bot. Afr., 17, pp. 253–378 (1929).
- Brölemann, H. W. Essai de Classification des Polydesmiens. Ann. Soc. Entom. France, **84**, pp. 523–608 (1916).
- Blaniulidæ. Arch. Zool. Paris, **61**, pp. 99–453 (1923). CARL, J. Die Diplopoden-Fauna von Celebes. Rev. Suisse Zool., **20**, pp. 75–206 (1912).
- Chamberlin, R. V. Julidæ and Isobatidæ of North America. Proc. Biol. Soc. Washington, **34**, pp. 81–84 (1921).
  - The Millipedes of Central America. Proc. U. S. Nat. Mus., **60**, art. 8, pp. 1–75 (1922).
- Cook, O. F. and G. N. Collins. Craspedosomatidæ of North America. Ann. New York Acad. Sci., 9, pp. 1–100 (1895).
- Рососк, R. J. Diplopoda in Biologia centrali-Americana, pp. 41–217 (1904–10).
- Verhoeff, K. W. Die Diplopoden Deutschlands. 8 parts, 640 pp. Leipsig (1910–14).

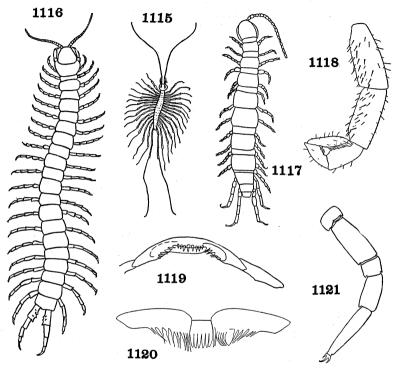
# CLASS CHILÓPODA

Body long, comparatively or very narrow, of nearly even width and dorsoventrally depressed; nineteen or more, sometimes many more body segments. Fifteen or more pairs of legs, inserted at the sides of the body and widely separated by sternal plates; never more than a single pair to any body segment; legs six- or seven-jointed, with a single apical claw. Head bearing a pair of long, many-jointed antennæ (14 joints or more); eyes composed of groups of ocelli, sometimes massed to form two apparently faceted eyes. Mandibles and two pairs of maxillæ present; appendages of first body segment greatly enlarged, forming a pair of large six-jointed poison-fangs (toxicognaths). Last two pairs of legs often greatly modified and directed backwards. Respiration through spiracles, the latter paired and located on the pleuræ or unpaired and located along the dorsal line. Genital ducts usually opening on the penultimate body segment. A widespread, moderately large group, of active carnivorous habits. Centipedes.

The system adopted here follows that of Attems in his recent account of "The Myriopoda of South Africa."

- - Twenty-five or more body segments with 21 or more pairs of legs, sometimes many more; nine or more pairs of spiracles, the tracheæ anastomosing; newly hatched animals with the full number of legs present in the adult. (Subclass EPIMÓRPHA).6
- Spiracles unpaired, seven in number; placed on the middle dorsal line near the posterior border of the tergites; eyes compound, with faceted surface; tracheæ not branched. (Fig. 1116). (Order SCUTIGEROMÓRPHA). (Scutigera, Thereuónema, Parascutigera, Thereuópoda, mainly tropical or subtropical).
- 3. Tergites of the leg-bearing segments alternately long and short, except at the middle of the body; those corresponding to legs 2, 4, 6, 9, 11 and 13 (not counting the jaws as legs) much smaller than the others. (Suborder LITHOBIOMORPHÍDEA)....4

	Tergites of the leg-bearing segments not noticeably different in
	size, except the one bearing the jaws, which is shorter; in front or
	segments 4, 6, 8, 9, 11 and 13 with a greatly reduced, inter-
	calary segment. (Suborder CRATEROSTIGMORPHIDEA)
	(Craterostigmus, austr.) CRATEROSTÍGMIDÆ
4.	From one to five of the posterior pairs of legs with coxal glands
	opening by pores, legs usually spinose; ocelli, when present
	placed near to the margin of the head. (Superfamily LITHO-
	BIÒIDEA)
	No coxal glands opening by pores; legs without spines; ocelli
	widely separated from the margin of the head. (Superfamily
	CERMATOBIÒIDEA). (Cermatòbius, ind.).
	CERMATOBÌID <i>A</i>
5.	Tibiæ of all legs without a spinose projection externally at tip.
	(Fig. 1117). (Lithòbus, holarc., ethiop.; Monotarsòbius,
	Alokòbius, Polybóthrus, palæarc.; Garíbius, nearc.; Aus-
	tralòbius, palæarc., indoaustr.; Bothrópolys, holarc., austro-
	mal.) LITHOBÌIDÆ
	Tibiæ of legs 11-14 with a spinose, tooth-like projection externally
	at tip. (Hénicops, austr.; Paralamýctes, Anopsòbius, neo-
	trop., ethiop., austr.; Zygethòbius, Am.) HENICÓPIDÆ
6.	· · · · · · · · · · · · · · · · · · ·
	more; 21–23 leg bearing segments; 9, 10, 11 or 19 pairs of spir-
	acles. (Order SCOLOPENDROMÓRPHA)
	Antennæ 14-jointed; ocelli absent; 31-170 leg-bearing segments
	all leg-bearing segments except the first and last with a pair of
_	spiracles. (Order GEOPHILOMÓRPHA)8
7.	Eyes present, very rarely (Mimops) indicated only by a pale spot
	tarsi two-jointed; sternites usually with a pair of longitudinal
	grooves, never with transverse groove. (Fig. 1115). (Scolo-
	péndra, Rhysídia, tropicopol.; Cùpipes, palæarc., ethiop., ind.;
	Pseudocrýptops, ethiop., ind.; Collària, palæarc., neotrop.
	Echmostigmus, palæarc., indoaust.). SCOLOPÉNDRIDÆ
	Eyes absent; tarsi of legs 1–19 almost always one-jointed; sternites
	usually with a single median groove, never with a pair of grooves
	often with a transverse groove. (Fig. 1118). (Crýptops, cos-
	mop.; Otocrýptops, widespr.; Trigonocrýptops, palæarc.
	ethiop., neotrop.; Scolopocrýptops, palæarc., Am.; Newpórtia neotrop.)
Q	Mandible with several pectinate lamellæ and with or without a
0.	dentate lamella
	uomaao lallitila



Figs. 1115-1121. Chilopoda and Symphyla

- 1115. Scolopendra (Newport) Scolopendridæ.
- 1116. Scutigera (Howard) Scutigeridæ.
- 1117. Monotarsobius. Lithobiidæ.
- 1118. Cryptops, last leg (Chamberlin) Cryptopidæ.
- 1119. Soniphilus, labrum (Chamberlin) Soniphilidæ.
- 1120. Suturodes, labrum (Chamberlin) Geophilidæ.
- 1121. Scutigerella, leg (Ewing) Scutigerellidæ.
- 9. Mandible with one dentate lamella in addition to the pectinate lamellæ; labrum consisting of one piece; antennæ short, thick at base and gradually tapering, the basal joints with short hairs, but without long bristles. (Himantàrium, palæarc., ethiop.;

	Mesocánthus, palæarc., ethiop., indomal.; Haplóphilus,
	Bothriogáster, palæarc.) HIMANTARÌIDÆ
	Mandible with pectinate lamellæ, but without any dentate lamella in addition
10	Labrum consisting of one piece, not especially broad; coxæ of
10.	first maxillæ completely fused. (Örya, palæarc.; Orphnæus,
	tropicopol.; Ctenòrya, ethiop.; Paròrya, nearc.) ORŸIDÆ
	Labrum tripartite; one small median tooth and two broad lateral
	pieces with strong edges at the sides of the cephalic pleuræ;
	coxæ of first maxillæ meeting at the median suture. (Mecisto-
	céphalus, ethiop., indoaustr.; Prolámmonyx, Dicellóphilus,
	palæarc.; <b>Árrup</b> , nearc.; <b>Tygárrup</b> , neotrop.).
	MECISTOCEPHÁLIDÆ
11.	Mandible with one dentate lamella; labrum consisting of one
	piece; antennæ filiform or clavate. (Schendyla, holarc., neo-
	trop., mal.; Escáryus, holarc.; Nannóphilus, ethiop., neo-
	trop.; Ballóphilus, ethiop., austromal.; Schendylùrus, pal-
	æarc., ethiop., neotrop.)
12	Coxè of the first maxillæ fused, each bearing a median process and
14.	a one- or two-jointed apical portion
	Coxæ of the first maxillæ completely separated, bearing a conical
	second joint; claws of anterior legs with a strong tooth. (Neo-
	geóphilus, Evallogeóphilus, neotrop.). NEOGEOPHÍLIDÆ
13.	Labrum consisting of a single piece; antennæ generally flattened
	at the base and tapering14
	Labrum consisting of three parts, rarely more or less fused (Fig.
	1119), the median piece sometimes partially fused with the
	lateral pieces; coxæ of the last legs not enlarged
14.	Coxæ of the last pair of legs much enlarged, extending forward
	at the sides sometimes to the antepenultimate leg-bearing
	segment; paratergites (one or several rows of small plates be-
	tween the tergites and the row of pleurites that bear the spiracles) generally present. (Gonibrégmatus, austromal.; Eu-
	crátonyx, indoaustr.; Himantosòma, indomal.; Macronicó-
	philus, neotrop.)
	Coxe of the last pair of legs not enlarged, not extending in front
	of the last leg-bearing segment; paratergites absent. (Sogona,
	Timpìna, nearc.; Garrina, neotrop.) SOGÓNIDÆ
15.	The median piece of the labrum, if present, not fused with the
	lateral pieces: in the middle the two larger and ventrally directed

teeth lacking. (Fig. 1120). (**Geóphilus**, widespr.; **Eurýtion**, ethiop., neotrop., austr.; **Scolióplanes**, holarc.; **Polygonàrea**, austr., ethiop.; **Aphílodon**, ethiop., neotrop.; **Hènia**, palæarc.; **Pachymèrium**, holarc., neotrop., ethiop.) . . **GEOPHÍLIDÆ** 

The median piece of the labrum at least partly fused with the lateral ones; two larger teeth in the middle, directed more or less ventrally. (Fig. 1119). (Soníphilus, Poáphilus, nearc.).

SONIPHÍLIDÆ

#### LITERATURE ON CHILOPODA

See also Literature on Myriapoda, p. 597

Attems, C. Myriapoda. Geophilomorpha. Das Tierreich, Lief. 52, 388 pp. (1929).

Chilopoda, Scolopendromorpha. Das Tierreich, Lief. 54, 308 pp. (1930).

Bailey, J. W. Chilopoda of New York State. Bull. New York State Mus., No. 276, 50 pp. (1928).

Chamberlin, R. V. Chilopoda of California. Pomona Journ. Zoöl. Entom., **2, 3, 4** (1910–12). (Three parts).

The Chilopoda of Brazil. Bull. Mus. Comp. Zoöl. Harvard,  ${\bf 58}$ , pp. 151–221 (1914).

The Centipedes of Central America. Proc. U. S. Nat. Mus., 60, Art. 7, pp. 1–17 (1922).

The Ethnopolidæ of North America. Bull. Mus. Comp. Zoöl. Harvard, 57, pp. 385–437 (1925).

Kraepelin, K. Revision der Scolopendriden. Jahrb. Hamburg Anst., Beiheft 2, Mitt. Mus. Hamburg, pp. 1–276 (1903).

Рососк, R. I. Chilopoda. Biologia centrali-Americana, pp. 1-40 (1896).

Verhoeff, K. W. Chilopoda. Bronn's Tierreich, 5, Abth. 2, Lief. 83–99, pp. 83–537 (1915–18).

# CLASS SÝMPHYLA

Small, delicate elongate species with thin unpigmented integument. Body long, composed of fifteen to twenty-two similar segments; twelve pairs of short, five-jointed legs. Mouthparts consisting of a pair of mandibles and two pairs of maxillæ. Antennæ many-jointed, the joints short, more or less moniliform. Cerci stout, lanceolate, one-jointed or very indistinctly annulated. Respiration by tracheæ opening by a single pair of spiracles on the head.

 2. An intercalated segment without legs present behind the fourth, sixth and eighth segments only; posterior margin of tergites rounded, the corners rarely angulate; first pair of legs fully half as long as the others. (Figs. 11, 1121). (Scutigerélla, Hanseniélla, Tasmaniélla) . . . . . . . SCUTIGERÉLLIDÆ An intercalated legless segment present behind segments four,

#### LITERATURE ON SYMPHYLA

- HANSEN, H. J. The Genera and Species of the Order Symphyla. Quart. Journ. Micr. Sci., 47, pp. 1–101 (1903).
- LATZEL, R. Die Myriapoden der Oesterreichisch-Ungarischen Monarchie. Vienna, Alb. Hölder (1880–84).

## GLOSSARY OF SPECIAL TERMS.1

This glossary is intended to include only such terms as are not easily understood from the figures referred to in the keys, and other words only when their meaning in the keys might not be readily ascertained from an ordinary English dictionary.

Abdòmen, the hindermost of the three main body divisions.

**Acróstichal bristles,** one, two or several longitudinal rows of minute bristles along the center of the mesonotum of some flies (cf. dorsocentral bristles).

Aculea (-ex), one of the numerous minute spines on the wing membrane of certain Lepidoptera.

**Aculeate**, (a) furnished with a sting; (b) beset with aculeæ.

Adventítious, not regular; accidental or additional.

Ædæagus, the male intromittent organ.

Álula (-læ), a small lobe, borne at the base of the wing (Diptera).

Amphipneustic, having the first and last pairs of spiracles open and functioning.

**Anal**, pertaining to the last abdominal segment or to the hind basal angle of the wing.

**Anal lobe**, the rounded posterior part of the wing which includes the anal veins.

**Anal veins,** in the typical wing the three most posterior main veins.

**Annulated**, incompletely divided into ring-like joints.

**Annulus** (-li), a ring or band.

Antecóxal sclerite, a part of the metasternum in front of the hind coxæ (Coleoptera).

Anténna (-næ), a pair of jointed sensory appendages of the head above the mouthparts.

Adfrontal plates, a pair of long oblique sclerites on the front of the head extending upwards from the base of the antennæ and meeting medially above (larvæ of Lepidoptera).

**Antenòdal crossveins**, crossveins along the costal border between the base and the nodus (Odonata).

Apneustic, having all the spiracles closed and not functioning.

**Appendiculate cell,** a small indistinct cell just beyond the apex of the marginal cell (Hymenoptera).

<sup>&</sup>lt;sup>1</sup> Where the plural form is unusual the differing termination is given in parentheses added to the last common letter of the root.

Apterous, wingless.

**Arcuate**, arched like a bow.

**Árculus**, a basal crossvein between the radius and cubitus (Odonata).

Arèola (-læ), areóle, a small closed cell in the forewing, enclosed by radial veins (Lepidoptera).

Arîsta, a bristle-like process at or near the end of the antennæ (Diptera).

Aròlium (-ia), a terminal pad of the foot between the claws.

Atrium (-ia), a cavity or enlarged entrance.

Atténuated, gradually tapering.

Auxiliary vein, the subcostal vein of Diptera, anterior to the radius.

Axilla (-læ), a triangular sclerite on each side of the scutellum (Hymenoptera).

Bàsal cells, the two cells proximal to the anterior crossvein and the discal cell (Diptera).

Bifid, split into two parts.

Bilobed, divided into two lobes.

**Calamistrum**, one or two rows of curved spines on the upper margin of the metatarsus of the hind legs (Araneida).

Calýpteres, small membranous disks under the base of the wings (Diptera).

**Cámerostome**, the opening through which the beak is extended in certain Acarina.

**Campodèiform**, having the form of Campodea (Thrysanura), said of certain active carnivorous larvæ.

Cápitate, with a distinct knob at the tip.

**Cárapace**, the more or less fused dorsal sclerites of the cephalothorax (Arachnida). Also any fused series of sclerites covering a part of the body.

Cárinate, ridged, or furnished with a raised line or keel.

**Cáruncle**, a fleshy papillate or sucker-like appendage at the tip of the tarsus in certain Acarina.

Caudal filaments or setæ, thread-like processes terminating the abdomen.

Cell, a space in the wing bounded by veins.

Cephalothòrax, the combined head and thorax in Arachnida.

Cérci, a pair of appendages near the end of the abdomen.

Chætosèma, a series of short bristles on the head of certain Lepidoptera.

Cheek, the lateral part of the head between the eyes and the mouth. Chèlate, pincers-shaped, having two opposable claws.

Cheliceræ, the first or most anterior pair of appendages in the Arachnida.

Chitin, the horn-like material forming the hard parts of the body wall.

Clavate, clubbed or enlarged at the tip.

Clàvus, an oblong basal part along the inner edge of the fore wings (Heteroptera, Homoptera).

Clýpeus, the sclerite bearing the labrum (Hymenoptera); a horseshoeshaped sclerite under the margin of the mouth (Diptera).

Coárctate, with narrowed base and enlarged tip.

Coarctate pupa, a type of pupa in certain Diptera, enclosed in a hardened shell formed from the last larval moult-skin.

**Cólulus**, a slender or pointed organ lying in front of the spinnerets (Araneida).

**Compréssed**, flattened from side to side, as distinguished from depressed.

Cónnate, immovably united, fused.

Constricted, narrowed in part.

Còrium, an elongate middle part of the fore wing (Hemiptera).

Córneous, horn-like in texture.

**Córnicle**, one of certain paired dorsal tubular processes on the posterior part of the abdomen of aphids.

Cósta, the front margin of the wing, considered as the first vein.

Costal area, the part of the wing immediately behind the front margin.

**Cóstal cell**, the space of the wing between the costa and the subcostal vein; formerly sometimes termed subcostal.

Cóxa (-xæ), the basal joint of the legs, sometimes quite fused with the body.

**Cribéllum**, a sieve-like spinning organ lying in front of the spinnerets (Araneida).

Cróchets, a series of spines on the prolegs of the larvæ of Lepidoptera.

Crossvein, any transverse vein connecting adjacent longitudinal veins.

Ctenídium (-ia), a comb-like row of bristles.

Cùbitus, the fifth of the main veins of the typical wing.

Cursorial, fitted for running.

**Declivity**, the abruptly bent apex of the elytra (Coleoptera).

Decúmbent, bending downward.

Dentículate, with minute tooth-like projections.

Dichóptic, eyes not touching above (Diptera).

Digitate, with finger-like processes.

Direct eyes, the anterior median pair of eyes in spiders.

Diúrnal eyes, in spiders, eyes that are dark colored.

**Dórsal**, pertaining to the upper surface or back of the body.

**Dorsocéntral bristles**, several longitudinal rows of bristles near the middle of the mesonotum, lateral to the smaller acrostichals (Diptera).

Ectopárasite, a parasite which lives on the exterior of animals.

Élytron (-ra), the horny fore wings, or wing covers, of beetles.

**Empòdium** (-ia), a single middle pad or bristle between the tarsal claws) (Diptera).

**Epimeron** (-ra), the posterior portion of the pleura of a thoracic segment (Insecta). The sclerite to which the basal segment of the leg is attached (Arachnida, Diplopoda).

Epíphysis (-ses), a lappet-like process.

Epipleura (-ræ), the infolded edge of the elytra (Coleoptera).

Epistérnum, the anterior portion of the pleura of a thoracic segment.

Épistome, the part of the face just above the mouth.

Epizòic, living as external parasites of animals.

**Erùciform**, having the body shaped like a caterpillar.

**Exuviæ**, the molt, or cast skin of an insect. Recently used as exuvia, exuviæ to differentiate the molts of a single from those of several instars, as in Coccoidea.

**Eye-cap**, a group of modified scales overhanging the eye (Lepidoptera).

Face, the front of the head, between the clypeus and antennæ.

Fàcial plate, the central part of the face (Diptera).

Fastigium, the upper edge or median ridge of the vertex (Orthoptera).

Fèmur (-émora), the thigh or third division of the legs.

Filiform, hair-like, or filamentous, longer than setaceous.

Flabéllate, with fan-like processes or projections.

Flabéllum (-la), a leaf-like or fan-like process.

Flagéllum, the distal part of the antenna when lash-like.

**Fontanél**, a small, depressed, pale spot on the front of the head between the eyes (Isoptera).

Foramen (-ámina), a small opening, orifice or puncture.

Fórcipate, bearing pincers, or pincers-shaped.

**Frénulum**, a strong spine or spines at the front basal angle of the hind wings (Lepidoptera).

Front, the forehead, between the antennæ, eyes and ocelli.

Frontàlia, the central strip of the front (Diptera).

Frontal lunule, a small crescent-shaped space just above the antennæ (Diptera)

**Frónto-órbital bristles,** several bristles along the front next to the eyes (Diptera).

Fundatrix, a stem-mother or female of the first generation which founds a new colony (Aphidoidea).

**Fúrcula**, the forked springing appendage below the end of the abdomen (Collembola).

**Gáster**, the rounded part of the abdomen behind the basal node or nodes in ants (Hymenoptera).

Gèna (-næ), the cheek.

Geniculate, abruptly bent, elbowed.

Genitàlia, the external sexual organs.

Gibbous, puffed out; hunch-backed.

Glàbrous, bald, smooth, free of hairs.

**Gnathochilàrium**, a plate-like structure formed by the mouthparts, exclusive of the mandibles (Diplopoda)

**Gónopod**, a modified leg, serving as a copulatory appendage, on the seventh or adjacent body segments (Diplopoda).

Gonapóphysis (-ses), each of the short conical paired egg-laying processes terminating the abdomen; also applied to certain paired genital appendages in the male.

Graduated crossveins, an oblique row of crossveins forming steps across the wing (Neuroptera).

Grávid, filled with eggs.

Gula, the median underpart of the head lying between the mouth and posterior foramen.

Gular suture, a longitudinal impressed line on each side of the gula or middle piece of the throat.

**Hálter**, a small knobbed appendage on each side of the thorax replacing the hind wings (Diptera).

Hàmus, a distinct abrupt spur-like vein in the hind wings of some Heteroptera.

Haustellate, formed for sucking, the mandibles not fitted for chewing.

Hemélytron (-ra), the heavily chitinized fore wing of Hemiptera.

Heterogèneous, of more than one type, as the eyes of certain spiders.

Heterómerous, differing in the number of joints in the three pairs of tarsi (e.g. 5, 5, 4), or in the structure of other repetitive parts.

Holóptic, having the eyes meeting above the antennæ (Diptera).

Homónomous, similar in form, function or development.

Hùmeral angle, the basal front corner of the wing.

Hỳaline, more or less transparent.

**Hypermetamórphosis**, development in which two or more different types of larvæ follow one another in succeeding stages of growth.

Hypopleural bristles, a more or less vertical row of bristles above each hind coxa (Diptera).

Hypopýgium, the last ventral plate; or the inflexed genitalia.

**Hypostigmátic cell,** a specialized cell located beneath the stigma of the wing of some Neuroptera.

Imago, the final, adult, or reproductive stage of an insect.

Inglùvial, pertaining to the crop.

**Inclinate**, bent or directed toward the median line.

**Inquiline**, an animal that occurs regularly in the nest or habitation of some other species.

**Instar**, any of the successive stages during metamorphosis, marked off by moltings.

Interfrontàlia, the central portion of the front of some flies when differentiated from the orbits.

Interfrontal bristles, minute bristles on the central part of the front (Diptera).

**Interstitial**, occurring between two segments, e.g. the trochanter, linking the coxa and femur; or coincident, as the ends of two veins.

Intra-àlar bristles, several bristles above the root of the wing lateral to the dorsocentrals (Diptera).

**Jùgum**, a lobe-like process at the base of the fore wings overlapping the hind wings (Lepidoptera).

Labéllum (-la), the expanded tip of the proboscis (Diptera).

Làbial pálpus (-pi), a jointed sensory appendage at each side of the labium.

Làbium, the lower lip.

Làbrum, the upper lip, lying just below the clypeus.

Lamélla (-læ), a leaf-like plate.

Lámellate, bearing or composed of lamellæ.

Láminate, composed of leaf-like plates.

Lánceolate, tapering at each end, spear-shaped.

Lárva (-væ), the growing stages of an insect having a complete metamorphosis, after hatching from the egg and before the pupal period.

Lateral, at, toward, or pertaining to the sides of the body.

Ligula, the central part of the labium, borne by the mentum (Coleoptera).

Lòra (-ræ), the cheek (Homoptera).

Lung-book; lung sac, a respiratory sac, opening by a slit-shaped aperture on the ventral surface of the abdomen in certain Arachnida.

Lûnule, a small crescent-shaped piece just above the antennæ (Diptera).

Macrotrichia, the larger hairs on the surface of the wings.

Mandibulate, with jaws fitted for chewing.

Marginal cell, one or more cells near the anterior margin of the wing, below the stigma (Hymenoptera).

Mask, the extensile labium of the nymphs of Odonata which forms an organ for grasping the prey.

Maxilla (-læ), the second pair of appendages belonging to the mouth, behind the mandibles or jaws.

Máxillary palpus, a finger-like jointed sensory appendage on each maxilla.

Mèdia, the fourth of the principal veins of the typical wing.

Mèdial, pertaining to the media.

Mèdian, lying along the middle line of the body.

Méntum, the part of the labium bearing the movable parts.

Mèsad, lying toward the median line.

Mesepistérnum (-na), the anterior of the oblique side pieces of the mesothorax.

Mesonòtum, the back or upper side of the mesothorax.

Mesopleura (-ræ), the space between the root of the wings and the middle coxe, consisting of episternum and epimeron (Diptera).

Mesostérnum, the middle part of the underside of the mesothorax.

Mesothèrax, the middle of the thoracic divisions, bearing the second legs and the fore wings.

Metamórphosis (-ses), the series of marked external changes through which an insect passes during its development, e.g. egg, larva, pupa, adult. Direct development entails no such changes. Incomplete metamorphosis lacks the pupal stage. Complete metamorphosis includes a pupal stage. See also hypermetamorphosis.

**Metapneustic**, having only the posterior pair of spiracles open and functioning.

Metastérnum, the middle piece of the under side of the metathorax.

Metatarsus (-si), the first joint of any tarsus, next to the tibia, also called basitarsus.

Metathòrax, the third division of the thorax, bearing the hind legs and the hind wings.

Micrópterous, with small or vestigial wings.

Microtrichia, the smaller abundant hairs that clothe the surface of the wings in some insects.

Molt, to cut off or shed the skin.

Moniliform, resembling a string of beads.

Nàiad, the nymph of aquatic insects.

Nàsus, the drawn-out forward part of the head of crane-flies.

Nasute, a form of worker termite in which the head bears a snout-like projection in front.

Neuration, the arrangement of the veins of the wings, the venation.

Noctúrnal eyes, in spiders, eyes of a pearly white color.

Node, a swelling or knot-like knob.

Nodus, a stout crossvein at or before the middle of the costal border of the wing (Odonata).

**Notauli**, a pair of longitudinal furrows on the mesonotum of certain Hymenoptera, lying nearer to the median line than the parapsidal furrows when both pairs of furrows are present.

Nôtum, the dorsal surface of the body, particularly of the thorax.

**Nymph**, the larval or growing stage of those insects that have no pupal period.

Océllus (-li), the simple eyes, usually three in number, on the upper part of the head; also the simple eyes of insect larvæ and of some other arthropods.

**Ócciput**, the back part of the head.

Onýchium (-ia), a pad between the tarsal claws.

**Órbit**, the part of the head immediately surrounding the eyes.

Óstioles, the paired lateral openings of the heart.

Ovipósitor, the egg-laying apparatus.

Pálpus (-pi), one or two pairs of jointed sensitive, finger-like processes borne by the mouth. See maxillary palpus and labial palpus.

**Parafàcials**, the sides of the face of some flies, when differentiated from the sunken central portion and the orbits.

Parápsidal furrow, a longitudinal groove between the median line and each side of the mesonotum, lying near to the lateral margin (Hymenoptera). See notauli.

Párasite, an animal that feeds on or in some other living animal.

Paronýchium (-ia), a bristle-like appendage of the claws.

Péctinate, with branches like a comb.

Péndulous, hanging from one end.

Pedipálpi, pédipalps, the second of the paired appendages of Arachnida.

**Peripneustic**, having a series of functional spiracles along each side of the body.

Péritreme, a chitinous plate surrounding the spiracle in certain mites.

Pétiolate, attached by a stalk or stem.

Phytóphagous, feeding on plants.

Plantula (-læ), a pad-like sole on the underside of the tarsi of certain insects.

Pleurite, one of the side pieces of the body.

Plùmose, feathery.

**Postèrior callósity**, a swelling between the root of the wings and the scutellum (Diptera).

**Postèrior cells**, a variable number of cells extending to the hind margin of the wings, the first bounded inwardly by the anterior crossvein (Diptera).

Posthumeral bristle, one or more bristles just behind the shoulder-swelling (Diptera).

Postnòdal crossveins, a series of short transverse veins next to the costal margin of the wing, beyond the nodus (Odonata).

Postscutéllum, a small transverse piece of the thorax immediately behind the scutellum.

Postvértical bristles, a pair of minute bristles behind the ocelli (Diptera).

**Preápical bristle**, a bristle on the outside of the tibiæ just before the apex (Diptera).

**Prebasilare**, a sclerite on the underside of the head, sometimes divided medially, lying behind the mentum (Diplopoda).

Prédatory, capturing living prey.

Prefúrca, the petiole of the second and third veins (R<sub>4</sub> and R<sub>5</sub>) of Diptera.

**Presutural bristle**, one or more dorsocentral bristles just in front of the transverse suture (Diptera).

Probóscis, the extended trunk-like or beak-like mouthparts.

Pròclinate, inclined forward.

Proepimeron (-ra), that part at the rear of the side of the prothorax next the coxæ.

Prógnathous, having the mouthparts projecting forwards.

Pròlegs, the fleshy abdominal legs of certain insect larvæ.

**Promentum**, a median sclerite in the gnathochilarium, lying in front of the mentum or stipites (Diplopoda).

Pronotum, the back or upper side of the prothorax.

Propleura (-ræ), the side portion of the prothorax.

**Propneustic,** having only the anterior pair of spiracles open and functioning.

**Propodeum**, the large apical portion of the thorax, behind the wings and scutellum (Hymenoptera).

**Prosoma**, an anterior clearly separated section of the cephalothorax in certain Phalangida.

Prostérnum, the middle of the underside of the prothorax.

Prothorácic bristle, a bristle above each of the front coxe (Diptera).

**Prothorax**, the first division of the thorax, bearing the front legs. **Prùinose**, coated with a hoary dust.

**Pseudopod**, an outgrowth or projection of the body of certain larvæ, assisting in locomotion.

Pteropleural bristles, bristles located on the sides of the body just beneath the root of the wings (Diptera).

**Ptilinum**, a temporary bladder-like structure above the antennæ of certain Diptera.

Pulvillus (-li), a pad beneath each tarsal claw.

**Pùpa** (-pæ), the resting stage of insects with complete metamorphosis, following the larva and preceding the adult.

Puparium, the oval, hardened covering of the pupa of the higher Diptera and some scale insects, formed of the larval or nymphal skin.

**Pupiparous**, giving birth to full-grown larvæ that are ready to pupate. **Pygidium**, the last dorsal segment.

Ràdial, of or pertaining to the radius.

Ràdial cell, any cell in the wing bordered in front by a branch of the radius.

Ràdial séctor, the posterior of the two main divisions of the radius.

Ràdius, the third of the principal veins of the typical wing.

Raptòrial, fitted for grasping prey.

Rastéllum, a structure on the cheliceræ of certain spiders bearing numerous tooth-like projections.

Réclinate, pointing backward.

Recurrent nervure, one or two transverse veins arising from the lower side of the submarginal cells (Hymenoptera).

Réniform, kidney-shaped.

Reticulate, meshed, like net work.

Rôstrum, a beak or snout.

Scape, the basal joint or joints of the antennæ.

Sclèrite, any piece of the body wall bounded by sutures.

**Scopa**, a brush on the underside of the abdomen, for collecting pollen (Hymenoptera).

Scópula, a brush of hairs or bristles (Araneida).

**Scrobe**, a groove or furrow, especially one on the mandible or one which receives the antenna.

Scutéllum, a somewhat triangular or crescentic sclerite just behind the mesonotum.

Sérrate, toothed along the edge, like a saw.

**Séssile**, (a) broadly attached; (b), incapable of movement from place to place.

Sèta (-tæ), a bristle or filament.

Setàceous, bristle-like, slender.

Shagreened, having a finely roughened surface.

Sínuous, S-shaped, winding back and forth.

**Small-crossvein**, a short crossvein extending from the base of the discal cell to the fifth posterior cell between M and Cu (Diptera).

Spátulate, broad at tip, narrowed at base.

Spinulated, furnished with very small spines.

Spiracles, breathing pores or external openings of the tracheal system.

Spurs, movable spines, usually two, at the end of the tibiæ.

**Spurious vein**, an adventitious longitudinal vein crossing the anterior (r-m) crossvein (Diptera).

**Squamopygidium**, a plate formed by the fusion of several apical abdominal segments (Dermaptera).

Sternauli, a pair of lateral furrows on the mesothorax below the base of the wings (Hymenoptera).

Stérnite, the ventral piece of each abdominal segment.

Sternopleural bristles, the bristles on the triangular pleural sclerite between the front and middle coxæ (Diptera).

Stigma, a thickening on the costal border of the wings near the apex. Stipites (sing. stipes), a pair of sclerites lying at each side of the gnathochilarium (Diplopoda).

Stridulation, a chirping or creaking noise.

**Style**, (a) a bristle-like process terminating the antennæ, thicker than the arista (Diptera); (b) short slender appendages on the underside of the abdomen (Thysanura).

Stỳliform, drawn out as a slender stiff process.

Subanténnal groove, a groove or grooves in the middle of the face (Diptera).

Subcosta, the second of the principal veins of the typical wing.

Submarginal cell, one or more cells lying behind the marginal cells, usually forming a second row below the anterior margin apically (Hymenoptera).

Submedian cell, a long basal cell near the middle of the wing (Hymenoptera).

Submentum, the basal part of the mentum.

Súlcate, grooved or furrowed.

Supraànal plate, a dorsal sclerite terminating the abdomen.

Suture, a line separating the parts of the body wall.

Társus (-si), the foot, the jointed portion of the legs beyond the tibia.

Tégmen (-mina), the toughened fore wings of grasshoppers, etc.

**Tégula** (-læ), a small convex plate over the root of the fore wings (Hymenoptera).

**Télepod**, a modified leg, serving as a copulatory appendage, on one of the posterior body segments of male Diplopoda.

**Télson**, the last abdominal segment.

**Tentòrial rods**, several diverging chitinous rods within the posterior part of the head of certain dipterous larvæ.

Térgite, the dorsal piece of an abdominal segment.

Térmen, the outer, or distal margin of the wing.

Thèca, a sheath or sac-like covering.

**Thòrax**, the second of the main divisions of the insect body, between the head and the abdomen, bearing the legs and wings.

Tibia (-iæ), the shin-joint of the legs, between the femur and the tarsus.

Triangle, a small triangular cell near the base of the wing (Odonata).

**Trichobóthria**, minute sensory hairs on the tarsal joints of the legs of certain spiders.

**Triúngulin**, the active first-stage larva of the Strepsiptera and certain hypermetamorphic beetles.

**Trochanter**, the small second joint of the legs between the coxa and the femur.

**Trochántin**, a small piece between the sternum and the front coxa (Coleoptera).

Trúncate, ending squarely with sharply cut-off edge.

**Týmpanal hoods, týmpanal bullæ**, a pair of tubercles or swellings at the base of the abdomen in certain Lepidoptera.

Venation, the course of the veins or rod-like thickenings of the wings.

Véntral, pertaining to the underside of the body.

**Véntral mémbrane**, the skin-like tissue connecting the tergites and the sternites (Diptera).

Véntral ségments, the sternites of the abdomen.

Vérrucose, covered with minute warts or tubercles.

Vértex, the crown of the head.

Verticillate, provided with whorls of fine hairs.

Vestígial, small, degenerate, not functional.

Vibrissa (-sæ), a bristle or bristles on each side of the mouth-opening in front (Diptera).

Viviparous, bringing forth living young, not laying eggs.

## INDEX TO GENERA AND HIGHER GROUPS

Illustrations are indicated by page numbers in bold face type.

Ababa, 435 Abavus, 286 Abedus, 154 Abisara, 239 Ablabesmyia, 282 Ablerus, 486 Abræinæ, 423 Abraxas, 209 Abrosoma, 63 Acalypta, 142 Acalyptratæ, 312, 321 Acanalonia, 118, 120 Acanaloniidæ, 120 Acanthaspis, 147 Acanthedra, 231 Acantheis, 563 Acanthiidæ, 143, 146 Acanthocera, 287 Acanthoceridæ, 451 Acanthocerus, 451 Acanthochermes, 127 Acanthocnema, 319 Acanthocneminæ, 299 Acanthocolonia, 63 Acanthocoris, 148 Acanthoctenidæ, 561 Acanthoctenus, 561 Acanthomera, 286 Acanthomeridæ, 286 Acanthophorus, 440 Acanthophrynus, 542 Acanthoplus, 53 Acanthops, 87 Acanthopsylla, 406 Acanthothrips, 76 Acarida, 565 Acaridæ, 574 Acarina, 565 Acaroidea, 572 Acartophthalminæ, 334 Acartophthalmus, 334, **335**, 341 Acarus, **573**, 574 Acattyma, 556 Acelyphus, 326 Acentropus, 214 Aceratogallia, 112 Acercus, 572 Acerentomidæ, 40 Acerentomon, 40, 41 Acerentulus, 40, 44 Acheles, 576

Acherontia, 208, **528** Acheta, 54 Achetidæ. 54 Achetinæ, 54 Achias, 331 Achilidæ, 119 Achilixia, 119 Achilixiidæ, 119 Achilus, 119 Achlæna, 87 Achlyodes, 235 Achlyodidæ, 236 Achoreutes, 46 Achrochordum, 591 Achroia, 215 Acicephala, 319 Acidalia, 210 Acidaliidæ, 210 Acidia, 341 Acilius, 411 Acinipe, 57 Aciura, 341 Aclista, 495 Acmæodera, 419 Acmæops, 441 Acmonotus, 187 Acnemia, 278 Acneus, 416 Acoloithus, 204 Acoma, 451 Acontista, 86 Acontistinæ, 86 Acontistoptera, 350, **351** Acordulecera, 474, 513 Acræa, 238 Acræidæ, 238 Acraga, 206 Acragidæ, 206 Acreioptera, 427 Acrida, **55** Acrididæ, 56 Acridiidæ, 56 Acridiinæ, 57 Acridinæ, 56 Acridoxena, 51 Acridodea, 49 Acritus, 436 Acrobasis, 215 Acroberotha, 186 Acrocera, **291**, 292 Acroceratidæ, 292, 375

Acroceratine, 292

Acrocercops, 232 Acroceridæ, 362 Acrodesmia, 285 Acrolepia, 221 Acrolepiidæ, 221 Acrolophidæ, 224, 245 Acrolophus, 224, **244** Acromantine, 91 Acromantis, 91 Acrometopia, 329 Acronycta, 218 Acrophylla, 64 Acrophyllinæ, 65 Acropsopilio, 552, 553 Acropsopilionidæ, 553 Acropteroxys, 432 Acrotrichis, 433 Acrotylus, 56 Acrydiidæ, 55, 56 Acrydium, 57 Actaletes, 45 Actaletidæ, 45 Actenoptera, 323 Actenotarsus, 99 Actias, 213 Actina, 284 Actininæ, 284 Actinoptera, 341 Actinopus, 556 Actinothrips, 75 Acucephalinæ, 114 Acucephalus, 114 Aculeata, 475 Acumontia, 552 Acutangulus, 588 Adæa, 214 Adæum, 552 Adalia, 434 Adanastus, 432 Adapsilia, 330 Addæa, 212 Adela, 224, **241**, **244** Adelges, **125**, 126 Adelgidæ, 126 Adelginæ, 126 Adelidæ, 224, 243 Adelocephala, 211 Adelocera, 421 Adelopteromyia, 350 Adelotopus, 409 Adelphomyia, 268 Adenophlebia, 172

Adephaga, 408 Aderidæ, 439 Aderus, 439 Adimeridæ, 431 Adimerus, 431 Adiscofiorinia, 130 Adonea, 558 Adoneta, 207 Adoretus, 452 Adoxus, 444 Adrama, 340 Adraminæ, 340 Adranes, 415, **416** Ædaleus, 56 Aedes, 272 Ægeria, 203 Ægeriidæ, 203, 243 Ægialia, 451 Ægialiidæ, 451 Ægialites, 436 Ægialitidæ, 436 Ægidium, 451Ægithus, 432 Ænictus, 490 Ænigmatias, 305, 350 Ænigmatiinæ, 305 Ænigmatistes, 305, 350 Ænigmatopæus, 350 Æolothripidæ, 73 Æolothrips, 73 Aepophilidæ, 143 Aepophilus, 143 Æschna, 164, **166** Æschnidæ, 164, 167 Æschnoidea, 163 Æthalochroa, 86 Æthialion, 111 Æthialionidæ, 111 Æthomorpha, 443 Aethus, 151 Ætioptera, 91 Agabus, 411 Agalena, 560 Agalenidæ, 560 Agallia, 112 Agalliaphagus, 470 Aganaidæ, 217 Aganais, 217 Agandecca, 119 Agaon, 483 Agaontidæ, 483, 487 Agapetidæ, 238 Agapetus, 193 Agarista, 219 Agaristidæ, 219, 248 Agathidinæ, 480

Agathinæ, 480 Agathis, 480 Agathomera, 64 Agathomyia, 310 Agdistidæ, 202 Agdistis, 202 Agenia, 497 Ageniaspis, 484 Aglais, 238 Aglaope, 207 Aglossa, 215 Aglycyderes, 444, **448** Aglycyderidæ, 444 Aglycyderoidea, 444 Agnatha, 169 Agnippe, 227 Agonopteryx, 229 Agonosomatinæ, 303 Agonoxena, 225, 233 Agonoxenidæ, 225, 233 Agraulis, 238 Agria, 316 Agriidæ, 162, 165 Agrilinæ, 420 Agrilus, 419 Agrioidea, 161 Agrion, 161, 162 Agrionoidea, 161 Agrionopsis, 86 Agriothera, 225 Agriotypidæ, 477 Agriotypus, 477 Agrœcia, 52 Agrœciinæ, 52 Agromyza, 338, 368 Agromyzidæ, 339, 343 Agrotera, 216 Agroterinæ, 216 Agrotinæ, 218 Agrypnetes, 192 Agrypnia, 193 Alaptus, 483 Alaus, 421 Albia, 572 Albiorix, 550 Alcestis, 120 Alcidis, 213 Aldrichia, 296 Aldrichiella, 345 Aldrovandiella, 276 Aleochara, 429 Aleocharinæ, 429 Aletia, 218 Aleurobius, 574 Aleurocanthus, **122**, 124 Aleurochiton, 124

Aleurodicinæ, 124 Aleurodicus, **122**, 124 Aleuropteryginæ, 182 Aleuropteryx, 182 Aleyrodidæ, 124 Aleyrodinæ, 124 Aleyrodes, 122, 124, 125 Alichus, 576 Alienidæ, 488 Alienus, 488 Allecula, 435 Alleculidæ, 435 Allobosca, 347, 352 Alloboscinæ, 347 Allochrysa, 185 Allocœlia, 493 Allocoryninæ, 447 Allocorynus, 447 Allocosa, 561 Allodahlia, 70 Allodape, 508 Allodia, 278 Allœostylus, 320 Allogaster, 164 Allognosta, 284 Allononyma, 228 Allopauropus, 584 Allophyla, 334 Allopocockia, 595 Alloporus, 596 Alloptes, 574 Allorhina, 452 Allostethella, 68 Allostethidæ, 68 Allostethus, 67, 68 Allotria, 483 Allotrichia, 192, **193** Allotrichoma, 343 Alofia, 581 Alokobius, 600 Alombus, 352 Alophora, 318 Altella, 558 Altica, 444 Alticidæ, 444 Alucitidæ, 202 Alydidæ, 148 Alydus, 148 Alypia, 219 Alypiodes, 220 Alysia, 477 Alysiidæ, 477 Alyson, 505 Alysonidæ, 505 Amantis, 90 Amara, 411

Amata, 219 Amathomyia, 299 Amathusia, 238 Amathuxidea, 238 Amatidæ, 219 Amaurobiidæ, 558 Amaurobius, 558 Amblyaspis, 494 Amblycera, 101 Amblyomma, 567 Amblypygi, 540 Amblythyreus, 147 Ambrysus, 154 Ameles, 90Ameletus, 172 Amelinæ, 90 Ametastegia, 474 Ametropodidæ, 172, 175 Ametropus, 172 Amicrocentrum, 480 Amictus, 296 Amiota, 345 Amiotinæ, 345 Amitermes, 95 Ammophila, 503 Ammoplanus, 506 Ammotrecha, 547 Ammoxenidæ, 561 Ammoxenus, 561 Amobia, 317 Amobiinæ, 317 Amœbaleria, 335 Amorphoscelinæ, 84 Amorphoscelis, 84 Amphiacusta, 54 Amphibolips, 483 Amphicoma, 451 Amphientomidæ, 98 Amphientomum, 98 Amphigerontia, 99 Amphigyrum, 558 Amphilecta, 285 Amphiopinæ, 414 Amphipsocus, 99 Amphipsylla, 406 Amphipterygidæ, 162 Amphipteryx, 162 Amphiscepa, 120 Amphiscepidæ, 120 Amphisternus, 434 Amphithera, 225 Amphitheridæ, 225 Amphorophora, 125 Amphix, 434 Amphizoa, 409 Amphizoidæ, 409

Amphypterygidæ, 166 Amplinus, 590 Ampulex, 503 Ampulicidæ, 503 Ampulicimorpha, 492 Amycle, 119 Amydria, 224 Amydriidæ, 224 Amytta, 51 Anabolia, 192 Anaborrhynchus, 294 Anabrus, **49**, 52 Anacampsis, 227 Anacampta, 331 Anacanthaspis, 289 Anacanthella, 286 Anacanthotermes, 95 Anacrabro, 506 Anadiastothele, 555 Anæa, 238 Anagarypus, 550 Anagenesia, 169 Anagnota, 339 Anagraphis, 562 Anajapyx, **41**, 44 Analcocerinæ, 286 Analcocerus, 285, 286 Analges, 574 Analgesidæ, 574 Analopteryx, 350 Aname, 556 Anamorpha, 599 Ananteris, 545 Anapausis, 276 Anaphes, 483 Anaphothrips, 74 Anaplecta, 79 Anaptycha, 81 Anareolatæ, 61 Anarmonia, 226 Anarsia, 227 Anasa, 148, **149** Anasigerpes, 91 Anaspis, 438 Anastatus, 484 Anastrepha, 341 Anatalanta, 350, **351** Anatelia, 66, **67** Anateliidæ, 66 Anatis, 434, **528** Anatopynia, **281**, 282 Anax, 164 Anaxarcha, 91 Anaxiphus, 54 Ancistrocerus, 497 Ancistrogaster, 67, 70

Ancistrogastridæ, 70 Ancylis, 226 Ancylolomia, 215 Ancylolomiinæ, 215 Ancylometis, 225 Ancyloxipha, 236 Ancyrona, 419 Andex, 69 Andrena, 508 Andrenidæ, 508 Andrenosoma, 299 Androloma, 219 Androniscus, 534 Androprosopa, 273 Anechura, 70 Anechuridæ, 70 Anepisceptus, 53 Anepitacta, 51 Anerastia, 215 Anerastiinæ, 215 Aneurus, 145 Angela, **85**, 86 Angelinæ, 86 Angituloides, 332 Angitula, 332 Anigrus, 119 Anisacantha, 63 Anisaspis, 556 Anisaspoides, 556 Anisocentropus, 194 Anisolabidæ, 68 Anisolabis, 68 Anisomorpha, 62, 64 Anisomorphinæ, 64 Anisopleura, 162 Anisoplia, 452 Anisopodidæ, 275, 359, 373 Anisopogon, 298 Anisops, 153 Anisoptera, 160, 165 Anisopus, **265**, 275, **359** Anisota, 211 Anisotamia, 295 Anisotomidæ, 428 Anisozygoptera, 160, 165 Ankothrips, 73 Anobiidæ, 426 Anobium, 426 Anogdus, **416** Anomala, 452 Anomalaphis, 125 Anomalempis, 301 Anomalochæta, 339 Anomologa, 225 Anomologidæ, 225

Anomophagus, 419 Anomoses, 201. 201 Anomosetidæ, 201 Anopheles, 272, 370, Anophelinæ, 272 Anophthalmus, 411 Anoplocnemis, 148 Anoplura, 104 Anopsobius, 600 Anorostoma, 335 Anosia, 237 Anostostoma, 50 Anostostomatinæ, 50 Anotia, 119 Anotogaster, 164 Anoura, 45 Antarctophthirius, 104 Antennomegistus, 568 Antennophorinæ, 568 Antennophorus, 568 Anteon, 493 Anthalia, 301 Anthaxia, 419 Anthela, 217 Anthelidæ, 217, 220 Antheræa, 212, 213 Anthericomma, 469, 470 Antherophagus, 424 Anthicidæ, 439 Anthicus, 439 Anthidium, 507 Anthobosca, 499 Anthoboscidæ, 499 Anthocharis, 236 Anthocoptes, 565 Anthocoridæ, 144 Anthocoris, 144 Anthogona, 591 Anthogonidæ, 591 Anthomyia, 319 Anthomyiidæ, 319, 366, 367 Anthomyiinæ, 319 Anthomyza, 339 Anthomyzidæ, 339 Anthophasia, 332 Anthophila, 502 Anthophora, 508 Anthophoridæ, 508 Anthracinæ, 294 Anthracomyia, 316 Anthrax, 294, 295 Anthreninæ, 425 Anthrenus, **416**, 425 Anthribidæ, 449

Anthroleucosoma, 591 Anthroleucosomatidæ 591 Aphidius, 482 Antichæta, 328 Antilochus, 150 Antineura, 331 Antipaluria, 92 Antiphonus, 590 Antisolabis, 69 Antispila, **230**, 230 Antissa, 286 Antissinæ, 286 Antliata, 264 Antocha, 269 Antochinæ, 269 Antongilia, 63 Antonia, 134, 295 Antrops, 350 Anurida. 46 Anuraphis, 125 Anuraptervx, 209 Anystidæ, 575 Anystis, 575 Apachvidæ, 66 Apachyoidea, 66 Apachyus, 66, **67** Apantesis, 219 Apatania, 192 Apatela, 218 Apanteles, 480 Apatelodes, 211 Apatelodinæ, 251 Apatidæ, 426 Apatolestes, 288 Apatura, 238 Apemon, 280 Apetenus, 351 Apfelbeckia, 594 Aphænocephalidæ, 434 Aphænocephalus, 434 Aphæreta, 477 Aphalara, 123 Aphalaroida, 123 Aphaniosoma, 336 Aphaniptera, 404 Aphantochilidæ, 562 Aphantochilus, 562 Aphelinidæ, 486 Aphelinus, 486 Aphelochiridæ, 154 Aphelonema, 118, 121 Aphelopus, 493 Aphelosetia, 229 Aphelosetiidæ, 230 Aphididæ, 124 Aphidiidæ, 477 Aphidiinæ, 482

Aphidinæ, 125 Aphidoidea, 124 Aphilanthops, 505 Aphilodon, 603 Aphiochœta, 306 Aphis, 125, **125** Aphodiidæ, 451 Aphodius, 451 Aphæbantinæ, 295 Aphœbantus, 295 Aphorista, 434 Aphorura, 46 Aphoruridæ, 46 Aphrastobraconinæ, 478 Aphria, 318 Aphrophora, 110, 111 Aphrosylinæ, 303 Aphrosylus, 303, 361, 362 Aphthonetus, 233 Apidæ, 506, 515 Apiocera, 294 Apioceratidæ, 294 Apiococcus, 134 Apoidea, 502 Apiomerus, 147, 587 Apiomorpha, 132 Apiomorphidæ, 132 Apion,  $4\overline{4}7$ Apioninæ, 447 Apioscelis, 55 Apis, 506, **506** Apistomvia, 270 Apistomyinæ, 270 Aplastus, 428 Aploneura, 126 Apneumonella, 557 Apochrysa, 186 Apochrysidæ, 186 Apocephalus, 306 Apocrita, 472 Apoda, 207 Apodacra, 316 Apodigona, 593 Apoidea, 514 Apolysis, 296 Apomidas, 294 Apophorhynchus, 325 Apoprogenes, 203 Apoprogenidæ, 203 Apospasma, 284 Apostraphia, 238 Apsinota, 344 Aptanogyna, 349 Aptericola, 103 Apterina, 350

Apterogyna, 489 Apterogynidæ, 489 Apteroscirtus, 51 Aptervgida, 70 Aptilotus, 350 Apyrrothrix, 235 Arachnida, 535 Arachnocampa, 279 Arachnomorphæ, 555 Aradidæ, 145 Aradoidea, 145 Aradus, 145, **146** Aranea, 560 Araneida, 554 Arbelidæ, 206 Archæa, 558 Archæidæ, 558 Archæolothrips, 73 Archæopsylla, 405 Archæopsyllidæ, 405 Archandra, 419 Archiblatta, 81 Archiblattidæ, 81 Archichauliodes, 179 Archidux, 70 Archihymen, 497 Archihymeidæ, 497 Archilepidoptera, 200 Archilestes, 162 Archimandrita, 82 Archimyia, 289 Archimantinæ, 90 Archimantis, 90 Archimyza, 286 Archips, 226 Archipsocidæ. 99 Archipsocus, 99 Archisometrus, 545 Archistratiomys. 285 Archytas, 318 Arctæcia, 196 Arctia, 219 Arctiidæ, 219, 239, 248, 250 Arctiscidæ, 583 Arctiscon, 583 Arctocoris, 150 Arctophila, 309 Arctophilinæ, 309 Arctosa, 561 Arctosyrphus, 309 Areolaria, 81 Areolariidæ, 81 Areolatæ, 61 Argantidæ, 567

Argas, 537, 567

Arge, 474 Argia, 161 Argidæ, 238, 474, 512 Argiolestes, 162 Argiope, 560 Argiopidæ, 560 Agrotis, 218 Argus, 238 Argynnis, 238 Argyra, 302, 304 Argyramæba, 294 Argyreidæ, 238 Argyresthia, 231, 247 Argyresthiidæ, 231 Argyria, 216 Argyrodes, 559Argyroneta, 560 Argyrotoxa, 226 Argyrotypidæ, 206 Argyrotypus, 206 Ariadna, 557 Ariasella, 350 Aricia, 320 Ariciinæ, 320 Arilus, 147 Aristotelia, 227 Arixenia, 66 Arixeniidæ, 66 Arixenina, 66 Armacia, 121 Armadillididæ, 534 Armadillidium, 534 Armadillo, 534 Armillifer, 581 Arphax, 64 Arphia, 56 Arrhenuridæ, 571 Arrhenurus, 571 Arria, 89 Arrup, 602 Arsenuridæ, 213 Arta, 215 Artematopus, 418 Artemita, 285 Arteriopteryx, 184 Arthria, 276 Arthroceras, 289 Arthroceratinæ, 289 Arthrolips, 431 Arthromacra, 436 Arthropeas, 289 Arthropleona, 45 Arthropterus, 408, 412 Arthrosphæra, 586 Arthrotardigrada, 583 Arthroteles, 289

Arthrotelinæ, 289 Articerus, 415 Aruanoidea, 64 Arytaina, 123 Arvtropteris, 51 Ascalaphidæ, 187 Ascalaphus, 187 Asceles, 64 Ascelis, 132 Ascepasma, 63 Ascepasminæ, 63 Aschiphasma, 63 Aschiphasmatinæ, 63 Ascia, 236 Asciidæ, 236, 253 Asclera, 437 Ascodipteridæ, 347 Ascodipteron, 346, 347, **351**, 352 Ascogaster, 481 Ashinaga, 224 Ashinagidæ, 224 Asida, 436 Asilidæ, 298, 363, 376, 377 Asilinæ, 298 Asilus, 298 Asindulum, 280 Asiphum, 126 Asota, 217 Asotidæ, 217 Asphalidesmus, 590 Asphondylia, 275 Aspicolpus, 480 Aspidiotinæ, 130 Aspidiotus, 129, 130 Aspidolopha, 443 Aspidomorpha, 444 Aspidoptera, 346, 352 Aspistes, 276 Aspistinæ, 276 Assamia, 552 Assamiidæ, 552 Astata, 503 Astatidæ, 503 Astega, 281 Asteia, 332 Asteromyia, 275 Asterolecaniidæ, 132 Asterolecanium, 132 Asthenida, 213 Asthenidæ, 269 Astia, 332, **337**, 340 Astiidæ, 332, 340 Astollia, 86 Astomella, 292 Astrodesmus, 590

Astroma, 55 Astylus, 429 Asyndetus, 304 Atænius, 451 Atalanta, 301 Atalantinæ, 301 Atalophlebia, 171, 172 Atarba, 269 Ateledera, 442 Atelura, 42 Atherigona, 320 Atherix, 290, 362, **363** Athyreus, 450 Athyroglossa, 343 Atissa, 343 Atlides, 239 Atomaria, 424 Atomosia, 298, 299 Atomosiinæ, 299 Atomus, 576 Atopocixius, 119 Atopodesmus, 588 Atopognathus, 332 Atopopus, 172 Atractides, 572 Atractididæ, 572 Atractoceridæ, 430 Atractocerus, 430 Atrichops, 290 Atropidæ, 98 Atropos, 98 Atrytene, 236 Atta, 490 Attacidæ, 213 Attacus, 213 Attageninæ, 425 Attagenus, 425 Attaphila, 78, 80 Attaphilidæ, 78 Attelabinæ, 447 Attemsia, 592 Attemsiidæ, 592 Atteva, 229 Attevidæ, 229 Atticola, 81 Atticolidæ, 81 Attidæ, 561 Attus, 561 Aturus, 572 Atylotus, 288 Atypidæ, 556 Atypus, 556 Auchenomus, 69 Auchenorrhyncha, 106 Augasma, 231 Augochlora, 508

Aulacaspis, 130, **131** Aulacidæ, 476 Aulacidea, 483 Aulacocentrum, 480 Aulacogaster, 333, 333 Aulacogastridæ, 333 Aulacophilus, 504 Aulacus, 476, **482** Aulicus, 416 Aulodesmus, 590 Aulonium, 432 Aulonogyrus, 412 Aulonothroscus, 421 Auromantis, 89 Australobius, 600 Australiosoma, 589 Austrogomphus, 164 Austrohelcon, 480 Austrolestes, 162 Austronymphes, 187 Austroperlidæ, 176 Austrophlebia, 164 Austrosciara, 349 Austrosialis, 180, 180 Austrostylops, 468 Austrozele, 480 Autocrates, 438 Autolyca, 64 Automeris, 213 Automolis, 219 Auximobasis, 232 Avellopsis, 558 Avicularia, 556 Aviculariidæ, 556 Avicularimorphæ, 555 Axia, 211 Axiidæ, 211 Axima, 486 Axymyia, 274, 277 Axysta, 343

Babia, 443
Baccha, 309
Bacchinæ, 309
Bacchinæ, 309
Bacilia, 346
Bacillidæ, 61
Bacillothrips, 76
Bacillothrips, 76
Bacillothrips, 63
Bacteria, 62, 64
Bacteriidæ, 64
Bacteriidæ, 64
Bacteriidæ, 64
Bacterioidea, 61
Bactridium, 64
Bactrocera, 341
Bactroceros, 472

Bactromantis, 84 Bacunculidæ, 64 Bacunculinæ, 64 Bacunculus, 64 Badumna, 558 Bæocera, 426 Baeotis, 239 Baetidæ, 170, 175 Baetis, 170, **171**, **174** Baetisca, 171, 172 Baetiscidæ, 172 Baetoidea, 170 Balaninus, 448 Balbillus, 116 Balioptera, 339 Ballophilus, 602 Bandakia, 572 Bantia, 84 Barce, 147 Bareogonalos, 494 Bargena, 571 Barychelidæ, 556 Barychelus, 556 Barypenthus, 195 Basiæschna, 164 Basilarchia, 238 Basilia, 352 Basilona, 211 Bassareus, 443 Bathyscia, 428 Batillipes, **582**, 583 Batocera, 441 Batoceridæ, 441 Batrachedra, 232 Batrisodes, 415 Bdella, **575**, 576 Bdellidæ, 576 Bebaiotes, 119 Bebelothrips, 74 Bedellia, 223, 223 Bedelliidæ, 223 Belgica, 349 Belidæ, 448 Bellardia, 288 Bellardiinæ, 288 Belomantis, 86 Belomicrus, 505 Beloniscus, 553 Belonogastechthrus, 470 Belonogaster, 497 Belostoma, 154 Belostomatidæ, 154 Belvosia, **317**, 318 Belyta, 495, **495** Belytidæ, 495, 500

Bembecia, 202

Bembex, 505Bembicidæ, 505 Bembidion, 410 Bembidula, 505 Bembix, **504**, 505 Benacus, 154 Bephratoides, 487 Beræa, 194 Beræodes, 194 Beridinæ, 284 Beridops, 284 Beris, 284 Berismyia, 284 Berosus, 414 Berotha, 186 Berothidæ, 186 Bertea, 349 Berytidæ, 148 Bethylidæ, 494, 501, 514 Bethylus, 494 Bezzia, 282 Bibio, **265**, 275, **276**, 370 Bibiocephala, 270, 353, **355**, 371 Bibiodes, 275 Bibionidæ, 275, 358, 370 Bibioninæ, 275 Bicalcar, 290 Bicalcarinæ, 290 Bicellaria, 301 Bicyrtes, 505 Bidessus, 411 Bimichælia, 576 Biosteres, 481 Bittacidæ, 189 Bittacomorpha, 267, 267, **355,** 373 Bittacomorphella, 267 Bittacomorphinæ, 267 Bittacus, 189 Bittacusidæ, 189 Blaberidæ, 82 Blaberus, 82 Blacinæ, 481 Blacus, 481 Blæsoxipha, 316 Blaniulidæ, 595 Blaniulus, **589**, 595 Blapidæ, 436 Blaps, 436 Blaptica, 82 Blasticotoma, 474 Blasticotomidæ, 474, 511 Blastobasidæ, 222, 232, 246Blastobasis, 232

Blastodacna, 232 Blastophaga, 483 Blatella, 78, 79, 80 Blatta, 78 Blattariæ, 77 Blattidæ, 78 Blattoidea, 77 Blennocampinæ, 512 Blepharocera, 269, 270 Blepharoceratidæ, 269, 353,371Blepharocerating, 270 Blepharodes, 84 Blepharopsis, 84 Blepharoptera, 335 Blissus, 149, **149** Blothrus, 549 Boarmia, 209 Boarmiid**æ,** 209 Bocchus, 493 Bochica, 550 Bogeria, 313 Bolbe, 88 Bolboceras, **416**, 450 Bolbomyia, 290 Bolbula, 88 Boletina, 278, **278** Bolitophagus, 436 Bolitophila, 279, **279** Bolitophilella, 279 Bolitophilidæ, 279, 358 Bolivaria, 86 Bolyphantes, 560 Bombidæ, 507, 515 Bomboptera, 471 Bombus, 507 Bombycidæ, 206, 212, 252 Bombyliidæ, 294, 363,376, 377Bombyliinæ, 297 Bombylius, **283**, **295**, 297 Bombyx, 205, 212 Bomolocha, 218 Bondia, 225 Boophthora, 281 Boopia, 102 Boopiidæ, 102 Borboridæ, 337, 338, 350 Borboroidea, 312 Borborus, 337 Boreidæ, 189 Boreodromia, 301 Boreoides, 349 Boreus, 189 Borkhausenia, 232 Bormansia, 68

Bostra, 64 Bostrychidæ, 426 Botanobia, **339**, 340 Botanobiinæ, 340 Bothrideres, 433 Bothrideridæ, 433 Bothriocera, 120  $\operatorname{Bothriogaster}$ , 602Bothriophorinæ, 417 Bothriuridæ, 543 Bothriurus, 543, **544** Bothropolys, 600 Bothynostethus, 505 Bourletiella, 46 Brachinus, 410, 418 Brachiosternus, 543 Brachistes, 481 Brachistinæ, 481 Brachybainosoma, 593 Brachybothrium, 556 Brachycentrus, 194 Brachycera, 265, 304, 347, Brachycercidæ, 171 Brachveerinæ, 447 Brachycerus, 447 Brachychæteuma, 591 Brachychæteumidæ, 591 Brachveistinæ, 499 Brachycistis, 499,502Brachycolus, 125 Brachycoma, 316 Brachydesmus, 588 Brachydeutera, 343 Brachygaster, 476 Brachylabidæ, 69 Brachylabis, 69 Brachymera, 317 Brachvnemurus, 187 Brachyneura, 275 Brachyopa, 309 Brachyopinæ, 309 Brachypalpus, 309 Brachypauropodidæ, 584 Brachypauropus, 584 Brachypelta, 151 Brachyplatys, 150 Brachypoda, 572 Brachypodidæ, 572 Brachypremna, 268 Brachypteridæ, 419 Brachypterus, 419 Brachyrhinidæ, 448 Brachyrhynchus, 145 Brachys, 419 Brachyscelidæ, 132

Brachysphænus, 432 Brachystoma, 301, **302** Brachystomatinæ, 301 Brachythele, 556 Brachytypus, 56 Brackenridgia, 534 Bracon, 480 Braconidæ, 477, 488 Braconinæ, 478, 480 Bradyporine, 52 Bradyporus, 52 Bradythrips, 74, 75 Brahmæa, **212**, 213 Brahmæidæ, 213 Brancsikia, 89 Brassolidæ, 238 Brassolis, 238 Brathinidæ, 429 Brathinus, 429 Braula, 345, **346**, 352 Braulidæ, 345, 352 Braunsia, 480 Bremidæ, 507 Bremus, 507 Brenthidæ, 447 Brenthis, 238 Brentidæ, 447 Brentus, 447, 448 Brephidæ, 209 Brephos, 209 Brevicoryne, 125 Brizoides, 64 Brochymena, 151 Brœlemanneuma, 592 Brœlemannia, 594 Bromophila, 330 Brontes, 416, 424 Broteas, 545 Brotheus, 447 Bruchelidæ, 449 Bruchidæ, 440 Bruchomorpha, 121 Bruchomyia, 271, **271** Bruchomyiinæ, 271 Bruchophagus, 486, 487 Bruchus, 439 Brunneria, 86 Bryaxis, 415, **416** Bryobia, 576 Bucculatrigidæ, 223 Bucculatrix, 222 Bucranium, 562 Buenoa, 153 Bulla, 56 Bundera, 117

Buplex, 288

Buprestidæ, 419 Buprestinæ, 420 Buprestis, 419 Burinia, 597 Burriola, **67**, 70 Butalidæ, 231 Butalis, 231 Butheolus, 545 Buthidæ, 545 Buthus, 536, 545 Byrrhidæ, 417 Byrrhinæ, 417 Byrrhus, 417 Byrsopidæ, 449 Byrsops, 449 Byssodon, 281 Bythoscopidæ, 112 Bythoscopus, 112 Byturidæ, 425 Byturus, 425

Cacoblatta, 82 Cacœcia, 226 Caddo, 553 Cadmon, 556 Cæcilliidæ, 99 Cæcilius, 97, 99 Cæculidæ, 574 Cæculus, 574, **575** Cælenopsis, 569 Cælopygus, 553 Cænidæ, 171, 175 Cænides, 236 Cænis, 170, 171 Cænocholax, 469 Cænocolax, 469 Cænoscelis, 424 Cænotus, 296 Calamacris, 57 Calamoceras, 194 Calamoceratidæ, 194a, 196 Calonyx, 571 Calamothespis, 86 Calandridæ, 449 Calendra, 449 Calendrinæ, 449 Calephelis, 234, 237, 239 Calidomantis, 89 Caligo, **237**, 238 Caligonidæ, 238 Caliope, 327 Caliridinæ, 89, 90 Caliris, 89 Caliscelis, 121 Callibaetis, 170

Callibia, 86

Callicera, 308

Calliceras, **493**, 494 Calliceratidæ, 494, 501 Calliceratine, 308 Callichroma, 441 Callidula, 221 Callidulidæ, 216, 221 Callimenus, 52 Callilepis, 562 Callimome, 485 Callimomidæ, 485, 487. 488 Callimorpha, 217 Callimorphidæ, 217 Callimyia, 364, **365** Callipappus, 128 Callipharixenidæ, 469 Callipharixenos, 469 Calliphora, **312**, **315**, 316, 367Calliphoridæ, 314, 367 Calliphorinæ, 316 Callipodella, 594 Callipodidæ, 593 Calliprobola, 309 Callipterus, 125 Callipus, 593 Callirhipis, 425 Callispa, 444 Callistoma, 295 Callistoptera, 99 Callopistria, 331 Callosamia, 213 Calobata, **324**, 325 Calobatella, 325 Calobatinæ, 325 Caloblatta, 79 Calocoris, 145 Caloctenus, 563 Calolampra, 79 Calommata, 556 Calophya, 123 Caloptenus, 55 Calopterella, 344 Calopterygidæ, 162 Calopteryx, 162 Calopus, 437 Calosima, 222 Calosoma, 410, **528** Calotarsa, 310, **310** Caloteleia, 494 Calotermes, 94, 94 Calotermitidæ, 94, 95 Calycopteryx, 351 Calvotinæ, 481 Calyptratæ, 311

Calyptus, 481 Cambala, 596 Cambalidæ, 596 Cambaloideæ, 596 Cambalomorpha, 597 Cambalopsidæ, 597 Cambalopsis, 597 Camilla, 344 Camillinæ, 344 Campodea, 41, 44, 44 Campodeidæ, 44 Campodeoidea, 43 Campœa, 209 Camponotus, 489, 491 Campsieneminæ, 304 Campsicnemus, 304 Campsomeris, 498 Campsurus, 170 Camptobrochis, 145 Camptocerus, **414**, **446**, 446 Camptocladius, 282, 357, 357 Camptonotus, 53 Camptoprosopella, 327 Campylocera, 330 Campylodes, 207 Campylomyza, 275 Canace, 336, **337**, 338 Canaceidæ, 336, 338 Canaria, 295 Canestrinia, **573**, 574 Canestriniidæ, 574 Cantacader, 142 Cantharidæ, 431, 439, 453 Cantharis, 431, 439 Cantharoctonus, 478 Cantharoidea, 428 Canthon, 450 Canthydrus, 411 Canthyloscelis, 276, 276 Canuleius, 63 Capitoniidæ, 477 Capnella, 178 Capnia, 178 Capniidæ, 178 Capnura, 178 Caponia, 557 Caponina, 557 Caponiidæ, 557

Capritermes, 95

Capsidæ, 145

Carabidæ, 411

Caraboctonus, 546 Carabus, 411

Capsus, 145

Caradrina, 218 Caradrinidæ, 218 Caradrininæ, 218 Carausius, 64 Carcinocoris, 147 Cardiocephala, 325 Cardiochiles, 480 Cardiochilinæ, 480 Cardiodactylus, 54 Caria, 239 Cardiophorus, 421 Caribolus, 595 Carnidæ, 345, 352 Carnus 345, 352 Carpocapsa, 226 Carpophilus, 419 Carposina, 225, **244** Carposinidæ, 225, 245 Carsidara, **122**, 123 Carsidarinæ, 123 Cartocere, 434 Carventus, 145 Caseya, 591 Casevidæ, 591 Cassida, **442**, 444 Cassididæ, 444 Castaneira, 563 Castnia, 203 Castniidæ, 203 Catacantha, 151, **151** Catageus, 542 Cataplectica, 231 Catara, 81 Catogenus, 424 Catharosoma, 589 Catocala, 218 Catocalinæ, 218 Catonia, 119 Catopochrotidæ, 424 Catopochrotus, 424 Catops, 428 Catopsidæ, 428 Catopsilia, 236 Catoptrichus, 416 Cebrio, **422**, 428 Cebrionidæ, 428 Cebriorhipis, 428 Cecidomyia, 275 Cecidomyiidæ, 274, 349, Ceratocampa, 211 369Cecidomyiinæ, 275 Cecidoses, 223, 224 Cecidosidæ, 224 Celama, 219

Celerio, 209

Celithemis, 164

Celcenorrhinus 236 Celvphidæ 326 Celyphus, 326, 326 Cemiostoma, 223 Cemiostomatidæ, 223 Cenchridobia 345, 352 Cenocœliinæ, 480 Cenocœlius, 480 Cenoloba, 202 Cenopogon, 298 Centistes, 481 Centistina, 481 Centris, 508 Centrophlebomyia 341 Centroptilum, 170 Centrotus, 108 Centruroides, **544**, 545 Cephalelinæ, 113 Cephalelus, 113, **113** Cephaliidæ, 331 Cephalobæna, 580 Cephalobænidæ, 579 Cephalobæninæ, 580 Cephalocera, 297 Cephalodonta, 444 Cephaloidæ, 438 Cephalolia, 444 Cephalomyia, 313 Cephaloon, 438 Cephalothrips, 72 Cephanium, 428 Cephidæ, 472, 513 Cephus, 472, **473** Ceramioides, 496 Cerambycidæ, 441 Cerambycoidea, 413 Ceramius, 496 Cerapachyinæ, 490 Cerapachys, 490 Ceraphronidæ, 494 Cerapterus, 408 Cerasommatidiidæ, 434 Cerataphis, 126 Ceratina, 508 Ceratinidæ, 508 Ceratipsocus, 99 Ceratitinæ, 341 Ceratitis, 341 Ceratocampidæ, 211 Ceratocombidæ, 143 Ceratocombus, 143 Ceratocrania, 86 Ceratomantis, 86 Ceratomerinæ, 301

Ceratomerus, 301

Ceratomontia, 552 Ceratophyllidæ, 406 Ceratophyllus, 406, 406 Ceratopogon, 282 Ceratopogonidæ, 282, 356, Ceratopogoninæ, 282 Ceratosolen, 483 Ceratosoma, 592 Ceratothripidæ, 74 Ceratothrips, 74 Cerceridæ, 505 Cerceris, 504, 505 Cercometus, 154 Cercopa, 111 Cercophana, 213 Cercophanidæ, 213 Cercophonius, 543 Cercopidæ, 111 Cercopoidea, 109 Cercyon, 414 Cercyonis, 238 Cerdistus, 298 Ceresa, 107, 108 Ceria, 308 Ceriacreminæ, 123 Ceriacremum, 122, 123 Cerioides, 308 Cerioidinæ, 308 Cermatobiidæ, 600 Cermatobioidea, 600 Cermatobius, 600 Cerococcus, 132, **133** Cerodonta, 339 Ceroma, 547 Cerometopon, 343 Ceropales, 497 Ceropalidæ, 497 Cerophytidæ, 421 Cerophytum, 421 Ceroplastes, 132 Ceroplatidæ, 280, 358,  $37\bar{0}$ Ceroplatus, 280, 358, **359**,  $37\overline{2}$ Ceropsylla, 123 Cerostoma, 228 Cerotelion, 280 Cerotoma, 444 Ceroxys, 331 Ceroys, 63 Ceruchus, 450 Cerura, 211

Ceruridæ, 211

Cerylon, 432

Cerynia, 120

Cethosia 238 Cetonia, 452 Cetoniidæ, 452 Ceuthophilus, 49, 50 Chactas, 545 Chactidæ, 545 Chactinæ, 545 Chærea, 558 Chæriliidæ, 545 Chæriliinæ, 545 Chærilius, 545 Chæteessa, 84, 88 Chæteessinæ, 84 Chætodacus, 341 Chætoneurophora, 305, 305 Chætopsis, 330, **330** Chætospania, 69 Chaitophorus, 125 Chalarus, 310 Chalastogastra, 471, 510 Chalceopla, 224 Chalcididæ, 485 Chalcidoidea, 482, 514 Chalcis, 485 Chalcodectus, 486 Chalcophorinæ, 420 Chalcopteryx, 162 Chalcosia, 207 Chalcosiidæ, 207 Chalepus, 444 Challia, 66 Chamæbosca, 350 Chamæmyia, 329 Chamæmyiidæ, 329 Chamæosoma, 591 Chamæosomatidæ, 591 Chamæpsila, 340 Chamæsyrphus, 308 Chanystis, 225 Chaoboridæ, 272 Chaoborinæ, 272 Chaoborus, 272 Chapuisia, 445 Chapuisiidæ, 445 Charagis, 201 Charidea, 207 Charideidæ, 207 Charinus, 542 Charon, 542 Charontidæ, 542 Chartergus, 497 Chasmia, 287 Chasmiinæ, 287 Chasmoptera, 183, **183** Chauliodes, 179, 180

Chauliodus, 231 Chauliognathus, 431 Cheirochela, 154 Cheiropachys, 486 Cheirotonus, 452 Chelepteryx, 217 Chelidura, 70 Chelidurella, 70 Cheliduridæ, 70 Chelifer, 536, 549 Chelifera, 301, 302 Cheliferidæ, 550 Chelipoda, 301 Chelisia, 320 Chelisodoches, 69 Chelisodochidæ, 69 Chelonariidæ, 417 Chelonarium, 417 Chelonella, 481 Chelonethida, 548 Cheloninæ, 481 Chelonogastra, 478 Chelonus, 481 Cherastus, 595 Chelymorpha, 444 Chermes, 123 Chermidæ, 122 Cherminæ, 123 Chernetidea, 548 Chevletia, 576 Cheyletidæ, 576 Cheyletiella, 576 Cheyletus, **575**, 576 Chiasmoneura, 280 Chiasmopes, 561 Chilocoris, 434 Chilognatha, 585 Chilopoda, 599 Chilosia, 308 Chilosiinæ, 308 Chimarrha, 192 Chimarrhometra 152 Chionæma, 219 Chionamœba. 294 Chionaspidinæ, 130 Chionaspis, 130, **131** Chionea, 348, **348** Chiracanthium, 563 Chiromyia, 336 Chiromyza, 286 Chironyzidæ, 286, 349 Chironomidæ, 347, 349, Chironominæ, 282 Chironomus, 281, 357, 357 Chirosia, 319

Chirothripoides, 75 Chirothripoididæ, 75 Chirotonetes, **171**, 172 Chitoniscus, 63 Chitonophora, 172 Chitra, 549 Chlænius, 410 Chlamydidæ, 443 Chlamys, 443 Chlidanota, 226 Chlidanotidæ, 226 Chlidonia, 227 Chlorion, 473, 491, 503 Chlorippe, 238 Chlorissa, 210 Chlorochara, 120 Chlorocnemis, 161 Chlorolestes, 161 Chloromioptervx, 90 Chloroperla, 176, **177** Chloropidæ, 340, 352 Chloropinæ, 340 Chloropisca, 340 Chlorops, 340 Chlorotettix, 114 Chœradodinæ, 90 Chœradodis, 90 Chœridium, 450 Choneiulus, 595 Chonocephalus, 306, 350 Choragidæ, 449 Chordeuma, 593 Chordeumella, 593 Chordeumidæ, 593 Chordeumoidea, 590 Chordonota, 286 Choreutidæ, 228 Choreutinæ, 228 Choreutis, 228 Chorioptes 574 Chorisoneura, 81 Chorisoneuridæ, 81 Chorista, 191 Choristella, 190 Choristima, 81 Choristinæ, 191 Chorcetypinæ, 56 Chorcetypus, 56 Chortophila, 319 Chremylus, 479 Chromagrion, 160 Chrotogonus, 57 Chrysantheda, 509 Chrysanthedidæ, 509 Chrysauge, 215 Chrysauginæ, 215

Chrysididæ, 493 Chrysis, 493, **493** Chrysobothrinæ, 420 Chrysobothris, 419, 421. Chrysochlora, 285, 286 Chrysochlorinæ, 286 Chrysochorixenos, 469 Chrysochroa, 419 Chrysochroinæ, 420 Chrysochroma, 286 Chrysochus, 444 Chrysoclista, 233 Chrysocoris, 150 Chrysogaster, 308 Chrysolampra, 444 Chrysolampus, 486 Chrysomela, 443 Chrysomelidæ, 443 Chrysomphalus, 130 Chrysomyia, 314, 336 Chrysomyiinæ, 314 Chrysomyza, 330 Chrysopa, 185, 186, 528 Chrysopeleia, 229 Chrysopeleiidæ, 230 Chrysophanus, 239 Chrysopidæ, 185 Chrysopila, 362, 363, 374, 375 Chrysopilinæ, 291 Chrysopilus, 291 Chrysopoloma, 207 Chrysopolom dæ, 207 Chrysops, 288 Chrysosoma, 303 Chrysosomatinæ, 303 Chrysotimus, 304 Chrysotoxinæ, 308 Chrysotoxum, 308 Chrysotus, 304 Chrysotypidæ, 206 Chrysotypus. 206 Chthoniidæ, 548 Chthonius, 548 Chyliza, **339**, 340 Chylizinæ, 340 Chymomyza, 345 Chyphotes, 499, 501

Chyromyiidæ, 336

Cicada, 107, 108

Cicadellidæ, 111

Cicadella, 111

Cicadidæ, 108

Cicadula, 114

Cicindela, 410

Cicindelidæ, 410 Cicinnus, 213 Ciidæ, 432 Cilix, 217 Cilliba, 568 Cimbex, 473 Cimbicidæ, 473, 512 Cimex, 143, 144 Cimicidæ, 143 Cimicoidea, 143 Cinara, 124 Cinglis, 210 Cinxia, 309 Cinxiinæ, 309 Cioidæ, 432 Cirphis, 218 Cirsia, 63 Cis, 432 Cisidæ, 432 Cissia, 238 Cissites, 439 Cistela, 435 Cistelidæ, 435 Cisthene, 219 Cithæron, 560 Citharomantis, 91 Citheronia, 211 Citheroniidæ, 211, 251,252 Cuilfina, 88 Cixiidæ, 120 Cixius, 113, 120 Cladiinæ, 513 Cladius, 473 Cladochæta, 345 Cladura, 268 Cladypha, 120 Clambidæ, 431 Clasiopa, 343 Clastes, 562 Clastoptera, **110**, 111 Clastopteridæ, 111 Claviger, 415 Clavigeridæ, 415 Clavimyia, 286, 288 Cleandrus, 51 Cledeobia, 215 Cleidogona, 592 Cleis, 221 Cleitamia, 331 Cleonistria, 64 Cleonotus, 451 Cleonymidæ, 486, 488 Cleonymus, 486 Cleptes, 494 Cleptidæ, 494 Cleridæ, 427, 428

Clerus, 427 Climacia, 185 Climaciella, 183 Clinidium, 411 Clinocentrus, 479 Clinocera, 301 Clinoceratinæ, 301 Clinocoridæ, 143 Cliomantis, 84 Cliopeza, 325 Clisiocampa, 221 Clistogastra, 472, 510 Clitellaria, 286 Clitellariinæ, 286 Clitumninæ, 64 Clitumnus, 64 Cloeon, 170 Clonia, 52 Clothoda, 92 Clubiona, 563 Clubionidæ, 563 Clunio, 282, 348, 349 Clunioninæ, 282 Clusia, 334, **335** Clusiidæ, 323, 334, 341 Clusiinæ, 334 Clusiodes, 334 Clydonopteron, 215 Clypeasteridæ, 431 Clythia, 310 Clythiidæ, 310 Clytiomyia, 318 Clytra, 443 Clytridæ, 443 Cnaphalodes, 126 Cnemidoptes, 574 Cnemidotus, 410 Cnemodon, 308 Cnephasia, 226 Cnephia, 281 Cnetha, 281 Cnethocampa, 211 Cobboldia, 313, **315** Cobboldiinæ, 313 Coboldia, 349 Coccidæ, 133 Coccinella, **433**, 434 Coccinellidæ, 434 Coccoidea, 121 Coccomptilus, 130 Coccophagus, 484 Coccus, 133 Cochlidiidæ, 207 Cochliomyia, 314 Cocoblatta, 80 Cocytia, 219

Cocvtiidæ, 219 Codionus, 296 Cœlambus, 411 Cœlidia, 114 Cœlididæ, 114 Celometopia, **330**, 336 Cœlomyia, 320 Cœlopa, 327 Cœlopidæ, 323, 327, 351 Cœlopinæ, 327 Cœlostomidia, 128 Cœlostomidiinæ, 128 Cœlotes, 560 Cœnagriidæ, 161, 165 Cœnagriodea, 161 Cœnagrion, 161 Cœnagrionidæ, 161 Cœnagrionoidea, 161 Cœnia, 343 Cœnobius, 443 Cœnomyia, 283, 289, 363, 375Cœnomyiidæ, 289, 360, 375 Cœnomyiinæ, 289 Cœnonympha, 238 Cœnosia, 320, 351 Cœnosiinæ, 320 Cœnura, 288 Cœnurinæ, 288 Colabris, 301 Colænis, 238 Colaspis, 443 Colasposoma, 444 Coleophora, 230, 232 240, Coleophoridæ, 232, 243 Coleoptera, 408 Coleopterophagus, 574 Coleoptratidæ, 73 Coleorrhyncha, 107 Colias, 236 Collaria, 600 Collembola, 44 Colletes, 507 Colletidæ, 507 Collops, 429 Colobaspis, 443 Colobathristes, 148 Colobathristidæ, 148 Colobodesmus, 590 Colobognatha, 587 Coloboneura, 301 Colobopterus, 187 Colocasia, 218 Colon, 416 Colpocephalum, 103

Colpodia, 275 Colydiidæ, 432, 435 Colydium, 432 Colymbetinæ, 412 Commophila, 227 Commophilidæ, 227 Commoptera, 350 Compsodes, 81 Compsomantinæ, 90 Compsomantis, 90 Compsomyia, 314 Compsothespinæ, 84 Compsothespis, 84, 88 Comptosia, 295 Conchaspididæ, 131 Conchaspis, 131, **133** Conchylidæ, 227 Conchylis, 227 Coniceps, 336 Conicera, 305 Coniocompsa, 182 Coniopterygidæ, 182 Coniopteryginæ, 182 Coniopterygoidea, 182 Coniopteryx, 182 Conistra, 218 Conmachærota, 109 Conocephalinæ, 52 Conocephalus, 49, 52 Conophorinæ, 296 Conophorus, 296 Conopidæ, 306, 324, 366 Conopinæ, 307 Conops, 307, **307**, **365**, 366 Conorhinus, 147 Conostigmus, 494 Conosyrphus, 309 Conotyla, 593 Conotylidæ, 593 Contarinia, 275 Conwentzia, 182 Copeognatha, 96 Copestylum, **307**, 309 Cophura, 298 Copidosoma, 484 Copophora, 52 Copophorinæ, 52 Copridæ, 450 Copris, 450 Coptocycla, 444 Coptodisca, 229 Coptosoma, 150 Coptosomidæ, 150 Coptotriche, 230 Coquillettia, 294

Cordax, 70 Cordulegaster, 163, 164, Cordulegastridæ, 164, 167 Cordulia, 164 Corduliidæ, 164, 167 Cordyla, 278 Cordylobia, 321 Cordylura, **315**, 319 Cordyluridæ, 319, 333 Cordylurinæ, 319 Coreidæ, 148 Coreoidea 148 Corethra, 272 Corethrinæ, 272 Coriarachne, 562 Corimelænæ, 150 Corimelænidæ, 150 Corinna, 563 Coriscidæ, 148 Coriscus, 147 Corixa, **151**, 152 Corixidæ, 152 Corizidæ, 148 Corizoneura, 288 Corizus, **146**, 148 Cornetidæ, 426 Corphyra, 439 Corrodentia, 96 Corsomyza, 297 Corticaria, 434 Corticariinæ, 435 Corticoris, 145 Corydalidæ, 179, 180 Corydalis, 179, 180 Corydia, 82 Corydiidæ, 82 Corylophidæ, 431 Corylophus, 431 Corymbites, 416 Corynephoria, 46 Corynephoridæ, 46 Corynetes, **416**, 426 Corynetidæ, 428 Corynodes, 444 Corynoneura, 282 Corynorhynchus, 55 Corynoscelinæ, 276 Corynoscelis, 276 Corynothripoides, 73 Corypherepsis, 588 Corythucha, 142, **144** Coscinoptera, 443 Cosmetidæ, 553 Cosmopepla, 151 Cosmophorus, 481

Cosmopterygidæ, 232,233, 246Cosmopteryx, 233 Cosmoscarta, 111 Cosmosoma, 219Cossidæ, 206, 243, 246 Cossoninæ, 449 Cossus, 206 Cossyphodes, 436 Cossyphodidæ, 436 Cossyphodinus, 436 Cossyphodites, 436 Cotalpa, 452 Cotinus, 452 Crabro, 506 Crabronidæ, 506 Crambinæ, 215, 216 Crambus, 216 Cramptonomyia, 273 Cranogona, 591 Cranopygia, 68 Cranothrips, 73 Craspedosoma, 592 Craspedosomatidæ, 592 Cratærrhina, 347 Craterostigmidæ, 600 Craterostigmorphidea,600 Craterostigmus, 600 Craticulina, 316 Cratomelus, 50 Cratyna, 278 Cratyninæ, 278 Creagris, 187 Cremastochilus, 452 Cremnops, 480 Crepidohamma, 332 Creobroter, 91 Creobrotinæ, 91 Crescentius, 143 Cricotopus, 282 Crioceridæ, 442 Criocerus, 442, **442** Criorrhina, 309, **377** Croce, 183 Crocidium, 296 Crocisa, 509 Crocothemis, 164 Crosbycus, 554 Crossotarsus, 445 Crustacea, 534 Crustulina, 559 Cryptarcha, 419 Cryptobethylus, 492 Cryptocellus, 536, 543 Cryptocephalidæ, 443 Cryptocephalus, 443

Cryptocerata, 141 Cryptocercidæ, 79 Cryptocercus, 79, 80 Cryptochætidæ, 342, 366 Cryptochætum, 339, 342, 366. **368** Cryptocorypha, 588 Cryptodesmidæ, 588 Cryptodesmus, 588 Cryptogenius, 450 Cryptognathidæ, 576 Cryptognathus, 576 Cryptokermes, 128 Cryptoparlatoria, 131 Cryptophagidæ, 424, 437 Cryptophagus, 424 Cryptophasa, 228 Cryptophasidæ, 228 Cryptopidæ, 600 Cryptops, 600, 601 Cryptorhopalum, 425 Cryptosiphum, 125 Cryptostemma, 543 143. Cryptostemmatidæ, 543 Cryptotermes, 94 Cryptothele, 560 Crypturgus, 446 Cryptus, 478 Ctenidæ, 563 Cteniopus, 435 Ctenisolabis, 69 Cteniza, 556 Ctenizidæ, 556 Ctenocephalides, 405, 406 Ctenocephalus, 405 Ctenophora, 268 Ctenophorinæ, 268 Ctenophthalmidæ, 405 Ctenophthalmus, 405 Ctenophyllus, 406 Ctenopsyllidæ, 405 Ctenorya, 602 Ctenostylidæ, 322 Ctenostylum, 322 Ctenota, 299 Ctenucha, 219 Ctenus, 563 Cubaris, 534 Cubobolus, 596 Cucujidæ, 424, 438 Cucujus, 424 Cucullia, 218 Culex, 272, **273**, 354, **528** Culicidæ, 272, 354, 373 Culicinæ, 272

Culicoides, 282 Culiseta, 272 Cummingsia, 102 Cunaxa, 576 Cunaxidæ, 576 Cuniculina, 64 Cupes, 422 Cupidæ, 422 Cupidinidæ, 239 Cupipes, 600 Curculoididæ, 543 Curculionidæ, 447 Curculioninæ, 449 Curculionoidea, 444 Cursoria, 77 Curu, 148 Curupira, 270 Cuterebra, 313, **315**, 366, Cuterebridæ, 313 Cuterebrinæ, 313 Cyamops, 331 Cyathocerus, 435 Cvathoceridæ, 435 Cyaniris, 443 Cybæus, 560 Cybister, 411, **412** Cybistrinæ, 412 Cybocephalini, 431 Cybocephalus, 431 Cycloberotha, 186 Cyclodesmus, 590 Cyclopididæ, 236 Cyclopides, 236 Cyclopodia, 346, 352 Cycloptilum, 54 Cyclorhabdus, 589 Cyclorrhapha, 304, 353, 364Cyclosia, 207 Cyclotorna, 225 Cyclotornidæ, 225 Cycnodia, 229 Cycnodiidæ, 230 Cydia, 226 Cydnidæ, 151 Cydnus, 151 Cyladidæ, 446 Cylas, **414**, 446 Cyliosoma, 586 Cylindracheta, 53, **53** Cylindrachetidæ, 53 Cylindrococcidæ, 134 Cylindrococcus, 134 Cylindrosella, 433 Cylindrotoma, 266, 268

Cylindrotomidæ, 268 Cyllene, 441, **528** Cyllenia, 296 Cylleniinæ, 296 Cymatodera, 427 Cymatophora, 211 Cymatophoridæ, 211 Cymatopsocus, 98 Cynipidæ, 483, 488, 514 Cynipimorpha, 285, 285 Cynipoidea, 482 Cynomyia, 316 Cynorrhina, 309 Cynorta, 553 Cynortula, 553 Cyphocallipus, 593 Cyphomyia, 286 Cyphon, 418 Cyphonidæ 418 Cyphophthalmi, 551 Cypsela, 337 Cypselidæ, 337 Cypselosoma, 338 Cyrpoptus, 119 Cyrtacanthacrinæ, 57 Cyrtacanthacrus, 57 Cyrtaspis, 51 Cyrtidæ, 292 Cyrtinæ, 292 Cyrtomenus, 151 Cyrtomorpha 296 Cyrtomorphus, 432 Cyrtonotinæ, 344 Cyrtonotum, **342**, 344 Cyrtopogon, 298 Cyrtosia, 296 Cyrtosiinæ, 296 Cyrtosternum, 145 Cyrtoxiphus, 54 Cyrtus 292 Cystococcus, 132 Cystocœlia, 56 Cystogaster, 318 Cyta, 576 Cytherea, 295 Cythereinæ, 295 Cytilus, 417 Cytodotidæ, 574 Cytoleichidæ, 574 Cytoleichus, 574 Dacinae, 341 Dacne, 424 Dacnusa, 477

Dactylispa, 444

Dactylopius, 134

Dactylopiinæ, 134 Dactylopteryx, 89 Dacus, 341, **342** Dæsia, 547 Dahlica, **348**, 349 Dalaca, 201 Dalcera, 206 Dalceridæ 206 Dalcerides, 206 Dalmannia, 307, 307 Dalmanniinæ, 307 Damasippoides, 64 Damon, **541**, 542 Dampetrus, 552 Danaidæ, 238, 253 Danais, 237, **237** Darthula, 111 Dartia, 572 Dascillidæ, 418 Dascillus, 418 Dasycera, 229 Dasycerinæ, 434 Dasycerus, 416 Dascyllidæ, 418 Dasyllis, 299, 375, 376. Dasymetopa, 331 Dasymutilla, 500 Dasyneura, 275 Dasyphlebomyia, 341 Dasyphora, 321 Dasypogon, 298 Dasypogoninæ, 298, 299 Dasysciara, 349 Dasytes, 429 Dasythrix, 299 Dasytidæ, 427, 429 Datames, 63 Datana, 211 Decatoma, 486 Decidia, 64 Decimia, 87 Decticinæ, 52 Decticus, 52 Degeeriella, 103 Deilephila, 208 Deiphobe, 86 Deipnopsocus, 98 Delena, 562 Delphacidæ, 119 Delphacixenos, 470 Delphax, 119 Delphinia, 331 Deltocephalus, 113, 114 Deltochilum, 450 Demacrida, 50

Demodex, 566, 566 Demodicidæ, 566 Demodicoidea, 565 Dendroiketes, 66 Dendroctonus, 416, 445, 445Dendroides, **416**, 439 Dendrolasma, 554 Dendroleon, 187 Dendromonomeron, 592 Dendryphantes, 561 Dendrosoter, 479 Depanidæ, 212 Depressaria, 229, 247 Depressariidæ, 229 Deraiophorus, 568  $\overline{\text{Derallimus}}, 52$ Derancistrus, 441 Derbe, 119 Derbidæ, 119 Dermacentor, 567 Dermanyssidæ, 568 Dermanyssus, 568 Dermaptera, 65 Dermatophilidæ, 407 Dermatophilus, 407 Dermestes, **422**, 425 Dermestidæ, 425 Dermestinæ, 425 Dermodermaptera, 71 Derodontidæ, 425 Derodontus, 425 Dermatobia, 313 Dermatobiinæ, 313 Deromantis, 89 Deromyia, 298 Deronectes, 411 Deropeltis, 78 Deroplatinæ, 89 Deroplatys, 85, 89 Desmatoneura, 295 Desmia, 216 Desmometopa, 342, 345 Desmonus, 590 Desmoptera, 57 Desmothrips, 73 Deuterophlebia, 269, 269, **370**. 371 269, Deuterophlebiidæ, 354, 371Devadatta, 162 Devillea, 589 Dexia, 318

Dexiidæ, 318

Diabantia, 87

Diabrotica, **442**, 444

Diachasma, 481 Diachlorinæ, 287 Diachlorus, 287 Diachorisia, 222 Diacrisia, 219 Diadasia, 508 Diadocidia, 279, **279** Diadocidiidæ, 279 Diagryphodes, 424 Dialeurodicus, 124 Dialineura, 294 Dialysis, 290 Dialyta, 320 Diamesa, 282 Diamesinæ, 282 Diamma, 499 Diamorus, 485 Diamusonia, 87 Diaperasticidæ, 70 Diaperasticus, 70 Diaperidæ, 436 Diaperis, 436 Diaphania, 216 Diapheromera, 64 Diaphorinæ, 304 Diaphorus, 304 Diaporus, **594**, 596 Diapriidæ, 491, 500 Diardia, 64 Diarthronomyia, 275 Diaspididæ, 129 Diaspidinæ, 130 Diaspis, **129**, 130 Diastata, 335, **342**, 344 Diastatidæ, 344 Diastatinæ, 344 Diastephanus, 476 Diastrophus, 483 Diathoneura, 345 Diatræa, 216 Diazosma, 267 Dibrachys, 484, **484** Dicallaneura, 239 Dicellophilus, 602 Dichæta, 343 Dichelocera, 288 Dichomeris, 227, **241** Dichomeridæ, 228 Dichoptera, 120 Dichronychidæ, 422 Dichronychus, 421 Dicrana, 68 Dicraneura, 113 Dicranolasma, 554 Dicranomyia, 268 Dicranoptycha, 269

Dicranota, 268, 354, 355 Dicterias, 163 Dictya, 328 Dictyna, 558 Dictynidæ, 558 Dictyophara, 119 Dictyopharidæ, 120 Dictyopsocus, 99 Dictyoptera, 181 Dicymolomia, 216, **241** Didea, 309, **377** Didymochæta, 334 Didymocorypha, 87 Diesingia, 581 Dieta, 562 Diglochis, 484 Diguetia, 559 Dilar, **183**, 184 Dilaridæ, 184 Dilophus, 275 Dilta, 42 Dimorpha, 503 Dimorphidæ, 503 Dimorphodes, 64 Dimorphothynnus, 499 Dina, 63 Dimera, 97 Dinapsidæ, 476 Dinapsis, 476 Dinelenchus, 470 Dinematocricus, 596 Dinetus, 505 Dineutes, 412, **412** Dinex, 70Dinocampus, 481 Dinopidæ, 558 Dinopis, 558 Dinorhax, 547 Dinotoperla, 177 Dinurothrips, 74 Dinychus, 568 Dioctria, 298 Diomonus, 277 Dione, 238 Dionycha, 557 Diopsidæ, 329 Dopisis, **328**, 329 Diopsiulus, 593 Dioptidæ, 207, 210, 249 Dioryctria, 215 Dioryctus, 443 Diospilinæ, 481 Diospilus, 481 Diozocera, 470 Diozoceratidæ, 470 Dipalta, 294

Diphadnus, 474 Diphlebia, 162 Diphleps, 145 Diphyllidæ, 424 Diphyllus, 424 Diplatyidæ, 66 Diplatys, 66 Diplectrona, 194 Diplocentra, 344 Diplocentridæ, 544 Diplocentrus, 544 Diplocelus, 424 Diplodontus, 571 Diploglena, 557 Diploglossata, 71 Diplomaragna, 593 Diplomaragnidæ, 593 Diploneura, 305 Diplopectron, 503 Diplopoda, 585 Diploptera, 79, 80 Diplopteridæ, 79 Diplosara, 233 Diplosaridæ, 233 Diplosis, 275 Diplosphyronida, 548 Diplotaxis, 451 Diplothele, 556 Diplura, 43, 556 Dipluridæ, 556 Diprion, 474 Diprionidæ, 474, 511, 513 Dipsocoridæ, 143 Dipsocoroidea, 143 Diptera, 264 Dircenna, 238 Dirrhinus, 485 Discobola, 268 Discocerina, 343 Discocyrtus, 553 Discoloma, 434 Discolomidæ, 434 Discomyza, 343 Discopoma, 568 Disholcaspis, 483 Disophrys, 480 Dissosteira, 55, 56 Ditænia, 328 Ditaxis, 183 Ditha, 548 Ditomyia, 279, 280 Ditomyiidæ, 280, 358 Dixa, 273, **273**, 356, 373 Dixidæ, 273, 356, 373 Dixippus, 64 Docosia, 278

Doderia, 587 Dodona, 239 Dolerus, 473, 474 Dolichocephala, 301 Dolichoderinæ, 489 Dolichoderus, 489 Dolichomyia, 296 Dolichopeza, 267, 268 Dolichopezinæ, 268 Dolichopoda, 50 Dolichopodidæ, 302, 350, 362, 377Dolichopodinæ, 303 Dolichopsylla, 406 Dolichopsyllidæ, 406 Dolichopus, 299, 303, 361, 362Dolichurus, 503 Doliomalus, 561 Dolistenus, 597 Dolomedes, 561 Domomyza, 339 Donacia, 441 Donaciidæ, 441 Donaconethis, 92, 92 Donusa, 64 Doratodesmus, 588 Dorcadion, 441 Dorcatoma, 416 Dorceus, 558 Dorcus, 450 Dores, 63 Doros, 309 Doru, 67, 70 Dorycallipus, 593 Dorycera, 331 Doryceridæ, 331 Doryctes, 479 Doryctinæ, 479 Dorylaidæ, 310 Dorylas, 310 Dorylinæ, 490 Dorylus, 490 Dorymyrmex, 489 Dorypetalidæ, 593 Dorypetalum, 593 Doubledaya, 432 Douglasia, 230 Douglasiidæ, 230 Doydirhynchidæ, 449 Dræculacephala, 111 Drapetes, 421 Drapetis, **299**, 300, 362, 363 Drapetisca, 560 Drassidæ, 562

Drassodes, 562 Drassus, 562 Drasterius, 421 Drepana, 217 Drepanepteryx, 184 Drepanicus, 183 Drepanidæ, 217, 221, 248 Drepanulidæ, 217 Dreyfusia, 126 Drilidæ, 431, 453 Drilocephalus, 431 Drilus, 431 Dromæolus, 422 Drosicha, 128 Drosophila, **342**, 345, 350, Drosophilidæ, 344, 350, 368 Drosophilinæ, 345 Drunella, 172 Dryas, 238 Dryinidæ, 493, 501, 502 Dryinopsis, 500 Dryinus, 493 Drymodromia, 301 Drymonia, 211 Drymusa, 559 Dryomyza, 327 Dryomyzidæ, 327, 351 Dryomyzinæ, 327 Dryoperia, 232 Dryopidæ, 418 Dryops, 418 Dyschirius, 410 Dyscoletes, 481 Dysdera, 557 Dysdercus, 149, **149** Dysderidæ, 557 Dysmachus, 298 Dyme, 64 Dynastes, 452 Dynastidæ, 452 Dynatosoma, 278 Dysodia, 214 Dysodiidæ, 145 Dysphæa, 162 Dyspteris, 210 Dystacta, 87 Dystactinæ, 87, 90 Dytiscidæ, 411 Dytiscinæ, 412 Dytiscus, 411, 412, 528 Duliticola, 453 Duomyia, 331 Duralimnesia, 571

Dusmetina, 350 Duvita, 227 Dziedzickia, 277

Eacles, 211 Earias, 219, **220** Earinus, 480 Eatoniana, 576 Eatonica, 170 Eburia, 441 Ecacanthothripidæ, 76 Ecacanthothrips, 76 Eccoptogaster, 446 Eccoptomera, 335 Ecdyonuridæ, 172 Ecdyonurus, 172 Echestypus, 347 Echidnophaga, 407 Echidnophagidæ, 407 Echinischodea, 583 Echiniscidæ, 583 Echiniscoides, 583 Echiniscus, 583 Echinolælaps, 568 Echinomegistus, 568 Echinopsalis, 68 Echinophthirius, 104 Echinophthiriidæ, 104 Echinopsocus, 98 Echinosoma, 68 Echinosomatidæ, 68 Echmepteryx, 98 Echmostigmus, 600 Ecitomyia, 306, 350 Eciton, 490, **491** Eclimus, 296 Ecphylus, 479 Ectadoderus, 54 Ectatops, 150 Ectatosticta, 557 Ectecephala, 340 Ectemnia, 281 Ectemniinæ, 281 Ectobia, 79 Ectobiidæ, 79 Ectœdemia, 221, **223** Ectognatha, 41 Ectotrophi, 41 Ectrephes, **422**, 423 Ectrephidæ, 423 Ectropa, 207 Ectropidæ 207 Edapteryx, 217 Edwardsellum, 281 Edwardsina, **269**, 270 Edwardsinæ, 270

Eginia, 321 Eginiinæ, 321 Egle, 319 Egropa, 117 Elacatidæ, 435 Elacatis, 435 Elachertidæ, 486 Elachiptera, 340 Elachista, 229, 230 Elachistidæ, 230 Elæa, 87 Elaphidion, **440**, 441 Elaphogonus, 590 Elaphomyia, 332 Elaphroptera, 499 Elasmidæ, 486 Elasmoscelis, 121 Elasmosoma, 480 Elasmus, **475**, 486 Elassogaster, 331 Elassoneuria, 170 Elater, 421 Elateridæ, 422 Elatobia, 224 Eleleis, 562 Elenchidæ, 470 Elenchoidea, 470 Elenchoides, 470 Elenchus, 470 Eleodes, 436, **437** Eleuterata, 408 Elgiva, 328 Elidoptera, 119 Elipsocus, 99 Elis, 498, **498**, 499 Ellampus, 493 Ellipoptera, 104 Ellipsidion, 79 Elliptoblatta, 82 Elophila, 216 Elythroptera, 408 Embia, 92 Embidopsocus, 98 Embidotroctes, 97, 98 Embiidæ, 92 Embiidina, 91 Embiodea, 91 Embioidea, 91 Embioptera, 91 Embolemidæ, 492, 500 Embolemus, 492 Emesa, 147 Emesidæ, 147 Emmenognatha, 180 Emperoptera, 350, 351 Empheriidæ, 98

Emphor, 508 Emphytinæ, 512 Emphytus, 474 Empididæ, 297, 300, 309, 362Empidideicus, 296 Empidinæ, 301 Empis, 301, **302** Empoa, 113 Empoasca, 114 Empusa, 84, 88 Empusinæ, 84 Enallagma, 161 Enalliapyx, 43 Enchenopa, 108 Encyrtidæ, 484, 488 Encyrtothynnus, 499 Encyrtus, 484 Endacusta, 54 Enderleinia, 109 Enderleiniinæ, 109 Endomia, 439 Endomychidæ, 434 Endomychus, 434 Endotrophi, 43 Endromidæ, 220 Endromididæ, 220 Endromis, 220 Endrosis, 232 Eneoptera, 54 Eneopterinæ, 54 Engycystis, 499 Engyophlebidæ, 204 Engyophlebus, 204 Enicita, 323 Enicmus, 344 Enicocephalidæ, 147 Enicophlæbia, 88 Enkrates, 69 Ennearthron, 432 Ennomos, 209 Enochrus, 414 Enodia, 238 Ensina, 341 Entedontidæ, 486 Entognatha, 43 Entomobrya, 46 Entomobryidæ, 46 Entylia, **107**, 108 Eoctenes, 154 Eomantis, 88 Eoperipatinæ, 533 Eoperipatus, 533 Eosentomidæ, 40 Eosentomon, 40 Epacmus, 295, **295** 

Epallagidæ, 162, 165 Epanerchodus, 588 Epaphrodita, 87 Epaphroditinæ, 87, 91 Eparchus, 70 Epargyreus, **234**, 235 Eparmene, 119 Epectris, 557 Epeira, 560 Epeiridæ, 560 Epeolus, **475**, 509 Epeorus, 172Epermenia, 231 Epermeniidæ, 231 Epexochlænoides 485 Ephedrus, 482 Ephemera, 170 Ephemerella, 171, 172,174 Ephemerellidæ, 172, 175 Ephemerida, 169 Ephemeridæ, 170, 173 Ephemeroidea, 169 Ephemeroptera, 169 Ephestia, 215, 217 Ephippiger, 53 Ephippigerinæ, 53 Ephippium, 286 Ephutomorpha, 500 Ephydra, 343, **368** Ephydridæ, 343, 350, 368 Ephydrinæ, 343 Ephygrobia, 343 Epibates, 296 Epiblemidæ, 226 Epicarsa, 123 Epicauta, **437**, 439 Epicimelia, 211 Epicnaptera, **220**, 221 Epicnopterygidæ, 208 Epicoccus, 133 Epicopeia, 212, 213 Epicopeiidæ, 213 Epidapus, 278, 349 Epierus, **416** Epilachna, **433**, 434 Epilampra, 79 Epilampridæ, 79 Epimarptidæ, 233 Epimarptis, 233 Epimartyria 200 Epimorpha, 599 Epinannolene, 596 Epinannolenidæ, 596 Epinotia, 226 Epiophlebia, 160 Epiophlebiidæ, 160, 165.

Epipaschia, 216 Epipaschiinæ, 216 Epiperipatus, 533 Epiphlœus, 426 Epiphragma, 268 Epiplatea, 336 Epiplema, 210 Epiplemidæ, 210, 251 Epipocus, 434 Epipsocus, 99 Epipsylla, 123 Epipyropidæ, 205, 249 Epipyrops, 205 Episcapha, 424 Epistenia, 486 Epitragus, 436 Epitrimerus, 565 Epochra, 341 Epomyia, 294 Eproboscidea, 311 Epuræa, 419 Epyris, 494 Equitidæ, 236 Erannis, 209 Erasmia, 207 Erastria, 218 Erax, 298, 298 Erechthiidæ, 223 Eremazus, 451 Eremiaphila, 87 Eremiaphilinæ, 86, 87 Eremobates, 547, **547** Eremocneminæ, 298 Eremopsocus, 99 Eremus, 53 Eresidæ, 558 Eresus, 558 Eretmoptera, 349 Eretmopteridæ, 349 Ereynetes, 576 Erginulus, 553 Erianthus, 56 Erineophilus, 446 Erinna, 289 Erinnidæ, 289 Eriocampoides, 474 Eriocephala, 200 Eriocephalidæ, 200 Eriocera, 268 Eriococcidæ, 134 Eriococcus, 134 Eriocrania, 200 Eriocraniidæ, 200, 242 Eriogaster, 221 Eriogone, 560 Eriophyes, 565, **566** 

Eriophyidæ, 565 Erioptera, 268 Eriopterinæ, 268 Eriosoma, 126 Eriosomatidæ, 126 Eriosomatinæ, 126 Eriozona, 309 Eristalinæ, 309 Eristalis, 307, 309, 365. 365Ernestia, 318 Ernobius, 426 Ero, 559 Erotylidæ, 424, 432 Erotylus, 432 Erpetogomphus, 164 Erycina, 239 Erycinidæ, 239 Erynninæ, 236 Erynnis, 235 Erythræidæ, 576 Erythræus, 576 Erythroneura, 114 Escaryus, 602 Eschatocerus, 52 Esenbeckia, 289 Esphalmenidæ, 68 Esphalmenus, 67, 68 Esthiopterum, 103 Estigmene, 219 Ethelum, 534 Ethmia, 229 Ethmiidæ, 229, 247, 250 Euacanthidæ, 117 Euacanthus, **115**, 117 Euaresta, 341 Eubadizon, 481 Eubelidæ, 534 Eubelum, 534 Eublemma, 218 Euborellia, 68 Eubrianax, 418 Eucampsipoda, 352 Eucarlia, 595 Eucentrobolus, 595 Eucera, 508 Eucerceris, **504**, 505 Eucessia, 295 Eucestia, 210 Eucestidæ, 232 Eucharididæ, 485 Euchera, 217, **217** Euchiridæ, 452 Euchirus, 452 Euchloris, 210 Euchomenella, 86

Euchromia, 219, 220 Euchromiidæ, 219, 250 Eucinetidæ, 418 Eucinetus, 418 Euclea, 207 Eucleidæ, 207, 241 Euclemensia, 231 Eucles, 64 Euclimacia, 183 Eucnemidæ, 422 Eucnemis, 422 Eucocytiidæ, 219 Eucoila, 483 Euconnus, 428 Euconocephalus, 52 Eucorethra, 272, 273 Eucoryphus, 304 Eucosma, 226 Eucosmidæ, 226 Eucratonyx, 602 Eucymatoge, 210 Eudamidæ, 236 Eudamus, 235 Eudarcia, 222 Eudermaptera, 66 Eudesmia, 219 Eudiospilus, 481 Eudmeta, 285 Eudohrnia, 70 Eudohrniidæ, 70 Eucididæ, 238, 253 Euemenophorus, 556 Eufernaldia, 215 Eugaster, 53 Euglenes, 439 Euglenidæ, 439 Euglossa, 508 Euglossidæ, 508 Eugnatha, 587 Eugonia, 209, 238 Euhybos, 302, **302** Eulachnus, 124 Eulalia, 286 Eulema, 508 Eulia, 226 Eulonchus, 292 Eulophidæ, 486, 488 Eulophus, 484, 486 Eumacrocentrus, 480 Eumastacinæ, 56 Eumastax, 56 Eumecacis, 284 Eumenes, 497, **498** Eumeninæ, 497 Eumenotes, 145 Eumerinæ, 309

Eumerus, 309 Eumolpidæ, 444 Eumyiidæ, 311 Euoxypilus, 86 Eupalus, 576 Euparagia, 496 Euparagiinæ, 496 Euparia, 451 Euparyphus, 286 Eupathocera, 470 Eupatithripidæ, 76 Eupatithrips, 76 Eupatra, 571 Eupelmidæ, 484 Eupelmus, 484 Euphalarus, 123 Euphoria, 452 Euphorinæ ,481 Euphorus, 481 Euphydryas, **234**, 238 Euphyllura, 123 Euphyonartex, 117 Eupines, 415 Eupistidæ, 232 Eupithecia, 210 Euplectrus, 486 Euplectus, 415 Euplecidæ, 238 Eupodes, 576 Eupodidæ, 576 Eupodoidea, 574 Euporismus, 185 Euprepia, 219 Euproctis, 218 Euprosopia, 331 Eupsalis, 447 Eupterote, 211Eupterotidæ, 211, 251 Eupterygidæ, 114 Euptoieta, 238 Eurema, 236 Eurhinocricus, 596 Euribia, 341 Euribiidæ, 340 Euricania, 121 Eurina, 340 Eurobius, 184 Eurosta, 341, **342** Eurrhypara, 216 Eurybata, 325 Eurybia, 239 Eurybrachidæ, 121 Eurybrachys, 121 Eurycantha, 64 Eurychoromyia, 322 Eurychoromyiinæ, 322

Eurycnema, 65 Eurycotis, 78 Eurycyttarus, 209 Eurygaster, 150 Euryischia, 486 Eurymela, 112 Eurymus, 236 Euryneura, 286 Euryophthalmus, 150 Eurypauropodidæ, 584 Eurypauropus, 584, 584 Eurypelma 556 Euryporyphes, 57 Eurystethidæ, 436 Eurystethus, 436 Eurytenes, 481 Eurytion, 603 Eurytoma, 486 Eurytomidæ, 486, 488,514 Euscaphurus, 418 Eusapyga, 500 Eusarcus, 553 Euscelidia, 298 Euscelis, 114 Euschemon, 203, 235 Euschemonidæ, 203, 235 Euschistus, 151, **151** Euscorpiinæ, 545 Euscorpius, 545 Euselasia, 239 Eusemia, 219 Eusiphona, 345 Eustalocerus, 481 Eusthenia, 176 Eustheniidæ, 176, 178 Eustrotia, 218 Eutardigrada, 582 Eutarsus, 304 Eutermes, 94 Eutettix, 114 Euthore, 162 Euthrips, 72 Euthycera, 328 Euthychæta, 344 Euthyneura, 301 Euthyphleps, 86 Euthyplocia, 169 Euthyrrhapha, 81 Euthyrrhaphidæ, 81 Euthysanius, 428 Eutreta, 341 Eutrichophilus, 103 Euvanessa, 238 Euxanthis, 227 Euxesta, 330, **330** Euxoa, 218

356,

Euzercon, 569 Evæsthetinæ, 430 Evagrus, 556 Evalces, 438 Evallogeophilus, 602 Evania, 476 Evaniidæ, 476 Evaniocera, 438 Evaniocerini, 438 Evasa, 285 Everes, 239 Evetria, 226 Evippa, 561 Ewingia, 572 Ewingiidæ, 572 Exallonyx, 492 Exechia, 278, 358, 359 Exeiridæ, 503 Exeirus, 503 Exema, 443 Exeuthyplocia, 170 Exoprosopa, 294 Exoprosopinæ, 295 Exorista, 318, **377** Exoristidæ, 318 Exothecinæ, 479 Exothecus, 479 Eylaidæ, 570 Eylais, 570

Fagina, 591 Faginidæ, 591 Falcaria, 217 Fallenia, 291 Fannia, 320 Fanniinæ, 320 Fasisuga, 131 Faventia, 119 Feaellidæ, 550 Fecenia, 558 Feltria, 572 Fenusa, 474 Fenusinæ, 513 Fessonia, 576 Fidena, 289 Fidia, **442**, 443 Figitidæ, 483, 488, 514 Filistata, 557 Filistatidæ, 557 Fioria, 597 Fiorinia, 130, **131** Fioriniinæ, 130 Fischeria, 86 Fischeriinæ, 86, 89 Flata, 120

Flatidæ, 120

Flatoides, 120 Flirtea, 553 Fontaria, 590 Fontariidæ, 590 282. Forcipomyia, 357 Forcipula, 68 Forda, 126 Fordinæ, 126 Forellia, 572 Forficula, **67**, 70 Forficulales, 69 Forficulidæ, 70 Forficulina, 65 Forficuloidea, 69 Formica, 489 Formicaleo, 187 Formicidæ, 489, 515 Formicinæ, 489 Formicoidea, 489 Formicomus, 439 Fornax, 422 Fracticipita, 404 Frankliniella, 74, 75 Franklinothripidæ, 73 Franklinothrips, 73, 75 Frenatæ, 200 Freyana, 574 Freysiula, 122 Friesia, 281 Frontina, 318 Frontipoda, 572 Fucellia, 319 Fucelliinæ, 319 Fulcidacidæ, 443 Fulciniella, 88 Fulgora, 119 Fulgoridæ, 119 Fulgoroidea, 108 Fumea, 207 Fungivoridæ, 277 Fungivorides, 278 Fustiger, 415

Gagrella, 553
Galeatus, 142
Galeodes, 536, 547
Galeodidæ, 547
Galepsus, 87
Galeruca, 444
Galerucella, 444
Galerucidæ, 491
Galloisiana, 47, 48
Galleria, 215
Gallerinæ, 215

Galgulidæ, 153 Galgulus, 153 Galumna, 570 Gamasomorpha, 557 Gamasus, 568 Ganonema, 194 Gardena, 147 Gargaphia, 142 Garibius, 600 Garrina, 602 Garypinus, 550 Garypodidæ, 550 Garypodoidea, 549 Garypus, 550 Gasteracantha, 560 Gasteruption, 476. 482 Gasteruptionidæ, 476 Gastropacha, 221 Gastrophilidæ, 322, 366 Gastrophilus, 315, 322, 366, **367** Gastropsis, 507 Gastrotheca, 478 Gastrotheus, 41 Gaurax, 340 Gayella, 496 Gayellinæ, 496 Gayenna, 563 Gayomyia, 184 Gekobia, 575 Gelanor, 559 Gelastocoridæ, 153 Gelastocoris, 153 Gelchossa, 142 Gelechia, 228, **528** Gelechiidæ, 228, 233, 241, 243, 246 Geocoridæ, 149 Geocoris, 149 Geocorisæ, 141 Geogarypus, 550 Geomantis, 86 Geometra, 209 Geometridæ, 209, 210, 248 Geometroidea, 240 Geomyza, 335, 339, **339** Geomyzidæ, 335, 339 Geophilella, 603 Geophilellidæ, 603 Geophilidæ, 603 Geophilomorpha, 600 Geophilus, 603 Georgella, 571 Georyssidæ, 432 Georyssus, 432

Geosarginæ, 286

Geosargus, **284**, 286 Geoscapheus, 80 Geotomus, 151 Geotrupes, 450 Geotrupidæ, 450 Gephyra, 215 Geranomyia, 268 Geron, 295, 296 Gerridæ, 152 Gerris, 151, **151** Gerroidea, 142 Gervaisiidæ, 587 Gervasia, 587 Gibbium, 427 Gigantodax, 281 Gigantothrips, 75 Gilletteella, 126 Gingla, 207 Giraffomyia, 332 Gitona, 345 Givira, 206 Glabellula, 296 Glabellulinæ, 296 Glaphyria, 216 Glaphyridæ, 451 Glaphyriinæ, 216 Glaphyrus, 451 Glaresis, 450 Glaucolepis, 221 Glaucopsyche, 239 Glenanthe, 343 Gliphypteryx, 227 Gliricola, **102**, 102 Glischrochilus, 418 Globotherium, 586 Gloma, 301 Glomerellina, 587 Glomerida, 587 Glomeridella, 587 Glomeridellidæ, 587 Glomeridesmidæ, 586 Glomeridesmus, 586 Glomeridia, 586 Glomeris, 587 Glossata, 198 Glossina, **299**, 321 Glossinidæ, 321 Glossosoma, 193 Glutops, 289 Glyciphagus, 574 Glyphidocera, 227 Glyphipterygidæ, 228, 247 Glyphipteryginæ, 228 Glyphipteryx, 228 Glyptocombus, 143 Glyptometopa, 499

Glyptomorpha, 478 Gmogala, 557 Gnaphiscus, 572 Gnaphosa, 562 Gnomeskelus, 589 Gnomulus, 552 Gnophomyia, 268 Gnorimus, 452 Gnoriste, 278 Gnorimoschema, 227, 228 Gnosippus, 547, **547** Gnostidæ, 417 Gnostus, 417 Goeldia, 272 Goliathus, **437**, 452 Gomophomastax, 56 Gomphidæ, 164, 166 Gomphocerus, 56 Gomphodesmidæ, 590 Gomphodesmus, 590 Gomphomastacinæ, 56 Gomphus, 164 Gonatista, 89 Gonatistella, 89 Gonatocerus, 483 Gonatopus, 493, **495** Gongrozus, 286 Gonia, **299**, 318 Goniastes, 418 Gonibregmatidæ, 602 Gonibregmatus, 602 Goniocotes, 103 Goniodes, 103 Goniodoma, 232 Goniops, 288 Goniosoma, 553 Goniozus, 494 Gonolabidura, 68 Gonolabina, 68 Gonolabis, 68 Gonophora, 444 Gonvleptes, 553 Gonyleptidæ, **552**, 553 Gonypeta, 90 Gonypetella, 87 Gorgora, 108 Gorytes, 503, **504** Gorytidæ, 503 Gossyparia, 134 Gracilaria, 232 Gracilariidæ, 232, 242 Grallipeza, 325 Grammodes, 218 Grammostola, 556 Graphocephala, 111 Graphomyia, 321

Graphosomatidæ, 150 Grapta, 238 Graptolithidæ, 226 Graptostethus, 149 Gratidia, 64 Gressoria, 61 Gripopterygidæ, 176, 177 Gryllacridæ, 53 Gryllacris, 49, 53 Gryllidæ, 54 Grylloblatta, **47**, 48 Grylloblattidæ, 48 Grylloblattodea, 47 Grylloblattoidea, 47 Gryllodes, 54 Gryllotalpa, **53**, 54 Gryllotalpidæ, 54 Gryllinæ, 54 Gryllus, **51**, 54 Gryocraspedum, 295 Gryon, 494 Gularostria, 107 Gylippus, 547 Gymnaspis, 131 Gymnocerata, 141 Gymnomyza, 323 Gymnopa, 343 Gymnophora, 306 Gymnopleurus, 450 Gymnopinæ, 343 Gymnoscelus, 480 Gymnosoma, 318 Gymnosomatidæ, 318 Gymnostreptus, 594, 596 Gyna, 82 Gynacantha, 164 Gynandrophthalma, 443 Gypona, **107**, 112 Gyponidæ, 112 Gypsonoma, 226 Gyretes, 412 Gyrinidæ, 413 Gyrinoidea, 413 Gyrinus, **412**, 412 Gyroneuron, 479 Gyropidæ, 102 Gyrophallus, 588 Gyropus, 102, **102** Gyrostoma, 497 Haania, 84

Haasea, 592

Habrobracon, 478

Habrocerinæ, 429

Habrodesmus, 589

Habrophlebia, 172, **174** 

Habropogon, 298 Habrosyne, 211 Hadena, 218 Hades, 239 Hadruroides, 546 Hadogenes, 545 Hadrobregmus, 426 Hadronotus, 494 Hadrotarsidæ, 557 Hadrotarsus, 557 Hadrurus, 546 Hæmaphysalis, 567 Hæmatobia, 321 Hæmatobosca, 321 Hæmatomyzidæ, 104 Hæmatomyzus, 104 Hæmatopinidæ, 105 Hæmatopinus, 105 Hæmatopinoides, 105 Hæmatopinoididæ, 105 Hæmatopota, 287 Hæmatopotinæ, 287 Hæmatosiphon, 143 Hæmogamasus, 568 Hæmolytis, 225, 233 Hæmonia, 441 Hæmorrhagia, 208 Hæterina, 162 Hahnia, 560 Halacaridæ, 570 Halacarus, 570 Halacogaster, 431 Halarachne, 568 Halarachnidæ, 568 Halechiniscidæ, 583 Halechiniscus, 583 Halesus, 192 Halictoides, 508 Halictophagidæ, 470 Halictophagoidea, 470 Halictophagus, 470 Halictophilus, 470 Halimococcus, 134 Haliplidæ, 410 Haliplus, 410, **412** Halisidota, 219 Halleinosoma, 590 Hallomenus, 438 Halobates, 151 Halterata, 264 Halteriptera, 264 Haltica, 444 Haltichella, 485 Halticidæ, 444 Halticus, **144**, 145, 508 Hamadryas, 238

Hamamelistes, 126 Hamataliva, 560 Hammatorrhina, 269, 270 Hammomyia, 319 Hamophthirius, 105 Hanseniella, 44, 604 Hapalotrichine, 270 Hapalothrix, 270 Haphospatha, 321 Haploa, 219 Haplobainosoma, 592 Haplobainosomatidæ, 592 Haplocampa, 44 Haploembia, 92 Haplocnemus, 429 Haplophilus, 602 Haplopeza, 88 Haploporatia, 591 Haploptilia, 232 Haploptiliidæ, 232 Haplophthalmus, 534, 534 Haplostomata, 312 Haplothele, 556 Haplothysanus, 596 Harmolita, 486 Harmologa, 226 Harmostes, 148 Harpactes, 557 Harpactor, 147 Harpagomantis, 91 Harpagophora, 596 Harpagophoridæ, 596 Harpalus, **409**, 410 Harpobittacus, 189 Harrisina, **208**, 208 Hartomyia, **281**, 282 Hasora, 235 Haustellata, 264 Hebecnema, 320 Hebridæ, 147, 150 Hebrus, 147, 150 Hecabolinæ, 479 Hecabolus, 479 Hectopsylla, 406 Hectopsyllidæ, 406 Hedroneura, 328 Hedychrum, 493 Helcomyza, 327 Helcomyzinæ, 327 Helcon, 480 Helconinæ, 480 Heleidæ, 282 Heleocoris, 154 Heleodromia, 301 Helice, 233 Helicobia, 316

Helicoconis, 182 Heliconiidæ, 238 Heliconius, 238 Helicopis, 239 Helicus, 418 Helina, 320 Heliocharis, 163 Heliodines, 231 Heliodinidæ, 229, 231,244. Heliothis, 218 Heliothrips, 72, 74 m Helle, 292Heliozela, 230 Heliozelidæ, 229, 230, 242Hellichia, 281 Hellichiinæ, 281 Hellula, 216 Helmidæ, 420 Helmis, 420 Helobia, 268 Helodes, 418 Helodidæ, 418 Helodon, 281 Helomyza, 334, 335 Helomyzidæ, 334 Helomyzinæ, 334, 335 Helorimorphinæ, 482 Helophilus, 309 Helophorinæ, 414 Helophorus, 414 Helopidæ, 436 Helops, 436 Heloridæ, 492 Helorimorpha, 482 Helorus, 492 Helota, 424 Helotidæ, 424 Helotrephes, 152 Helotrephidæ, 152 Helotrephinæ, 152 Hemaris, 208 Hemerodromiinæ, 301 Hemerobiidæ, 184 Hemerobioidea, 183 Hemerobius, 184 Hemerocampa, 218 Hemerodromia, 301 Hemeromyia, 345 Hemerophilidæ, 228 Hemiceras, 208 Hemichionaspis, 130 Hemiclœa, 562 Hemiclonia, 52 Hemicoccinæ, 134 Hemifænus, 476

Hemileuca, 213 Hemileucidæ, 213 Hemileucinæ, 251 Hemicampa, 44 Hemimene, 226 Hemimeridæ, 71 Hemimerus, 71 Hemineura, 99 Heminyctibora, 79 Hemiodœcus, 134 Hemipauropus, 584 Hemipenthes, 294 Hemipeplus, 424 Hemiphlebia, **160**, 161 Hemiphlebiidæ, 161 Hemiphrynus, 542 Hemipsocus, 99 Hemiptera, 140 Hemipterota, 81 Hemithea, 210 Hemitheidæ, 210 Hemisaga, 52 Hemisarcoptes, 574 Hemiscorpion, 544 Hemisia, 508 Hemisphærius, 121 Hemistephanus, 476 Hendecatomus, 432 Henia, 603 Henica, 296 Henicocephalidæ, 147 Henicocephalus, 146, 147 Heniconeura, 296 Henicops, 600 Henicopidæ, 600 Henopidæ, 292 Heodes, 239 Hepialidæ, 201, 245 Hepialoidea, 200 Hepialus, 201 Heptagenia, 172 Heptageniidæ, 172, 173 Heptathela, 555 Heptatoma, 287 Hera, 320 Hercostomus, 303 Herculia, 215 Heretomera, 413 Heriades, 507 Herina, 331 Heringia, 308 Hermannia, 570 Hermarchus, 64 Hermes, 179 Hermetia, 285, 360, 361 Hermetiinæ, 285

Herminia, 218 Hermippus, 559 Hersilia, 560 Hersiliidæ, 560 Hersiliola, 560 Hesperempis, 301 Hesperia, 235, 236 Hesperiidæ, 235, 252 Hesperiinæ, 236 Hesperinidæ, 273 Hesperinus, 274 Hesperioidea, 233 Hesperoctenes, 154 Hesperoleon, 187 Hesperophylum, 144 Hestiasula, 91 Hetæriinæ, 423 Heteracanthia, 284 Heterocampa, 211 Heterocera, 201 Heteroceridæ, 432 Heterocerus, 416, 432 Heterochætula, 90 Heterochila, 327 Heterochroa, 238 Heteroconis, 182 Heterocopus, 62, 63 Heterodoxus, 102 Heterogenea, 207 Heterogeneidæ, 207 Heterogyna, 489 Heterogynidæ, 208, 239 Heterogynis, 208 Heterojapyx, 43 Heterolampra, 79 Heterolatzella, 592 Heterolatzellidæ, 592 Heterometrus, 545 Heteroneura, 200, 334 Heteroneuridæ, 334 Heteropeza, 275 Heteropezinæ, 275 Heterophrynus, 542 Heteroplectron, 194 Heteropoda, 562 Heteropogon, 298 Heteroporatia, 591 Heteroporatiidæ, 591 Heteropsylla, 123 Heteroptera, 140 Heteropteryginæ, 63 Heteropteryx, 62, 63 Heterosphyronida, 548 Heterospilus, 479 Heterostylum, 297 Heterotardigrada, 582

Heterothripidæ, 74 Heterothrips, 74 Heterotropinæ, 296 Heterotropus, 296 Hetrodes, 53 Hetrodinæ, 53 Hexacentrus, 52 Hexagenia, 170 Hexalebertia, 572 Hexamitocera, 319 Hexatoma, 268 Hexatominæ, 268 Hexisopodidæ, 547 Hexisopus, 547 Hexura, 556 Hibrildes, 221 Hieroxestidæ, 223 Hilara, 301 Hilarella, 316 Hilarempis, 301, 302 Hilarimorpha, 290, 290 Hilarimorphine, 290 Hilda, 117 Himantariidæ, 602 Himantarium, 601 Himantopteridæ, 204 Himantopterus, 204, 205 Himantosoma, 602 Hindola, 109 Hindolinæ, 109 Hippasa, 561 Hippelates, 340 Hippiscus, 56 Hippobosca, 347 Hippoboscidæ, 347, 352 Hippoboscinæ, 347 Hippodamia, 434 Hirmoneura, 291 Hirmoneurinæ, 291 Hirtea, 286 Hispa, 444 Hispaniosoma, 592 Hispella, 442 Hispidæ, 444 Hister, 423 Histeridæ, 418, 423, 436 Histerinæ, 423 Histiosoma, 574 Histriciidæ, 318 Hodotermes, 94 Hodotermitidæ, 95 Hofmannia, 231 Hollandidæ, 206 Holcocera, **230**, 232 Holocompsa, 81 Hololampra, 79

Hololepta, 423 Hololeptinæ, 423 Holoparamecinæ, 434 Holopogon, 298 Holorusia, 268 Holosiro, 566 Holotergidæ, 543 Holothyrus, 567 Holopodostreptus, 597 Holothyridæ, 567 Holurothrips, 76 Homæmus, 150 Homaledra, 233 Homalocneminæ, 301 Homalocnemis, 301 Homalomyia, 366, 367 Homalonychidæ, 562 Homalonychus, 562 Homalopterus, 79 Homalota, 429 Homilops, 470 Homœocerus, 148 Homœogamia, 81 Homœogamiidæ, 81 Homœoxiphus, 54 Homoneura, 200, 327 Homoneuria, 170 Homoptera, 106 Homorocoryphus, 52 Hoplacantha, 284 Hoplocampa, 474 Hoplocampinæ, 512 Hoploclonia, 63 Hoplocorypha, 85, 87 Hoplocyrtoma, 301 Hoploderma, **569**, 570 Hoplodermatidæ, 570 Hoplodictya, 328 Hoplistes, 286 Hoplodonta, 286 Hoplopleura, **105**, 105 Hoplopsyllus, 405 Hoplionota, 444 Hoplopria, 491 Hormaphidinæ, 126 Hormaphis, 126 Hormetica, 82 Hormiinæ, 479 Hormiopterus, 479 Hormius, 479 Hormopeza, 301 Hormosomyia, **274**, 275 Hormurus, 545 Hornia, 439 Horus, 550 Howardia, 130

Huitfeldtia, 572 Humbertiella, 87 Huttonella, 284 Hya, 549 Hyadina, 343 Hyalanthrax, 295 Hyalopterus, 125 Hyarinus, 549 Hybalus, 451 Hyblæa, 207 Hyblæidæ, 207 Hybosoridæ, 451 Hybosorus, 451 Hybotidæ, 302 Hybotinæ, 302 Hybris, 187 Hydaticinæ, 412 Hydnocera, 427 Hydrachna, 571 Hydrachnidæ, 571 Hydrachnoidea, 570 Hydræna, 414 Hydræninæ, 414 Hydrellia, 343 Hydrelliidæ, 343 Hydrelliinæ. 343 Hydrina, 343 Hydriomena, 210 Hydriomenidæ, 210 Hydrobates, 572 Hydrobatidæ, 152 Hydrobiinæ, 414 Hydrobiosis, 193, **193** Hydrocampa, 216 Hydrocampinæ, 216 Hydrochoreutes, 572 Hydrocorisæ, 141 Hydrometra, 142, **144** Hydrometridæ, 142 Hydromyza, 319 Hydrophilidæ, 414 Hydrophilinæ, 414 Hydrophiloidea, 414 Hydrophilus, 410, 414, Hydrophoria, 319 Hydrophorinæ, 304 Hydrophorus, 304 Hydroporinæ, 411 Hydroporomorpha, 409 Hydroporus, 411 Hydropsyche, **193**, 194 Hydropsychidæ, 194, 195 Hydropsychodes, 194 Hydroptila, 192 Hydroptilidæ, 192, 195

Hydroscapha, 434 Hydroscaphidæ, 434 Hydrotæa, 320 Hydrotæinæ, 320 Hydrous, 414 Hydrovolzia, 570 Hydrovolziidæ, 570 Hydryphantes, 571 Hydryphantidæ, 571 Hygrobatidæ, 572 Hygrobia, 410 Hygrobiidæ, 410 Hyidæ, 549 Hylæidæ, 507 Hylæoides, 507 Hylæus 507 Hylastes, 445, 445 Hylastinus, **528** Hylecetus, 427, 429 Hylechthridæ, 470  ${
m Hylecthrus},\,470$ Hylemyia, 319 Hylemyiinæ, 319 Hylesinidæ, 446 Hylesinus, 445 Hyloicus, 208 Hylophila, 219 Hylophilidæ, 219, 439 Hylophilus, 439 Hylorus, 284 Hylotoma, 474 Hylotomidæ, 474 Hymenochimæra, 499 Hymenopharsalia, 477 Hymenopodinæ, 91 Hymenoptera, 471 Hymenorus, 435 Hyocephalidæ, 149 Hyocephalus, 149 Hypatus, 239 Hypena, 218 Hypeninæ, 218 Hyperalonia, 295 Hyperaspis, 434 Hyperetes, 98 Hyperhomala, 51 Hyperothrix, 588 Hyphantria, 219 Hyphoraia, 219 Hypnorna, 81 Hypocephalus, 427 Hypocera, 305 Hypochilidæ, 557 Hypochilus, 557 Hypochthonius, 570 Hypoctonus, 541

Hypoderma, 313 Hypoderminæ, 313 Hyponomeutidæ, 229 Hyponotidæ, 229 Hypoprepia, 219, **220** Hypoptidæ, 206 Hyporhicnoda, 80 Hyposemata, 555 Hyposmocoma, 233 Hyposmocomidæ, 233 Hypsa, 217, 217 Hypselophidæ, 229 Hypselosoma, 143 Hypsibius, 583 Hypsidæ, 217 Hypsimetopus, 534 Hypsopygia, 215 Hypsotropa, 215 Hypsotropinæ, 215 Hyptia, 476 Hyptiotes, 558 Hypurgus, 70 Hyrmophlæba, 291 Hyrophicnoda, 79 Hysterosia, 227 Hystichopsyllidæ, 405 Hystrichothripidæ, 76 Hystrichothrips, 76 Hystricophora, 227

Ibalia, 483 Ibaliidæ, 483, 514 Icaria, 497 Icaridion, 351 Icasma, 301 Icerya, 128, **129**, **133** Ichneumon, 475 Ichneumonidæ, 477, 488 Ichneumonoidea, 475, 514 Ichneutes, 481 Ichneutidea, 481 Ichneutinæ, 481 Icosidesmus, 589 Icosta, 347 Icterica, 341 Ictinus, 164 Incurvariidæ, 222 Idana, 331 Idarnes, 485 Idarnini, 487 Ideoroncidæ, 550Ideoroncus, **549**, 550 Idiocerus, 112 Idiococcidæ, 134 Idiocorinæ, 152 Idiocoris, 152

Idiogastra, 472 Idioglossa, 231 Idiops, 556 Idolomorpha, 84 Idolopsalis, 68 Idolothrips, 75 Idolothripidæ, 75 Idolum, 84 Illice, 219 Illinoia, 125 Ilythea, 343 Incisalia, 239 Incubinæ, 482 Incubus, 482 Incurvaria, 222 Incurvariidæ, 243 Inocellia, **181**, 181 Inostemma, 494 Integricipita, 404 Iomachus, **544**, 545 Iphiaulax, 478 Iphiclides, 236 Ipidæ, 446 Ipo, 112Ips, 445, **445**, 446 Irbisia, 145 Iridomyrmex, 489 Iridopteryginæ, 88 Iridopteryx, 88 Iris, 86 Iron, 172Isaphe, 589 Ischnomantis, 86 Ischnomyia, 339 Ischnopsyllidæ, 405 Ischnopsyllus, 405 Ischnopteryx, 99 Ischnothrix, 267 Ischnura, 161 Ischnuridæ, 545 Ischnurus, 545 Ischyropsalidæ, 554 Ischyropsalis, 554 Ischnocera, 101 Ischnogaster, 497 Ischnoptera, 79 Ishiana, **47**, 48 Ismene, 235 Ismeninæ, 235Isocorypha, 222 Isodermidæ, 145 Isodermus, 145 Isomerocera, 285 Isometopidæ, 145 Isometopus, **144**, 145 Isometroides, 545

Isometrus, 545 Isophya, 50 Isopoda, 534 Isopogon, 298 Isopsera, 50 Isoptera, 93 Isopteryx, 176 Isoscelipteron, 186 Isosoma, 486 Isosticta, **166** Isotoma, **45**, 46 Issidæ, 121 Issus, 121 Iteaphila, 301 Ithomia, 238 Ithomiidæ, 238 Ithone, 182, **183** Ithonidæ, 182 Ithonoidea, 182 Ithycerinæ, 448 Ithycerus, 448 Itonida, 275 Itonididæ, 274 Iurus, 546 Ixodes, 566, 567 Ixodidæ, 567 Ixodoidea, 567

Jacobsiella, 349 Jacobsoniidæ, 435 Jacobsonium, 435 Jalysus, 148 Janiodes, 212, 213 Janus, 472 Japanosoma, 593 Japygidæ, 43 Japyx, **41**, 43 Jassidæ, 112 Jassinæ, 114 Jassoidea, 109 Jassus, 114 Jemadia, 235 Joppeicidæ, 148 Joppeicus, 148 Joblotia, 272 Johannsenomyia, 282 Johannsenomyiidæ, 282 Jugatæ, 200 Jugofrenatæ, 200 Juliformia, 587 Julidæ, 595 Julodinæ, 420 Juloidea, 594 Julomorpha, 596 Julus, 595 Junonia, 238

Kalissus, 430 Kalobittacus, 189 Kalocrania, 68 Kana, 117 Kapala, 485, **487** Karschiella, 67, 68 Karschiellidæ, 68 Karumia, 431 Karumiidæ, 431 Karschia, 547 Kasserota, 121 Katastylops, 470 Kelloggina, 270 Kermesia, 119 Kermesidæ, 134 Kermes, 134 Kermococcus, 134 Kinnara, 119 Kinnaridæ, 119 Kiricephalus, 580, 581 Koebelea, **115**, 116 Koebeleidæ, 116 Koenenia, **539**, 540 Koeneniidæ, 540 Kolba, 111 Konowiella, 497 Konowiellidæ, 497 Koroana, 120 Kosmetor, 70 Kuwania, 128 Kleiduchus, 69 Krœberia, 325

Labia, 69 Labiales, 69 Labidarge, 474 Labidocarpus, 574 Labidostomma, 569 Labidostommatidæ, 569 Labidura, **67**, 68 Labidurales, 66 Labiduridæ, 68 Labiduroidea, 66 Labiidæ, 69 Labioidea, 69 Labulla, 560 Labus, 496 Laccifer, 132 Lacciferidæ, 132 Laccobius, 414 Laccophilinæ, 411 Laccophilus, 411 Laccotrephes, 154 Laches, 560 Lachlania, 170 Lachneidæ, 221

Lachninæ, 124 Lachnosterna, 451 Lachnus, 124 Lacinius, 553 Lacon, 421 Lacosoma, 213 Lacosomatidæ, 213, 250 Lacosomidæ, 213 Lactistomyia, 302 Lælaps, 568 Læmobothriidæ, 103 Læmobothrion, 103 Læmophlæidæ, 424 Læmophlæus, 424 Læmopsylla, 405 Laertias, 236 Laglaisia, 331 Lagoidæ, 204 Lagria, 436 Lagriidæ, 436 Lamarckiana, 57 Lamellicornia, 413 Lamenia, 119 Laminitarsus, 480 Laminosioptes, 574 Lampetia, 309 Lampria, 299 Lamprima, 450 Lamprocera, 431 Lamprochromus, 304 Lamprogaster, 331 Lampromyia, 290 Lampronia, 222 Lamproniidæ, 222 Lamprophthalma, 331 Lamprosoma, 443 Lamprosomatidæ, 443 Lamprotatus, 486 Lampyridæ, 431 Lampyris, 431 Lamyra, 299 Languria, **422**, 432 Languriidæ, 432 Laniatores, 551 Lanternaria, 119 Laphria, 299 Laphriinæ, 298, 299 Laphystia, 298 Larentia, 210 Larentiidæ, 210 Larex, 69 Laria, 439 Laricobius, 425 Lariidæ, 440 Larra, 505 Larridæ, 505

Larvævora, 318 Larvævoridæ, 318 Lasia, 292 Lasiocampa, 221 Lasiocampidæ, 221, 251 Lasioderma, **422**, 426 Lasiopa, 286 Lasiopinæ, 286 Lasiopogon, 298 Lasiopsocus, 99 Lasioptera, 275 Lasiopteryx, 275 Lasiosepsis, 323 Lasius, 489 Laspeyresia, 226 Lathelmis, 420 Lathrobium, 429 Lathys, 558 Latindia, 81 Latindiidæ, 81 Latridiidæ, 434 Latridiinæ, 434 Latridius, 434 Latrodectus, 559 Latumcephalum, 102 Lauxania, 327 Lauxaniidæ, 327 Laverna, 233 Lavernidæ, 233 Lebertia, 572 Lebertiidæ, 571 Lebia, 410 Lecaniidæ, 132 Lecaniodiaspis, 132 Lecaniopsis, 132 Lecanium, **129**, 132 Ledra, 112 Ledridæ, 112 Ledropsis, 112 Legnotomyia, 297 Leia, 278, **359**, 370, **372** Leiophron, 481 Leiophroninæ, 481 Leiotheidæ, 103 Leiotheum, 103 Leiperia, 581 Lelaps, 486 Lema, 442 Lemodes, 439Lemonia, 212 Lemonias, 238 Lemoniidæ, 212, 239 Leocrates, 63 Leonardius, 124 Lepidanthrax, 295 Lepidilla, 98

Lepidillidæ, 98 Lepidophora, 296 Lepidophthirius, 104 Lepidopsocidæ, 98 Lepidopsocus, 98 Lepidoptera, 198 Lepidosaphes, 130, 133 Lepidosaphinæ, 130 Lepinotus, **97**, 98 Lepiselaga, 287 Lepiselaginæ, 287 Lepisma, **41**, 42 Lepismatidæ, 42 Lepismatoidea, 42 Leprolepis, 98 Leptacis, 494 Leptidæ, 290, 374 Leptinæ, 290 Leptinidæ, 427 Leptinillus, 427 Leptinotarsa, 414, 443 Leptinus, 427 Leptis, 290 Leptisolabis, 69 Leptocella, 194, **196** Leptocera, **337**, 344 Leptoceratidæ, 344 Leptoceridæ, 194, 197 Leptocerus, 194 Leptocorisa, 148 Leptodesmidæ, 589 Leptodesmus, 589 Leptofænidæ, 485 Leptofænus, 485 Leptogaster, 298 Leptogastrinæ, 298 Leptoglossus, 148, 149 Leptohyphes, 171 Leptohyphodes, 171 Leptomantis, 89 Leptomastax, 428 Leptomeris, 210 Leptomydas, 297, **297** Leptoneta, 559 Leptonetidæ, 559 Leptopanorpa, 190 Leptopelma, 556 Leptoperla, 177 Leptoperlidæ, 177, 178 Leptopeza, 301 Leptophlebia, 172 Leptophlebiidæ, 172, 173 Leptopodinæ, 303 Leptorethrum, 303 Leptorhaptus, 495 Leptotyphlinæ, 430

Leptoxyda, 341 Leptura, 441 Leptynoptera, 123 Leria, 335 Leriinæ, 335 Lestes, **160**, 162 Lestidæ, 162, 165 Lestoidea, 162 Lestophonus, 342 Lestremia, **274**, 275 Lestremiinæ, 275 Lethocerus, **153**, 154 Lethrus, 450 Leto, 201Leucania, 218 Leucaspidinæ, 131  $L_{eucaspis}$ , 131  $L_{eucauge, 560}$ Leucomele, 222 Leucophæa, 82  $L_{eucophenga, 345}$ Leucophthalmia, 210 Leucopis, **326**, 329 Leucopomyia, 329 Leucoptera, 223 Leucorrhinia, 164 Leucospididæ, 485 Leucostola, 304 Leucostoma, 318 Leucozona, 309 Leuctra, 178 Leuctridæ, 178 Leurolestes, 79 Liancalus, 304 Libellaginidæ, 163 Libellago, 163 Libellula, **163**, 164 Libellulidæ, 164, 167 Libelluloidea, 159, 163 Libethra, 64 Libitoides, 553 Liburnelenchus, 470 Liburnia, **113**, 119 Liburniella, **113**, **118** Libythea. 239 Libytheidæ, 239, 253 Lichenomima, 99 Lidar, 184 Liesthes, 432 Ligaria, 90 Ligia, 534 Ligidium, 534, **534** Ligiidæ, 534 Ligydidæ, 534 Ligyrocoris, 149, **149** Lilæa, 289

Limacodidæ, 207 Limacomorpha, 586 Limatus, 272 Limenitis, 238 Limnephilidæ, 192, 197 Limnephilus, 192, **193** Limnesia, 571 Limnesiella, 571 Limnesiidæ, 571 Limnia, 328 Limnichinæ, 417 Limnichus, 417 Limnobates, 142 Limnobatidæ, 142 Limnobia, 268 Limnobiidæ, 268 Limnochares, 570 Limnocharidæ, 570 Limnœcia, 233 Limnophila, **267**, 268 Limnophilinæ, 268 Limnophora, **317**, 320 Limnophorinæ, 320 Limnospila, 320 Limonia 268, 354, **355** Limonicola, 270 Limoniidæ, 268, 348, 354, 374Limoniinæ, 268 Limosina, 344 Limulodes, 433 Lindenia, 164 Linguatula, **580**, 581 Linguatulida, 579 Linguatulidæ, 581 Linguatulinæ, 581 Linobia, 574 Linognathus, 105 Linopodes, 576 Linyphia, 560 Linyphiidæ, 560 Liocranum, 563 Liodes, **416**, 428 Liodidæ, 428 Liodrosophila, 345 Liogma, 268 Liomyza, 332 Liosomaphis, 125 Liothrips, 76 Liparidæ, 218 Liparididæ, 218 Liparis, 218 Liparochrus, 451 Lipeurus, **102**, 103 Liphistiidæ, 555 Liphistiomorphæ, 555

Liphistius, **537**, 555 Liphochæta, 343 Lipochætinæ, 343 Lipocosma, 216 Liponeura, 270 Liponeuridæ, 269 Liponyssus, 568 Lipoptena, 347, 352 Lipopteninæ, 347 Lipoptera, 101 Liposcelidæ, 98 Liposcelis, 98 Liriomyza, 339 Liriope, 267 Liriopeidæ, 266 Lispa, 320 Lispinæ, 320 Lispocephala, 320 Lissa, 329 Lissoderma, 437 Listrocelinæ, 52 Listrocelis, 52 Listrophoridæ, 574 Listrophorus, 574 Litaneutria, 90 Litargus, 432 Lithobiidæ, 600 Lithobioidea, 600 Lithobiomorpha, 599 Lithobus, 600 Lithocolletidæ, 232 Lithocolletis, 232 Lithosia, 219 Lithosiidæ, 209, 219, 248 Lithurgus, 507 Litorimus, 432 Liturgusa, 89 Liturgusinæ, 89 Livia, 123 Liviinæ, 123 Llaveia 128 Loberus, 424 Loboceratidæ, 474 Lobogaster, 275 Loboptera, 79 Locusta, 52, 56 Locustidæ, 50, 56 Locustinæ, 52 Locustodea, 48 Lœmopsylla, 405 Loewinella, 299 Lomanyia, 186 Lomatia, 295 Lomatiinæ, 295 Lonchæa, 334, 336, **337** Lonchæidæ, 334, 336

Lonchodes, 64 Lonchodinæ, 64 Lonchoptera, **305**, 306,364 Lonchopteridæ, 306, 364, 365 Longurio, 268 Lonomiidæ, 213 Lophocarenum, 560 Lophopoda, 435 Lophopidæ, 121 Lophoproctus, 585 Lophops, 121 Lophoptilus, 233 Lophoteles, 285 Lophotelinæ, 285 Lophyridæ, 474 Lophyrus, 474 Lorditomæus, 451 Lordotus, 297 Loxocera, 340 Loxomantis, 86 Loxoneura, 331 Loxosceles, 559 Loxostege, 216 Loxotropa, 493 Lucanidæ, 450 Lucanus, **416**, 450 Lucidota, 431 Lucilia, 316 Luciola, 431 Ludius, **416** Luffia, 205 Lule, 331 Luperina, 218 Luperus, 444 Lutica, 559 Lutrochus, 418 Lutzia, 272 Lycæna, **234**, 239 Lycænidæ, 239, 252 Lycenopsis, 239 Lycidæ, 430 Lycinus, 555 Lycnophanes, 443 Lycogaster, 494 Lycoperdina, 434 Lycoria, 278 Lycoriidæ, 278 Lycosa, 561 Lycosidæ, 561 Lyctidæ, 426 Lyctus, **422**, 426, **528** Lyda, 472 Lydidæ, 472 Lygæidæ, 149 Lygæoidea, 148

Lygæosoma, 149 Lygæus, 149 Lygistopterus, 410 Lygistorrhina, 277 Lygistorrhininæ, 277 Lygocerus, 494 Lygus, **144**, 145 Lymantria, **217**, 218 Lymantriidæ, 217, 218, 240, 249Lymexylidæ, 427, 429 Lymexylon, 427 Lymexylonidæ, 427 Lymnadidæ, 238 Lymnas, 239 Lynchia, 347 Lyonetia, 222 Lyonetiidæ, 223, 243, 244 Lype, 194 Lyperosia, 321 Lyroda, 505 Lysiognatha, 477 Lysiognathidæ, 477 Lysiopetalidæ, 594 Lysiopetaloidea, 593 Lysiopetalum, 594 I ysiphlebus, **478**, 482 Lysozum, 286 Lyssomanes, 561 Lytogaster, 343 Lytta, 439 Lyttidæ, 439

Macaria, 209 Machærium, 304 Machærota, 109, **110** Machærotidæ, 109 Machærotinæ, 109 Macheiriophoron, 592 MacLilidæ, 42 Machilinus, 42 Machilis, 42, **44** Machiloidea, 42 Machiloides, 42 Macquartia, 317 Macratria, 439 Macrobasis, 439 Macrobiotidæ, 583 Macrobiotus, 582, 583 Macrocentrinæ, 480 Macrocentrus, 480 Macrocephalidæ, 147 Macrocephalus, 147 Macrocera, **279**, 280 Macroceratidæ, 280 Macroceratogonia, 116 Macroceratogoniinæ, 116 Macrochæteuma, 591 Macrocheles, 569, **569** Macrochelinæ, 569 Macrodampetrus, 552 Macrofrenatæ, 204 Macroglossa 208 Macrogyrus, 413 Macrojugatæ, 200 Macromantis, 86 Macromia, 164 Macronema, 194 Macronicophilus, 602 Macronychus, 420, **421** Macroperipatus, 533 Macrophya, 474 Macropsis, 112 Macropsyllidæ, 405 Macrosiagon, 439 Macrosiphum, 125, **125** Macrotheca, 215 Macrothecinæ, 215 Macrothele, 556 Macrotoma, 325, 440 Macroxenus, 585 Macroxyela, 472 Macrurothrips, 74 Madiza, 345 Madizinæ, 345 Magalomus, 184 Magrettia, 50 Maindronia, 42 Majanga, 89 Majangella, 89 Majanginæ, 89 Majella, 562 Malachiidæ, 429 Malachius, 429 Malacodermata, 428 Malacomyza, 327 Malacopoda, 532 Malacopsylla, 406 Malacopsyllidæ, 406 Malacosoma, 221 Malcus, 148 Mallochella, 281 Mallochiola, 145 Mallodon, 440, **528** Mallophaga, 101 Mallophora, 298 Mallota, 309 Malthacotricha, 296 Malthodes, 429 Mamersa, 571 Manidiidæ, 209 Maniolidæ, 238

Manota, 278 Manotinæ, 278 Mansonia, 272 Mantichora, 410 Mantidæ 84 Mantillica, 87 Mantinæ, 89, 90 Mantis, 89 Mantispa, 183, **183** Mantispidæ, 183 Mantodea, 83 Mantoida, 88 Mantoidea, 83 Maorigarypus, 550 Marava, 69 Marbenia, 331 Marchalina, 128 Margarodes, 128 Margarodidæ, 127 Margarodinæ, 128 Margaronia, 216 Margaropus, 567 Marilia, 195 Mariobezzia, 295 Mariobezziinæ, 295 Marmara, 232 Marmessoidea, 64 Marpissa, 561 Maruina, 271 Masaridinæ, 496, 514 Masariella, 496 Masaris, 496 Masarygidæ, 308 Masicera, 318 Masiceridæ, 318 Maso, 560Mastigonodesmidæ, 588 Mastigonodesmus, 588 Mastigoproctus, 541 Mastogeninæ, 420 Mastostethus, 443 Mastotermes, 93, 94 Mastotermitidæ, 93, **94**, 95Masyntes, 56 Matachia, 558 Matsucoccus, 128, **133** Maura, 57 Maurina, 371 Maxudeinæ, 109 Mayetiola, **274** Mazarredia, 55 Mecaptera, 189 Mecistocephalidæ, 602 Mecistocephalus, 602 Mecistogaster, 160, 162

Meconema, 51 Meconeminæ, 51 Mecopoda, **51**, 51 Mecoptera, 189 Mecopodinæ, 51 Mecostethus, 56 Mecysmauchenius, 558 Medetera, 303 Medeterinæ, 303 Medusia, 210 Meenoplidæ, 119 Meenoplus, 119 Megachile, 507 Megacilissa, 507 Megacorminæ, 545 Megacormus, 545 Megalestes, **160**, 161 Megallanes, 184 Megaloblatta, 79 Megalodacne, 414, 424 Megalodontes, 473 Megalodontidæ, 473, 511 Megalopodidæ. 443 Megaloprepus, 162 Megaloptera, 179 Megalopus, 443 Megalopyge, 204 Megalopygidæ, 204, 248 Megalosphyinæ, 278 Megalosphys, 278 Megalothoracidæ, 46 Megalothorax, 46 Megalothrips, 76 Megalybus, 292 Megalyra, 476 Megalyridæ, 476 Megamerina, 329 Megamerinidæ, 329 Megapodagriidæ, 162, 166 Megapodagrion, 162 Megapsychops, 184 Megapsyllidæ, 406 Megapus, 572 Megarhininæ, 272 Megarhinus, 272 Megascelidæ, 442 Megascelis, 442 Megaselia, 305, 306 Megaspilus, 494 Megaspis, 309 Megastigmus, 485 Megathripidæ, 76 Megathrips, 76 Megathymidæ, 235, 252 Megathymus, 235 Megatomine, 425

Megatrioza, 123 Meghyperus, 302 Megilla, 434 Megisthanus, 569 Megisto, **234** 238 Megistocera, 268 Megistopoda, 347, 352 Megistorhynchus, 291 Megophthalma, 319 Megophthalmidæ, 114 Megophthalmus, 114 Melanchrinæ, 218 Melandrya, 438 Melandryidæ, 438 Melanolestes, 147 Melanophora, 317 Melanophthalma, 434 Melanoplus, 57 Melanostoma, 309 Melanothripidæ, 73 Melanothrips, 73 Melanotus, 421, 421, 528 Melaphe, 590 Melaphis, 126 Melasidæ. 422 Melasis, 422 Meleaba, 210 Melecta, 509Melectidæ, 509 Meleoma, 185 Melieria 331 Meligethes, 419 Melina, 328 Melipona, 506 Melissa, 509 Melissodes, 508 Melissopus, 226 Melitæa, 238 Melittia, **202** Melittobia, 486 Melittomma, 427 Melliera, 89 Mellieriella, 89 Mellierinæ, 89 Mellinidæ, 503 Mellinus, 503 Meloe, 439Meloidæ, 439 Melolontha, 451 Melolonthidæ, 451 Melophaginæ, 347 Melophagus, 347, 352 Melpia, 288 Melpiinæ, 288 Melusinidæ, 280 Melyridæ, 429

Melyris, 429 Membracidæ, 108 Membracis, 108 Menesta, 228 Menneus, 558 Mengea, 468 Mengeidæ, 468 Mengenilla, 468 Mengenillidæ, 468 Mengeoidea, 468 Menolepis, 272 Menopon, 103 Menoponidæ, 103 Menoxemus, 64 Menthidæ, 550 Menthus, 550 Meoneura, **342**, 345 Meracantha, 436 Meracanthomyia, 340  ${
m Meridogastra}, 542$ Merizocera, 559 Mermerus, 552 Merodon, 309 Meromyza, 340 Merope, 191 Meropidæ, 191 Meropliinæ, 323 Meroplius, 323 Merothripidæ, 74 Merothrips, 74, 75 Merragata, 147 Mesargus, 116 Mesasiobia, 67 Mesembrina, 321 Messembrinella, 314 Mesembrinellinæ, 314 Mesitius, 494 Meskea, 214 Mesobiobia, 70 Mesobracon, 479 Mesocanthus, 602 Mesochelidura, 70 Mesochria, 280 Mesolophora, 570 Mesomyza, 286 Mesoparopia, 114 Mesoperipatus, 533 Mesopsocidæ, 99 Mesopsocus, 97, 99 Mesorhaga, 303 Mesosemia, 239 Mesotermitidæ, 95 Mesothea, 210 Mesothelæ, 555 Mesovelia, 146 Mesoveliidæ, 146

Messena, 121 Messicobolus, 595 Meta, 560 Metabiantes, 553 Metacanthus, 148 Metachanda, 225 Metachandidæ, 225 Metacynorta, 553 Metæopsephenus, 427 Metagynodes, 553 Metallea, 314 Metallyticinæ, 85 Metallyticus, 85, 88 Metalype, 194 Metapelma, 484 Metaperipatus, 532 Metaphara, 288 Metaphoricus, 589 Metarbela, **205**, 206 Metarbelidæ, 206 Metarhaucus, 553 Metassamia, 552 Metatermitidæ, 95 Metelasmus, 352 Metentella, 90 Meteorinæ, 481 Meteorus, 481 Methana, 78 Methlinæ, 411 Methoca, 500 Methocidæ, 500, 501 Metilia, 87 Metisolabis, 69 Metopia, 316 Metopidothrigidæ, 593 Metopidothrix, 593 Metopiinæ, 316 Metopina, 306 Metopininæ, 306 Metoponia, 284 Metoponiinæ, 284 Metriocnemis, 282 Metrioptera, 52 Metriotes, 232 Metrocampa, 209 Metropus, 172 Mezira, 145 Meziridæ, 145 Mezium, 422, 427 Miagrammopes, 558 Miastor, 275 Micillus, 432 Micracis, 446 Micrambe, 424 Micrempis, 301 Microbembex, 505

Microbisium, 549 Microbracon, 478, **478** Microcerotermes, 95 Microcerus, 447 Microchordeuma, 593 Microchrysa, 286 Microdixa, 273 Microdon, **307**, 308 Microdontinæ, 308 Microdus, 480 Microfrenatæ, 203 Microgaster, 480 Microgastrinæ, 480 Microĥexura, 556 Microjugatæ, 200 Micromalthidæ, 429 Micromalthus, 429 Micrommata, 562 Micromorphus, 304 Micromus, 184, **184** Micromyia, 275 Micronecta, 152 Micropeplidæ, 430 Micropeplus, 430 Micropeza, 325 Micropezidæ, 325, 351 Micropezinæ, 325 Microphorus, 301 Microphysidæ, 145 Microplitis, 480 Microprosopa, 319 Micropsylla, 405 Micropterygidæ, 200, 240 Micropterygina, 200 Micropterygoidea, 200 Micropteryx, 200 Microsania, 310 Microspirobolus, 595 Microstigma, 162 Microstylum, 298 Microtermes, 95 Microthelyphonida, 539 Microvelia, 152 Mictis, 148 Midea, 572 Mideopsis, 572 Migadidæ, 556 Migas, 556 Migidæ, 556 Migonitis, 238 Milesia, 309 Milesiinæ, 309 Milichia, 345 Milichiella, 345 Milichiidæ, 344, 345 Milichiinæ, 345

Milnesiidæ, 583 Milnesium, **582**, 583 Miltinus, 297 Miltogramma, 316 Miltogrammatinæ, 316 Mimallo, 213 Mimallonidæ, 207, 213 Mimesidæ, 505 Mimetidæ, 559  ${f Mimetus},\ 559$ Mimnerminæ, 50 Mimnermus, 50 Mimoscorpius, 541 Minanga, 481 Mindarinæ, 125 Mindarus, 125, **125** Minettia, **326**, 327 Miobantia, 87 Mionyx, 84 Miopsalis, 551 Miopteryginæ, 90 Miopteryx, 90 Mirax, 480 Miridæ, 145, 150 Mirientomata, 40 Miris, 145 Mirophasma, 63 Mischocyttarus, 497 Miscogaster, 486 Miscogastridæ, 486, 488 Miscophidæ, 505, 506 Miscophus, 505 Misumena, 562 Mitopus, 553 Mitoura, 239 Mixogaster, 308 Mnemonica, 200 Mnesarchæa, 200 Mnesarchæidæ, 200 Mochlonyx, 272 Modisimus, 559 Moggridgea, 556 Mogoplistes, 54 Mogoplistinæ, 54 Molanna, 194 Molannidæ, 194, 197 Molannodes, 194 Molophilus, 268 Moma, 218 Mominæ, 218 Mompha, **230**, 233 Monachus, 443 Monardia, 275, **370**, 371 Monecphora, 111 Monedula, 505 Moneta, 559

Moneiziella, 574 Monistria, 57 Monobia, 497 Monochamus, 441 Monoclona, 277 Monocrepidius, 421 Monocteniidæ, 209 Monodes, 218 Monodontomerus, 485 Monodromia, 301 Monædidæ, 431 Monœdus, **422**, 431 Monogyropus, 102 Monomachidæ, 477, 495 Monomachus, 477 Monomma, 437 Monommatidæ, 437 Monommidæ, 437 Monomorium 490 Mononychidæ, 153 Mononyx, 153 Monophadnoides, 474 Monophlebidæ, 127 Monophlebinæ, 128 Monophlebus, 128 Monopidæ, 224 Monopis, 224 Monopsis, 120 Monosphyronida, 548 Monotarsobius, 600, 601 Monotomidæ, 435 Monoxia, 444 Moonia, 116 Mordella, 438 Mordellidæ, 438 Mordellistena, 437, 438 Morellia, 321 Moritziella, 127 Mormolyce, 411 Mormolycidæ, 411  ${
m Morpho}, 238$ Morphoidæ, 238 Mortoniella, 192 Mortoniellus, 52Mosillus, 330 Mosoia, 552 Muirixenos, 470 Mumetopia, 339 Mundamella, 571 Mundochthonius, 548 Munichryia, 217 Murgantia, 151 Murmidiidæ, 433 Murmidius, 433 Murricia, 560 Musca, **299**, **317**, 321, **367** 

Muscaridæ, 311 Muscidæ, 321, 366, 367 Muscina, 321, **367**, 362 Muscinæ, 321 Muscoidea, 300, 311, 319, 365Musidora, 306 Musidoridæ, 306 Musonia, 87 Musoniella, 87 Musotima, 216 Mutilla, 500 Mutillidæ, 499, 500, 501, Mycetæa, 432 Mycetæidæ, 432 Mycetaulus, 334 Mycetobia, 279, 280, 359, Mycetobiidæ, 280, 359 Mycetophagidæ, 432 Mycetophagus, 432 Mycetophila, 278, 278 Mycetophilidæ, 277, 349, 358, 370Mycetophilinæ, 278 Mychocerus, 433 Mycodrosophila, 345 Mycomyia, 277 Mycophaga, 319 Mycterus, 437 Mycotretus, 432 Mydæa, 320 Mydæinæ, 320 Mydaidæ, 297, 362, 376 Mydas, **297**, 297, 362, **363**, 375 Mydasidæ, 297 Myennis, 331 Myersiidæ, 477 Mygalomorphæ, 555 Myielaphus, 298 Myiocera, 318 Myiodactylidæ, 187 Myiodactylus, 187 Myiodaria, 311 Myiolepta, 309 Myiomma, 145 Myjophanes, 147 Myiospila, 321 Mylabridæ, 440 Mylabris, 439, **440**, **528** Mymaridæ, 483 Myndus, 120 Myobia, 576 Myocoptes, 574

Myodaria inferiora, 312 Myodaria superiora, 311 Myodina, 330  ${f Myodites}, 439$ Myodochidæ, 149 Myonia, 210 Myopa, 307 Myopina, 319 Myopinæ, 307 Myopsocidæ, 99 Myopsocus, **97**, 99 Myrcinus, 90 Myrientomata, 33 Myrmacicelus, 446 Myrmarachne, 561 Myrmecia, 490  ${
m Myrmecodesmus}, 588$ Myrmecolacidæ, 469 Myrmecolax, 469 Myrmecopterina, 497 Myrmecothea, 331 Myrmecomorpha, 352 Myrmecomorphus, 492 Myrmecomyia, 331 Myrmecomyiinæ, 331 Myrmecophila, 54 Myrmecophilinæ, 54 Myrmeleon, 187 Myrmeleonidæ, 187 Myrmeleonoidea, 183 Myrmeleontidæ, 187 Myrmeleontoidea, 183 Myrmica, 490, **491** Myrmicinæ, 490 Myrmosa, 500 Myrmosidæ, 500, 502 Myropsis, 558 Myrsidea, 103 Myscelus, 235 Mysoria, 235  ${
m Mystacides}, 194$ Mystalides, 595 Mystrocnemis, 492 Mythicomyia, **295**, 296 Mythicomyline, 296 Mytilaspis, 130 Myxodesmus, 588 Myxosargus, 286 Myzine, 499 Myzinidæ, 499 Myzus, 125 Nabidæ, 147 Nabis, 147 Nacerda, 437

Nachaba, 215

Næogæidæ, 147 Nala, 68 Nallachius, 184 Nannisolabis, 69 Nannochorista, 190 Nannochoristinæ, 190 Nannolene, 596 Nannolenidæ, 596Nannophilus, 602 Nanophyinæ, 449 Nanophyllium, 63 Napæa, 343 Napæinæ, 343 Napomyza, 339 Nasonia, 484 Nasutitermes, 95 Nathalis, 236 Naucoridæ, 154 Naucoris, 154 Nauphœta, 82 Naupoda, 331 Nausibius, 424 Nausigaster, 308 Nausigastrinæ, 308 Nautarachna, 572 Neandra, 419 Neaspilota, 341 Neatractosoma, 592 Neatractosomatidæ, 592 Nebo, 544 Nebria, 411 Nebrobia, 426 Necrophorus, 409, 410,428 Necroscia, 64 Necrosciinæ, 64 Nectarina, 497 Nedyopus, 589 Neelidæ, 46 Neelus, 46 Negritomyia, 286 Neĥallenia, 161 Neides, 148 Neididæ, 148 Nelipoda, 555 Nemastosoma, 554 Nemastosomatidæ, 554 Nemastosomatoidea, 553 Nematocera, 265, 347,353, 369Nematoceropsis, 289 Nematophora, 587 Nematinæ, 474, 513  ${
m Nematus}, 474$ Nemeobiidæ, 239 Nemeobiinæ, 239

Nemeobius, 239

Nemeophila, 219 Nemestrinidæ, 291, 361 Nemestrininæ, 291 Nemestrinus, 291 Nemobiinæ, 54 Nemobius, 54 Nemognatha, 439 Nemonychidæ, 449 Nemonyx, 449 Nemopalpinæ, 271 Nemopalpus, 271 Nemophora, 222 Nemopistha, 183 Nemopoda, 323 Nemopodinæ, 323 Nemoptera, 183 Nemopterella, 183 Nemopteridæ, 183 Nemopteroidea, 183 Nemoræa, 316 Nemoria, 210 Nemosoma, 419 Nemostira, 436 Nemotelus, 286 Nemotois, 224 Nemoura, 178 Nemouridæ, 178 Neoalticomerus, 338 Neoascia, 309 Neobisiidæ, 549 Neobisioidea, 548 Neobisium, 549 Neobittacus, 189 Neocapritermes, 95 Neocastnia, 203 Neocastniidæ, 203 Neochauliodes, 179 Neoconocephalus, 52 Neocurupira, 270 Neodiospilus, 481 Neodiprion, 510 Neodixa, 273 Neoempheria, 277 Neoexaireta, 284 Neogæus 150 Neogeophilidæ, 602 Neogeophilus, 602 Neoglaphyroptera, 278 Neohermes, 179Neolecanium, 132 Neolobophora, 70 Neolobophoridæ, 70 Neomaskiella, 124 Neoneurinæ, 480 Neoneuromus, 179  ${
m Neoneurus}, 480$ 

Neonympha, 238 Neopanorpa, 190 Neopelomyia, 337 Neopseustidæ, 200 Neopseustis, 200 Neopsyllidæ, 405 Neorhacodes, 480 Neorhacodinæ, 480 Neorhacodidæ, 477 Neorhynchocephalus, 291 Neoscutops, 331 Neosticta, 161 Neostylops, 470 Neotermes, 94 Neotettix, 55 Neottiophilidæ, 323, 334 Neottiophilum, 333, 334 Neozele, 480 Nepa, **153**, 154 Nepal, 184 Nephesa, 120 Nephila, 560 Nephoneura, 187 Nephroceros, 310 Nephropteryx, 215 Nephrotoma, 268 Nepidæ, 154 Nepticula, 221 Nepticulidæ, 221, 242 Neriidæ, 325 Neriocephalus, 325 Nerius, **324**, 325 Nerophilus, 195 Nerthra, 153 Nerthridæ, 153 Nervijuncta, 280 Nesiope, 123 Nesiotinidæ, 103 Nesiotinus, 103 Nesobasis, 160 Nesogaster, 69 Nesogastridæ, 69 Nesomachilis, 41 Neuraphes, 428 Neureclipsis, 194 Neurigona, 304 Neurigoninæ, 304 Neumannia, 572 Neurobasis, 162 Neuroctena, 327, **328** Neuroctenus, 145 Neuromachærota, 109 Neurometa, 120 Neuromus, 179 Neuronia, 193 Neuroptera, 181

Neuroptynx, 187 Neurorthrus, 185 Neurotoma, 472 Nevermannia, 281 Nevermanniinæ, 281 Newportia, 600 Nicagus, 450 Nicocles, 298 Nicodamus, 559 Nicoletia, 42 Nicoletiella, 569 Nicsara, 52 Nigetia, **220** Nilio, 435 Nilionidæ, 435 Nina, 183 Ninetis, 559 Nionia, 114 Niphas, 182 Niponiidæ, 423 Niponius, 423 Nirvana, **115**, 117 Nirvanidæ, 116, 117 Nisitra, 54 Nitela, 506 Nitelidæ, 505 Niteliopsis, 504 Nitidula, 419 Nitidulidæ, 419, 431, 435 Nocticola, 78 Nocticolidæ, 78 Noctua, 218 Noctuidæ, 218, 248, 249, 250Noctuinæ, 218 Nodonota, 444  ${f Nodostoma}, 444$ Noeza, 302 Nogodina, 120 Nogodinidæ, 120 Nola, 219 Nolidæ, 219, 248 Nomada, 509 Nomadidæ, 509 Nonacris, 286 Nonagria, 218 Nopoiulus, 595 Nops, 557Norape, 204 Norellia, 319 Norelliinæ, 319 Nosodendridæ, 417 Nosodendron, 417 Nososticta, 161 Notanatolica, 194 Noterinæ, 411

Nothoblatta, 81 Nothoblattidæ, 81 Nothochrysa, 185 Nothomyia, 286 Nothotrichocera, 267 Nothra, 292 Nothybidæ, 325 Nothybus, 325 Notiophygidæ, 434 Notiophygus, 434 Notiothauma, **190**, 191 Notiothaumidæ, 191 Notiphila, 343 Notiphilidæ, 343 Notiphilinæ, 343 Notophallus, 576 Notodonta, 211 Notodontidæ, 211, 249 Notolophus, 218 Notonaulax, 340 Notonecta, 153, **153** Notonectidæ, 153 Notonemoura, 177 Notoptera, 47 Notoxus, 439 Noya, 170 Nuncia, 552 Nusa, 299 Nyctalemon, 213 Nycteribia, 346, **346**, 352 Nycteribiidæ, 346, 352 Nycterophila, 347 Nyctibora, 79 Nyctiboridæ, 79 Nyctobates, 436 Nygmia, 218 Nymphalidæ, 237, 238,251, 253Nymphes, 187 Nymphidæ, 187 Nymphidion, 187 Nymphipara, 311 Nymphula, 216 Nymphulinæ, 216 Nysius, 149

Oarces, 559 Obalonianus, 553 Oberea, 441 Obriminæ, 63 Obrimus, 63 Obrussa, 221 Ochledes, 236 Ochlerotatus, 272

Nysson, 504

Nyssonidæ, 504

Ochodæidæ, 451 Ochodæus, 451 Ochteridæ, 153 Ochterus, **151**, 153 Cchthebius, 414 Ochtheroidea, 343 Ochthiphila, 329 Ochthiphilidæ, 329 Ochyrocera, 559 Ocnæa, **291**, 292 Ocneria, 218 Ocneriidæ, 218 Ocnophila, 64 Ocydromia, 301 Ocydromiinæ, 301 Odagmia, 281 Odiellus, 553 Odinia, 338 Odiniidæ, 338 Odo, 563 Odonata, 159 Odontobracon, 479 Odontoceridæ, 195, 196 Odontocerum, 195 Odontolabis, 450 Odontomachus, 490 Odontomantis, 91 Odontomera, 336 Odontomyia, 284, 286 Odontophyes, 472 Odontopyge, 596 Odontopygidæ, 596 Odontopygideæ, 596 Odontoxenia, 349 Odontoxiphidium 52 Odynerus, 497 Œcanthinæ, 54 Œcanthus, **51**, 54 Œcetis, 194 Œclidius, 119 Œcobiidæ, 557 Œcobius, 557 Ecophora, 229 Œcophoridæ, 229, 232,246 CEcothea, 335, 335;Œdalea, 301 Œdaspis, 341 Œdemagena, 313 Œdemera, 437 Œdemeridæ, 437 Œdipoda, 56 Œdipodinæ, 56

Œdosmylus, 185

Œnochromatidæ, 210

Œneis, 238

Enoe, 222

Enophila, 222 Enophilidæ, 232 Œstridæ, 312, 366 Œstrinæ, 313 Estromyia, 313 Œstrus, 313, **315** Ogmophasmus, 478 Ogovea, 551 Oiclus, 544 Oinophilidæ, 222 Olbiogaster, 275 Olethreutes, 226 Olethreutidæ, 226 Olfersia, 347 Olfersiinæ, 347 Oliarces, 182 Oliarus, 120 Olibrus, 423 Oliera, 224 Oligarces, 275 Oligochætus, 303 Oligochrysa, **186**, 186 Oligoneura, 91 Oligoneuria, 170, **171** Oligoneuriidæ, 170, 173 Oligoneurus, 480 Oligonychinæ, 84 Oligonyx, 84, **85** Oligotoma, **92**, 92 Oligotomidæ, 92 Olina, 337 Olliffia, 132 Olpiidæ, 550 Olpium, 550 Olynthidæ, 92 Omaliinæ, 430 Omaloptera, 311 Omma, 422 Ommatius, 298 Omomantis, 86 Omophron, 411 Omophronidæ, 411 Omosita, 419 Omphalocera, 215 Omphalophora, 291 Omphrale, 294 Omphralidæ, 294 Omus, 410 Oncideres, 441 Oncocephalus, 147 Oncocerus, 451 Oncodes, 292 Oncodesmus, 588 Oncodidæ, 292 Oncodinæ, 292 Oncodocera, 295

Oncometopia, **110**, 111 Oncomyia, 307 Onconotus, 52 Oncopeltus, 149 Oncopodidæ, **552**, 552 Oncopsis, 112 Oncopus, **552**, 552 Oncopygius, 304 Oncozygia, 150 Oneida, 216 Onesia, 316 Oniella, 117 Oniscidæ, 535 Oniscigaster, 172 Oniscodesmidæ, 588 Oniscodesmus, 588 Oniscoidea, 534 Oniscomorpha, 586 Oniscomyia, 350 Oniscosoma, 82 Oniscus, 535 Onthophagus, 450 Onukia, 117 Onychiura, 45, 46 Onychiuridæ, 46 Onychophora, 532 Onychotrechus, 152 Oonopidæ, 557 Conops, 557 Ooperipatus, 532 Oophthora, 486 Ootetrastichus, 486 Oothecaria, 77 Opatridæ, 436 Opharus, 219 Ophiogomphus, 164 Ophion, **482** Ophionellidæ, 477 Ophionellus, 477 Ophionyssus, 568 Ophiuchus, 117 Ophrynopus, 472 Ophyra, 320 Opiinæ, 481 Cpilio, 553 Opiliones, 551 Opilo, 427 Opisthacanthus, 545 Opisthandria, 585 Opisthocheiridæ, 592 Opisthocheiron, 592 Opisthocosmia, 70 Opisthocosmiidæ, 70 Opisthopatus, 532 Opisthophthalmus, 545 Opisthoporodesmus, 588

Opisthoscelis, 132 Opisthothelæ, 555 Opisthotretus, 588 Opogona, 222 Opomyza, 339 Opomyzidæ, 339, 352 Opostega, 221 Opostegidæ, 221, 242 Opsebius, 292 Opsidia, 316 Opsiphanes, 238 Opsolasia, 319 Opsomantis, 90 Orasema, 485 Orchelimum, 52 Crchesella, 46 Orchesia, 438 Orchestina, 557 Orchestinæ, 449 Orectochilus, 412 Orectogyrus, 413 Oreella, 583 Orellia, 341 Oreothalia, 301, **302** Oreta, 221 Orgamara, 120 Orgerius, 120 Orgilus, 480 Orgyria, 218 Oribata, 570 Oribatella, **569**, 570 Oribatidæ, 570 Oribatoidea, 569 Oricia, 210 Ormenis, **118**, 120 Ormothrips, 76 Ormyrus, 485 Ormyridæ, 485 Ornebius, 54 Orneodes, 202, 202 Orneodidæ, 202, 243 Ornithoctonus, 556 Ornithodoros, 567 Ornithœca, 347 Ornithomyia, 347 Ornithomyiinæ, 347 Ornithopertha, 347 Ornithopsylla, 405 Ornithoptera, 236 Ornix, 232 Orobainosoma, 593 Orobainosomatidæ, 593 Orobia, 63 Orocharis, 54 Orodesmus, 589 Oroniscus, 535

Oroperipatus, 533, **533** Orophius, 423, 432 Orothripidæ, 73 Orothrips, 73 Orphnæus, 602 Orphilinæ, 425 Orphnephila, 273 Orphnephilidæ, 273 Orphnidæ, 451 Orphnus, 451 Orsiboe, 597 Orsodacne, 442 Orsodacnidæ, 442 Ortalidæ, 368 Ortalididæ, 329, 331 Ortalis, 331 Orthezia, 127 Ortheziidæ, 127 Orthoceratium, 304 Orthochæta, 319 Orthochile, 303 Orthochordeumella, 593 Orthocladiinæ, 282 Orthocladius, 282 Orthoderella, 86 Orthoderinæ, 87 Ortholasma, **552**, 554 Orthoneura, 308 Orthoperidæ, 431 Orthoperus, 431 Orthopleura, 426 Orthopodomyia, 272 Orthoporus, **594**, 596 Orthoptera, 48 Orthorrhapha, 353, 369, 374Orthosia, 218 Orthosoma, 441 Orthostegana, 345 Orthotælia, 229 Orthotæliidæ, 229 Orussidæ, 472 Orussus, 472 Orya, 602 Orygma, **326**, 327 Oryidæ, 602 Orygminæ, 327 Oryssidæ, 472, 513 Oryssoidea, 472 Oryssus, 472 Osča, 288 Oscinidæ, 340 Oscinis, 340 Oscinosoma, 340 Oscinosominæ, 340 Osmia, 507

Osmoderma, 452 Osmylidæ, 185 Osmylops, 187 Osmylus, 185 Osphya, 438 Ostomatidæ, 419 Otiocerus, **118**, 119 Othniidæ, 435 Othnius, 435 Otiorhynchinæ, 448 Otiothops, 559 Otocryptops, 600 Otomantis, 91 Ourococcus, 134 Oxacis, 437 Oxus, 572 Oxybelidæ, 505 Oxybelus, 505 Oxycera, 286 Oxychila, 410 Oxychiris, 452 Oxychirota, 208 Oxychirotidæ, 202, 208 Oxycoryninæ, 447 Oxycorynus, 447 Oxydactylon, 592 Oxydesmidæ, 589 Oxydesmus, 589 Oxyethira, 192 Oxyhaloa, 82 Oxyhaloidæ, 82 Oxylabis, 495 Oxyomus, 451 Oxyopeidon, 560 Oxyopes, 560 Oxyopidæ, 560 Oxyoposis, 86, 571 Oxyothespinæ, 90 Oxyothespis, 90 Oxypilinæ, 86, 89 Oxypilus, 86 Oxyporinæ, 430 Oxyposcus, 97, 98 Oxyptila, 562 Oxyptilus, 202 Oxytelinæ, 430 Oxytelus, 429 Oxytenidæ, 213 Oxytenis, 212, 213

Pachybolidæ, 595 Pachybolus, 595 Pachybrachys, 443 Pachychelonus, 481 Pachycondyla, 490 Pachydesmus, 590

Pachygaster, 285 Pachymantis, 86 Pachygastrinæ, 285 Pachyloides, 553 Pachylomerus, 556 Pachylomma, 482 Pachylommatidæ, 477 Pachymenes, 497 Pachymerium, 603 Pachymerus, 439 Pachymorpha, 64 Pachynematus, 474 Pachyneres, 296 Pachyneura, 277 Pachyneuridæ, 277 Pachyneuron, 484 Pachyophthalma, 316 Pachypodidæ, 452 Pachypsylla, **122**, 123 Pachypus, 452 Pachyrrhina, 268, 370,373 Pachysima, 490 Pachytarsus, 145 Pachythelia, 207 Pachytroctes, 98 Pachytylus, 56 Pactopus, 421 Pæderinæ, 430 Pagasa, 147 Palæococcus, 128, 129 Palæoplatyura, 279, 280 Palæoses, 201, 201 Palæosetidæ, 201 Palæotropidæ, 238 Palarus, 505 Paleacrita, 209 Palicidæ, 68 Palimpsestia, 211 Palingenia, 169, **174** Palingeniidæ, 169, 173 Palistes, 562 Palloptera, 336, **337** Pallopteridæ, 336 Palophus, 64 Palpares, 187 Palpatores, 552 Palpicornia, 414 Palpigrada, 539 Palpigradi, 539 Palpimanidæ, 559 Palpimanus, 559 Palpinidæ, 553 Palpomyia, 282, 356, **357** Paltostoma, **269**, **27**0 Paltostomatinæ, 270 Pambolinæ, 479

Pambolus, 479 Pammegischia, 476 Pammene, 226 Pamphaginæ, 57 Pamphagus, 57 Pamphila, 236 Pamphiliidæ, 472, 511 Pamphilinæ, 236 Pamphilius, 472, **510** Panacris, 285 Pancalia, 231 Panchætothripidæ, 74 Panchætothrips, 74 Panchlora, 82 Panchloridæ, 82 Pandemis, 226 Pandinus, **544**, 545 Pandora, 322, **324** Pandorinæ, 322 Panesthia, 80 Panesthiidæ, 80 Pangæus, 151 Pangonia, 289 Pangoniinæ, 289 Paniscomima, 498, 399 Panisus, 571 Panopinæ, 292 Panops, 292 Panorpa, 190, **190** Panorpatæ, 189 Panorpidæ, 190 Panorpina, 189 Panorpinæ, 190 Panorpodes, 190 Pantala, 164 Pantarbes, 295 Pantoclis, 495 Pantographa, 216 Pantophthalmidæ, 286 Pantophthalmus, 286, 287 Panurgica, 91 Panurgidæ, 508 Panurginus, 508 Panurgus, 508 Papilio, 236, **237** Papilionidæ, 236, 252 Papilionoidea, 235 Papiriidæ, 46 Papirius, 46 Parablepharis, 87 Parabuthus, 545 Paracalobata, 325 Paracladura, 266, 267 Paraclemensia, 222 Paraclius, 303 Paracodrus, 501

Paracosmia, 70 Paracrias, 487 Paradermaptera, 66 Paradorydium, 113 Paradrapetes, 421 Paradryinus, 493 Paradyschiria, 352 Paragaleodes, 547 Paragia, 496 Paragryllacris, 53 Paragus, 309 Paraiulus, **594**, 595 Parajapyx, 41, 43 Paralamyctes, 600 Paralevrodes, 124 Paralimna, 343 Paralispa, 215 Parallelomma, **315**, 319 Paralogistis, 231 Paramacronychia, 316 Paramacronychiinæ, 316 Paramadiza, 345 Paramasaris, 496 Paramesius, 491 Paramyia, 345 Parandra, 419 Parandridæ, 419 Paraneuroptera, 159 Paranthomyza, 339 Paranthrene, 203 Parantissa, 286 Paraperipatus, 532, **533** Parapsinota, 344 Parapsyllus, 406 Pararctophila, 309 Parascutigera, 599 Parasemia, 219 Parasemidalis, 182 Parasimulium, 281 Parasita, 104 Parasitidæ, 568 Parasitinæ, 568 Parasitoidea, 567 Parasitus, 568, **569** Parasparatta, 69 Parasphæria, 82 Paraspiniphora, 305, 305 Parastenopsyche, 192 Parastylops, 469, 470 Paratettix, 55 Paratiphia, 499 Paratoxopoda, 323 Paratrioza, 123 Paratropididæ, 556 Paratropis, 556 Paravanones, 553

Parcelyphus, 326 Pardosa, 561 Parechiniscus, 583 Parectatosoma, 63 Parectopa, 232 Parepiscopus, 87 Paria, 444 Parisolabidæ, 68 Parisolabis, 68 Parlatoria, 131 Parlatoriinæ, 131 Parnara, 236 Parnassiidæ, 236, 252 Parnassius, 236 Parnidæ, 418 Parnopes, 493 Paromalus, 423 Paropia, 114, 115 Paropiidæ, 114 Parornix, 232 Parorya, 602 Partnunia, 571 Parydra, **342**, 343, 368 Pasimachidæ, 411 Pasites, 509 Paskia, 152 Passalidæ, 449 Passalœcus, 505 Passandra, 424 Passandridæ, 424 Paurocephala, 122, 123 Pauropoda, 583 Pauropodidæ, 584 Pauropsylla, 123 Pauropsyllinæ, 123 Pauropus, 584, 584 Paussidæ, 408 Paussoidea, 408 Paussus, 408 Paxylomma, 482 Paxylommatinæ, 482 Pazius, 189 Pectinophora, 227 Pediasiomyia, 316 Pedicia, 268 Pediciinæ, 268 Pedicinus, 105 Pediculidæ, 105 Pediculoides, 573, **573** Pediculus, 105, **105** Pedilidæ, 439 Pedilophorus, 417 Pedilus, 439 Pedinomma, 492 Pedipalpi, 540 Pedipalpida, 540

Pegomyia, 319, 367, **367** Pegomyiinæ, 319 Pelastoneurus, 303 Pelecinella, 485 Pelecinellidæ, 485 Pelecinidæ, 492, 514 Pelecinus, 492, 493 Pelecocera, 308 Pelecoceratinæ, 308 Pelecorhynchinæ, 288 Pelecorhynchus, 288 Pelecotoma, 438 Pelecotomoides, 439 Peleteria, 318 Peletophila, 336 Pelidnota, 452 Pelina, 343 Pelitnus, 552 Pellenes, 561 Pelmatops, 341 Pelobiidæ, 410 Pelobius, 410 Pelocoris, 154 Pelogonidæ, 153 Pelogonus, 153 Pelomyia, 337 Pelomyza, 327 Pelonium, 426 Pelopia, 282 Pelopiinæ, 282 Pelopœus, 503 Pelops, 570 Pelorempis, 272 Peloridiidæ, 134, 152 Peloridium, 134, 152 Peltacanthina, 331 Peltastica, 425 Peltis, 419 Peltodytes, 410 Pemphegostola, 203 Pemphigella, 126 Pemphigus, 126 Pemphredon, 505 Pemphredonidæ, 505, 506 Penguistus, 352 Penicillidia, 346, 352 Pentacladocera, 470 Pentacladus, 99 Pentacora, 146, **146** Pentagenia, 170 Pentagrammaphila, 470 Pentaneura, 282 Pentaphlebia, 162 Pentarthron, 486 Pentastomida, 579 Pentastomum, 580

Pentatoma, 151 Pentatomidæ, 151 Pentatomoidea, 150 Penthe, 438 Penthesilea, 309 Penthetria, 275 Pentheus, 534 Penthimiidæ, 112 Penthoptera, 268 Pentozocera, 470 Peodes, 304 Peoria, 215 Pepsis, 497 Perasis, 298 Perdita, 508 Perga, 474 Pergidæ, 474, 513 Peribalus, 151 Pericambala, 596 Pericambalidæ, 596 Pericoma, 271, **271** Pericomidæ, 69 Pericomus, 69 Pericopidæ, 217, 250 Peridontodesmidæ, 588 Peridontodesmus, 588 Peridromia, 217 Perientomidæ, 98 Perientomum, 97, 98 Periglischrus, 567 Peringueyomyina, 266 Perilampidæ, 486, 514 Perilampus, 486, **487** Perilestes, 161 Perilitus, 481 Perimede, 233 Periommatus, 445 Peripatidæ, 533 Peripatinæ, 533 Peripatoides, 532, **533** Peripatoidinæ, 532 Peripatopsidæ, 532 Peripatopsinæ, 532 Peripatopsis, 532, 533 Peripatus, 533, **533** Periplaneta, 78, **78** Periplanetinæ, 78 Periscelidæ, 331 Periscelis, 331 Perisierola, 494 Perisphæria, 82 Perisphæriidæ, 82 Peritheates, 270 Perittia, 230 Perkinsiella, 119 Perla, 176, **177** 

Perlamantinæ, 84 Perlamantis, 84 Perlaria, 176 Perlidæ, 176, 178 Perlodidæ, 176 Peronea, 226 Perophora, 213 Perophoridæ, 213 Perreyia, 474 Perreyidæ, 474 Perrisia, 265 Peruda, 148 Petalura, **163**, 164 Petaluridæ, 164, 167 Petaurista, 267 Petauristidæ, 267 Petiolata, 472 Petria, 435 Petriidæ, 435 Petrobius, 42 Petrorossia, 295 Pettalus, 551, **552** Peucetia, 560 Pezomyia, 351 Phægoptera, 219 Phænacantha, 148 Phænocarpa, 477 Phænocephalidæ, 431 Phænocephalus, 431 Phænomeridæ, 452 Phænomeris, 452 Phænopria, 491 Phænoserphus, 492 Phæochrous, 451 Phæodesmus, 589 Phæophasma, 64 Phæophyllacris, 54 Phæoses, 222 Phalacridæ, 423 Phalacrocera, 268 Phalacrus, 423 Phalænoides, 219 Phalænoididæ, 219 Phalangida, 551 Phalangiidæ, **552**, 553 Phalangioidea, 553 Phalangium, 553 Phalangodes, 553 Phalangodidæ, 553 Phalangopsis, 54 Phalera, 211 Phalonia, 227, **247** Phaloniidæ, 227, 245 Phanæus, 450 Phaneroptera, 50 Phaneropterinæ, 50

Phanerotoma, 481 Phantia, 120 Phanurus, 494 Phaonia, 320 Phaoniinæ, 320 Phara, 289 Pharmacis, 227 Pharnacia, 65 Pharyngomyia, 313 Phasgonophora, 485 Phasgonura, 52 Phasgonuridæ, 50 Phasgonurinæ, 52 Phasiidæ, 318 Phasma, 64 Phasmatidæ, 63 Phasmatinæ, 64 Phasmatodea, 30, 61 Phasmatoidea, 61 Phasmida, 61 Phasmodea, 61 Phasmodes, 53 Phasmodidæ, 53 Phasmoidea, 61 Phassus, 201, **201** Phatnoma, 142 Pheidole, 490 Phelister, 423 Phenacaspis, 130 Phenacoccus, 134 Phenacoleachia, 132 Phenacoleachiidæ, 132 Phenacoporus, 588 Phenes, 164 Phengodidæ, 431 Pheomantis, 90 Pherbellia, 328 Phibalosominæ, 64 Phidippus, 561 Philandesia, 102 Philanthidæ, 505 Philanthus, 505 Philharmostes, 451 Philodicus, 298 Philodromus, 562 Philonthus, 429 Philopotamidæ, 195 Philopotamus, 192, 196 Philopotamidæ, 192 Philopteridæ, 103 Philopterus, **102**, 103 Philorus, 270 Philosamia, 213 Philoscia, **534**, 535 Philotarsus, 99 Philpota, 292

Philpotinæ, 292 Philygria, 343 Phisis, 52 Phlebonotus, 79 Phlebotominæ, 271 Phlebotomus, 271, 271 Phlegethontius, 208 Phlepsius, 114 Phlœobiinæ, 430 Phlœocharinæ, 430 Phlæothripidæ, 76 Phleothrips, 76 Phlœotryia, 438 Phlugis, 52 Phlyctænia, 216 Phobetinus, 559 Phobetron, 207 Phœnicococcus, 127 Pholcidæ, 559 Pholcus, 559 Pholeomyia, 345 Pholisora, 235 Phora, 305 Phorantha, 318 Phoraspididæ, 79 Phoraspis, 79 Phorbia, 319 Phoridæ, 305, 350, 364 Phorinæ, 305 Phormia, 314 Phormiinæ, 314 Phorocera, 318 Phorodon, 125 Phorodonta, 278 Phortica, 345 Phorticinæ, 345 Photodes, 99 Photina, 86 Photininæ, 86, 87 Photinus, 431 Photopsis, 500Photuris, 431 Phragmatæcia, 206 Phreatoicidæ, 534 Phreatoicoidea, 534 Phreatoicopsis, 534 Phreatoicus, **534** Phrixothrix, 431 Phronia, 278 Phryacaces, 491 Phryganea, 193 Phryganeidæ, 192, 193 Phryganidia, 207, 210 Phryganoidea, 191 Phryne, 275

Phryneidæ, 275

Phrynichinæ, 542 Phrynichus, 542 Phrynidæ, 542 Phrynus, 542 Phtheochroa, 227 Phthinia, 278 Phthiracarus, 570 Pthiraptera, 104 Phthiria, 296 Phthiriidæ, 105 Phthiriinæ, 296 Phthiripedicinus, 105 Phthirius, 105, **105** Phthorimæa, 228 Phyciodes, 238 Phycita, 215 Phycitinæ, 215 Phycodromia, 327 Phycodromidæ, 327 Phycus, 294 Phylliidæ, 63 Phyllipsocidæ, 98 Phyllipsocus, 98 Phyllium, **62**, 63 Phyllobænus, 426 Phyllobrostis, 223 Phylloceridæ, 428 Phyllocerus, 428 Phyllocnistidæ, 223, 230 Phyllocnistis, 223 Phyllocoptes, 565 Phyllocrania, 87 Phyllodromia, 79 Phyllodromiidæ, 79 Phylloicus, 194 Phyllomimus, 51 Phyllomyza, 345 Phyllonorycter, 232 Phyllophaga, **416**, **437**, 451, **528** Phyllophora, **51**, 51 Phyllophorinæ, 51 Phylloporia, 222 Phyllorheithrus, 193, 194 Phyllorycteridæ, 232 Phyllotoma, 510 Phyllotominæ, 511 Phylloxera, **125**, **127**, 127 Phylloxeridæ, 127 Phylocentropus, 196 Phyma, 120 Phymaphora, **416** Phymata, 147 Phymatidæ, 147 Phymatodes, 441 Phymatostetha, 111

Physiostreptidæ, 597 Physiostreptus, 597 Physocephala, 307, **307** Physoglenes, 559 Physokermes, 132 Physopelta, 150 Physopoda, 72 Physothrips, 74 Phytalmia, 332 Phytalmiidæ, 332 Phytalmodes, **330**, 332 Phyto, 317 Phytodecta, 443 Phytomyza, 339, 339 Phytophaga, 275, 413, 471 Phytorptidæ, 565 Phytoptus, 565 Pialeoidea, 292 Piara, 331 Picobia, 576 Pieltania, 350 Pieridæ, 236 Pieris, 236 Piersigia, 571 Piesma, **141**, 142 Piesmidæ, 142 Piezura, 320 Pigmeophorus, 573 Pigritia, 232 Pilolebertia, 572 Pimeliidæ, 436 Pinconia, 206 Pineinæ, 126 Pineodes, 126 Pineus, 126 Pinnaspis, 130 Pinophilinæ, 430 Pinotus, 450 Piona, 572 Piophila, 334 Piophilidæ, 334, 368 Piophilosoma, 341 Pipiza, 308 Pipunculidæ, 310, 364 Pipunculus, 310, **310**, 364, 365 Pirata, 561 Pirates, 147 Pisaura, 561 Pisauridæ, 561 Pison, 504 Pisonopsis, 504 Pissonotus, 119 Pityogenes, **445** Pityophthorus, 445 Plagiodera, 443

Plagioneurinæ, 303 Plagioneurus, 303 Plastoceridæ, 428 Plastocerus, 428 Plastosciara, 278 Platamodes, 296 Plataspidæ, 150 Plataspididæ, 150 Plataspis, 150 Plateumaris, 441 Plathemis, 166 Plator, 561 Platoridæ, 561 Platybrachys, 121 Platybracon, 478 Platycerus, 450 Platychirus, 309 Platycnemidæ, 161 Platycnemis, 161 Platydascyllus, 418 Platydema, 436 Platydesmidæ, 597 Platydesmus, **594**, 597 Platygaster, 494 Platygastridæ, 494, 501 Plea, 134, 154 Plebejus, 239 Plebejidæ, 239 Plecia, 275, **276** Pleciinæ, 275 Plectoptera, 81, 169, 176 Plectreurys, 559 Plectrocnemia, 194 Pleidæ, 134, 154 Pleistodontes, 483 Plenoculus, 505 Pleocoma, 451 Pleocomidæ, 451 Pleonaraius, 589 Pleisocera, 295 Plethochæta, 319 Plethogenesia, 169 Pleurota, 229 Plicatoperipatus, 533 Plodia, 215 Ploiaria, 147 Ploiariidæ, 147 Plumariidæ, 497 Plumarius, 497 Plusiidæ, 218 Plusiocampa, 44 Plusiotis, 452 Plutella, 229 Plutellidæ, 228, 229 Pneumonyssus, 568, 569 Pneumora, 56

Pneumoridæ, 56 Pnyxia, **348**, 349 Poaphilus, 603 Pococera, 216 Pococerinæ, 216 Podabrus, 431 Podagrion, 485 Podaliriidæ, 508 Podalirius, 508 Podisma, 57 Podisminæ, 57 Podisus, 151 Podocera, 331 Podolestes, 162 Podopidæ, 150 Podops, 150 Podura, 46 Poduridæ, 46 Pœcilocapsus, 145 Pœcilocoris, 150 Pœcilographa, 328 Pecilomigas, 556 Pogonomyia, 320 Pogonomyrmex, 490 Pogonosoma, 299 Pogonostoma, 410 Pogonota, 319 Platylabia, 68 Platylabiidæ, 68 Platymetopius, 114 Platyna, 285 Platyomopsis, 441 Platypalpus, 301, **302** Platyparea, 341 Platypedia, 108 Platypeza, 310, **310** Platypezidæ, 310, 364 Platyphora, 305, 350 Platyphorinæ, 305 Platypodidæ, 445 Platypsyllidæ, 427 Platypsyllus, **422**. 427 Platyptilia, 202, **202** Platypus, 445, **445**, **446**, 448 Platypygus, 296 Platyrhacidæ, 590 Platyrhacus, 590 Platyrrhinidæ, 449 Platysamia, 213 Platysoma, 423 Platystoma, 331 Platystomatidæ, 331 Platystomatinæ, 331 Platystomidæ, 449

Platyura, 280

Platyuridæ, 280 Polia, 218 Poliaspis, 130 Poliinæ, 218 Poliocheridæ, 543 Polistes, 497, **498** Polistinæ, 497 Polistomorpha, 485 Pollenia, 316, **317** Polleniinæ, 316 Polphopeza, 324 Polyaspis, 568 Polybia, 497 Polybiinæ, 497 Polybioides, 497 Polybothrus, 600 Polycentropodidæ, 194, Polycentropus, 194 Polycestinæ, 420 Polychrosis, 226 Polyconoceras, 596 Polyctenes, **153**, 154 Polyctenidæ, 154 Polydesmidæ, 588 Polydesmidea, 587 Polydesmoidea, 587 Polydesmus, 588, **589** Polygnotus, 494 Polygonarea, 603 Polygonia, 238 Polylepis, 590 Polylepta, 277 Polymitarcidæ, 170, 173 Polymitarcys, 170, **174** Polynema, 483 Polyphaga, 82, 408 Polyphagidæ, 82 Polyphylla, 451 Polyplax, 105 Polyplectropus, 194 Polyploca, 211 Polyplocidæ, 211 Polypoda, 532 Polyrhachis, 489 Polyspila, 443 Polyspolota, 89 Polystechotes, 185 Polystæchotidæ, 185 Polythoridæ, 162, 165 Polyxenidæ, 585 Polyxenus, 585 Polyzoniidæ, 597 Polyzonium, 597 Polyzosteria, 78 Pompilidæ, 497

Pompilus, 497 Pomposa, 62, 64 Ponera, 490, **491** Ponerinæ, 490 Ponjadia, 215 Pontania, 474 Pontia, 236 Pontomyia, 347, 349 Popa, 86 Popillia, 452 Poratophilus, 596 Porcellio, **534**, 535 Porina, 201, Porismus, 185 Porocephalidæ, 581 Porocephalinæ, 581 Porocephalus, 581 Porohalacarus, 570 Porsenus, 334 Porthetria, 218 Potamanthidæ, 170, 173 Potamanthodes, 170 Potamanthus, 170, **174** Povilla, 170 Praon, 482 Praos, 70 Prebistus, 63 Prenolepis, 489 Presibylla, 89 Preta, 117 Prexaspes, 64 Prionapteryx, 215 Prionellus, 275 Prionidæ, 441 Prionocyphon, 416 Prionopeltis, 589 Prionopetalum, 596 Prionoxystus, 205, 206 Prionus, 440 Prisopus, 64 Prista, 289 Pristaulacus, 476 Pristocera, 494 Privesa, 121 Procecidochares, 341 Prochilidæ, 50 Prochilus, 50 Prochyliza, 334 Prociphilus, 126 Procladius, 282 Proconia, 111 Proconiidæ, 111 Procris, 207 Proctacanthus, 298 Proctotrypes, 492 Proctotrypidæ, 492

Procubitermes, 95 Procympiutus, 145 Prodicus, 591 Prodida, 562 Prodidomidæ, 562 Prodidomus, 562 Prodoxidæ, 222, 241 Prodoxus, 222, 241 Projapygidæ, 44 Projapyx, 44 Prolabia, 69 Prolammonyx, 602 Prolimacodes, 207 Promachus, 64, 298, 363, 363 Promioptervx, 90 Pronuba, 222 Prophalangopsidæ, 53 Prophalangopsinæ, 50 Prophalangopsis, 50 Propocera, 286 Propomacrus, 452 Propsocus, 99 Propyragra, 67, 68 Prorates, 296 Proreus, 69 Prosacantha, 494 Prosarthria, 55, **55** Proscopia, 55 Proscopiidæ, 55 Prosimuliinæ, 281 Prosimulium, 281 Prosomera, 64 Prosopididæ, 507 Prosopis, 507 Prosopistoma, 173, 174 Prosopistomatidæ, 171, 173Prosopochrysa, 284 Prosopochrysinæ, 284 Prosotropis, 119 Prosparatta, 69 Prostemmiulus, 593, 594 Protapteron, 40 Proteininæ, 430 Protenor, 148 Protenthes, 282, 357, 357 Proterandria, 585 Proterhinidæ, 444 Proterhinus, 444, **448** Protermitidæ, 94, 95 Proteropoides, 481 Proterops, 481 Protobiella, 186 Protocalliphora, 314

Protodermaptera, 66 Protogyropus, 102 Protohermes, 179 Protolophora, 570 Protolophus, **537** Protoneura, 161 Protoneuridæ, 161, 165 Protoparce, 208 Protophormia, 314 Protoplasa, 266, **266** Protoplectron, 187 Protopsychidæ, 213 Protosialis, 180 Protothemira, 323 Prototheora, 201 Prototheoridæ, 201 Protracheata, 532 Protura, 40 Protzia, 571 Protziidæ, 571 Psacaphora, 233 Psalidæ, 68 Psalidia, 288 Psalididæ, 68 Psalidiidæ, 448 Psalis, 68 Psallus, 145 Psammobius, 451 Psammochares, 497 Psammocharidæ, 497, 501 Psarinæ, 308 Psarus, 308 Psechridæ, 558 Psechrus, 558 Psectra, 184 Psednura, 56 Psednurinæ, 56 Psegmomma, 285 Pselaphidæ, 415 Pselaphus, 415 Pselapognatha, 585 Psen, 505 Psenidæ, 505 Psenobolus, 478 Psephenidæ, 427 Psephenops, 427 Psephenus, 427 Psephiocera, 285 Pseudagrion, 161 Pseudatrichia, 294 Pseudechiniscus, 583 Psedectobia, 79 Pseudisolabis, 68 Pseudochelidura, 70 Pseudoclididæ, 592 Pseudoclis, 592

Pseudococcidæ, 134 Pseudococcus, 134 Pseudocorvlophidæ, 434 Pseudocryptops, 600 Pseudodatames, 63 Pseudodinia, 329 Pseudofænus, 476 Pseudogametes, 313 Pseudogametinæ, 313 Pseudohazis, 213 Pseudohydryphantes, 571 Pseudohydryphantidæ, 571Pseudolfersia, 346 Pseudomantis, 90 Pseudomasaris, 496 Pseudomeryle, 62 Pseudomesophalia, 444 Pseudomilichia, 345 Pseudomiopteryginæ, 87 Pseudomiopteryx, 87 Pseudomopidæ, 79 Pseudomops, 79 Pseudomorpha, 409 Pseudomorphidæ, 409 Pseudomusonia, 84 Pseudomyrma, 490 Pseudomyrminæ, 490 Pseudonannolene, 596 Pseudonannolenidæ, 596 Pseudonirvana, 117 Pseudopachymerus, 440 Pseudophæa, 162 Pseudophyllinæ, 51 Pseudophyllus, 51 Pseudopomyza, 344 Pseudopyrochroa, 439 Pseudorhynchota, 104 Pseudoscorpiones, 548 Pseudoscorpionida, 548 Pseudosperchon, 571 Pseudospirobolellus, 595 Pseudostenopsyche, 192 Pseudostigma, 162 Pseudostigmatidæ, 162, 165 Pseudotephritis, 331 Pseudotorrenticola, 572 Pseudotremia, 592 Pseudoxenos, 470 Pseudoxyops, 86 Pseudoxypilus, 86 Pseudoyersinia, 90 Psiathalassius, 297 Psila, 339, 340 Psilidæ, 339

Psilinæ, 340 Psilocephala, 294, 362, **375**, 376 Psilochorema, 193 Psilochorus, 559 Psilocurus, 298 Psiloderces, 559 Psilonyx, 298 Psilopa, 343 Psilopinæ, 343 Psiloplagia, 335 Psilopodinæ, 303 Psilopodinus, 302, 303 Psilopsocus, 99 Psiloptera, 419 Psilosciara, 278 Psilota, 308 Psilotreta, 195, **196** Psithyrus, 507 Psocidæ, 99 Psocinella, 98 Psocoptera, 96 Psocus, 97, 99 Psoquilla, 98 Psorophora, 272 Psoquillidæ, 98 Psoroptes, 574 Psychidæ, 207, 227, 239, Psychoda, 271, 356, 370, 373Psychodidæ, 271, 349, 356 373Psychodinæ, 271 Psychomyia, 194 Psychomyiidæ, 194, 195 Psychophasis, 184 Psychopsella, 184, 184 Psychopsidæ, 184 Psychopsis, 184 Psylla, 123 Psyllidæ, 122 Psyllinæ, 123 Psyllomyia, 305, 350 Psylloneura, 98 Ptecticus, 286 Pterellipsis, 347, 352 Pternaspatha, 281 Pterocalla, **330**, 331 Pterocallidæ, 331 Pterochilus, 497 Pterocolinæ, 447 Pterocolus, 447 Pterocomma, 125 Pterodectes, 574 Pterodela, 99

Pterodontia, 291, 292, 292 Pterolichus, 574 Pteromalidæ, 484, 488 Pteromalus, 484, 484 Pterombridæ, 499 Pterombrus, 499 Pteromicra, 328 Pteronarcidæ, 176, 178 Pteronvarcys, 176 Pteronidea, 474, 510, 510 Pteronus, 474 Pterophoridæ, 202, 249 Pterophorus, 202, 202, 217 Pteroptinæ, 567 Pteroptus, 567 Pterorthochætes, 451 Pterostichus, 410 Pterythysanidæ, 221 Pterothysanus, 220, 221 Pterygida, 70 Pterygophoridæ, 474 Pterygophorus, 474 Pterygosoma, 575 Ptiliidæ, 433 Ptilium, 433 Ptilocera, 285 Ptilodexia, **317**, 318 Ptilonyssus, 567 Ptilosphen 325 Ptinella, 433 Ptinidæ, 427 Ptinobius, 486 Ptinus, 427 Ptiolina, 291 Ptochoryctis, 228 Ptychopoda, 210 Ptychoptera, 267, 267 Ptychopteridæ, 266, 356, 373Ptychopterinæ, 267 Ptyelus, 111 Pucnoscellus, 82 Pulex, 405, **528** Pulicidæ, 405 Puliciphora, 306, 350 Puliciphoridæ, 306 Pulvinaria, 132 Pupipara, 311, 345 Purex, 69 Pycnogaster, 53 Pycnoglossa, 319 Pycnopogon, 298 Pycnothele, 555 Pycnothelidæ, 555 Pycnotropis, 590

Pvgidicrana, 68 Pygidicranales, 66 Pygodicranidæ, 68 Pygidicranoidea, 66 Pygirhynchinæ, 63 Pygirhynchus, 63 Pygodon, 595 Pygostolus, 481 Pygothripidæ, 76 Pygothrips, 75, 76 Pyragra, 68 Pyragridæ, 68 Pyragropsis, 68 Pyralididæ, 204, 214, 243 Pyralidinæ, 215, 216 Pyralis, 215 Pyrameis, 238 Pyrausta, 216 Pyraustinæ, 216 Pyrellia, 321 Pyrgidæ, 236 Pyrgo, 443 Pyrgomantis, 87 Pyrgomorpha, 57 Pyrgomorphinæ, 57 Pyrgota, 330, **330** Pyrgotidæ, 330 Pyrgus, 235 Pyrilla, **118**, 121 Pyrochroa, 439 Pyrochroidæ, 439 Pyroderces, 232 Pyrodes, 441 Pyromorpha, 204 Pyromorphidæ, 204, 208, 249Pyrophæna, 309 Pyrophorus, 421

Pyrophæna, 309 Pyrophorus, 421 Pyrops, 119 Pyrrhocoridæ, 150 Pyrrhocoris, 150 Pyrrhopyge, 235 Pyrrhopyginæ, 235 Pyrtaniinæ, 298 Pyryganeidæ, 196 Pythamidæ, 117 Pythamus, 115, 117 Pythidæ, 437 Pytho, 437

#### Quedius, 429

Ræcius, 561 Raillietiella, **580**, 580 Raillietiellidæ, 579, 580 Ramila, 214

Ranatra, **153**, 154 Raphidia, 181, **181** Raphidiidæ, 181 Raphidiodea, 180 Raphiglossa, 496 Raphiglossinæ, 496 Raphiochæta, 317 Raphiptera, 215 Rapisma, 182 Ratarda, 206 Ratardidæ, 206 Ravinia, 316 Recurvaria, 227 Reduviidæ, 147 Reduvioidea, 145 Reduviolus, 147 Reduvius, 146, 147 Reichertella, 276 Reighardia, 580 Reighardiinæ, 580 Renocera, 328 Reticulitermes, 94, 95 Retinia, 226 Retinodiplosis, 353, **355** Rhabdophaga, 275 Rhabdopselaphus, 296 Rachicerinæ, 289, 374 Rhachicerus, 275, 283, 289, Rhachidomorpha, 588 Rhacodesmidæ, 588 Rhacodesmus, 588 Rhadalidæ, 429 Rhadalus, 429 Rhadinopsylla, 405 Rhadinus, 298 Rhagadochir, 92, 92 Rhagidia, **575**, 576 Rhagigaster, 499 Rhagio, **290**, 290 Rhagionidæ, 290, 362 Rhagium, 441 Rhagodes, 547 Rhagoletis, 341 Rhagophthalmidæ, 431 Rhagophthalmus, 431 Rhagovelia, 152 Rhammatopoda, 51 Rhamphidia, 269 Rhamphomyia, 301, **375**, 376,377Rhampsinitus, 553 Rhanis, 432 Rhantus, 411 Rhaphidolabis, 268 Rhaphidophora, 50

Rhaphidophorinæ, 50 Rhaphidorrhynchus, 447 Rhaphignathidæ, 576 Rhaphignathus, 576 Rhaphiinæ, 304 Rhaphiocera, 286 Rhaphioceratinæ, 286 Rhaphiochætinæ, 317 Rhaphiomydas, 294 Rhaphiorrhynchus, 286 Rhaphium, 304 Rhegmoclema, 276 Rheomantis, 90 Rheumatobates, 152 Rhicnoessa, 337 Rhignogastra, 542 Rhingia, 309 Rhingiopsis, 286 Rhinia, 314 Rhiniinæ, 314 Rhinocola, 123 Rhinocricidæ, 596 Rhinocricus, **589**, **594**, 596 Rhinocypha, 163 Rhinœstrus, 313 Rhinomacer, 449 Rhinomaceridæ, 449 Rhinophora, 317 Rhinophoridæ, 317 Rhinoplus, 481 Rhinopsis, 503 Rhinopsylla, 123 Rhinotermes, 95 Rhinotermitidæ, 95 Rhinotora, 325, 326, 338 Rhinotoridæ, 325, 338 Rhiodinidæ, 239 Rhipicephalus, 566 Rhipicera, 425 Rhipiceratidæ, 425, 428 Rhipiceridæ, 425 Rhipidia, 268 Rhipidius, 439 Rhipidoceridæ, 425 Rhipidothrips, 73 Rhipiphoridæ, 438, 439,  $4\overline{5}2$ Rhipiphorus, 439 Rhipiptera, 467 Rhipipteryx, 54 Rhithrogena, 172 Rhizococcus, 133 Rhizoglyphus, 574 Rhizophagidæ, 419, 436 Rhizophagus, 419 Rhodacarus, 569

Rhodesiella, 345 Rhodites, 483 Rhodoneura, 214 Rhœnanthus, 170 Rhogadinæ, 479 Rhogas, 478, 479 Rhoicinus, 560 Rhopalidiinæ, 497 Rhopalocampa, 235 Rhopalocera, 201 Rhopalodontus, 432 Rhopalomera, 325 Rhopalomeridæ, 325 Rhopalomeris, 587 Rhopalosiphum, 125 Rhopalosoma, 499 Rhopalosomatidæ, 476, 499,514Rhopalosyrphus, 308 Rhopalum, 506 Rhopobota, 226 Rhotana, 119 Rhyacionia, 226 Rhyacophila, 193 Rhyacophilidæ, 193, 195 Rhymbus, 434 Rhymosia, 278 Rhynchitinæ, 447 Rhynchocephalus, 291, 297Rhynchodexia, 318 Rhyncolophidæ, 576 Rhyncholophus, 576 Rhyncophora, 413 Rhynchoprionidæ, 407

Rhynchoproctus, 596 Rhynchota, 106, 140 Rhyncomyia, 314 Rhyncophoridæ, 449 Rhyncophoromyia, 306 Rhyopsocus, 98 Rhyphidæ, 275 Rhyphus, 275 Rhysidia, 600 Rhysodiastes, 411 Rhysodes, 411, **412** Rhysodesmus, 590 Rhysodidæ, 411 Rhyssalus, 479 Ricania, 121 Ricaniidæ, 121 Richardia, 336 Richardiidæ, 336 Ricinidæ, 103 Ricinoides, 543 Ricinoididæ, 543

Ricinulei, 542 Ricinus, 103 Ridiaschina, 223 Ridiaschinidæ, 223 Rileyia, 486 Riodina, 239 Riodinidæ, 239, 252 Riodininæ, 239 Riolus, 420 Ripidosyrma, 293, 294 Risama, 214 Rivellia, 331 Rivelliinæ, 331 Rivetina, 86 Rivetinæ, 86, 88 Rodhainomyia, 313 Rœselia, 219 Rogenhofera, 313 Roncus, 549 Ropalidia, 497 Ropalidinæ, 497 Rophoteira, 404 Ropronia, 492 Roproniidæ, 492 Rothenbuehleria, 592 Rothenbuehleriidæ, 592 Ruba, 286 Ruralidæ, 239 Rutelidæ, 452

Sabacon, 554 Sabatinca, 200 Sabethes, 272 Sabethinæ, 272 Sabine, 207 Sacchiphantes, 126 Saccopheronta, 303 Sacium, 431 Saga, 52 Saginæ, 52 Sagra, **442**, 442 Sagridæ, 442 Saissetia, 132 Salagena, 206 Salda, 146 Saldidæ, 146 Saldula, 146 Salganea, 80 Salomona, 52 Salpingidæ, 437 Salpingus, 437 Saltatoria, 48 Saltelliseps, 322 Salticus, 561 Saltusaphis, 125 Sambonia, 581

Samia, 213 Samichus, 596 Sandalidæ, 425 Sandalus, 425 Saperda, 414, 414 Sapho, 162 Saprininæ, 423 Saprinus, 423 Sapromyza, 327 Sapromyzidæ, 327 Sapromyzosoma, 327 Saprosites, 451 Sapyga, 500 Sapygidæ, 500 Sarax, 542 Sarcophaga, 316, 366, **367**, 367,377Sarcophagidæ, 316, 366, 367Sarcophila, 316 Sarcopsylla, 407 Sarcopsyllidæ, 407 Sarcoptes, 574 Sarcoptidæ, 574 Sarcotachina, 316 Sarginæ, 286 Sargus, 286 Saronomus, 547 Saropogon, 299 Saroxenus, 585 Sarrothripus, 219 Sason, 556 Satorystia, 425 Saturnia, 213 Saturniidæ, 213, 251 Saturniinæ, 251 Satyridæ, 238, 249, 253 Satyrodes, 238 Satyrus, 238 Sayomyia, 354, **355**, 372 Scalidia, 424 Scalidiidæ, 424 Scaphidiidæ, 426, 428 Scaphidium, 426 Scapheutes, 505 Scaphiella, 557 Scaphiostreptus, 596 Scaphoideus, 114 Scaphosoma, 426 Scapteriscus, 54 Scaptolenus, 428 Scaptomyza, 345, 368, 368 Scarabæidæ, 450 Scarabæus, 450 Scardia, 224, 244 Scarites, 410, **412** 

Scarphia, 288 Scarphiinæ, 288 Scatella, 343 Scatophaga, **315**, 319 Scatophagidæ, 319 Scatophila, 343 Scatophora, 337 Scatomyzidæ, 319 Scatopse, **276**, 276, 358, Scatopsidæ, 349, 358, 373, Scatopsinæ, 276 Scelio, 494 Scelionidæ, 494, 501, 514 Sceliphron, 503 Scellus, 304 Scenopinidæ, 294, 362, 376 Scenopinus, 294, **297** Scepsis, 219 Schænophilus, 304 Schauinslandia, 480 Schedoleiodesmus, 588 Schedorhinotermes, 95 Schedotrigona, 593 Schendyla, 602 Schendylidæ, 602 Schendylurus, 602 Schindalmonotus, 585 Schistocerca, 57 Schizaspidia, 485 Schizocarpus, 574 Schizocephala, 86 Schizocephalinæ, 86, 87 Schizocera, 474 Schizomidæ, 541 Schizomus, 541, **541** Schizoneura, 126 Schizopinæ, 420 Schizopteridæ, 143 Schizotus, 439 Schizura, 211 Schoenbaueria, 281 Scheenobiinæ, 214 Scheenobius, 214 Schenomyza, 320 Scheenophilus, 350 Schreckensteinia, 231 Scholastes, 331 Sciara, 278, 278, 358, 371, 372Sciaridæ, 278, 349, 358, 371Sciarinæ, 278 Sciomyza, 328, **328** 

Sciomyzidæ, 328

Sciomyzinæ, 328 Scione, 289 Sciophila, 277, 278 Sciophilidæ, 277 Sciopus, 303 Scipopeza, 324 Scipopus, 325 Scirtes, 418, 421 Scirtotypus, 56 Scirula, 576 Sciadocera, 300, **310** Sciadoceratidæ, 300 Sciapodinæ, 303 Sclerobunus, 552 Scleroderma, 494 Sclerogibba, 492 Sclerogibbidæ, 492, 500 Scleropauropus, 584 Scolia, 498 Scoliaula, 221 Scoliidæ, 498, 514 Scoliocentra, 335 Scolioneurinæ, 512 Scolioplanes, 603 Scolopendra, 600, **601** Scolopendrella, 604 Scolopendrellidæ, 604 Scolopendridæ, 600 Scolopendromorpha, 600 Scolopendropsis, 604 Scolopocryptops, 600 Scolops, 118, 120 Scolytidæ, 446 Scolytoidea, 444 Scolytoplatypodidæ, 446 Scolytoplatypus, 446, 446 Scolytopsis, 446 Scolytus, 446 Scoparia, 216 Scopariinæ, 216 Scopeuma, 319 Scopeumidæ, 319 Scopeuminæ, 319 Scorpio, 545 Scorpiones, 543 Scorpionida, 543 Scorpionidæ, 544, 545 Scorpiops, 546 Scotolathys, 558 Scotolemon, 553 Scraptia, 438 Scraptiidæ, 438 Scrupulaspis, 130 Scudderia, 50 Scutacarus, 573 Scutare, 131

Scutellera, 150 Scutelleridæ, 150 Scutelleroidea, 150 Scutigera, 599 Scutigera, 601 Scutigerella, 601, 604 Scutigerellidæ, 604 Scutigeridæ, 599 Scutigeromorpha, 599 Scutogona, 591 Scutops, 331 Scymænidæ, 428 Scydmænus, 428 Scymnus, 434 Scyphacella, 534 Scyphacidæ, 534 Scyphax, 534 Scythridæ, 231 Scythrididæ, 231, 250 Scythris, 230, 231 Scytodes, 559Scytodidæ, 559 Scytonotus, 589 Sebekia, **580**, 581 Sebekiinæ, 581 Sedulothrips, 76 Segestria, 557 Selachops, 343 Selandria, 474 Selandriinæ, 512 Selasius, 431 Selenocosmia, 556 Selenopidæ, 562 Selenops, 562Selidopogon, 299 Selidosema, 209 Selidosematidæ, 209 Sematura, 209 Sematuridæ, 209 Semidalis, 182 Seminota, 494, **495** Semioptila, 204 Semiotellus, 486 Semiothisa, 209 Semiramis, 296 Semniinæ, 215 Semnosoma, 589 Senoculidæ, 560Senoculus, 560 Senotainia, 316 Seoptera, 330 Sepedon, **328**, 328 Sepsidæ, 322 Sepsidomorpha, 323 Sepsinæ, 323 Sepsis, 323, **324** 

Serianus, 550 Serica, 451 Sericomyia, 309 Sericomyiinæ, 309 Sericophorus, 505 Sericosoma, 295 Sericostomatidæ, 192, 194, 197Serinetha, 148 Serphidæ, 492, 501 Serphoidea, 514 Serphus, 492, 493 Serropalpidæ, 438 Serropalpus, 438 Sesiidæ, 203 Sessiliventres, 471 Sessinia, 437 Setellia, 336 Setiostoma, 228 Setodes, 194 Setomorpha, 223, 224 Setomorphidæ, 224 Shravana, 550 Sialidæ, 180 Sialis, 180, **528** Sialoidea, 179 Sibylla, 89 Sibyllinæ, 89 Sicariidæ, 559 Sicarius, 559 Sicus, 307 Sierola, 494 Sierolomorpha, 500 Sierolomorphidæ, 500 Sigaloessa, 332, 340 Sigalphinæ, 481 Sigalphus, 481 Sigerpes, 91 Siginæ, 214 Signiphora, 484 Signiphoridæ, 484 Signoretia, **115**, 117 Signoretiidæ, 117 Sigrisma, 442 Silpha, 428 Silphidæ, 428 Silphomorpha, 409 Silvanidæ, 424 Silvanus, **422**, 424 Silvestria, 593 Silviinæ, 288 Silvius, 288 Simaethidæ, 228 Simaethis, **205**, 228 Simuliidæ, 280, 355, 371 Simuliinæ, 281

Simulium, **265** 281, 355, 371, **528** Sinea, 147 Sinella, 46 Sinodendridæ, 450 Sinodendron 450 Sinophthalmus, 345 Siphlonisca, 172Siphlonuridæ 172 Siphlonurus, 172 Siphluridæ, 172, 175 Siphlurus, 172 Siphona, 318 Siphonaptera, 404 Siphoniulidæ, 597 Siphoniulus, 597 Siphlopteryx, 350 Siphonocryptidæ, 597 Siphonocryptus, 597 Siphonophora, 125, Siphonophoridæ, 597 Siphonorhinus, 597 Sipnonotus, 597 Siphunculata, 104 Sipyloidea, 64 Sira, 46 Sirex, 473 Siricidæ, 473, 513 Siricoidea, 472 Siro, 551 Sironidæ, 551 Sisyra, 185, **186** Sisyridæ, 185 Sisyrella, 185 Sitodrepa, 426 Sitones, 448 Sitophilus, 448 Sitotroga, 227 Skalistes, 70 Smaridæ, 576 Smaris, **575**, 576 Smerinthus, 208 Smicra, 485 Smicrides, 194 Smicriptini, 435 Sminthuridæ, 46 Sminthurinus, 46 Sminthurus, 45, 46 Sobarocephala, 334, 335 Soerensenella, 552 Sogata, 113 Sogona, 602 Sogonidæ, 602 Soldanellonyx, 570 Solenius, 506

Solenobia, 205, **208** Solenopsis, 490 Solenosoma, 69 Solifugæ, 546 Solinus, 550 Solpuga, 547, **547** Solpugida, 546 Solpugidæ, 547 Solva, 289 Solygia, 90 Solygiinæ, 90 Somabrachys, 208 Somatochlora, 164 Soniphilidæ, 603 Soniphilus, **601**, 603 Sophronia, 228 Sostea, 418 Spadobius, 184 Spalangia, 484 Spalangiidæ, 484 Spania, 291 Spaniocelyphus, 326 Spaniophlebia, 170 Spaniotoma, 282 Sparassidæ, 562 Sparatta, 69 Sparattidæ, 69 Sparganothidæ, 226 Sparganothis, 226 Sparianthis, 562 Sparnapolius, 297, 363, **375**, 376, 377 Spathlinæ, 478 Spathiogaster, 309 Spathiophora, 319 Spathius, 478 Spelæoblatta, 78 Spelæorhynchidæ, 567 Spelæorhynchus, 567 Spercheinæ, 414 Sperchon, 571 Sperchonidæ, 571 Spermophagus, 439 Spermophora, 559 Sphæricus, 427 Sphæridium, 414 Sphæriidæ, 433 Sphæriodesmidæ, 590 Sphæriodesmus, 586, 590 Sphæristes, 437 Sphærites, 427Sphæritidæ, 427 Sphærius, 433 Sphærocera, 337, **337** Sphæroceridæ, 337 Sphærodema, 154

Sphæroniscus, 534 Sphærophoria, 309 Sphærophthalma, 498 Sphæropæus, 586 Sphæropyx, 481 Sphærosoma, 434 Sphærotheria, 586 Sphærotheriidæ, 586 Sphærotherium, 586, 586 Sphærotrichopidæ, 589 Sphærotrichopus, 589 Sphallomorpha, 409 Sphecidæ, 503 Sphecius, 503 Sphecodes, 508 Sphecoidea, 502, 514 Sphecomyia, 309 Sphegina, 309 Spheginæ, 309 Sphenophorus, 449 Sphenopterinæ, 420 Sphex, 503 Sphinididæ, 436 Sphindus, 436 Sphingidæ, 208, 252 Sphinx, 208 Sphodromantis, 89 Sphodropoda, 89 Sphyracephala, 329 Sphyroperiscelis, 331, 333 Sphyximorpha, 308 Spilochalcis, 485 Spilochroa, 335 Spilogaster, 320 Spilomicrus, 491 Spilomyia, 309 Spilonota, 226 Spilosmylus, 185 Spinaria, 479 Spinotarsus, 596 Spintharus, 559 Spinturnicidæ, 567 Spinturnix, 567, **569** Spirobolellus, 595 Spirobolidæ, 595 Spiroboloidea, 595  ${f Spirobolus}, 595$ Spiromimidæ, 595 Spiromimus, 595 Spirostreptoidea, 596 Spirostreptidæ, 596 Spirostreptomorpha, 595 Spirostreptus, 596 Spogostylum, 294 Spongifera, 69 Spongiphoridæ, 69

Spongovostox, 69 Sponidium, 438 Stachyocnemus, 148 Stagmatophora, 233 Stagmatoptera, 86 Stagmomantis, 85, 89 Staianus, 562 Stalita, 557 Staphylinidæ, 429 Staphyliniformia, 415 Staphylininæ, 429 Staphylinus, **418**, 429 Statira, 436 Stauronotus, 56 Stauropus, 211 Steatococcus, 128 Steatoda, 559 Stegana, 345 Steganinæ, 345 Steganopsis, 327 Stegodyphus, 558 Stegomyia, 272 Stegopterna, 281 Stegopterninæ, 281 Steingelia, 128, **133** Steingeliinæ, 128 Stelididæ, 508 Stelis, 508 Stemmiulidæ, 593 Stemmiuloidea, 593 Stemmiulus, 593 Stenelmis, 420 Stenichnus, 428 Steninæ, 429 Steniolia, 505 Stenobiella, 186 Stenobothrus, 56 Stenocephalus, 148 Stenochilus, 559 Stenocotidæ, 115 Stenocotis, 115 Stenocranus, 119 Stenogaster, 497 Stenogastrinæ, 497 Stenolæmus, 147 Stenoma, 228 Stenomacra, 336 Stenomatidæ, 228 Stenometopiinæ, 116 Stenometopius, 115, 116 Stenomicra, 332 Stenomidæ, 228, 246 Stenopelmatidæ, 49 Stenopelmatinæ, 50 Stenopelmatus, 49, 50 Stenoperla, 176, 177

Stenophragma, 277 Stenophylax, 192 Stenophylla, 86 Stenopilema, 82 Stenopogon, 298 Stenopsyche, 192 Stenopsychidæ, 192, 194, 195Stenopsychodes, 194 Stenopteridæ, 74 Stenopterina, 331 Stenopterinæ, 331 Stenopteryx, 347 Stenosialis, 180 Stenotabaninæ, 287 Stenotabanus, 287 Stenotortor, 116 Stenovates, 86 Stenoxenidæ, 282 Stenoxeninæ, 282 Stenoxenus, 281, 282 Stenus, 429 Stephanidæ, 476 Stephaniscinæ, 478 Stephaniscus, 478 Stephanopsis, 562 Stephanothrips, 74 Stephanus, 476 Stereopalpus, 439 Sternorrhyncha, 107 Steropleurus, 53 Sterrhidæ, 210 Stethopathidæ, 306, 350 Stethopathus, 306 Sthenopis, 201, **241** Stibadocera, 268 Stichopogon, 298, 298 Stichotrema, 469 Stichotrematidæ, 469 Stichotrematoidea, 469 Stictia, 505 Stigmacoccus, 128, 133 Stigmæus, 576 Stigmatomma, 490 Stigmatopathus, 98 Stigmellidæ, 221 Stigmodera, 419 Stigmoderinæ, 420 Stigmus, 505Stilbopterygidæ, 187 Stilbopteryx, 187 Stilbula, 485 Stilbus, 423 Stilpon, 301 Stilpnochlora, 49 Stilpnotia, 218

Stiphidion, 558 Stiphrosoma, 339 Stizidæ, 503 Stizus, 503 Stolidosoma, 304 Stolidosomatinæ, 304 Stomacoccus, 128 Stomatorhina, 314 Stomatothrips, 73 Stomoxydinæ, 321 Stomoxys, 317, 321 Stonyx, 295 Storena, 560 Strangalia, 441 Strathmopoda, 231 Stratioleptinæ, 289 Stratioleptis, 289 Stratiomyia, 286, 360, **361** Stratiomyiidæ, 284, 360, 374Stratiomyiinæ, 286 Stratiomys, 286 Stratocles, 64 Straussia, 341 Strebla, 346 Streblidæ, 347, 352 Strepsimana, 228 Strepsimanidæ, 228 Strepsiptera, 467 Striaria, 590 Striariidæ, 590 Striarioidea, 590 Striglina, 214 Strobliella, 275 Strongylium, 436 Strongylocephalus, 114 Stronglylodesmus, 588 Strongylophthalmyia, 340 Strongylophthalmyiinæ, 340 Strongylopsalididæ, 69 Strongylopsalis, 69 Strongylosoma, 589 Strongylosomatidæ, 589 Strongylosomatidea, 587 Strophius, 562 Strymon, 239 Sturmia, 318 Stygophrynus, 542 Stylipauropus, 584 Stylocellus, 551 Stylodesmidæ, 588 Stylodesmus, 588 Stylogaster, 306, 324

Stylogastrinæ, 306

Stylophthalmyia, 331

Stylopidæ, 470 Stylops, **469**, 470 Subtriquetra, 581 Subulonia, **283**, 289 Suctoria, 404 Suhpalacsa, 187 Suillia, 334, **335** Suilliinæ, 334 Suturaspis, 131 Suturodes, 601 Suva, 119 Swammerdamella, 276 Syarinidæ, 549 Syarinus, 549 Sybota, 558 Sycophaga, 487 Sycorax, 271, 271 Sylepta, 216 Symmerus, 280 Symmictus, 291 Symmoca, 228 Symmorphus, 497 Symperipatus, 532 Sympetrum, 164 Sympherobiidæ, 184 Sympherobius, 184 Symphoromyia, 291 Symphrasis, 183 Symphyla, 603 Symphylella, 604 Symphypleona, 45 Symphyta, 471 Sympyona, 162 Sympycnus, 304 Synæma, 562Synagris, 497 Synallagma, 233 Synamphotera, 301 Syncalypta, 417 Synchloe, 236 Syndyas, 302 Syneches, 302 Synemon, 203 Syneura, 306 Syngenaspis, 131 Syngonopodium, 592 Synistata, 181 Synlestes, 161, **166** Synlestidæ, 161, 165 Synneuron, 276 Synoptura, 588 Synsphyronus, 550 Syntelia, 426 Synteliidæ, 426 Syntelopodeuma, 593 Synthesiomyia, 321

Syntomidæ, 219 Syntomididæ, 219 Syntomis, 219 Syntormon, 304Syntormoneura, 304 Synuropus, 535 Syringogaster, 328, 329 Syritta, 309 Syrittomyia, 329 Syrphidæ, 308, 365 Syrphinæ, 309 Syrphus, **307**, 309, **365**,365 Syssphingidæ, 211 Syssphinx, 211 Systelloderes, 147 Systenus, 304 Systechus, 297 Systropinæ, 296 Systropus, 296 Syzeton, 439

Tabanidæ, 287, 360, 374 Tabaninæ, 288 Tabanus, **283**, **287**, 288 Tabaria, 51 Tabuda, 294 Tæniochorista, 191 Tæniopterna, 281 Tæniopterygidæ, 178 Tæniorhynchus, 272 Tæniostigma, 99 Tachardia, 132 Tachardiella, 132 Tachardiidæ, 132 Tachardina, 132 Tachina, 318 Tachinidæ, 318, 366 Tachinisca, 332, **333** Tachiniscidæ, 332 Tachiniscidia, 332 Tachinæstrus, 332 Tachinoidea, 313 Tachopteryx, 164, **166** Tachydromia, 300 Tachydromiinæ, 301 Tachyempis, 301 Tachygoninæ, 447 Tachygonus, 447 Tachypeza, 301 Tachyporinæ, 429 Tachyporus, 429 Tachypus, 410 Tachys, 410 Tachysphex, 505 Tachytes, **504**, 505 Tachytrechus, 303

Tagiades, 235 Talæporia, 205 Talæporiidæ, 205, 227,239 245Tama, 560 Tambinia, 120 Tanaostigmatidæ, 484 Tanyderidæ, 266 Tanyderus, 266 Tanypeza, 324, **324** Tanypezidæ, 324 Tanypoda, 325 Tanypodinæ, 282, 325 Tanypremna, 268 Tanypus, 282, 357, **357** Tanytarsus, 282 Tapinogalos, 494 Tapinopa, 560 Tapponia, 560 Tarachina, 88 Tarachodes, 87 Tarachodinæ, 87 Tarachodula, 87 Taracticus, 299 Taracus, 554 Tarantula, 542 Tarantulidæ, 542 Tardigrada, 581 Targionia, 130 Tarraga, 50 Tarsonemidæ, 573 Tarsonemus, 573 Tarsotomus, 575 Tascina, 203 Tascinidæ, 203 Tasmaniella, 604 Tasmanoperla, 176 Tegenaria, **536**, 560 Tegeticula, 222, **223** Tegonotus, 565 Teichomyza, 343 Teichophrys, 56 Telamona, 108 Telea, 213 Teleas, 494 Teleconidæ, 236 Telegeusidæ, 430 Telegeusis, 430 Telegonus, 235 Telema, 557 Telemidæ, 557 Telenomus, **493**, 494 Telephoridæ, 431 Telephorus, 431 Telethera, 225 Telmatogeton, 282

Telmatoscopus, 271 Teloganodes, 172 Telomantis, 87 Telostylus, 325 Telphusa, 228 Temnaspis, 443 Temnocera, 309 Temnochila, 419 Temnochilidæ, 419 Temnochilus, 416 Temnopteryx, 79 Temnostoma, 309 Tenaga, 224 Tendipedidæ, 282 Tendipedinæ, 282 Tendipes, 282 Tenebrio, 433, 436 Tenebrioides, 419, 421 Tenebrionidæ, 436 Teneriffa, 303 Teneriffia, 576 Teneriffiidæ, 576 Tengella, 558 Tengellidæ, 558 Tenodera, 89 Tenthredella, 474 Tenthredinidæ, 474, 511 Tenthredininæ, 512 Tenthredinoidea, 472 Tenthredo, 474 Tentyria, 436 Tenuopus, 303 Tephritidæ, 340 Tephritinæ, 341 Tephritis, 341 Tephrochlamys, 335 Tephroclystis, 210 Tephromyia, 316 Tephronota, 331 Teragra, 206 Teragridæ, 206 Terastiomyia, 332 Teratembia, 92 Teratembiidæ, 92 Terebrantia, 73, 474 Terellia, 341 Termatophylidæ, 144 Termatophylum, 144 Termes, **94**, 95 Termitadelphus, 271, 349 Termitaphididæ, 154 Termitaphis, 154 Termitaradus, 153, 154 Termitidæ, 95 Termitocoridæ, 154 Termitocoris, 154

Termitodesmus, 586 Termitodipteron, 349 Termitomastinæ, 274, 348 Termitomastus, 274, 348. 348 Termitomyia, 349 Termitoxenia, 349, **351** Termitoxeniidæ, 349 Termopsis, 95  ${
m Tesserocerus}, 445$ Tessinosoma, 591 Tetanocera, 328 Tetanoceratidæ, 328 Tetanoceratinæ, 328 Tetanops, 331 Tetanura, 328 Tetanurinæ, 328 Tethina, 337 Tethinidæ, 337 Tetrabalius, 541 Tetracanthina, 286 Tetracha, 410 Tetraconus, 477 Tetragnatha, 560 Tetragoneuria, 164 Tetrakentron, 583 Tetralopha, 216 Tetranychidæ, 576 Tetranychus, 537, 575,576 Tetraopes, 441 Tetrapus, 483 Tetrastichidæ, 486 Tetrastichus, 486 Tetrasticta, 556 Tetratoma, 438 Tetrigidæ, 55 Tetrix, **55**, 55 Tetrodontophora, 46 Tetrozocera, 468 Tettigidæ, 55 Tettigidea, 55, **55** Tettigometra, 117 Tettigometridæ, 117 Tettigoniellidæ, 111, 113 Tettigoniidæ, 50, 111 Tettigoniinæ, 52 Tettigoniodea, 48 Tettigoxenos, 470 Teucholabis, 269 Teuthraustes, 545 Teutonia, 571 Thalperus, 70 Thalassius, 561 Thamnotettix, 114 Thamyrididæ, 235 Thanaos, 235

Thanasimus, 427 Thanatidius, 561 Tharsalea, 239 Thaumalea, 273, 274 Thaumaleidæ, 273, 356 Thaumasia, 561 Thaumaspis, 51 Thaumastocoridæ, 146 Thaumastophila, 344 Thaumastoscopidæ, 112 Thaumastotheriidæ, 146 Thaumatoneura, 162 Thaumatopsis, 216 Thaumatoxena, 350, 351 Thaumatoxenidæ, 350 Thaumetopæa, 211 Thaumetopœidæ, 211 Thecesterninæ, 449 Thecesternus, 449 Thecla, 239 Thecomyia, 307 Thecostomata, 311 Theisoa, 233 Thelaira, 318 Thelaxes, 125 Thelaxinæ, 125 Thelyphonellus, 541 Thelyphonidæ, 541 Thelyphonus, **537**, 541 Thermacarus, 570 Themacrys, 558 Themira, 323, 324 Themirinæ, 323 Theobaldia, 272 Theomantis, 91 Theopompa, 87 Theopompula, 87 Theraphosa, 556 Theresia, 318 Thereuonema, 599 Thereuopoda, 599 Thereva, 283, 293, 294 Therevidæ, 294, 362, 376 Theridiidæ, 559 Theridion, 559 Theridiosoma, 560 Therioplectes, 288 Thermacaridæ, 570 Thermobia, 42, 44 Thespinæ, 87 Thespis, 87 Thessitus, 121 Thestylus, 543 Thevenetimyia, 296 Thinophilinæ, 304 Thinodromia, 350

Thinophilus, 304 Thiotricha, 228 Thomisidæ, 562 Thoracantha, 485 Thoracophoracarus, 571 Thore, 162 Thorictidæ, 426 Thorictus, 426 Thraulus, 172 Thrincopyginæ, 420 Thripidæ, 74 Thripoidea, 73 Thripomorpha, 348 Thrips, 72, 73, 74 Throscidæ, 421 Throscus, 421 Thrypticus, 303 Thyas, 571 Thyatira, **212**, 216 Thyatiridæ, 211, 216, 247, Thyllis, **291**, 292 Thymalus, 419 Thymele, 235 Thymelicus, 236 Thymelidæ, 236 Thynnidæ, 499, 502 Thyreocoridæ, 150 Thyreocoris, 150 Thyreophora, **337**, 341 Thyreophoridæ, 341 Thyridanthrax, 295 Thyrididæ, 214, 245 Thyridopteryx, 209, 247 Thyris, 214 Thyropygus, 596 Thyrsophoridæ, 99 Thyrsophorus, 97, 99 Thysanoptera, 72 Thysanura, 41 Tibicen, 108 Tillus, 427 Timarcha, 443 Timena, 64 Timeodes, 196 Timia, 330 Timomenus, **67**, 70 Timpina, 602 Tinægeriidæ, 231 Tinagma, 230 Tinda, 285 Tinea, 223, 224 Tineidæ, 222, 224, 244

Tineinæ, 215

Tineola, 224

Tineoidea, 205

Tineomorpha, 98 Tingidæ, 142 Tingididæ, 142 Tingidoidea, 142 Tingitidæ, 142 Tinodes, 194 Tiphia, 499 Tiphiidæ, 499, 514 Tipula, **267**, 268, 354, **528** Tipulidæ, 267, 348, 354, Tipulinæ, 268 Tipuloidea, 266 Tisamenus, 63 Tischeria, 231 Tischeriidæ, 231, 242 Titanodamon, 542 Titanœca, 558 Tithanetes, 534 Tithraustes, 210 Tithrone, 86 Tityus, 545 Tmetocera, 226 Tolida, 438 Tolmerus, 298 Tolype, 221 Tomaspididæ, 110 Tomaspis, 110 Tomicomerus, 554 Tomocerus, 45, 46 Tomoderus, 439 Tomomyza, 295 Tomomyzinæ, 295 Tomopyga, 68 Tomoxia, 438 Toosa, 207 Tortricidæ, 225, 239, 246 Tortrix, 226 Torymidæ, 485 Torymus, 485 Toumeyella, 132 Toxocerus, 451 Toxodera, **85**, 86 Toxoderinæ, 86 Toxoneuron, 480 Toxophora, 296 Toxophorinæ, 296 Toxopoda, 323 Toxopodinæ, 323 Toxoptera, 125 Toxotrypana, 341 Trachelas, 563 Trachelizus, 447 Trachelomegalus, 595 Tracheoniscus, 535 Trachusa, 507

Trachviulidæ, 597 Trachviulus, 597 Trachypus, 505 Trachysoma, 590 Trachysomatidæ, 590 Trachytes, 568 Trachyzona, 590 Traginops, 338 Tragocephala, 441 Trama, 124 Tramea, 164 Traphera, 331 Trapherinæ, 331 Traumatomutilla, 500 Trechona, 556 Trechus, 411 Tremex, 473, 473, 510 Trepidaria, 325 Triaspidinæ, 481 Triaspis, 481 Triatoma, 146, 147 Tribæus, 485 Tribolium, 436 Tricentrus, 108 Trichardis, 298 Trichiidæ, 452 Trichilogaster, 486 Trichina, 301 Trichiosoma, 239, 473 Trichius, 452 Trichobius, **346**, 347 Trichocera, 267, 358, **359** Trichoceratidæ, 267, 358 Trichodectes, 103 Trichodectidæ, 103 Trichodes, 427 Trichœcius, 574 Trichogramma, 482, 486 Trichogrammatidæ, 486 Trichoma, 186, 186 Trichomatidæ, 186 Trichomyia, 271 Trichomyiinæ, 271 Trichoniscidæ, 534 Trichoniscus, 534 Trichonta, 278 Trichopetalum, 593 Trichopeza, 301 Trichophaga, 224 Trichophilopteridæ, 103 Trichophilopterus, 103 Trichophyinæ, 429 Trichopoda, 318 Trichopolydesmus, 588 Trichopria, 491 Trichoproctus, 585

Trichoptera, 191 Trichopterygidæ, 433 Trichopteryx, 210, 433 Trichoscelidæ, 335 Trischoscelis, 324, 335, 335, 339 Trichotaphe, 228 Trichotarsus, 574 Trichothrips, 76 Tricladellus, 99 Tricliona, 444 Triclis, 298 Triclonus, 297 Tricondyla, 410 Tricopalpus, 319 Tricoryphodes, 170 Tricorythus, 171, **174** Trictenotoma, 438 Trictenotomidæ, 438 Tricyphona, 268 Tridactylidæ, 54 Tridactylus, 53, 54 Tridymidæ, 486 Trifurcula, 221 Trigonalidæ, 476, 494 Trigonalys, 494 Trigonidiinæ, 54 Trigonidium, 54 Trigoniulidæ, 595 Trigoniulideæ, 595 Trigoniulus, 595 Trigonocryptops, 600 Trigonoderus, 487 Trigonometopidæ, 327 Trigonometopus, 327 Trigonostylus, 588 Triænonychidæ, 552 Triænonyx, 552 Trimenopon, 102 Trimenoponidæ, 102 Trimera, 97 Trimeria, 496 Trimerina, 343 Trimerophoron, 592 Trimerotropis, 56 Trimicra, 268 Trineura, 305 Trinodinæ, 425 Trinoton, 103 Triogma, 268 Trionycha, 557 Trionymus, 134 Trionyxella, 552 Trioxys, 482 Trioza, **122**, 123 Triozinæ, 123

Triozocera, 468, 469 Triphæna, 218 Triphleps, 144, 144 Triplasius, 297 Triplax, 432 Tripomorpha, 349 Triptotricha, 290 Trirhabda, 444 Tristanella, 70 Tristega, 475 Trithyreus, 541 Tritoma, 432 Tritomidæ, 432 Tritoxa, 331, 367, 368 Trixagidæ, 421 Trixoscelidæ, 335 Trizetes, 570 Trochilium, 203 Trochilecetes, 103 Troctes, 97, 98 Troctidæ, 98 Trogidæ, 450 Trogiidæ, 98 Trogium, 98 Troglophilus, 50 Trogoderma, 425 Trogositidæ, 419 Trogulidæ, 552, 554 Trogulus, 554 Troides, 236 Trombidiidæ, 575 Trombidium, 575, 575 Trombidoidea, 574 Tropæa, 213 Tropidia, 309 Tropidomyia, 307 Tropiduchidæ, 120 Tropiduchus, 120 Tropisternus, 414 Tropusia, 98 Trotomma, 438 Trotommidea, 438 Trox, 450 Trullifiorinia, 130 Truxalinæ, 56 Truxalis, 56 Trycopeplus, 62 Trymaltis, 226 Trypanæinæ, 423 Trypanea, 341 Trypaneidæ, 340 Trypanus, 206 Trypeta, 341 Trypetidæ, 340, 368 Trypetinæ, 341 Trypoxylidæ, 504

Trypoxylon, 504, **504** Tryptochæta, 344 Tryxalinæ, 56 Tubercularium, 590 Tubulifera, 73 Tunga, 406, 407 Tungidæ, 407 Tychepsephenus, 427 Tychus, 415 Tydeus, 576 Tygarrup, 602 Tylidæ, 534 Tylobolus, 595 Tylopsis, 50 Tylos, 325, 534 Tylozygus, 111 Tympanophora, 52 Tympanophorinæ, 52 Typhloceratidæ, 405 Typhlocyba, 113, 113 Typhlocybinæ, 114 Typhloglomeris, 587 Typhloperipatus, 533 Typitium, 438 Typopeltis, 541 Tyrannochthonius, 548 Tyroglyphidæ, 574 Tyroglyphus, 573, 574

Udamacantha, 286 Udamoscelis, 122. 124 Udamoselinæ, 124 Ula, 268 Ulidia, 330 Ulidiidæ, 330 Uloboridæ, 558 Uloborus, 558 Ulodesmus, 590Ulomorpha, 268 Ulonata, 48 Ulopa, **115**, 116 Ulopidæ, 116 Ululodes, 187 Underwoodia, 591 Underwoodiidæ, 591 Unionicola, 572 Uraba, 219 Urania, 213 Uraniidæ, 213 Uranotænia, **273**, 273 Uranotæniinæ, 273 Urapteroides, **212**, 213 Urbaninæ, 236 Urbanus, 235 Urellia, 341 Uroblaniulus, 595

Urocerus, 473 Uroceridæ, 473 Uroctea, 557 Urocteidæ, 557 Urodacus, 544 Urodus, 229 Uromantis, 89 Uromenus, 53 Uropetala, 164 Urophora, 341 Uroplectes, 545 Uroplitella, 568 Uropoda, 568 Uropodias, 534 Uropodidæ, 568 Urophonius, 543 Uropsylla, 406 Uropsyllidæ, 406 Uropygi, 540 Urothripidæ, 74 Uropthrips, 74 Urothripoidea, 74 Usia, 297Usiinæ, 297 Usofila, 559 Utetheisa, 219 Uzuchidæ, 228

Valentinia, 232 Valgus, 452 Vandex, 69 Vandicidæ, 69 Vanessa, 238 Vanhæffenia, 588 Vanhœffeniidæ, 588 Vanhornia, 492, **493** Vanhorniidæ, 492 Varnia, 182 Vatellinæ, 411 Vates, 86, **88** Vatinæ, 86 Vectius, 561 Vejovidæ, 546 Vejovis, **544**, 546 Velia, 152 Veliidæ, 152 Velocipeda, 143 Velocipedidæ, 143 Venata, 119 Verhœffia, 591 Verhœffiidæ, 591 Vermileo, **290**, 290 Vermileoninæ, 290 Vermipsylla, 406 Vermipsyllidæ, 406 Veronatus, 418

Verrallia, 310 Verrucaditha, 548 Vespa, 496, **498** Vespaxenos, 470 Vespidæ, 496, 514 Vespinæ, 496 Vespoidea, 494, 514 Vespula, 496 Vestalis, 162 Vetillia, 65 Vietsia, 571 Villa, 295 Vipiinæ, 478 Vipio, 478 Vipionidæ, 477 Viticicola, 490 Vlax, 70 Volucella, **299**, 309 Volucellinæ, 309 Vostox, 69 Vulturops, **97**, 98

Waddycephalus, 581 Walshia, 233 Walterella, 570 Wandesia, 571 Wasmanniella, 349 Weyhia, 553 Wohlfartia, 316 Wiedemannia, 301 Wilhelmia, 281 Willistoniella, 325, 326 Winthemia, 318 Wuria, 571 Wyeomyia, 272

Xanthocanace, 336, 338 Xanthochlorinæ, 304 Xanthochlorus, 304 Xanthogramma, 309 Xantholininæ, 429 Xanthomelanus, **487**  $\mathbf{X}$ anthorhoe, 210Xanthrochroa, 437 Xenaspis, 331 $\mathbf{X}$ enidæ, 470 Xenoidea, 469 Xenolpium, 550Xenomorpha, 286 Xenophyes, 134 Xenopsylla, 405, 406 Xenos, **469**, 470 Xenylla, 46 Xeris, 473 Xerophlœa, 112 Xerophylla, 127

Xeroscopa, 216 Xestemvia, 294 Xestocephalus, 114 Xestozona, 590 Xiphandrium, 304 Xiphidiinæ, 52 Xiphidion, 52 Xiphochætoporatia, **589** Xiphosura, 268 Xiphydria, 472 Xiphyriidæ, 472, 513 Xiria, 331 Xyelá, 472, **473** Xvelidæ, 472, 511 Xylastodoris, 146, **149** Xyleborus, **445** Xylica, 63 Xylococcinæ, 128 Xylococcus, 128 Xylocopa, 508 Xylocopidæ, 508 Xylomyia, 288, 289, 360, Xylomyiidæ, 289, 360,374 Xylomyiinæ, 289 Xyloperga, 474 Xylophagidæ, 289, 361, 375 Xylophilidæ, 439 Xylophagus, 288, 289,361

Xylorictes, 228 Xylorictidæ, 228 Xyloryctidæ, 250 Xylota, 309 Xylotinæ, 309 Xylotrechus, 441 Xysticus, 562

Yersinia, 90 Yponomeuta, 229 Yponomeutidæ, 229, 245, 247

Zabalius, 51 Zabrachia, 285 Zagonia, 335 Zaitha, 154 Zalmoxis, 553 Zalusa, 348 Zaluscodes, 348 Zaprochilinæ, 50 Zaprochilus, 50 Zele, 480 Zelima, 309 Zelleria, 231 Zelmira, 280 Zelotes, 562 Zemeros, 239 Zenoa, 425 Zenillia, 318

Zephroniodesmus, 586 Zerene, 236 Zethinæ, 496 Zethus, 496 Zeugloptera, 200 Zeuzera, 206 Zeuzeridæ, 206, 246 Zimiris, 562 Zodariidæ, 560 Zodarion, 560 Zodion, 307 Zonites, 439 Zonosema, 341 Zopherus, 436 Zoraida, 119 Zoraptera, 100 Zorocrates, 561 Zoropsidæ, 561 Zoropsis, 561 Zorotypidæ, 101 Zorotypus, 100, 101 Zygæna, 207 Zygænidæ, 204, 207 Zygethobius, 600 Zygothrips, 76 Zygomyia, 278 Zygoneura, 279 Zygoneurinæ, 279 Zygonopus, 593 Zygoptera, 160, 165 Zygothrica, 345

#### INDEX TO COMMON NAMES

Agricultural ants, 490 Ailanthus moth, 213 Ambush-bug, 147 American cockroach, 78 Angle-wings, 238 Angoumois grain-moth,  $2\overline{27}$ Ant Lions, 187 Ants, 489 Aphids, 124 Aphis-lions, 185 Apple fruit-miner, 231 Apple leaf-hopper, 114 Apple leaf-miner, 231 Apple maggot, 341 Apple-skin miner, 232 Apple-skin worm, 226 Argentine ant, 489 Army worm, 218 Asparagus miner, 339 Assassin bugs, 147 Atlas moth, 213 Australian apple leafhopper, 113 Australian cockroach, 78

Back swimmers, 153 Bagworm moths, 207 Bark-beetles, 445 Bark lice, 96 Bat flies, 346 Bat parasites, 352 Bat-tick fly, 346 Bean weevils, 439 Bear animalcules, 581 Bedbug, 143Bee assassin, 147 Bee-louse, 345Bee moth, 215Bees, 475, 502Beet-fly, 319 Beetles, 408  $\operatorname{Big}$  bedbug, 147 Bird lice, 101 Biting lice, 101 Black-flies, 280 Black peach aphis, 125 Black scale, 132 Blow-flies, 316 Blue-bottle flies, 316 Blues, 239 Body louse, 105

Book lice, 96, 98 Book-scorpions, 537, 548 Botflies, 312, 322 Box-elder bug, 148 Bristle tails, 42 Brown scale, 132 Browntail moth, 218 Budmoth, 226 Buffalo-gnats, 280 Buffalo tree hopper, 108 Bull-dog ants, 490 Bumble-bees, 507 Burrowing webworms, 224 Butterflies, 201, 235

Cabbage butterfly, 236 Cabbage maggot, 319 Caddice flies, 191 Caddis flies, 191 Caddis worms, 191 Carolina mantis, 89 Carpenter moths, 206 Carrion-beetles, 428 Carrot rust fly, 340 Case-bearers, 240 Case-bearing clothes  $\mathrm{moth}, 224$ Cat-flea, 405 Cat-tail moth, 233 Cave crickets, 50 Cecropia moth, 213 Centipedes, 599 Cereal psocid, 98 Chalcis-flies, 482 Cheese skipper, 334 Cherry fruit sawfly, 474 Cherry maggot, 341 Chicken louse, 103 Chicken mite, 568 Chigger, 575 Chigoe flea, 407 Chinch-bug, 149 Chinese mantis, 89 Chrysanthemum gallmidge, 275Chrysanthemum leafminer, 339 Cicadas, 108 Cigar case-bearer, 232 Citrus purple scale, 130 Clear-wing moths, 203 Click beetles, 421

Clothes moth, 224 Clover-hay worm, 215 Clover leaf midge, 275 Clover mite, 576 Clover-seed worm, 226 Cluster-fly, 316 Coccids, 128 Cochineal insects, 133 Cockchafers, 451 Cockroach, 78 Codling moth, 226 Colorado potato-beetle, 443 Cocties 104

Cooties, 104 Coppers, 239 Corn aphis, 125 Corn-borer, 216 Corn blotch leaf-miner, 338 Corn earworm, 218 Cotton aphis, 125 Cotton boll-worm, 218 Cotton stainers, 149 Cottony maple scale, 132 Crab louse, 105 Cranberry fireworm, 226 Crescent-spots, 238 Crickets, 30 Crotonbug, 79 Cuckoo-wasps, 493 Currant fruit-fly. 341 Currant fruit worm, 225 Current sawfly, 474 Cutworms, 248

Daddy-long-legs, 538, 551 Damsel-bugs, 147 Damsel flies, 159 Date palm scale, 131 Death's head moth, 208 Death watch, 98 Deer-flies, 287, 288 Dengue-fever mosquito, 272Diamond-back cabbage moth, 229Digger wasps, 563 Diving beetles, 411 Dobson, 179 Dock sawfly, 474 Dog-flea, 405 Domesticated silkworm, 212

Doodle bug, 187 Dragon flies, 159 Driver ants, 490

Earwigs, 30, 34, 65
Elephant stomach bots, 309
Elm aphis, 126
Elm bark louse, 134
Emperors, 238
European corn borer, 216
European earwig, 70

False chinch-bug, 149 False-scorpions, 548 Feather-wing moths, 201 Fig-insects, 483 Fire-flies, 431 Flannel moths, 204 Fleas, 404 Flesh-flies, 316 Flies, 264 Flower-flies, 308 Follicle mite, 566 Forester moths, 219 Frittillaries, 238 Frog hoppers, 109 Fruit-flies, 340 Fruit tree leaf-roller, 226 Fungus-gnats, 277

Gad-flies, 287 Gall gnats, 274 Gall midges, 274 Gall wasps, 482 Giant skippers, 235 Gipsy moth, 218 Gnats, 264 Goatweed butterflies, 238 Goldenrod gallmaker, 341 Goldenrod gall moth, 228 Goliath beetle, 452 Gooseberry sawfly, 474 Grape-berry moth, 226 Grape blossom-midge, 275 Grape leaf-hopper 114 Grape phylloxera, 127 Grasshoppers, 30, 56 Green apple aphis, 125 Green bug of wheat, 125 Green lacewings, 185 Green peach aphis, 125 Ground beetles, 410 Grouse locusts, 55

Hair-streaks, 239

Harlequin cabbage-bug, Harvest-flies, 108 Harvestmen, 535, 551 Harvest mites, 575 Hawk moth, 208 Head louse, 105 Hellgrammite, 179 Hen flea, 407 Hessian fly, 275 Hickory phylloxeran, 127 Hog louse, 105 Honey-bee, 506 Hop aphis, 125 Hornets, 496 Horn fly, 321 Horntails, 473 Horse botflies, 322 Horse-flies, 287 Horseshoe crabs, 26 Housefly, 321 Humming bird moth, 208

Ichneumon-flies, 475 Imperial moth, 211 Indian-meal worm, 215 Indian wax scale, 132 Itch mites, 574

Jesus-bugs, 142 Jigger flea, 407 Joint-worms, 486 Jumping plant-lice, 122 Jumping spiders, 561 June-bugs, 451

Katydids, 30, 50 King crabs, 26 Kissing-bug, 147

Lace-bugs, 142 Lacewings, 185 Lac insects, 132 Larch sawfly, 474 Leaf-beetles, 440 Leaf blotch-miners, 232 Leaf-bugs, 145 Leaf-cutter ants, 490 Leaf-cutter bees, 507 Leaf-footed bug, 148 Leaf hoppers, 109 Leaf insects, 61, 63 Leaf miners, 221, 338 Leaf-rollers, 227 Leather-jackets, 354 Legionary ants, 490 Leopard moth, 206

Lesser apple worm, 226 Lice, 37 Lilac leaf-miner, 232 Locusts, 50, 108 Longicorns 440 Louse, 105 Long-horned locusts, 50 Luna moth, 213

Malaria mosquitoes, 272
Mantes, 30
Mantids, 84
March flies, 275
Marsh-treaders, 142
May-beetles, 451
Mayflies, 169
Meal moth, 215
Mealy bugs, 134
Medfly, 341
Mediterranean fruit-fly, 341
Melon and pickle worms, 216
Melon fly, 341
Mexican jumping bean

moth, 226 Midges, 264 Milkweed butterfly, 237 Millipedes, 585 Mites, 535, 565 Mosquitoes, 264, 272 Moth-flies, 271 Moths, 201

Narcissus bulb-fly, 309 Natal fruit-fly, 341 Negro-bugs, 150 No-see-ums, 282 Nun-moth, 218

Oblique-banded leaf-roller 226
Olive fly, 341
Onion maggot, 319
Orange maggot, 341
Orange-tips, 236
Orange tortrix, 226
Orb-weavers, 560
Oriental cockroach, 78
Oyster-shell scale, 130

Papaya fly, 341 Parnassians, 236 Peach scale, 132 Peach twig-borer, 227 Pear blister mite, 565 Pear midge, 275 Pear psylla, 123 Pear scale, 130 Pear-slugs, 474 Pea weevils, 439 Peg-top coccids, 132 Periodical cicada, 108 Periwinkles, 36 Phylloxera, 127 Pine aphids, 124 Pineapple scale, 130 Pine leaf scale, 130 Pine sawflies, 474 Pine twig moths, 226 Pink bollworm, 227 Pirate-bugs, 144 Pistol case-bearer, 232 Plague flea, 405 Plant-bugs, 145 Plant lice, 124 Polyphemus moth, 213 Pomace fly, 345 Potato aphis, 125 Potato tuber moth, 228 Praying mantids, 84 Predatory flower-bug, 144 Promethea moth, 213 Pseudoscorpions, 537 Psocids, 96 Pubic louse, 105 Punkies, 282

Raspberry cane maggot, 319Raspberry sawfly, 474 Red humped apple worm, Red spiders, 576 Rhinoceros beetles, 452 Ribbed cocoon makers, 222Roaches, 35, 77 Robber-flies, 298 Rose scale, 130 Rose leaf-hopper, 114 Rose midge, 275 Royal-palm bug, 146

Sacred scarabæus, 450 Salmon-flies, 176 Sand-flies, 271, 282 San José scale, 130 Satin moth, 218 Saw-flies, 472 Scale insects, 37, 128 Scorpion flies, 189 Scorpions, 535, 537, 543

Screw-worm, 314 Scurfy scale, 130 Seedcorn maggot, 319 Serpent-flies, 181 Serpentine leaf-miner, 338 Seventeen-year locust, 108 Sharp shooters, 109 Sheep-tick, 347, 352 Sheet-web weavers, 560 Shield-back bugs, 150 Shield-bearers, 229 Shore-bugs, 146 Short-horned locusts, 56 Silk worm, 212 Skin-beetles, 450 Skippers, 201, 233 Slug-caterpillars, 241 Snake-flies, 181 Snipe-flies, 290 Snout butterflies, 239 Snow-flea, 348 Soft brown scale, 132 Soldier flies, 284 Solitary wasps, 514 Solpugids, 538 Soothsayers, 84 Sovereigns, 238 Sowbug, 534 Sphinx caterpillars, 252 Spiders, 535, 555 Spittle insects, 109 Spongilla-flies, 185 Spring canker-worm, 209 Spring tails, 45 Spruce bud-worm, 226 Squash-bug, 148 Stable fly, 321 Stag-beetles, 450 Stick insects, 30, 61 Sticktight, 407 Stilt bug, 148 Stingless bees, 506 Stink-bugs, 151 Stink-flies, 185 Stoneflies, 176 Strawberry crown-miner, 227Strawberry leaf-roller, 226 Sucking lice, 104 Sugarcane hopper, 119 Swallow-tails, 236 Sweet-potato weevil, 446 Tachina flies, 318

Tarantula hawks, 497

Tarantulas, 556

Tarnished plant-bug, 145 Tent caterpillars, 221 Termites, 93 Thread-legged bugs, 147 Thrips, 73 Ticks, 535, 565 Tiger beetles, 410 Toad-bugs, 153 Toe-biters, 154 Tongue-worms, 535, 579Trap-door spiders, 556 Tree crickets, 54 Tree hoppers, 108 True bugs, 141 True lice, 104 Tse-tse flies, 321 Tumble-bugs, 450 Tumbu-fly, 321 Tussock moth, 218

Velvet ants, 500 Walking sticks, 61 Wasps, 32 Water Bears, 582 Water boatmen, 152 Water creepers, 154 Water scavenger beetles, 414Water-scorpions, 154Water striders, 142 Water tigers, 411 Web-worms, 216, 219 Weevils, 408Wheat bulb fly, 319 Wheat-stem saw-fly, 472Wheel-bug, 147 Whip-scorpions, 537, 540 Whirligig beetles, 412 White ants, 93 White flies, 124 Willow gall sawflies, 474 Wind-scorpions, 538 Winter-gnats, 267 Wireworm beetles, 421 Wolf spiders, 561 Wood-ants, 489 Wood-wasps, 472 Woolly aphis, 126

Yellow-fever mosquito, 272Yellow-jackets, 496 Yellow necked apple worm, 211Yucca moths, 222

# Bibliography of the Neuropterida

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